

Physiological Basis for Differential Selectivity of Four Grass Species to Aminocyclopyrachlor

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Aminocyclopyrachlor (AMCP) is a synthetic auxin herbicide used for broadleaf weed control in pasture and rangeland. The tolerance and fate of AMCP within pertinent grass species is not well understood. Research was conducted to establish the tolerance of four grass species to AMCP application and observe their absorption, translocation, and metabolism. Results indicate that tall fescue is the most tolerant of AMCP at rates required for weed control. Bahiagrass and bermudagrass are marginally tolerant, and cogongrass is the most sensitive. Tall fescue and bahiagrass absorbed more AMCP than bermudagrass and cogongrass, but cogongrass absorption is the most rapid and complete within 2 days after treatment (DAT). Cogongrass and bermudagrass translocated the least amount out of the target area, whereas bahiagrass and tall fescue translocated the most. Radioisotope imaging revealed that tall fescue may sequester absorbed AMCP in leaf tips. This sequestering may be the basis of the greater tolerance to AMCP by tall fescue relative to the other species evaluated. No metabolism of AMCP was detected in any grass species out to 42 DAT.

Nomenclature: Aminocyclopyrachlor; bahiagrass, *Paspalum notatum* Flueggé; bermudagrass, *Cynadon dactylon* (L.) Pers.; cogongrass, *Imperata cylindrica* (L.) Beauv.; tall fescue, *Lolium arundinaceum* (Schreb.) S.J. Darbyshire.

Key words: Absorption, herbicide fate, metabolism, synthetic auxin, translocation.

Synthetic auxin herbicides are widely used in turfgrass, pasture, roadside, and grass cropping systems for selective control of broadleaf weeds. Although broadleaf weeds are typically the targets of synthetic auxin herbicides, many warm-season grasses can be injured by their application (Bell et al. 2000; Cudney et al. 1997; Doroh et al. 2009; Grossmann 2003; Johnson 1995; Kaufman 1955; McElroy et al. 2005; Patton et al. 2010). Synthetic auxins are typically applied for POST weed control and subsequently absorbed by foliage and roots (Bovey et al. 1983; Ross and Lembi 1999; Senseman 2007). Synthetic auxin compounds function by mimicking indole-3-yl-acetic acid (IAA), which stimulates cell division, differentiation, and ultimately plant growth at meristematic plant tissues (Grossmann 2010; Ross et al. 2002). Symptoms of these herbicides include leaf epinasty, uncontrolled root growth, changes in nucleic acid levels, and

increased biosynthesis of ethylene (Anderson 1996; Deshpande and Hall 2000; Devine et al. 1993).

Currently, aminocyclopyrachlor (AMCP) is the only pyrimidine carboxylic acid herbicide to be registered and commercialized. It is structurally similar to the pyridine carboxylic acid herbicides, which include picloram, clopyralid, and aminopyralid (Senseman 2007). AMCP is characterized by comparatively low use rates and low mammalian toxicity (Claus et al. 2009; Strachan et al. 2010; Turner et al. 2008; U.S. Environmental Protection Agency [USEPA] 2010). AMCP exhibits the synthetic auxin herbicide mode of action (MOA) (Grossmann 2003; Woodward and Bartel 2005). Between fall 2010 and August 2011, the potassium salt formulation of AMCP was marketed as Imprelis[®] by E.I. du Pont de Nemours and Company (DuPont) for the selective control of broadleaf weeds in both cool-season and some warm-season turfgrasses. AMCP was labeled for use on lawns, golf courses, sod farms, and athletic fields. AMCP rates for select established warm-season grasses were 52.5 to 79.0 g ae ha⁻¹, whereas established cool-season grasses such as tall fescue were labeled as high as 105.0 g at ha^{-1} (Anonymous 2010).

Despite previous issues of off-target damage, AMCP has the potential to be widely used for broadleaf weed control in various grass crops as an alternative to currently labeled phenoxyacetic and pyridine herbicides (Anonymous 2012, 2014; Lewis

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et al. 2013; Patton et al. 2013). AMCP controls a number of troublesome and invasive weeds with minimal injury to desired grass species (Belcher and Walker 2010; Curtis et al. 2009; Flessner et al. 2011a; Gannon et al. 2009; Jenks and Walter 2012; Mansue and Hart 2010; Montgomery et al. 2009). Some grass species can be injured and exhibit differing levels of tolerance to AMCP application (Anonymous 2010; Belcher and Walker 2010; Brecke et al. 2010; Curtis et al. 2009; Flessner et al. 2011a, 2011b; Harmoney et al. 2012; Kniss and Lyon 2011; Reed et al. 2013). It is possible that AMCP could control some troublesome monocot species within grass crops (Enloe et al. 2012; Reed et al. 2013, Vargas et al. 2014). AMCP is currently being explored for use in pastures, roadsides, and forestry situations alone and in combination with other chemistries (Ezell and Self 2013; Isreal et al. 2013; Meredith et al. 2013; Sellers et al. 2013).

Previous research has revealed that AMCP is readily absorbed in tolerant and susceptible species (Bell et al. 2011; Bukun et al. 2010; Lewis et al. 2013; Lindenmayer et al. 2013, Vargas et al. 2014). Absorption of AMCP in broadleaf species has been classified as rapid, with complete absorption occurring within 24 to 48 h after treatment (HAT) (Bell et al. 2011; Bukun et al. 2010; Lindenmayer et al. 2013). Lewis et al. (2013) reported that absorption of AMCP into tall fescue, a tolerant Poaceae species, was also rapid with maximum absorption at 48 HAT. This study reported that maximum translocation away from the treated leaf occurred within 96 HAT, indicating limited but rapid movement throughout aboveground tissues. Vargas et al. (2014) reported that absorption of AMCP-methyl ester into large crabgrass [Digitaria sanguinalis (L.) Scop.], a sensitive Poaceae species, was rapid, with > 90%absorption at only 1 HAT. Studies indicate that AMCP movement in broadleaf species reached a peak between 24 and 192 HAT, depending on the species (Bell et al. 2011; Bukun et al. 2010; Lindenmayer et al. 2013). Thus, it appears that both susceptible and tolerant species exhibit similar absorption and translocation of AMCP, which did not correlate to susceptibility within broadleaf species (Bell et al. 2011). There is no documentation of AMCP metabolism in either susceptible or tolerant species out to 192 HAT (Bell et al. 2011; Bukun et al. 2010; Lewis et al. 2013; Lindenmayer et al. 2013). This correlates well with the lack of metabolism found with similar pyridine carboxylic acid herbicides (Bukun et al. 2009; Lym and Moxness 1989).

In light of the damage to off-target species, the tolerance of important grass species to AMCP and its fate within these plants must be examined. If AMCP is absorbed by grasses and remains as the active form, there is a possibility that it can be returned to the soil through mowing, grazing, or root exudation (Lewis et al. 2013). If AMCP remains as the active form within the plant where it is protected from microbial degradation and photolysis (USEPA 2010; Finkelstein et al. 2008), it is likely that AMCP activity can be prolonged.

A number of studies on grass tolerance to AMCP have been published. However, none directly compare tolerant and susceptible grass species (Flessner et al. 2011a, 2011b; Kniss and Lyon 2011; Lewis et al. 2013). Our first objective was to compare the tolerance of four grass species to AMCP. The four species were bahiagrass, bermudagrass, cogongrass, and tall fescue. The second objective was to utilize radiotracing techniques to determine the role of absorption, translocation, and metabolism as possible mechanisms of the observed variance in these species.

Materials and Methods

Tolerance Studies. Grasses selected for evaluation were 'Pensacola' bahiagrass (Pennington[®] Seed), common bermudagrass, cogongrass, and 'Kentucky 31' tall fescue (Pennington[®] Seed). Experiments use the free-acid formulation of AMCP (aminocyclopyrachlor, DPX-MAT28, E. I. du Pont de Nemours and Company). Experiments used a randomized complete block design with four replications per treatment. Two experiments each were conducted for bahiagrass, bermudagrass, cogongrass, and tall fescue for a total of eight experiments. All experiments were repeated in time and applied on February 3 and March 17, 2014. Herbicide-containing treatments also included a nonionic surfactant (NIS) (Activator 90 Non-Ionic Surfactant, Loveland Products Inc.) at $0.25\% \text{ v v}^{-1}$. This adjuvant was included because previous research involving AMCP and other related synthetic auxins indicates it substantially increased herbicide absorption (Bell et al. 2011; Bukun et al. 2009; Lewis et al. 2013). The greenhouse was located in Auburn, AL. Greenhouse temperatures were maintained between 23 and 26 C for the duration of the experiments. Normal sunlight was supplemented with sodium-halide growth lamps producing 150 μ mol m⁻² s⁻¹ at the grass canopy. Total peak irradiance was $< 800 \ \mu mol \ m^{-2} \ s^{-1}$ throughout the experiments. All grass species were grown simultaneously from seed in separate plastic flats containing a potting substrate (Scott's Miracle-Gro® Potting Mix, The Scott's Company, LLC). Grasses were then prepared for treatment by transplanting five individual plants into 700-cm³ pots with the use of Marvyn loamy sand (native) soil. Grasses were allowed to resume active growth for 2 wk prior to treatment application. Grasses were maintained with weekly fertilizer applications (Miracle-Gro® All Purpose Plant Food, The Scott's Company, LLC), daily irrigation, and monthly insecticide applications. AMCP rates were 52.5 and 105.0 g are ha^{-1} , and a nontreated check was included. These rates were based on the lowest and highest labeled AMCP rates for effective weed control (Anonymous 2010). Treatments were applied at a spray volume of 187 L ha⁻¹ in an enclosed spray chamber with a single nozzle (TeeJet[®] TP8002EVS, Spraying Systems Co.).

Visual ratings of each grass species were taken at 10 and 20 d after treatment (DAT) and were chosen to bracket a typical synthetic auxin rating date of 2 wk after treatment (WAT) (Sciumbato et al. 2004). Ratings were not extended beyond 20 DAT because previous research has indicated that tall fescue is tolerant of AMCP, and the other species studied are injured, but recover from AMCP applications (Anonymous 2010; Enloe et al. 2012; Flessner et al. 2011b; Montgomery et al. 2009). It should also be noted that because absorption and translocation occurs rapidly (< 8 DAT), plant injury within that time was of primary interest. Visual injury ratings were based on a scale of 0 to 100, with 100 corresponding to dead plants and 0 indicating healthy plants. All plants were clipped to a uniform height of approximately 7.5 cm following the 10 DAT rating date to simulate mowing and allow for measurement of regrowth. Plant heights and both fresh and dry weights were then taken at 20 DAT. Plant foliage was then oven dried at 85 C for 1 wk and weighed. All species were analyzed together. Experimental units consisted of a single pot.

AMCP Fate Studies. The free-acid formulation of ¹⁴C-AMCP (aminocyclopyrachlor DPX-MAT28) with a specific activity of 1522 kilobecquerels (kBq) mg⁻¹ was used in both experiments. Greenhouse conditions were maintained as in previously mentioned studies. Grasses were grown from seed in plastic flats containing a potting substrate (Scott's Miracle-Gro[®] Potting Mix). Plants were prepared for

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treatment by transplanting single plants into 66-cm^3 cell pack pots with the use of Marvyn loamy sand (native) soil. Grasses were allowed to resume active growth for 2 wk prior to treatment. Grasses were maintained by subirrigation to prevent removal of any adsorbed herbicide from the leaf surface. AMCP at 52.5 g ha⁻¹ was used for all studies (Anonymous 2010). All herbicide-containing treatments included a NIS as previously described.

Absorption and Translocation. Ten microliters of ¹⁴C-AMCP was applied to the adaxial side of a fully mature leaf for a total of 1.67 kBq plant⁻¹. The herbicide was applied beginning at one end of the marked treated area on the leaf then extending the solution across the treated area. This created a thin film that increased surface area contact, allowed for rapid drying, and reduced the chance of losing droplets to runoff. The physiology of the AMCP was of primary interest, therefore no broadcast application was applied prior to treatment with the radiolabeled AMCP. Lack of broadcast application allowed the detection of movement absent the confounding effects produced by herbicide injury. For each species, four individual plants were harvested at 1, 2, 4, and 8 d after treatment (DAT) then divided into the following sections: leaf wash, target area (a 2-cm leaf section to which the herbicide had been directly applied), treated leaf, remainder foliage, and crown plus roots. The leaf wash was obtained by placing the target area in a 20ml plastic vial containing 1 ml of a $90:10 \text{ v v}^{-1}$ methanol: deionized water solution and agitating it for 30 s to remove adsorbed ¹⁴C-AMCP. The leaf wash was combined with 15 ml of UniversolTM fluid (MP Biomedicals, LLC) after harvest. Radioactivity of the leaf wash fluid was quantified via liquid scintillation spectroscopy (LSS) (Beckman CoulterTM LS-6500 Multipurpose Scintillation Counter, Beckman Coulter, Inc.). This allowed quantification of unabsorbed herbicide relative to the amount applied. Upon removing the washed target area from the scintillation vial, the remainder of the plant was dissected as indicated above. Plant material was oven dried at a temperature of 85 C, combusted with the use of a tissue oxidizer (OX501 Biological Oxidizer, R.J. Harvey Instrument Corporation), and quantified via LSS.

Percentage of ¹⁴C-AMCP absorption was calculated by summing the total amount of radioactivity recovered in harvested plant parts minus the leaf wash. Translocation was calculated by dividing the radioactivity recovered in each plant part by the total radioactivity absorbed. Percent recovery was calculated as total radiation recovered across all plant parts and the leaf wash divided by total radiation applied. Radiation in the form of percent recovered of applied was the response variable for both absorption and translocation studies.

Metabolism. Two applications of 10 µl of ¹⁴C-AMCP totaling 3.34 kBq plant⁻¹ were applied to separate leaves in order to increase the likelihood of detecting potential metabolism. Only the target areas and remainder of aboveground foliage were analyzed for metabolism. Plants were harvested at 7, 14, 21, and 42 DAT to examine the possibility that persistence of the parent compound could lend itself to gradual metabolism by select grass species. Immediately after harvest, plant sections were placed in 7 by 12-cm plastic bags and stored at -20 C until processing. Root material was placed in aluminum trays for oxidizing and subsequent ¹⁴C quantification for recovery calculation. Plant parts were ground with a mortar and pestle with liquid nitrogen. Ground material was next moved into a glass grinding tube with 3 ml of methanol in order to improve extraction. Ground material was then placed into 14-ml glass tubes and centrifuged at $2000 \times g$ for 10 min. Supernatants were decanted into 20-ml scintillation vials and concentrated with the use of a nitrogen evaporator (N-EVAPTM122, OA-SYSTM, Organomation Associates, Inc.). The concentrated solution was immediately re-eluted in 1 ml of methanol for spotting onto silica gel chromatography plates (Silica gel G 20 by 20 cm TLC plates, Analtech, Inc.). The resulting pellet from centrifuging was oxidized along with harvested roots and the dried remainder of the 1 ml concentrated solution for recovery calculation.

Analysis of possible AMCP metabolites was done by normal-phase thin-layer chromatography (TLC). Silica gel plates were used as the stationary phase, and a solvent system of methanol, isopropanol, ethyl acetate, and acetic acid $(7:1:1:1 v v^{-1})$ was used as the mobile phase (Bell et al. 2011). Plates were manually scored into 19 individual 1cm lanes, and a 100-µl aliquot of concentrated solution was applied 3 cm above the bottom of the plate, leaving a blank lane between each replicate. The area of application was maintained to $\sim 10 \text{ mm}$ by successive spotting and drying. The first lane of each plate was left blank to use as a background. A 10-µl drop of ¹⁴C-AMCP was spotted on the next lane for metabolite comparison. Plates were developed for approximately 75 min or until the

mobile phase had moved 150 mm above the origin, then allowed to air dry prior to scanning.

Radioactive positions, percentages, and corresponding retardation factor (R_f) value of the parent compound and possible metabolites were determined by scanning TLC plates with the use of radio-chromatogram scanner (AR-2000 imaging scanner, Bioscan, Inc.). Radioactive trace peaks were manually integrated. Peaks below 5% of total radioactivity were rejected. Peaks were identified by comparing their R_f values with those from the corresponding ¹⁴C-AMCP standard. Data collected consisted of the percentage of the parent herbicide, the percentage of all metabolites detected that were more polar than the parent herbicide, and the percentage of all metabolites that were less polar than the parent herbicide (Bell et al. 2011).

Radioisotope Imaging. To confirm the biodistribution of radiolabeled material within plants visually, radioisotope images (RI) were taken with the use of a multipurpose image analyzer (Fujifilm FLA-5100, Fujifilm Holdings Corporation). Plants used for imaging were treated identically to those in absorption and translocation experiments prior to harvest. Upon harvest, plant roots were washed and entire intact plants were manually pressed between layers of newspaper, and dried at 85 C for 2 wk. Plants were removed and glued onto 8 by 10-cm cardstock mounts. Mounts were covered in plastic wrap then placed in exposure cassettes (Amersham Biosciences) and pressed against phosphor imaging plates for approximately 24 h, then scanned with the use of the image analyzer, in a procedure described in depth by Wehtje et al. (2007).

Experimental Design and Data Analysis. Separate experiments were conducted to determine the absorption and translocation, and the metabolism of AMCP within four grass species. Each experiment was repeated in time between February 3 and March 17, 2014. A randomized complete block design in a five by four factorial arrangement (five plant parts across four harvest intervals) was used for absorption and translocation studies, and a three by four factorial (three plant parts across four harvest intervals) was used for metabolism studies. All studies had four replications and two experimental runs.

Data from tolerance studies were subjected to ANOVA in SAS (SAS[®] Institute Inc., v. 9.2) with PROC GLIMMIX to test for significance of replication-in-time, herbicide rate, and grass species, and allow for lack-of-fit testing. Experimental run

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Table 1. Response of four graminaceous species to foliar-applied aminocyclopyrachlor at two rates

Grass species	Foliar height reduction ^{a,b}		Visual injury ^c	
		Foliar weight reduction	10 DAT ^d	20 DAT
		%		
52.5 g ae ha^{-1}				
Bahiagrass	17 a	59 с	7 bc	21 b
Bermudagrass	17 a	28 b	3 b	1 c
Cogongrass	67 b	85 d	13 c	72 a
Tall fescue	6 a	5 a	0 a	0 c
Mean	27 A	44 A	6 B	24 B
105.0 g ae ha^{-1}				
Bahiagrass	44 b	65 bc	20 a	56 b
Bermudagrass	37 b	56 b	6 b	17 c
Cogongrass	66 c	85 c	22 a	73 a
Tall fescue	14 a	(+) 5 a	3 b	3 d
Mean	40 B	50 A	13 A	37 A

^a Means within a common rate and column followed by similar letters are not significantly different according to Fisher's protected $LSD_{(0.05)}$ comparison. Rate main effect means within a column followed by different upper-case letters are statistically different according to $LSD_{(0.05)}$ comparison.

^b Reduction relative to appropriate nontreated control.

^c Visual injury rated on a scale of 0 to 100 with 0 equivalent to the nontreated control, and 100 corresponding to death.

^d Abbreviation: DAT, days after treatment.

and replication were considered random effects with treatment, species, and harvest interval being considered fixed effects. All visual rating data were transformed for statistical analysis with the use of square-root transformation, then backtransformed for presentation. Means were separated with the use of Fisher's protected least significant difference (LSD) and considered significant if $P \leq 0.05$. Data from fate studies were analyzed with the use of PROC GLIMMIX and multivariate analysis techniques to test for significance of replication-in-time, DAT, plant part, grass species, and allow for lack-of-fit testing. Means of fate studies were separated with the use of Duncan's multiple range test at the 0.05 level.

Results and Discussion

Tolerance Studies. ANOVA revealed that repetition in time and replications within the experiment were not significant. Therefore data were pooled accordingly. An interaction between treatment and species was significant (P < 0.005) for height and weight reduction data, whereas a three-way interaction of DAT, treatment, and species was significant (P < 0.005) for visual-injury data; this indicates that injury to most species increases with time. Cogongrass was clearly the most sensitive species, as indicated by all four response variables collected (Table 1). Visual injury symptoms on cogongrass included stunting of growth and reddening and weakening of crown tissues, followed by necrosis of

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plant foliage. Conversely, tall fescue was the most tolerant species, with only temporary leaf epinasty regardless of rate. The tolerance of tall fescue is in agreement with previous research (Anonymous 2010; Lewis et al. 2013). Bahiagrass and bermudagrass were intermediate to these two extremes; but between these two grasses, bahiagrass was more sensitive than bermudagrass. Bahiagrass was significantly more sensitive at the high rate than the low rate, and was injured at an unacceptable level for use in either turfgrass or forage systems (Johnson 1995; Johnson and Murphy 1995). Symptoms of bahiagrass injury were brittle crowns, reddening of stems, leaf epinasty, and in some cases complete plant necrosis. Like bahiagrass, bermudagrass was more sensitive to the high rate, whereas the low rate had little effect. In time, AMCP had an effect on bermudagrass growth resulting in significant swelling at nodes and leaf epinasty. These symptoms, however, did not prove fatal to treated plants or compromise grass quality. Ranking sensitivity (highest to lowest) when all four response variables are taken collectively is as follows: cogongrass > bahiagrass > bermudagrass > tall fescue. The differences in sensitivity are sufficiently great that AMCP may have potential for the selective control of cogongrass within other grass species.

As previously mentioned, it has been documented that tall fescue is tolerant of AMCP application. Our results agree with previous research, as well as the general trend that warm-season grasses are more sensitive to synthetic auxin herbicides than cool-

Table 2. Absorption of foliar-applied aminocyclopyrachlor at 1, 2, 4, and 8 d after foliar application by selected grass species. Greenhouse experiment repeated in time and data pooled.^a

Grass species	1 DAT ^b	2 DAT	4 DAT	8 DAT		
	% of applied					
Bahiagrass	16 a	28 a	29 a	45 a		
Bermudagrass	21 a	22 a	33 a	38 ab		
Cogongrass	24 a	25 a	24 a	25 b		
Tall fescue	25 a	33 a	42 a	46 a		

^a Means within a column followed by the same letter are similar according to Duncan's multiple range test at the 0.05 level.

^b Abbreviation: DAT, days after treatment.

season grasses (Bell et al. 2000; Flessner et al. 2009, 2011b; Lewis et al. 2013; Senseman 2007; Wehtje 2008). Treatment data indicate the expected outcome that greater rates of AMCP application result in impaired plant growth across sensitive species.

Absorption and Translocation. ANOVA revealed that repetition in time and replications within the experiment were not significant. Therefore data were pooled accordingly. Absorption of foliarapplied AMCP was sufficiently variable at both the 1 and 2 DAT harvests such that neither meaningful differences of absorption over time, nor differences between the grass species were

evident (Table 2). All species absorbed greater than 30% of the total uptake within 24 HAT. This rapid absorption is consistent with past research (Bell et al. 2011; Bukun et al. 2010; Lewis et al. 2013), as was total percent recovery, which averaged 89% across all four species and harvest intervals. At 4 DAT, the amount of applied AMCP that had been absorbed ranged from 24% with cogongrass to 42% in tall fescue. However, no differences between the species could be detected (P > 0.05). At 8 DAT, amount absorbed ranged from 25% (cogongrass), to at least 45% (tall fescue and bahiagrass), a difference that was statistically significant. Absorption by bermudagrass was intermediate to these extremes. The most sensitive of all species tested, cogongrass, absorbed the least ¹⁴C-AMCP, likely because of rapid uptake and desiccation of plant tissues, hindering further absorption. Across all species, no more than half of the applied AMCP was absorbed. Consequently, the overall highest-to-lowest ranking of absorption for foliar-applied AMCP is as follows: tall fescue \sim bahiagrass > bermudagrass > cogongrass. This ranking is nearly the opposite of the ranking of overall of sensitivity presented above. Therefore, differential absorption is not likely a contributing factor in differential response between the four grass species.

Table 3. Distribution of absorbed aminocyclopyrachlor over time by four grass species after foliar application. Greenhouse experiment, repeated in time and data pooled.^a

Tissue of grass plant	Bahiagrass	Bermudagrass	Cogongrass	Tall fescue	
	% of absorbed				
1 DAT ^b					
Target area	41 a	60 a	28 a	40 a	
Remainder of treated leaf	35 a	10 bc	29 a	36 a	
All remaining foliage	18 ab	29 b	21 a	20 ab	
Crown and roots	5 b	0 c	21 a	3 b	
2 DAT					
Target area	21 a	40 a	27 a	29 ab	
Remainder of treated leaf	28 a	50 a	26 a	50 a	
All remaining foliage	30 a	9 b	26 a	18 b	
Crown and roots	20 a	0 b	20 a	2 b	
4 DAT					
Target area	15 a	23 cb	30 a	28 a	
Remainder of treated leaf	40 a	40 a	20 a	42 a	
All remaining foliage	30 a	27 ab	28 a	26 a	
Crown and roots	16 a	9 c	21 a	3 b	
8 DAT				-	
Target area	15 b	14 b	29 a	26 b	
Remainder of treated leaf	23 ab	65 a	28 a	53 a	
All remaining foliage	37 a	17 b	26 a	19 b	
Crown and roots	25 ab	4 b	17 a	2 c	

^a Means within a column of a common exposure time and followed by the same letter are similar according to Duncan's multiple range test at the 0.05 level.

^o Abbreviation: DAT, days after treatment.

For translocation data, radioactivity recovered within the tissue samples harvested were normalized to 100, and presented as a percentage of the total amount absorbed (Table 3). Translocation varied considerably between the species. Extensive translocation occurred in cogongrass. The percentage of the foliar-absorbed AMCP that was detected in all four tissue samples were equivalent at both the 4 and 8 DAT harvests (Table 3). Cogongrass crown and root tissue had 21 and 17% of the amount absorbed at 4 and 8 DAT, respectively. Therefore foliar-absorbed AMCP had been readily translocated basipetally out of the treated leaf and into other tissues including the crown and root. Crafts (1961) stated that translocation of foliarabsorbed 2,4-D to all meristem tissues in the plants, particularly the roots, was required for treated plant death. Thus the translocation pattern exhibited by cogongrass is in agreement with its comparative sensitivity to AMCP. Translocation of foliarabsorbed AMCP was completely different in tall fescue compared to what was observed in cogongrass. No more than 3% of absorbed AMCP was recovered in the crown and root tissue in tall fescue at both harvest intervals. This percentage was significantly less than the amount recovered in the foliar portions of the plant. Furthermore, the "remainder of treated leaf' sample consistently (i.e., both 4 and 8 DAT) had the highest numerical percentage, and statistically the highest percentage (8 DAT only) recovered. Thus, in tall fescue, the propensity for absorbed AMCP to remain in the treated leaf indicated either minimal translocation, or at least a translocation pattern that precludes basipetal movement and contact with meristem tissues.

Translocation in bahiagrass and bermudagrass are intermediate to the extremes exhibited by cogongrass and tall fescue. Between bahiagrass and bermudagrass, more basipetal translocation occurred in bahiagrass. In bahiagrass the percentage recovered in the crown and root tissues was also statistically equivalent to the other three tissues samples collected. However, at 8 DAT recovery of radioactivity in the target area was less compared to the other tissue samples. The overall highest-tolowest ranking in translocation of foliar-absorbed AMCP is a follows: cogongrass > bahiagrass > bermudagrass > tall fescue. This ranking is in agreement the overall sensitivity of the grass species to foliar-applied AMCP, indicating that differential translocation is a possible mechanism of selectivity between these species.

Data indicate that the majority of all AMCP applied to grass species, whether tolerant or susceptible, remains on the leaf surface or within aboveground tissues. Herbicide remaining within aboveground tissues may be moved off target by mowing or grazing. Results are similar to previous reports involving many other synthetic auxin herbicides, which indicate that these herbicides may be released from treated tissues in quantities large enough to elicit plant injury (Branham and Lickfeldt 1997; Miltner et al. 2003).

It is clear that all species absorb and translocate ¹⁴C-AMCP in amounts sufficient to elicit injury (Rensburg and Breeze 1990; Strachan et al. 2011). Although AMCP is moved to the crown and roots of susceptible species, it is unlikely to result in prolonged control of either cogongrass (Enloe et al. 2012) or bahiagrass (Anonymous 2010) in turf, pasture, or forestry systems.

Metabolism. TLC analysis revealed no metabolites of ¹⁴C-AMCP out to 42 DAT across all four species tested. Comparisons of means determined that all peaks detected from treated plants had an Rf value not significantly different from the AMCP-only check (data not shown). Percent recovery in metabolism studies averaged 86% across species and harvest intervals. This recovery is within an acceptable range, considering losses due to sample harvesting and processing, and is consistent with previous research involving synthetic auxin absorption and fate (Bukun et al. 2009, 2010; Lewis et al. 2013). The absence of metabolism in our studies is similar to other findings regarding AMCP and synthetic auxin metabolism within plants. Previous research involving AMCP metabolism in field bindweed (Convolvulus arvensis L.), prickly lettuce (Lactuca serriola L.), rush skeletonweed (Chondrilla juncea L.), and yellow starthistle (Centaurea solstitialis L.) found no metabolites out to 8 DAT (Bell et al. 2011; Lindenmayer et al. 2013). Grossman and Kwiatkowski (2000) found that ~90% of quinclorac, another synthetic auxin, remained as the parent compound within treated grasses. They concluded that differences in susceptibility were not a result of this limited metabolism. Alternatively, dicamba was found to be rapidly metabolized in both wheat (Triticum aestivum L.) and bluegrass (*Poa* spp.) (Broadhurst et al. 1966). Studies reported that at 2 DAT, only 31% of dicamba remained as the parent material and that by 10 DAT, over 83% of dicamba was present as primary or secondary metabolites. The previous body of



Figure 1. Images of four grass species treated with foliar-applied 14 C-aminocyclopyrachlor at 8 d after treatment. Arrows indicate the location at which the 10 μ l of herbicide was applied. Images from left to right: cogongrass, bahiagrass, bermudagrass, and tall fescue.

research shows variable amounts of metabolism across synthetic auxins, but very limited or no metabolism of both pyridine and pyrimidine carboxylic acid herbicides (Bukun et al. 2009; Lewis et al. 2013; Lym and Moxness 1989).

Radioisotope Imaging. Radioisotope images (Figure 1) correlate well with absorption and translocation data. Both sets of data indicate that the majority of applied ¹⁴C-AMCP remained in the target area and treated leaf for the duration of the experiment. It is interesting to note that tall fescue appears to be sequestering the radiolabeled material in leaf tips. Sequestering to this degree is not evident in the other three species. This could be a possible method that tolerant species, namely some grasses, use to protect vital meristematic tissues such as the crown and roots. The collection of ¹⁴C-AMCP in leaf tips supports the argument that AMCP will likely experience "cyclic" movement in grass systems as treated material is mowed or grazed, returned to the soil profile, and then reabsorbed by plants. Future research may be in order to explore the role this sequestration of AMCP plays in plant tolerance and the ultimate fate of AMCP within grass weed control systems.

Our results indicate that the differential sensitivity of these four grass species to foliar-applied AMCP appears to be primarily related to differential translocation. Specifically, the tolerance of tall fescue is likely the result of minimal (less than 3% of applied AMCP) translocation. Furthermore, the limited translocation that does occur serves to transport the herbicide to leaf tips and away from herbicide-sensitive meristem tissues. Conversely, results indicate that neither the amount of absorption nor metabolism contribute to tolerance. It is

interesting to note that the tolerance to foliarapplied AMCP appears to be the result of limited translocation in conjunction with translocation that serves to isolate the herbicide away from meristematic tissues. This effect was most clearly evident in tall fescue. Tall fescue is a C₃ species; the other three grasses are C₄ species. Leaves of C₄ species have a higher CO₂ exchange rate, a higher ratio of crosssectional phloem area to leaf area, and greater translocation rates relative to C₃ species (Gardner et al. 1985). Leaves of C₄ species also export a larger percentage of their assimilate within a few hours than C₃ species. This improved export of assimilate by leaves of C₄ species may be because of their specialized anatomy, in which vascular sheath cells have chloroplasts, or the result of a greater crosssectional phloem area. Our conjecture is that these differences may also inadvertently allow for greater symplastic and apoplastic translocation of foliarabsorbed AMCP, increasing herbicide-meristem contact, and thus increasing phytotoxicity. Absence of AMCP metabolism paired with a potentially long soil half-life and low soil sorption (Conklin and Lym 2013; Oliveira et al. 2011) is a double-edged sword. Prolonged exposure to AMCP may offer extended control of hard-to-eliminate weeds, such as cogongrass, with repeated applications, but there is also risk involved when a chemical remains in the soil profile for extended periods of time. Prolonged exposure to AMPC by bahiagrass will likely result in thinning of the stand and extensive plant injury. Because the majority of ¹⁴C-AMCP remains in aboveground foliage as the parent compound, it is likely that AMCP will be returned to the soil profile in a cyclic manner, as proposed by Lewis et al. (2013). It has also been reported that AMCP exudes from grass roots (Lewis et al. 2013; Parker et al. 2014).

Altogether, these observations indicate that future AMCP use will be limited in some weed control systems, and highly effective in others. Applications near sensitive species should be avoided, as AMCP may be released deep into the soil profile via decaying roots or exudation from healthy tissues. In the future, the role of auxin binding proteins and their possible mutations should be explored as an additional mechanism of the observed tolerance of tall fescue to AMCP (Grossmann 2010).

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Literature Cited

- Anderson W (1996) Weed Science. 3rd edn. Long Grove, IL: Waveland Press. Pp 193–195
- Anonymous (2010) Imprelis[®] herbicide product label. DuPont Publication No. H-65717. Wilmington, DE: DuPont. Pp 1–7
- Anonymous (2012) Streamline[®] herbicide product label. DuPont Publication No. SL-1722. Wilmington, DE: DuPont. 2 p
- Anonymous (2014) Perspective[®] herbicide product label. DuPont Publication No. SL-1794. Wilmington, DE: DuPont. 2 p
- Belcher JL, Walker RH (2010) Cogongrass response to aminocyclopyrachlor. P 63 *in* Proceedings Southern Weed Science Society. Little Rock, AR: Southern Weed Science Society
- Bell GE, Martin DL, Kuzmic R, Stone ML, Solie JB (2000) Herbicide tolerance of two cold-resistant bermudagrass (*Cynodon spp.*) cultivars determined by visual assessmen tand vehicle-mounted optical sensing. Weed Technol 14:635–641
- Bell JL, Burke IC, Prather TS (2011) Uptake, translocation and metabolism of aminocyclopyrachlor in prickly lettuce, rush skeletonweed and yellow starthistle. Pest Manag Sci 67:1338–1348
- Bovey RW, Hein H, Meyer RE (1983) Absorption and translocation of triclopyr in honey mesquite (*Prosopis juliflora* var. glandulosa). Weed Sci 31:807–812
- Branham BE, Lickfeldt DW (1997) Effects of pesticide-treated grass clippings used as a mulch on ornamental plants. HortScience 32:1216–1219
- Brecke BJ, Unruh JB, Partridge-Telenko DE (2010) Aminocyclopyrachlor for weed management in warm-season turfgrass Page 132 *in* Proceedings of the Southern Weed Science Society. Little Rock, AR: Southern Weed Science Society
- Broadhurst NA, Montgomery ML, Freed VH (1966) Metabolism of 2-methoxy-3,6-dichloro-benzoic acid (dicamba) by wheat and bluegrass plants. J Agric Food Chem 6:585–588

Bukun B, Gaines TA, Nissen SJ, Westra P, Brunk G, Shaner DL, Sleugh BB, Peterson VF (2009) Aminopyralid and clopyralid

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absorption and translocation in Canada thistle (*Cirsium arvense*). Weed Sci 57:10–15

- Bukun B, Lindenmayer RB, Nissen SJ, Westra P, Shaner DL, Brunk G (2010) Absorption and translocation of aminocyclopyrachlor and aminocyclopyrachlor-methyl ester in Canada thistle (*Cirsium arvense*). Weed Sci 58:96–102
- Claus JS, Holliday MJ, Turner RG, Meredith JH, Stephen C (2009) Aminocyclopyrachlor development and registration update. Page 128 *in* Proceedings of the North Central Weed Science Society. Champaign, IL: North Central Weed Science Society
- Conklin KL, Lym RG (2013) Effect of temperature and moisture on aminocyclopyrachlor soil half-life. Weed Technol 27:552–556
- Crafts, AS (1961) The Chemistry and Mode of Action of Herbicides. New York: Wiley (Interscience). Pp 16–65
- Cudney D, Elmore C, Giceault V, Reints J (1997) Common bermudagrass (*Cynadon dactylon*) management in cool-season turfgrass. Weed Technol 11:478–483
- Curtis DW, Hinds-Cook BJ, Brewster BD, Hulting AG, Mallory-Smith CA (2009) Tolerance of perennial ryegrass and tall fescue grown for seed to aminocyclopyrachlor (DPX KJM-44). Pages 70–71 p *in* Proceedings of The Western Society of Weed Science. Albuquerque, NM: Western Society of Weed Science
- Deshpande S, Hall JC (2000) Auxinic herbicide resistance may be modulated at the auxin-binding site in wild mustard (*Sinapis arvensis* L.): a light scattering study. Pestic Biochem Physiol 66:41–48
- Devine M, Duke S, Fedtke C (1993) Physiology of Herbicide Action. Englewood Cliffs, NJ: Prentice-Hall. Pp 295–309
- Doroh M, McElroy J, Guertal E (2009) Triclopyr enhances metamifop and clodinafop control of bermudagrass and reduces zoysiagrass injury. Page 21 *in* Proceedings of the Southern Weed Science Society. Orlando, FL: Southern Weed Science Society
- Enloe SF, Belcher J, Loewenstein N, Aulakh JS, van Santen E (2012) Cogongrass control with aminocyclopyrachlor in pastures. Forage Grazinglands DOI: 10.1094/FG-2012-0828-02-RS
- Ezell AW, Self AB (2013) Loblolly pine tolerance to ground spraying applications of aminocyclopyrachlor. Page 102 *in* Proceedings of the Southern Weed Science Society. Houston, TX: Southern Weed Science Society
- Finkelstein BL, Armel GR, Bolgunas SA, Clark DA, Claus JS, Crosswicks RJ, Hirata CM, Hollingshaus GJ, Koeppe MK, Rardon PL, VA Wittenbach, Woodward MD (2008) Discovery of aminocyclopyrachlor (proposed common name) (DPX-MAT28): a new broad-spectrum auxinic herbicide. *In* Proceedings of the 236th American Chemical Society National Meeting Philadelphia, PA: American Chemical Society
- Flessner ML, McCurdy JD, McElroy JS (2011a) Tolerance of six zoysiagrass cultivars to aminocyclopyrachlor. Weed Technol 25:574–579
- Flessner ML, McElroy JS, Walker RH (2009) Quantification of warm season turfgrass phytotoxicity from broadleaf control herbicides. Page 20 *in* Proceedings of the Southern Weed Science Society. Orlando, FL: Southern Weed Science Society
- Flessner ML, McElroy JS, Wehtje GR (2011b) Quantification of warm-season turfgrass injury from triclopyr and aminocyclopyrachlor. Weed Technol 25:367–373
- Gannon TW, Yelverton FH, Warren LS, Silcox CA (2009) Broadleaf weed control with aminocyclopyrachlor (DPX-

KJM44) in fine turf. Page 394 *in* Proceedings of the Southern Weed Science Society. Orlando, FL: Southern Weed Science Society

- Gardner FP, Pierce B, Mitchell R (1985) Physiology of crop plants. Ames, IA: Iowa State University Press. Pp 60–61
- Grossmann K (2003) Mediation of herbicide effects by hormone interactions. J Plant Growth Regul 22:109–122
- Grossmann K (2010) Auxin herbicides: current status of mechanism and mode of action. Pest Manag Sci 66:113–120
- Grossman K, Kwiatkowski J (2000) The mechanism of quinclorac selectivity in grasses. Pestic Biochem Physiol 66:83–91
- Isreal TD, Rhodes N, Mueller TC, Bates GE, Waller JC (2013) Effects of aminocyclopyrachlor plus metsulfuron on tall fescue growth and forage quality. Page 147 *in* Proceedings of the Southern Weed Science Society. Houston, TX: Southern Weed Science Society
- Jenks BM, Walter TD (2012) Yellow toadflax control in rangeland with DPX-MAT28. Page 44 *in* Proceedings of the North Central Weed Science Society. St. Louis, MO: North Central Weed Science Society
- Johnson BJ (1995) Tolerance of four seeded common bermudagrass (*Cynodon dactylon*) types to herbicides. Weed Technol 9:794–800
- Johnson BJ, Murphy TR (1995) Effect of paclobutrazol and flurprimidol on suppression of *Poa annua* spp. reptans in creeping bentgrass (*Agrostis stolonifera*) greens. Weed Technol 9:182–186
- Kaufman P (1955) Histological responses of the rice plant (Oryza sativa) to 2,4-D. Am J Bot 42:649–659
- Kniss AR, Lyon DJ (2011) Winter wheat response to preplant applications of aminocyclopyrachlor. Weed Technol 25:51–57
- Lewis DF, Roten RL, Everman WJ, Gannon TW, Richardson RJ, Yelverton FH (2013) Absorption, translocation, and metabolism of aminocyclopyrachlor in tall fescue (*Lolium arundinaceum*). Weed Sci 61:348–352
- Lindenmayer RB, Nissen SJ, Westra PP, Shaner DL, Brunk G (2013) Aminocyclopyrachlor absorption, translocation and metabolism in field bindweed (*Convolvulus arvensis*). Weed Sci 61:63–67
- Lym R, Moxness K (1989) Absorption, translocation, and metabolism of picloram and 2,4-D in leafy spurge (*Euphorbia esula*). Weed Sci 37:498–502
- Mansue C, Hart S (2010) Broadleaf weed control and coolseason turfgrass safety with aminocyclopyrachlor applied on a fertilizer granular. Page 73 *in* Proceedings of the Northeastern Weed Science Society. Cambridge, MA: Northeastern Weed Science Society
- McElroy JS, Breeden GK, Yelverton FH, Gannon TW, Askew SD, Derr JF (2005) Response of four improved seeded bermudagrass cultivars to postemergence herbicides during seeded establishment. Weed Technol 19:979–985
- Meredith JH, Medlin CR, Rupp RN, Castner EP, Williams RW (2013) Weed control in southern pastures with aminocyclopyrachlor. Page 266 *in* Proceedings of the Southern Weed Science Society. Houston, TX: Southern Weed Science Society
- Miltner E, Bary A, Cogger C (2003) Clopyralid and compost: formulation and mowing effects on herbicide content of grass clippings. Compost Sci Util 11:89–99
- Montgomery D, Evans C, Martin D (2009) Control of kochia with DPX-KJM44 along Oklahoma highway rights-of-way. Page 471 *in* Proceedings of the Southern Weed Science Society. Orlando, FL: Southern Weed Science Society

- Oliveira RS, Alonso DG, Koskinen WC (2011) Sorptiondesorption of aminocyclopyrachlor in selected Brazilian soils. J Agric Food Chem 59:4045–4050
- Parker ET, McElroy JS, Wehtje GR, Price AJ (2014) Bioassay techniques for detecting root leakage of auxinic herbicides. Page 84 *in* Proceedings of the Southern Weed Science Society. Orlando, FL: Southern Weed Science Society
- Patton A, Creswell T, Ruhl G, Weller S (2011) A Turf Professional's Guide to Imprelis[®] Herbicide Injury in the Landscape. Purdue University Plant & Pest Diagnostic Laboratory publication. http://www.ppdl.purdue.edu/PPDL/ pubs/briefs/ImprelisLCO.pdf. Accessed April 18, 2014
- Patton A, Trappe J, Doroh M, McElroy J (2010) Evaluation of herbicides and tank-mixtures for suppression of bermudagrass in zoysiagrass. Page 225 *in* Proceedings of the Southern Weed Science Society. Little Rock, AR: Southern Weed Science Society
- Patton AJ, Ruhl GE, Creswell TC, Wan P, Scott DE, Becovitz JD, Weisenberger DV (2013) Potential damage to sensitive landscape plants from wood chips of aminocyclopyrachlor damaged trees. Weed Technol 27:803–809
- Reed TV, Yu J, McCullough PE (2013) Aminocyclopyrachlor efficacy for controlling Virginia Buttonweed (*Diodia virginiana*) and smooth crabgrass (*Digitaria ischaemum*) in tall fescue. Weed Technol 27:488–491
- Rensburg E van, Breeze VG (1990) Uptake and development of phytotoxicity following exposure to vapour of the herbicide ¹⁴C 2,4,-D butyl by tomato and lettuce plants. Environ Exp Bot 30:405–414
- Ross JJ, O'Neill DP, Wolbang CM, Symons GM, Reid JB (2002) Auxin-gibberellin interactions and their role in plant growth. J Plant Growth Regul 20:336–353
- Ross M, Lembi C (1999) Applied Weed Science. 2nd edn. Upper Saddle River, NJ: Prentice Hall. Pp 159–166
- Sciumbato AS, Chandler JM, Senseman SA, Bovey W, Smith KL, Bovey RW (2004) determining exposure to auxin-like herbicides. I. Quantifying injury to cotton and soybean. Weed Technol 18:1125–1134
- Sellers BA, Abe DG, Ferrell JA (2013) Control of various pasture weeds with aminocyclopyrachlor premixes. Page 265 *in* Proceedings of the Southern Weed Science Society. Houston, TX: Southern Weed Science Society
- Senseman SA (2007) Herbicide Handbook. 9th edn. Lawrence, KS: Weed Science Society of America. Pp 322–361
- Strachan SD, Casini MS, Heldreth KM, Scocas J, Nissen SJ, Bukun B, Lindenmayer RB, Shaner DL, Westra P, Brunk G (2010) vapor movement of synthetic auxin herbicides: aminocyclopyrachlor, aminocyclopyrachlor-methyl ester, dicamba, and aminopyralid. Weed Sci 58:103–108
- Strachan SD, Nanita SC, Ruggiero M, Casini MS, Heldreth KM, Hageman LH, Flanigan H, Ferry NM, Pentz AM (2011) Correlation of chemical analysis of residual levels of aminocyclopyrachlor in soil to biological responses of alfalfa, cotton, soybean, and sunflower. Weed Technol 25:239–244
- Turner RG, Claus JS, Hidalgo E, Holliday MJ, Armel GR (2008) Technical introduction of the new DuPont vegetation management herbicide aminocyclopyrachlor. *In* Proceedings of the Weed Science Society of America. Orlando, FL: Weed Science Society of America [Abstract 405]
- [USEPA] U.S. Environmental Protection Agency (2010) Registration of the new active ingredient aminocyclopyrachlor for use on non-crop areas, sod farms, turf, and residential

Parker et al.: ¹⁴C-aminocyclopyrachlor in four grass species • 797

lawns. Washington, DC: Office of Pesticide Programs, Registration Division. Pp 1–23

- Vargas JJ, Brosnan JT, Mueller TC, Kopsel DA, Klingeman WE, Armel GR (2007) Biokinetics and efficacy of aminocyclopyrachlor-methyl ester as influenced by difkufenzopyr. Weed Sci 62:538–547
- Wehtje G, Miller ME, Grey TL, Brawner WR (2007) Comparisons between x-ray film- and phosphorescence imaging-based autoradiography for the visualization of herbicide translocation. Weed Technol 21:1109–1114
- Wehtje GR (2008) Synergism of dicamba with diflufenzopyr with respect to turfgrass weed control. Weed Technol 22:679–684
- Woodward AW, Bartel B (2005) Auxin: regulation, action, and interaction. Ann Bot 95:707–735

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