Interictal psychosis following temporal lobe surgery: dentate gyrus pathology

M. Thom^{1,2}*, M. Kensche^{1,3}, J. Maynard^{1,2}, J. Liu¹, C. Reeves¹, J. Goc¹, D. Marsdon², D. Fluegel^{1,3} and J. Foong^{1,3}

¹Department of Clinical and Experimental Epilepsy, University College London, Institute of Neurology, London, UK

² Division of Neuropathology, National Hospital for Neurology and Neurosurgery, Queen Square, London WC1N 3BG, UK

³ Division of Neuropsychiatry, National Hospital for Neurology and Neurosurgery, Queen Square, London WC1N 3BG, UK

Background. *De novo* interictal psychosis, albeit uncommon, can develop in patients following temporal lobe surgery for epilepsy. Pathological alterations of the dentate gyrus, including cytoarchitectural changes, immaturity and axonal reorganization that occur in epilepsy, may also underpin co-morbid psychiatric disorders. Our aim was to study candidate pathways that may be associated with the development of interictal psychosis post-operatively in patients with hippocampal sclerosis (HS).

Method. A total of 11 patients with HS who developed interictal psychosis (HS-P) post-operatively were compared with a matched surgical HS group without psychosis (HS-NP). Resected tissues were investigated for the extent of granule cell dispersion, mossy fibre sprouting and calbindin expression in the granule cells. We quantified doublecortin, minichromosome maintenance protein 2 (MCM2) and reelin-expressing neuronal populations in the dentate gyrus as well as the distribution of cannabinoid type 1 receptor (CBR1).

Results. The patterns of neuronal loss and gliosis were similar in both groups. HS-P patients demonstrated less mossy fibre sprouting and granule cell dispersion (p < 0.01) and more frequent reduction in calbindin expression in granule cells. There were no group differences in the densities of immature MCM2, doublecortin and reelin-positive cells. CBR1 labelling was significantly lower in Cornu ammonis area CA4 relative to other subfields (p < 0.01); although reduced staining in all hippocampal regions was noted in HS-P compared with HS-NP patients, the differences were not statistically significant.

Conclusions. The alterations in dentate gyrus pathology found in HS-P patients could indicate underlying differences in the cellular response to seizures. These mechanisms may predispose to the development of psychosis in epilepsy and warrant further investigation.

Received 5 August 2013; Revised 3 February 2014; Accepted 4 February 2014; First published online 8 April 2014

Key words: Dentate gyrus, epilepsy, hippocampal sclerosis, psychosis.

Introduction

Temporal lobe surgery for intractable temporal lobe epilepsy (TLE) can result in many patients being rendered seizure free or having a significant reduction in seizure frequency (Choi *et al.* 2008). Psychiatric symptoms, however, can develop for the first time following temporal lobe surgery, most commonly depression and anxiety (Foong & Flugel, 2007; Cleary *et al.* 2012). *De novo* interictal psychosis (Stevens, 1990; Mace & Trimble, 1991; Christodoulou *et al.* 2002; Shaw *et al.* 2004) following surgery is uncommon, with a mean incidence of 7% (Trimble, 1992; Christodoulou *et al.* 2002; Nadkarni *et al.* 2007) but tends to follow a chronic course. The development of psychotic symptoms is not closely related to postoperative seizure outcome (Mace & Trimble, 1991; Manchanda *et al.* 1993; Leinonen *et al.* 1994). *De novo* interictal psychosis is of particular interest both as a potentially serious complication of the surgery and for the insights it might provide into primary psychosis such as schizophrenia (Shaw *et al.* 2004).

The underlying pathophysiological mechanisms of psychosis in epilepsy are unclear. Structural and functional neuroimaging studies have implicated left (dominant) temporal lobe pathology with greater reductions in hippocampal volumes (Maier *et al.* 2000), magnetization transfer ratio (Flugel *et al.* 2006) and single-photon emission computerized tomography (SPET) perfusion (Marshall *et al.* 1993; Mellers *et al.* 1998) in TLE patients with interictal psychosis. Hippocampal sclerosis (HS) is the most common

^{*} Address for correspondence: M. Thom, Ph.D., Department of Clinical and Experimental Epilepsy, University College London, Institute of Neurology, Queen Square, London WC1N 3BG, UK.

⁽Email: M.Thom@ucl.ac.uk)

pathology (Roberts et al. 1990; Briellmann et al. 2000), although low-grade tumours such as dysembryoplastic neuroepithelial tumours have also been reported (Andermann et al. 1999). Diminished mossy fibre sprouting (Kandratavicius et al. 2012), dysplasias of the dentate gyrus (Briellmann et al. 2000) and excess corpora amylacea (Radhakrishnan et al. 2007) have been detected in resected tissues of TLE/HS patients with pre-operative interictal psychosis. Detailed histological analysis in patients with de novo interictal psychosis following epilepsy surgery, however, is sparse; one study that explored the density of hippocampal calbindin-immunoreactive neurons in three patients with de novo post-operative and three with preoperative psychosis compared with controls did not find significant differences (Suckling et al. 2000). The neuropathological changes in de novo interictal psychosis may be similar to primary schizophrenia, given the common clinical symptomatology and pathological findings in the temporal lobe (Perez & Trimble, 1980; Trimble, 1992; Toone, 2000).

In contrast, the neuropathology of schizophrenia has been more widely researched. High-field neuroimaging studies have highlighted structural alterations involving the hippocampal dentate gyrus (Kirov et al. 2013). Hippocampal changes reported in post-mortem studies include altered neuronal densities (Knable et al. 2004), distribution (Zhang & Reynolds, 2002) and dendritic/synaptic organization (Cotter et al. 2000; Rosoklija et al. 2000), neuropil abnormalities (Garey, 2010) as well as dentate gyrus immaturity (Walton et al. 2012). Altered glutamatergic transmission in the dentate gyrus, including abnormalities of the mossy fibre pathway (Tamminga et al. 2010) and the endocannabinoid receptor system, have been associated with psychosis (Ujike & Morita, 2004; Luzi et al. 2008; Fernandez-Espejo et al. 2009). Post-mortem studies of the neocortex and hippocampal dentate gyrus have also confirmed a reduction in reelin-expressing cells in schizophrenia (Impagnatiello et al. 1998; Eastwood & Harrison, 2006); reelin has a critical role in the orchestration of normal cortical development as well as ongoing roles in the mature brain, regulating structural plasticity such as axonal growth and synaptogenesis (Lakatosova & Ostatnikova, 2012; Stranahan et al. 2013). Dysregulation of the reelin pathway is implicated in psychosis (Folsom & Fatemi, 2013).

HS is the most common and studied pathology in epilepsy surgical series (Blumcke, 2009; Blumcke *et al.* 2012). In addition to the neuronal loss and accompanying gliosis, frequent epilepsy-specific alterations of the dentate gyrus include granule cell dispersion (GCD) (Blumcke *et al.* 2009), mossy fibre sprouting (Sutula *et al.* 1989), altered calbindin expression (Martinian *et al.* 2012), and regenerative capacity of granule cells (Thom *et al.* 2005*a*) and reelin-expressing cell populations (Haas & Frotscher, 2010). Some pathological features have been associated with epilepsy comorbidities, e.g. hippocampal regenerative capacity and calbindin loss have been correlated with memory dysfunction (Coras *et al.* 2010; Karadi *et al.* 2012). Furthermore, alterations to the endocannabinoid presynaptic receptor system have recently been reported in HS. This system modulates both glutamatergic and GABAergic synaptic transmission (Katona & Freund, 2008), and decreases in cannabinoid type 1 receptor (CBR1), CBR1 binding protein mRNA and protein levels have been observed in human HS tissue (Ludanyi *et al.* 2008).

The aim of this study was to explore relevant cellular alterations gathered from a large series of patients with HS who underwent temporal lobe surgery for epilepsy and to determine whether specific pathological changes in the dentate gyrus could be associated with the development of *de novo* interictal psychosis.

Method

Subjects

Patients who had developed de novo interictal psychosis following temporal lobe surgery for drugresistant TLE over a 21-year period (1991 to 2012) were assessed for inclusion in the study. A total of 20 patients who met Diagnostic and Statistical Manual of Mental Disorders, fourth edition, text revision (DSM IV-TR; APA, 2000) diagnosis criteria for a psychotic disorder due to a general medical condition were identified from the hospital records in the neuropsychiatry clinics at the National Hospital for Neurology Neurosurgery (NHNN). Of the 20 patients, 15 had a diagnosis of HS, four patients had low-grade glioneuronal tumours (dysembryoplastic neuroepithelial tumours) and one patient had an epidermoid cyst involving the mesial temporal lobe. Of the 15 patients with HS and psychosis (HS-P), consent was available in 11 for further study. A total of 10 HS patients who also underwent temporal lobe surgery but without either a history of pre- or post-operative psychosis were selected as control cases (HS-NP). They were matched to the HS-P group with respect to age and the year of surgery. The study was approved by the Joint Research and Ethics committee of the Institute of Neurology and NHNN.

Histopathological techniques

Immunohistochemistry

For each patient, histological sections from all specimens from the temporal lobe surgical resection

[neocortex (anterior 3-4 cm of temporal lobe and pole), parahippocampal gyrus, amygdala and hippocampus] were reviewed with haematoxylin and eosin and Luxol fast blue stain for myelin. The tissue block from the mid-body of the hippocampus, to include the most complete contour of the dentate gyrus, was selected for an immunohistochemistry panel including: glial fibrillary acidic protein (GFAP) (1:1500; Dako, UK), NeuN (neuronal nuclei; 1:1500; Chemicon International); calbindin D28K (1:10000; Swant, Switzerland); reelin (gift from Professor Goffinet, University of Namur, Belgium; dilution. 1:200, microwave pre-treatment); mini-chromosome maintenance protein 2 (MCM2) (1:900; BD Biosciences, USA); dynorphin (1:100; AbD Serotec MorphoSys UK Ltd, UK); CBR1 (1:1000; Abcam, UK); doublecortin (DCX) (1:250; Cell Signaling, UK); nestin (1:1000; Abcam, UK); and GFAP- δ (1:5000; Chemicon International, USA). For this purpose, sections were cut at $7 \mu m_r$ dewaxed, and immersed in a 3% hydrogen peroxide solution to block endogenous peroxidase activity prior to antibody incubation. Dako EnVision™ diaminobenzidine (DAB) was used as the chromogen and slides were counterstained with haematoxylin.

Qualitative analysis and measurements

On the NeuN sections, the pattern of HS was classified as International League Against Epilepsy (ILAE) type 1, 2 or 3 (Blumcke et al. 2013). The presence of neuronal loss in the dentate gyrus, as assessed by the presence of gaps of depletion or discontinuity in the cell layer, was semi-quantified in four grades: <25% (not detectable); 25–50% (mild); 50–75% (moderate); and >75% (severe) neuronal loss. All assessments were made independently by two observers (J.M. and M.T.) with good agreement. The extent of GCD was assessed as in a previous study (Thom et al. 2010). Using an image analysis program (Image Pro Plus, version 6; Media Cybernetics, UK) linked to a Zeiss microscope (Carl Zeiss, UK), between six and 10 non-overlapping images per case were taken at 10x magnification representing the entire length of the dentate ribbon. For each image frame, the position of the 10 outermost cells in the granule cell layer (GCL) was measured relative to the basal layer in the perpendicular axis providing a maximum of 100 measurements per case. For each case a mean of all these measurements and a mean of the maximum measurement recorded per field were obtained as measures of the overall and maximal severity of GCD. These measurements were repeated by one observer (J.M.) in five cases for reproducibility, with an intra-class correlation coefficient (ICC) of 0.99.

The presence of mossy fibre sprouting (MFS) in the dentate gyrus on dynorphin-labelled sections was

graded as previously (Thom *et al.* 2009): grade 0 (no MFS); grade 1 (focal MFS in the inner molecular layer; ML); grade 2 (extensive MFS throughout ML). Calbindin expression in the dentate gyrus granule cells was also graded as previously (Martinian *et al.* 2012): grade 0 (normal expression through the GCL); grade 1 (loss of expression in basal GCL); and grade 2 (extensive loss of expression throughout the GCL).

Quantitative analysis

Reelin. A region of interest (ROI) was drawn at ×2.5 objective magnification using Histometrix image analysis software (Kinetic Imaging, UK) in the hippocampal white matter, with the dentate GCL and the pyramidal cell layer of Cornu ammonis (CA) area CA3–1 as the boundaries. This included the stratum moleculare, lacunosum and radiatum of CA1–3 in addition to the stratum moleculare of the dentate gyrus. The rationale to include all the white matter was that it was not feasible to confidently separate these layers on the reelinstained sections. All reelin-positive cells within this ROI, regardless of size or morphology (Abraham & Meyer, 2003), were counted at ×40 objective magnification by a single observer (M.K.).

MCM2. A ROI was drawn with Histometrix at ×2.5 magnification around the dentate gyrus GCL as in a previous study (Thom *et al.* 2002). Within this ROI (mean area 3.2×10^2 mm²), all fields were examined at ×40 magnification and all positive nuclei counted by a single observer (J.M.) and the mean cell density (per unit area) recorded. These measurements were repeated in eight cases with an ICC of 0.92.

DCX. Sequential images were captured along the entire length of the dentate gyrus at ×40 objective magnification with a Nikon (80i) microscope (average length of dentate gyrus studied per case=3.6 mm). Using Image Pro Plus analyser (version 6.3), positive cells were manually tagged and the mean cell density (per unit area) recorded.

CBR1. Representative, non-overlapping images from the dentate gyrus (GCL and ML), CA4 and the subiculum were taken at ×20 objective magnification (four to seven images per subfield) with a Nikon 80i microscope. Using RGB thresholding and Image Pro Plus (version 6), the field fraction of synaptic staining (percentage labelling) was quantified for each image. The same thresholds were applied for each image within a case and a mean labelling index for each subfield was obtained. Both the image capture and measurements were carried out by a single observer (J.M.)

Group	HS-P (<i>n</i> =11)	HS-NP (n=10)
Mean age at surgery, years (range)	39.7 (14–50)	38.72 (24–47)
Mean age of onset of epilepsy ^a , years (range)	10.9 (1-35)	5.3 (1-16)
Mean duration of epilepsy at time of surgery, years (range)	20.6 (5-40)	28.7 (23-36)
Gender, <i>n</i>		
Male	10	9
Female	1	1
Presurgical seizure type, number of patients		
CPS	11	10
SGTS	9	8
SE	2	2
Antiepileptic medications at time of surgery, number of patients		
On monotherapy	4	4
On polytherapy	7	6
Surgical laterality, <i>n</i>		
Left	10	9
Right	1	1
Post-operative seizure outcome, number of patients		
ILAE=class 1 (seizure free) at 12 months	3	3
Mean interval between surgery and onset of psychosis, months (range)	10.3 (2–36)	-

HS-P, hippocampal sclerosis and *de novo* post-surgical interictal psychosis; HS-NP, hippocampal sclerosis and epilepsy alone with no psychosis (control); CPS, complex partial seizures; SGTS, secondary generalized seizures; SE, episode of status epileptic; ILAE, International League Against Epilepsy.

^a Exact age of onset of seizures unknown for three HS-NP cases.

and repeated in six cases with good inter-observer reproducibility (ICC=0.96).

Statistical comparisons

Statistical comparisons between HS-P and HS-NP patients were carried out using SPSS for Windows (version 20; IBM, USA) and non-parametric tests, including the Mann–Whitney test and Spearman's correlation.

Results

Clinical characteristics

The clinical characteristics of both groups are summarized in Table 1. The two groups did not differ in age, presurgical seizure history or number of antiepileptic medications. Only one patient who was in the HS-P group had a history of cannabis use but no patient was prescribed anti-psychotic medications prior to surgery. There was also no family history of psychosis for any of the subjects. The number of patients who were seizure free (ILAE class 1) at 12 months did not differ between the groups.

In the HS-P group, the psychotic symptoms were predominantly persecutory delusions and auditory hallucinations, resembling schizophrenia. The onset of psychosis ranged from 2 to 36 months following surgery, with a mean of 10.3 months. The psychosis followed a chronic course (duration ranging from 3 to 20 years) and all patients were maintained on antipsychotic medications at the time of our study.

Histopathological findings

The patterns of HS were similar in the HS-P and HS-NP groups, with HS ILAE type 1 being the commonest pattern in both groups (Table 2 and Fig. 1A). In one of the HS-P patients, gliosis alone was confirmed in the hippocampus and this subject was reclassified as a no-HS case according to the new criteria (Blumcke et al. 2013). The severity of granule cell loss, as assessed semi-quantitatively, was also similar between the groups with no perceived loss in around half of the cases in each group. Nestin- and GFAP- δ -positive glial cells were noted in CA4, particularly in the subgranular zone, and CA1 in all HS cases as previously described (Martinian et al. 2009), but with no clear discrimination between groups on qualitative evaluation (Fig. 1B, C). GCD was less severe in the HS-P group than in HS-NP cases (Fig. 1D and Fig. 2); statistically significant differences were shown

Pathological feature ^a	Patterns/grades/region	HS-P	HS-NP	р
HS subtype represented ^b , $\%$ (<i>n</i> =21)	ILAE type 1	82	89	_
	ILAE type 2	_	-	
	ILAE type 3	9	-	
	ILAE type 1, probable ^b	_	11	
	No HS	9	_	
Granule cell depletion ^c , % ($n=20$)	None	55	50	_
	Mild	9	25	
	Moderate	18	12.5	
	Severe	18	12.5	
GCD				
Mean value of group, μ m (s.d.)		57.8 (33.3)	101.6 (59)	0.06
Mean of maximum GCD values, μ m (s.d.)		105.7 (45.7)	203.7 (72.4)	0.004
Mossy fibre sprouting ^c , % ($n=17$)	Grade 0	11	0	_
	Grade 1	44.5	16	
	Grade 2	44.5	84	
Calbindin expression in granule cells ^c , % ($n=16$)	Grade 0	0	0	_
	Grade 1	16	40	
	Grade 2	84	60	
Mean MCM2 in dentate gyrus x $10^{-5}/\mu m^2$ (s.d.) (<i>n</i> =18)		2.1 (1.7)	1.8 (1.4)	0.4
Mean reelin-positive cells (HWM) x $10^{-5}/\mu m^2$ (s.d.) (n=20)	4.02 (1.8)	4.43 (2.8)	0.8
DCX-positive cells in dentate gyrus x $10^{-5}/\mu m^2$ (s.d.)	(n=10)	7.59 (8.41)	7.67 (5.69)	0.6
Cannabinoid type 1 receptor	Dentate gyrus	2.7 (1.0)	3.2 (2.6)	0.9
Mean field fraction % (s.d.) $(n=12)$	CA4	0.3 (0.2)	1.8 (2.7)	0.4
	Subiculum	2.3 (1.2)	5.1 (2.9)	0.8

Table 2. Neuropathological findings in HS-P and HS-NP patients

HS-P, hippocampal sclerosis and *de novo* post-surgical interictal psychosis; HS-NP, hippocampal sclerosis and epilepsy alone with no psychosis (control); HS, hippocampal sclerosis; ILAE, International League Against Epilepsy; GCD, granule cell dispersion; S.D., standard deviation; MCM2, mini-chromosome maintenance protein 2; HWM, hippocampal white matter; DCX, doublecortin; CA4, Cornu ammonis area 4.

 a^{a} *n*=Number of cases on which assessment was possible; in some cases assessment was not possible due to technical reasons or insufficient representation of the dentate gyrus on further sections.

^b Classification of the HS subtype was based on ILAE system of 2013 (Blumcke *et al.* 2013) with probable HS referring to cases with incomplete subfields available for analysis.

^c See text for details of grading schemes for assessment of immunostaining.

for the maximum GCD values but not for the mean GCD values (Table 2). This significance remained when the no-HS case was excluded from the analysis. Whereas grade 2 mossy fibre sprouting was confirmed in over 80% of HS-NP cases, it was noted in less than half of HS-P cases (Fig. 1E and Table 2). All HS cases showed abnormal calbindin expression in the dentate gyrus, with 84% in the HS-P group and 60% in the HS-NP group showing a marked reduction (grade 2); the remaining cases showed reduced expression restricted to the basal granule cells only (grade 1) (Fig. 1F).

The morphology of reelin-positive cells in the hippocampal white matter was similar in both groups and included small round cells with a thin peripheral rim of cytoplasmic labelling in addition to larger cells with neuronal morphology and bipolar orientation, reminiscent of Cajal–Retzius cells (Fig. 1G). Quantitative analysis revealed no significant difference in the density of reelin-labelled cells between the HS-P and HS-NP groups (Table 2). Frequent MCM2-positive nuclei of small immature cells were observed in the GCL in most cases (Fig. 1H); we have previously reported that these cells were mainly negative with mature glial or microglial markers (Thom *et al.* 2005*b*). Cells of similar morphology and distribution in the dentate gyrus, with bipolar or delicate multipolar cytoplasmic process, were highlighted with DCX immunohistochemistry through the dentate gyrus and subgranular zone (Fig. 1I). Quantitative analysis, however, revealed no difference in MCM2 or DCX cell densities in the dentate gyrus between the groups (Fig. 2).

CBR1 immunohistochemistry revealed dense networks of processes in the hippocampus (Fig. 1J–M), with fibres and synaptic-like staining in the neuropil,



Fig. 1. (A) NeuN (neuronal nuclear antigen)-stained section from a patient with hippocampal sclerosis (HS) and post-operative psychosis (HS-P) with the classical pattern of neuronal loss in Cornu ammonis (CA) area CA4 and CA1 with sparing of the CA2 region (International League Against Epilepsy type 1). (B) Glial fibrillary acidic protein (GFAP)-δ labelling confirmed cells with astrocytic morphology in the regions of neuronal loss, particularly in the subgranular zone of the dentate gyrus (C). (D) Varying degrees of granule cell dispersion were measured in the HS-P cases with mild dispersion shown on the left and moderate dispersion on the right. (E) Dynorphin staining for mossy fibre sprouting was graded as grade 1 (mild sprouting into the inner molecular layer of the dentate gyrus) and grade 2 (extensive sprouting throughout the molecular layer). (F) Reduction of calbindin expression in the granule cell layer (GCL) was graded as grade 1 (reduction in basal cells only) and grade 2 (extensive loss of calbindin in granule cells): in these figures interneurons remain labelled, confirming technical efficacy of the immunostaining protocol. (G) Reelin immunohistochemistry reveals distinctively positive cytoplasmic labelling of a population of small round cells and larger cells with horizontal processes in the hippocampal white matter. (H) Mini-chromosome maintenance protein 2 (MCM2) immunostaining highlights positive nuclear labelling of small immature cells (compared with the size of the granule cell neuron); similar small cells also show cytoplasmic labelling for doublecortin (DCX) with bipolar processes (I), through the GCL and subgranular zone (arrowheads in both). (J) Cannabinoid type 1 receptor (CBR1) staining in the dentate gyrus and molecular layer shows diffuse synaptic-like staining surrounding the neurones in a HS case without psychosis (HS-NP); in some cases, and more frequent in HS-P cases, diminished staining was



Fig. 2. Scaled bar charts of measurements of dentate gyrus (DG) parameters for patients with hippocampal sclerosis (HS) who developed interictal psychosis (HS-P) post-operatively and a matched surgical HS group without psychosis (HS-NP). On the y axis the units for granule cell dispersion (GCD) mean and maximum (Max) are μ m×10¹. Reelin, mini-chromosome maintenance protein 2 (MCM2) and doublecortin (DCX) are expressed as cells per×10⁻⁵/ μ m². The cannabinoid type 1 receptor (CBR1) labelling index is shown as the percentage of immunolabelling over each subfield (DG, Cornu ammonis (CA) area CA4 and the subiculum). Values are means, with standard deviations represented by vertical bars. * Mean values were significantly different (*p*<0.05).

sometimes observed surrounding residual negatively stained neuronal cell bodies (Fig. 1M). This pattern is in keeping with previous descriptions of CBR1 expression in HS (Magloczky et al. 2010). In all HS samples, there was an impression of reduced CBR1 labelling in the CA4 subfield compared with the dentate gyrus and subiculum. Quantitative analysis confirmed significant differences in CBR1 labelling between the three subfields over all cases (p < 0.01). The mean CBR1 labelling fractions in all three regions (dentate gyrus, CA4 and subiculum), although lower in the HS-P group compared with the HS-NP group (Fig. 1G, H and Fig. 2), were not statistically significantly different (Table 2). There was no significant difference in the CBR1 labelling in the dentate gyrus in relation to the severity of granule cell loss, suggesting that this trend for group differences was independent of the degree of sclerosis.

Clinical-pathological correlations revealed an inverse correlation between the time interval to the development of psychosis post-operatively to both the mean value of GCD (p<0.05, Spearman's correlation) and CBR1 expression in the subiculum (p=0.05, Spearman's correlation). There was a negative correlation between the CBR1 index in the CA4 region and age of onset of seizures in the HS-P (p=0.005, Spearman's correlation) but not in the HS-NP group.

Discussion

Neuropathological studies of resected tissues from patients developing *de novo* psychosis following temporal lobe surgery for epilepsy are limited. Importantly, they provide a unique opportunity to capture potential cellular responses that may predispose to the development of psychosis without the confounding factor of anti-psychotic treatment-induced changes. The patterns of hippocampal neuronal loss in the HS-P and HS-NP patients in our study were typical of those encountered in epilepsy (82% and 89%, respectively, of patients showing classical or ILAE type 1 HS: neuronal loss and gliosis primarily

noted, and confirmed with quantitative analysis (K). A similar labelling pattern was noted in the subiculum (L) but with less pronounced labelling in the CA4 region in many HS cases (M). Original magnification objectives in (A) and (B)×1.6; in (C) and $(G-M)\times40$; and in $(D-F)\times10$.

involving CA1 and CA4), consistent with reports of 60–80% in large epilepsy series (Blumcke *et al.* 2013).

We found less prominent MFS in HS-P patients compared with the HS-NP group. This is in keeping with recent findings of diminished MFS, as visualized with Timms staining, in TLE patients with interictal psychosis compared with patients with HS and epilepsy alone or depression (Kandratavicius et al. 2012). The mechanisms promoting axonal sprouting in epilepsy are uncertain; experimental models demonstrate MFS after electrical stimulation but it also occurs in association with loss of hilar neurones and their projection pathways to the dentate gyrus (Nadler, 2003). MFS may therefore represent an aberrant reparative mechanism to restore function, but with evidence that its overall effect is to enhance synchronization and facilitate seizures. Local growth factors and components of the extracellular matrix, as GAP-43 and tenascin-C, may promote MFS (Proper et al. 2001; Heck et al. 2004). The maturity of granule cells may also govern the extent of MFS (Cameron et al. 2011; Martinian et al. 2012). Several of these factors may have influenced the diminished axonal sprouting observed in the HS-P patients. It has also been suggested that changes in the dynamics of axonal sprouting or synaptic reorganization in the remaining brain may contribute to the development of psychosis following temporal lobe surgery (Stevens, 1990). It is possible that in our HS-P group, a tendency for diminished axonal sprouting as a reparative response may extend beyond the resected hippocampus or extratemporally and may predispose to the development of psychosis following surgery.

There was no family history of psychosis in our HS-P group but as our sample size was small, the possibility that genetic or epigenetic factors may also contribute to the diminished capacity for axonal sprouting cannot be excluded. For example, the disrupted-in-schizophrenia 1 (*DISC1*) gene has been reported to be associated with major psychiatric disorders including psychosis and schizophrenia (Enomoto *et al.* 2009). Interestingly, DISC1 has a wide range of regulatory functions, including roles in axonal guidance and neurite extension. Clearly, further research into genetic risk factors in patients who develop psychosis following epilepsy surgery is warranted.

GCD is a well-recognized and relatively specific feature of HS in epilepsy. It is associated with hippocampal (typically hilar) neuronal loss, is observed in around 50% of cases in large surgical series and is associated with early onset or long duration of epilepsy (Blumcke *et al.* 2013). A small series of six patients with TLE and interictal psychosis confirmed GCD in 20% of cases compared with 64% of TLE patients without psychosis, although the differences were not significant (Suckling et al. 2000). We also noted significantly less GCD in the HS-P group, but based on the maximal dispersion measurements only, and this was independent of the pattern of HS. Experimental epilepsy models propose that seizure activity itself induces migration of mature granule cells (Chai et al. 2013) and cell 'ectopia' is influenced by y-aminobutyric acid (GABA) activity following febrile seizures (Koyama et al. 2012). Studies in human epilepsy tissues have also proposed that acquired hippocampal reelin deficiency arising in HS has a primary mechanistic role in GCD (Haas & Frotscher, 2010). Decreased reelin protein and expressing cells have been shown in HS in mesial MLE patients (Frotscher et al. 2003) and, in experimental studies, diminished reelin follows induction of seizures with the subsequent development of GCD (Haas & Frotscher, 2010). In this study, we therefore investigated reelin-expressing cell populations in view of this observation as well as the accumulating evidence also implicating the reelin-signalling pathway in schizophrenia. Variations of the reelin gene have been linked to increased risk of schizophrenia in women (Ben-David & Shifman, 2010; Kuang et al. 2011). Reduced reelin-expressing cells have been shown in multiple brain regions including the frontal cortex and hippocampus in schizophrenia (Impagnatiello et al. 1998; Fatemi, 2005; Eastwood & Harrison, 2006, Habl et al. 2012) and, in a meta-analysis, this reduction only approached significance in the ML of the dentate gyrus and declined with disease progression (Knable et al. 2004). Abnormalities in the reelin-signalling pathway could potentially link the association of epilepsy and psychosis. We found lower Cajal-Retzius cell numbers in the HS-P group but the differences were not statistically significant. There was also no association between cell number and the period to the onset of psychosis post-operatively. This lack of significance could be attributed to the relatively small sample size in our study and does not exclude a role for reelin in interictal psychosis.

An 'immature dentate gyrus' has been proposed as an underlying template of both experimental and human studies of schizophrenia, characterized by increased numbers of immature neuronal progenitors and a lack of calbindin-positive mature neurons (Walton *et al.* 2012; Shin *et al.* 2013). Studies have identified reduced levels of neural stem cell proliferation in the dentate gyrus as quantified by Ki-67 immunohistochemistry in post-mortem studies in schizophrenia (Reif *et al.* 2006). In patients with epilepsy and HS, the residual proliferative capacity of the dentate gyrus and isolation of stem cells has been shown to directly correlate with memory function (Coras *et al.* 2010). We utilized MCM2 as a marker of regenerative capacity in the dentate gyrus, which labels cells licenced for replication, expressed through the cell cycle, and therefore is a more sensitive marker than Ki-67 in fixed tissue samples (von Bohlen und Halbach, 2011). It has been used in previous studies of HS (Thom et al. 2005b; Fahrner et al. 2007) where small immunopositive nuclei were observed to co-localize with immature nestin-positive cells, and a minority co-localizing with mature glial/microglial markers (Thom et al. 2005b). We confirmed MCM2positive cells in both patient groups which were morphologically similar to DCX-positive cells in the dentate gyrus. Similar DCX-positive cells have been recently described in the infant dentate gyrus (Paine et al. 2013) but such immature-appearing DCX-positive cells have not previously been reported in the adult human hippocampus or in HS (D'Alessio et al. 2010). There were, however, no group differences in the numbers of DCX or MCM2 cells to suggest altered dentate maturation or neurogenesis in HS-P relative to HS-NP patients. It is possible that seizures and duration of epilepsy have a more over-riding influence on the numbers of DCX-positive cells.

Calbindin expression in the granule cells reaches full maturation by 11 years in humans (Abraham et al. 2011). Loss of calbindin expression in granule cells has been reported in TLE/HS with complete loss varying between 13% and 35% of cases in studies (Magloczky et al. 1997; Arellano et al. 2004; Martinian et al. 2012); it is also documented in experimental models of epilepsy (Shin et al. 2013). Loss of calbindin expression in granule cells has been directly correlated with memory impairment (Karadi et al. 2012) and a reduction noted in patients with schizophrenia and bipolar disorder (Walton et al. 2012). Gene expression studies of dentate granule cells in post-mortem schizophrenic brains have shown significant reductions in calbindin (Altar et al. 2005). A previous study of only three patients with psychosis following epilepsy surgery did not identify a significant reduction in calbindin in the dentate granule cells compared with controls (Suckling et al. 2000). In our study, marked loss of calbindin was more common in HS-P than HS-NP patients (84% v. 60%); although this was not statistically analysed this could suggest that loss of calbindin expression may be a risk factor for developing interictal psychosis following epilepsy surgery.

The endocannabinoid system has been implicated in schizophrenia (Fernandez-Espejo *et al.* 2009). Cannabis use is associated with acute psychotic episodes and an increased risk of developing schizophrenia in the long term. The main receptor, CBR1, is widely and abundantly expressed in the CNS, primarily at pre-synaptic terminals. Released endocannabinoids act as retrograde signal molecules, mediating glutamatergic and GABAergic neurotransmission, with an overall net effect considered to be inhibitory (Katona & Freund, 2012). Genetic alterations in the CNR1 gene which codes for CBR1 have been shown in hebephrenic schizophrenia (Chavarria-Siles et al. 2008) and polymorphisms associated with pharmacological responses antipsychotic medications in schizophrenia to (Hamdani et al. 2008). Elevated binding of CBR1 has also been shown in photon emission computerized tomography (PET) imaging in schizophrenic patients (Wong et al. 2010). Post-mortem findings in schizophrenia have reported an up-regulation of CBR1 in the frontal (Dean et al. 2001) and cingulate cortex (Zavitsanou et al. 2004) using autoradiography, although no tissue studies, to the best of our knowledge, have been carried out on the hippocampus. In post-mortem studies, antipsychotic medications may influence receptor-binding studies (Fernandez-Espejo et al. 2009) whereas, in contrast, all patients in our study were naive to antipsychotic medications at the time of tissue resection. Furthermore, only one patient in the HS-P group had a history of cannabis use.

In epilepsy, the endocannabinoid system is considered to have both anticonvulsive and neuroprotective effects (Monory et al. 2006). PET studies using the ligand [¹⁸F]MK-9470 showed an increase in CBR1 receptor availability associated with HS, suggesting an adaptive protective mechanism of neurons against hyperexcitability and seizure activity (Goffin et al. 2011). In tissue resections from HS patients there is evidence for an overall reduction in CBR1 (Ludanyi et al. 2008) although relative increases in CBR1 in the dentate gyrus in HS have been shown, primarily associated with GABAergic cell terminals (Magloczky et al. 2010) and confirmed in experimental models (Karlocai et al. 2011). These findings therefore support alteration of the endocannabinoid system in HS and epilepsy. In our study, we also noted a similar pattern of alteration of the CBR1 receptor system in all HS cases including significant increases in the dentate gyrus relative to CA4. There was a qualitative impression of a reduction in hippocampal subfield CBR1 in HS-P compared with HS-NP patients although the differences were not statistically significant. Further studies at a molecular-genetic level, as well as receptor affinity studies to explore the role of CBR1 receptor in epilepsy with co-morbid psychosis, are warranted.

The clinical characteristics of our HS-P group are in keeping with previous reports, with the majority being male with left-sided HS and not seizure free following surgery (Flor-Henry, 1969; Lindsay *et al.* 1979; Glosser *et al.* 2000; Maier *et al.* 2000; Inoue & Mihara, 2001; Kanner *et al.* 2009). Our observations therefore support the preponderance of involvement of the dominant hemisphere in psychotic patients with epilepsy which, in the current series, revealed itself only following surgical intervention. We can speculate the existence of lateralized neuronal networks that become altered in TLE/HS, but with different responses following surgery (as has been shown for lateralization of dynorphin levels following traumatic brain injury as an example; Hussain *et al.* 2012) which ultimately, in these patients, predisposes to the psychotic state. However, we did not identify any clear relationships between the clinical variables, laterality and pathology measurements in the HS-P group.

Several limitations to our study should be considered. The number of cases in our series was small due to the rarity of the condition. A normal (nonepilepsy) hippocampal surgical control group was not available. With quantitative immunohistochemistry, as always, variations in tissue processing and fixation can influence the intensity of immunolabelling; we aimed to minimize this by closely matching our control group for both clinical demographics as well as the year of surgery. Furthermore, pre-operative medications, including anti-epileptic drugs, may have influenced CBR1 expression although both patient groups would have been similarly affected as most patients were on polytherapy.

In summary, we report here the largest published series of patients with HS who developed interictal psychosis following epilepsy surgery. We identified a pattern of reduced mossy fibre axonal sprouting, dispersion of granule cells and calbindin expression as well as qualitative differences in hippocampal CBR1 expression. These findings suggest that subtle differences in the hippocampal circuitry and cellular response to seizures may predispose to the development of psychosis following epilepsy surgery.

Acknowledgements

This work was undertaken at University College London Hospital/University College London, which received a proportion of funding from the Department of Health's National Institute for Health Research (NIHR) Biomedical Research Centre's funding scheme. Neuropathology components of this work and J.L. and C.R. are supported by a grant from the Medical Research Council (MR/J01270X/1).

Declaration of Interest

None.

References

Abraham H, Meyer G (2003). Reelin-expressing neurons in the postnatal and adult human hippocampal formation. *Hippocampus* **13**, 715–727.

- Abraham H, Richter Z, Gyimesi C, Horvath Z, Janszky J, Doczi T, Seress L (2011). Degree and pattern of calbindin immunoreactivity in granule cells of the dentate gyrus differ in mesial temporal sclerosis, cortical malformationand tumor-related epilepsies. *Brain Research* 1399, 66–78.
- Altar CA, Jurata LW, Charles V, Lemire A, Liu P, Bukhman Y, Young TA, Bullard J, Yokoe H, Webster MJ, Knable MB, Brockman JA (2005). Deficient hippocampal neuron expression of proteasome, ubiquitin, and mitochondrial genes in multiple schizophrenia cohorts. *Biological Psychiatry* **58**, 85–96.
- Andermann LF, Savard G, Meencke HJ, McLachlan R, Moshe S, Andermann F (1999). Psychosis after resection of ganglioglioma or DNET: evidence for an association. *Epilepsia* **40**, 83–87.
- **APA** (2000). *Diagnostic and Statistical Manual of Mental Disorders*, 4th edn, text-revision. American Psychological Association: Washington, DC.
- Arellano JI, Munoz A, Ballesteros-Yanez I, Sola RG, DeFelipe J (2004). Histopathology and reorganization of chandelier cells in the human epileptic sclerotic hippocampus. *Brain* **127**, 45–64.
- **Ben-David E, Shifman S** (2010). Further investigation of the association between rs7341475 and rs17746501 and schizophrenia. *Americal Journal of Medical Genetics, Part B Neuropsychiatric Genetics* **153B**, 1244–1247.
- Blumcke I (2009). Neuropathology of focal epilepsies: a critical review. *Epilepsy Behaviour* 15, 34–39.
- Blumcke I, Coras R, Miyata H, Ozkara C (2012). Defining clinico-neuropathological subtypes of mesial temporal lobe epilepsy with hippocampal sclerosis. Brain Pathology 22, 402–411.
- Blumcke I, Kistner I, Clusmann H, Schramm J, Becker AJ, Elger CE, Bien CG, Merschhemke M, Meencke HJ, Lehmann T, Buchfelder M, Weigel D, Buslei R, Stefan H, Pauli E, Hildebrandt M (2009). Towards a clinicopathological classification of granule cell dispersion in human mesial temporal lobe epilepsies. *Acta Neuropathologica* **117**, 535–544.
- Blumcke I, Thom M, Aronica E, Armstrong DD,
 Bartolomei F, Bernasconi A, Bernasconi N, Bien CG,
 Cendes F, Coras R, Cross JH, Jacques TS, Kahane P,
 Mathern GW, Miyata H, Moshe SL, Oz B, Ozkara C,
 Perucca E, Sisodiya S, Wiebe S, Spreafico R (2013).
 International consensus classification of hippocampal
 sclerosis in temporal lobe epilepsy: a Task Force report from
 the ILAE Commission on Diagnostic Methods. *Epilepsia* 54, 1315–1329.
- Briellmann RS, Kalnins RM, Hopwood MJ, Ward C, Berkovic SF, Jackson GD (2000). TLE patients with postictal psychosis: mesial dysplasia and anterior hippocampal preservation. *Neurology* 55, 1027–1030.
- Cameron MC, Zhan RZ, Nadler JV (2011). Morphologic integration of hilar ectopic granule cells into dentate gyrus circuitry in the pilocarpine model of temporal lobe epilepsy. *Journal of Comparative Neurology* 519, 2175–2192.
- Chai X, Munzner G, Zhao S, Tinnes S, Kowalski J, Haussler U, Young C, Haas CA, Frotscher M (2013). Epilepsy-induced motility of differentiated neurons.

Cerebral Cortex. Published online 15 March 2013. doi:10.1093/cercor/bht067.

- Chavarria-Siles I, Contreras-Rojas J, Hare E, Walss-Bass C, Quezada P, Dassori A, Contreras S, Medina R,
 Ramirez M, Salazar R, Raventos H, Escamilla MA (2008).
 Cannabinoid receptor 1 gene (*CNR1*) and susceptibility to a quantitative phenotype for hebephrenic schizophrenia. *American Journal of Medical Genetics Part B Neuropsychiatric Genetics* 147, 279–284.
- Choi H, Sell RL, Lenert L, Muennig P, Goodman RR, Gilliam FG, Wong JB (2008). Epilepsy surgery for pharmacoresistant temporal lobe epilepsy: a decision analysis. *Journal of the American Medical Association* 300, 2497–2505.
- Christodoulou C, Koutroumanidis M, Hennessy MJ, Elwes RD, Polkey CE, Toone BK (2002). Postictal psychosis after temporal lobectomy. *Neurology* **59**, 1432–1435.
- Cleary RA, Thompson PJ, Fox Z, Foong J (2012). Predictors of psychiatric and seizure outcome following temporal lobe epilepsy surgery. *Epilepsia* 53, 1705–1712.
- Coras R, Siebzehnrubl FA, Pauli E, Huttner HB, Njunting M, Kobow K, Villmann C, Hahnen E, Neuhuber W, Weigel D, Buchfelder M, Stefan H, Beck H, Steindler DA, Blumcke I (2010). Low proliferation and differentiation capacities of adult hippocampal stem cells correlate with memory dysfunction in humans. *Brain* 133, 3359–3372.
- Cotter D, Wilson S, Roberts E, Kerwin R, Everall IP (2000). Increased dendritic MAP2 expression in the hippocampus in schizophrenia. *Schizophrenia Research* **41**, 313–323.
- D'Alessio L, Konopka H, Lopez EM, Seoane E, Consalvo D, Oddo S, Kochen S, Lopez-Costa JJ (2010). Doublecortin (DCX) immunoreactivity in hippocampus of chronic refractory temporal lobe epilepsy patients with hippocampal sclerosis. *Seizure* **19**, 567–572.
- **Dean B, Sundram S, Bradbury R, Scarr E, Copolov D** (2001). Studies on [³H]CP-55940 binding in the human central nervous system: regional specific changes in density of cannabinoid-1 receptors associated with schizophrenia and cannabis use. *Neuroscience* **103**, 9–15.
- Eastwood SL, Harrison PJ (2006). Cellular basis of reduced cortical reelin expression in schizophrenia. *American Journal* of Psychiatry 163, 540–542.
- Enomoto A, Asai N, Namba T, Wang Y, Kato T, Tanaka M, Tatsumi H, Taya S, Tsuboi D, Kuroda K, Kaneko N, Sawamoto K, Miyamoto R, Jijiwa M, Murakumo Y, Sokabe M, Seki T, Kaibuchi K, Takahashi M (2009). Roles of disrupted-in-schizophrenia 1-interacting protein girdin in postnatal development of the dentate gyrus. *Neuron* 63, 774–787.
- Fahrner A, Kann G, Flubacher A, Heinrich C, Freiman TM, Zentner J, Frotscher M, Haas CA (2007). Granule cell dispersion is not accompanied by enhanced neurogenesis in temporal lobe epilepsy patients. *Experimental Neurology* **203**, 320–332.
- Fatemi SH (2005). Reelin glycoprotein in autism and schizophrenia. International Review of Neurobiology 71, 179–187.

Fernandez-Espejo E, Viveros MP, Nunez L, Ellenbroek BA, Rodriguez de Fonseca F (2009). Role of cannabis and endocannabinoids in the genesis of schizophrenia. *Psychopharmacology (Berlin)* **206**, 531–549.

Flor-Henry P (1969). Psychosis and temporal lobe epilepsy. A controlled investigation. *Epilepsia* **10**, 363–395.

- Flugel D, Cercignani M, Symms MR, Koepp MJ, Foong J (2006). A magnetization transfer imaging study in patients with temporal lobe epilepsy and interictal psychosis. *Biological Psychiatry* **59**, 560–567.
- Folsom TD, Fatemi SH (2013). The involvement of reelin in neurodevelopmental disorders. *Neuropharmacology* 68, 122–135.
- Foong J, Flugel D (2007). Psychiatric outcome of surgery for temporal lobe epilepsy and presurgical considerations. *Epilepsy Research* 75, 84–96.
- **Frotscher M, Haas CA, Förster E** (2003). Reelin controls granule cell migration in the dentate gyrus by acting on the radial glial scaffold. *Cerebral Cortex* **13**, 634–640.
- Garey L (2010). When cortical development goes wrong: schizophrenia as a neurodevelopmental disease of microcircuits. *Journal of Anatomy* 217, 324–333.
- Glosser G, Zwil AS, Glosser DS, O'Connor MJ, Sperling MR (2000). Psychiatric aspects of temporal lobe epilepsy before and after anterior temporal lobectomy. *Journal of Neurology, Neurosurgery and Psychiatry* **68**, 53–58.
- **Goffin K, Van Paesschen W, Van Laere K** (2011). *In vivo* activation of endocannabinoid system in temporal lobe epilepsy with hippocampal sclerosis. *Brain* **134**, 1033–1040.
- Haas CA, Frotscher M (2010). Reelin deficiency causes granule cell dispersion in epilepsy. *Experimental Brain Research* 200, 141–149.
- Habl G, Schmitt A, Zink M, von Wilmsdorff M, Yeganeh-Doost P, Jatzko A, Schneider-Axmann T, Bauer M, Falkai P (2012). Decreased reelin expression in the left prefrontal cortex (BA9) in chronic schizophrenia patients. *Neuropsychobiology* 66, 57–62.
- Hamdani N, Tabeze JP, Ramoz N, Ades J, Hamon M, Sarfati Y, Boni C, Gorwood P (2008). The *CNR1* gene as a pharmacogenetic factor for antipsychotics rather than a susceptibility gene for schizophrenia. *European Neuropsychopharmacology* **18**, 34–40.
- Heck N, Garwood J, Loeffler JP, Larmet Y, Faissner A (2004). Differential upregulation of extracellular matrix molecules associated with the appearance of granule cell dispersion and mossy fiber sprouting during epileptogenesis in a murine model of temporal lobe epilepsy. *Neuroscience* **129**, 309–324.
- Hussain ZM, Fitting S, Watanabe H, Usynin I, Yakovleva T, Knapp PE, Scheff SW, Hauser KF, Bakalkin G (2012). Lateralized response of dynorphin A peptide levels after traumatic brain injury. *Journal of Neurotrauma* 29, 1785–1793.
- Impagnatiello F, Guidotti AR, Pesold C, Dwivedi Y, Caruncho H, Pisu MG, Uzunov DP, Smalheiser NR, Davis JM, Pandey GN, Pappas GD, Tueting P, Sharma RP, Costa E (1998). A decrease of reelin expression as a putative vulnerability factor in schizophrenia. *Proceedings of the National Academy of Sciences USA* 95, 15718–15723.

Inoue Y, Mihara T (2001). Psychiatric disorders before and after surgery for epilepsy. *Epilepsia* **42** (Suppl. 6), 13–18.

Kandratavicius L, Hallak JE, Young LT, Assirati JA, Carlotti CG Jr., Leite JP (2012). Differential aberrant sprouting in temporal lobe epilepsy with psychiatric co-morbidities. *Psychiatry Research* **195**, 144–150.

Kanner AM, Byrne R, Chicharro A, Wuu J, Frey M (2009). A lifetime psychiatric history predicts a worse seizure outcome following temporal lobectomy. *Neurology* **72**, 793–799.

Karadi K, Janszky J, Gyimesi C, Horvath Z, Lucza T, Doczi T, Kallai J, Abraham H (2012). Correlation between calbindin expression in granule cells of the resected hippocampal dentate gyrus and verbal memory in temporal lobe epilepsy. *Epilepsy Behaviour* 25, 110–119.

Karlocai MR, Toth K, Watanabe M, Ledent C, Juhasz G, Freund TF, Magloczky Z (2011). Redistribution of CB1 cannabinoid receptors in the acute and chronic phases of pilocarpine-induced epilepsy. *PloS ONE* 6, e27196.

Katona I, Freund TF (2008). Endocannabinoid signaling as a synaptic circuit breaker in neurological disease. *Nature Medicine* 14, 923–930.

Katona I, Freund TF (2012). Multiple functions of endocannabinoid signaling in the brain. *Annual Review of Neuroscience* 35, 529–558.

Kirov II, Hardy CJ, Matsuda K, Messinger J, Cankurtaran CZ, Warren M, Wiggins GC, Perry NN, Babb JS, Goetz RR, George A, Malaspina D, Gonen O (2013). *In vivo* 7 Tesla imaging of the dentate granule cell layer in schizophrenia. *Schizophrenia Research* 147, 362–367.

Knable MB, Barci BM, Webster MJ, Meador-Woodruff J, Torrey EF (2004). Molecular abnormalities of the hippocampus in severe psychiatric illness: postmortem findings from the Stanley Neuropathology Consortium. *Molecular Psychiatry* 9, 609–620.

Koyama R, Tao K, Sasaki T, Ichikawa J, Miyamoto D, Muramatsu R, Matsuki N, Ikegaya Y (2012). GABAergic excitation after febrile seizures induces ectopic granule cells and adult epilepsy. *Nature Medicine* **18**, 1271–1278.

Kuang WJ, Sun RF, Zhu YS, Li SB (2011). A new single-nucleotide mutation (rs362719) of the reelin (*RELN*) gene associated with schizophrenia in female Chinese Han. *Genetetics and Molecular Research* **10**, 1650–1658.

Lakatosova S, Ostatnikova D (2012). Reelin and its complex involvement in brain development and function. *International Journal of Biochemistry and Cell Biology* 44, 1501–1504.

Leinonen E, Tuunainen A, Lepola U (1994). Postoperative psychoses in epileptic patients after temporal lobectomy. *Acta Neurologica Scandinavica* **90**, 394–399.

Lindsay J, Ounsted C, Richards P (1979). Long-term outcome in children with temporal lobe seizures. III: Psychiatric aspects in childhood and adult life. *Developmental Medicine and Child Neurology* **21**, 630–636.

Ludanyi A, Eross L, Czirjak S, Vajda J, Halasz P, Watanabe M, Palkovits M, Magloczky Z, Freund TF, Katona I (2008). Downregulation of the CB1 cannabinoid receptor and related molecular elements of the endocannabinoid system in epileptic human hippocampus. *Journal of Neuroscience* **28**, 2976–2990.

Luzi S, Morrison PD, Powell J, di Forti M, Murray RM (2008). What is the mechanism whereby cannabis use increases risk of psychosis? *Neurotoxicity Research* 14, 105–112.

Mace CJ, Trimble MR (1991). Psychosis following temporal lobe surgery: a report of six cases. *Journal of Neurology, Neurosurgery and Psychiatry* 54, 639–644.

Magloczky Z, Halasz P, Vajda J, Czirjak S, Freund TF (1997). Loss of calbindin-D28 K immunoreactivity from dentate granule cells in human temporal lobe epilepsy. *Neuroscience* 76, 377–385.

Magloczky Z, Toth K, Karlocai R, Nagy S, Eross L,
Czirjak S, Vajda J, Rasonyi G, Kelemen A, Juhos V,
Halasz P, Mackie K, Freund TF (2010). Dynamic changes of CB1-receptor expression in hippocampi of epileptic mice and humans. *Epilepsia* 51 (Suppl. 3), 115–120.

Maier M, Mellers J, Toone B, Trimble M, Ron MA (2000). Schizophrenia, temporal lobe epilepsy and psychosis: an *in vivo* magnetic resonance spectroscopy and imaging study of the hippocampus/amygdala complex. *Psychological Medicine* **30**, 571–581.

Manchanda R, Miller H, McLachlan RS (1993). Post-ictal psychosis after right temporal lobectomy. *Journal of Neurology, Neurosurgery and Psychiatry* **56**, 277–279.

Marshall EJ, Syed GM, Fenwick PB, Lishman WA (1993). A pilot study of schizophrenia-like psychosis in epilepsy using single-photon emission computerised tomography. *British Journal of Psychiatry* **163**, 32–36.

Martinian L, Boer K, Middeldorp J, Hol EM, Sisodiya SM, Squier W, Aronica E, Thom M (2009). Expression patterns of glial fibrillary acidic protein (GFAP)-delta in epilepsy-associated lesional pathologies. Neuropathology and Applied Neurobiology **35**, 394–405.

Martinian L, Catarino CB, Thompson P, Sisodiya SM, Thom M (2012). Calbindin D28 K expression in relation to granule cell dispersion, mossy fibre sprouting and memory impairment in hippocampal sclerosis: a surgical and post mortem series. *Epilepsy Research* **98**, 14–24.

Mellers JD, Adachi N, Takei N, Cluckie A, Toone BK, Lishman WA (1998). SPET study of verbal fluency in schizophrenia and epilepsy. *British Journal of Psychiatry* **173**, 69–74.

Monory K, Massa F, Egertova M, Eder M, Blaudzun H, Westenbroek R, Kelsch W, Jacob W, Marsch R, Ekker M, Long J, Rubenstein JL, Goebbels S, Nave KA, During M, Klugmann M, Wolfel B, Dodt HU, Zieglgansberger W, Wotjak CT, Mackie K, Elphick MR, Marsicano G, Lutz B (2006). The endocannabinoid system controls key epileptogenic circuits in the hippocampus. *Neuron* **51**, 455–466.

Nadkarni S, Arnedo V, Devinsky O (2007). Psychosis in epilepsy patients. *Epilepsia* 48 (Suppl. 9), 17–19.

Nadler JV (2003). The recurrent mossy fiber pathway of the epileptic brain. *Neurochemistry Research* **28**, 1649–1658.

Paine S, Willsher A, Nicholson S, Sebire N, Jacques T (2013). Characterisation of a population of neural progenitor cells in the infant hippocampus. *Neuropathology* *and Applied Neurobiology*. Published online 6 June 2013. doi:10.1111/nan.12065.

Perez MM, Trimble MR (1980). Epileptic psychosis– diagnostic comparison with process schizophrenia. *British Journal of Psychiatry* **137**, 245–249.

Proper EA, Jansen GH, van Veelen CW, van Rijen PC, Gispen WH, de Graan PN (2001). A grading system for hippocampal sclerosis based on the degree of hippocampal mossy fiber sprouting. *Acta Neuropathologica* **101**, 405–409.

Radhakrishnan A, Radhakrishnan K, Radhakrishnan VV, Mary PR, Kesavadas C, Alexander A, Sarma PS (2007). Corpora amylacea in mesial temporal lobe epilepsy: clinico-pathological correlations. *Epilepsy Research* 74, 81–90.

Reif A, Fritzen S, Finger M, Strobel A, Lauer M, Schmitt A, Lesch KP (2006). Neural stem cell proliferation is decreased in schizophrenia, but not in depression. *Molecular Psychiatry* **11**, 514–522.

Roberts GW, Done DJ, Bruton C, Crow TJ (1990). A "mock up" of schizophrenia: temporal lobe epilepsy and schizophrenia-like psychosis. *Biological Psychiatry* 28, 127–143.

Rosoklija G, Toomayan G, Ellis SP, Keilp J, Mann JJ, Latov N, Hays AP, Dwork AJ (2000). Structural abnormalities of subicular dendrites in subjects with schizophrenia and mood disorders: preliminary findings. *Archives of General Psychiatry* **57**, 349–356.

Shaw P, Mellers J, Henderson M, Polkey C, David AS, Toone BK (2004). Schizophrenia-like psychosis arising *de novo* following a temporal lobectomy: timing and risk factors. *Journal of Neurology, Neurosurgery and Psychiatry* 75, 1003–1008.

Shin R, Kobayashi K, Hagihara H, Kogan JH, Miyake S, Tajinda K, Walton NM, Gross AK, Heusner CL, Chen Q, Tamura K, Miyakawa T, Matsumoto M (2013). The immature dentate gyrus represents a shared phenotype of mouse models of epilepsy and psychiatric disease. *Bipolar Disorder*. Published online 6 April 2013. doi:10.1111/ bdi.12064.

Stevens JR (1990). Psychiatric consequences of temporal lobectomy for intractable seizures: a 20–30-year follow-up of 14 cases. *Psychologcal Medicine* 20, 529–545.

Stranahan AM, Erion JR, Wosiski-Kuhn M (2013). Reelin signaling in development, maintenance, and plasticity of neural networks. *Ageing Research Reviews* **12**, 815–822.

Suckling J, Roberts H, Walker M, Highley JR, Fenwick P, Oxbury J, Esiri MM (2000). Temporal lobe epilepsy with and without psychosis: exploration of hippocampal pathology including that in subpopulations of neurons defined by their content of immunoreactive calcium-binding proteins. *Acta Neuropathologica* **99**, 547–554.

Sutula T, Cascino G, Cavazos J, Parada I, Ramirez L (1989). Mossy fiber synaptic reorganization in the epileptic human temporal lobe. Annals of Neurology **26**, 321–330.

Tamminga CA, Stan AD, Wagner AD (2010). The hippocampal formation in schizophrenia. *American Journal* of Psychiatry 167, 1178–1193. Thom M, Liagkouras I, Elliot KJ, Martinian L, Harkness W, McEvoy A, Caboclo LO, Sisodiya SM (2010). Reliability of patterns of hippocampal sclerosis as predictors of postsurgical outcome. *Epilepsia* 51, 1801–1808.

Thom M, Martinian L, Catarino C, Yogarajah M, Koepp MJ, Caboclo L, Sisodiya SM (2009). Bilateral reorganization of the dentate gyrus in hippocampal sclerosis: a postmortem study. *Neurology* 73, 1033–1040.

Thom M, Martinian L, Sisodiya SM, Cross JH, Williams G, Stoeber K, Harkness W, Harding BN (2005a). Mcm2 labelling of balloon cells in focal cortical dysplasia. *Neuropathology and Applied Neurobiology* **31**, 580–588.

Thom M, Martinian L, Williams G, Stoeber K, Sisodiya SM (2005b). Cell proliferation and granule cell dispersion in human hippocampal sclerosis. *Journal of Neuropathology and Experimental Neurology* **64**, 194–201.

Thom M, Sisodiya SM, Beckett A, Martinian L, Lin WR, Harkness W, Mitchell TN, Craig J, Duncan J, Scaravilli F (2002). Cytoarchitectural abnormalities in hippocampal sclerosis. *Journal of Neuropathology and Experimental Neurology* **61**, 510–519.

Toone BK (2000). The psychoses of epilepsy. *Journal of Neurology, Neurosurgery and Psychiatry* **69**, 1–3.

Trimble M (1992). Neurological models of schizophrenia – regional implications. *Clinical Neuropharmacology* 15 (Suppl. 1 Pt. A), 395A–396A.

Ujike H, Morita Y (2004). New perspectives in the studies on endocannabinoid and cannabis: cannabinoid receptors and schizophrenia. *Journal of Pharmacological Science* 96, 376–381.

von Bohlen und Halbach O (2011). Immunohistological markers for proliferative events, gliogenesis, and neurogenesis within the adult hippocampus. *Cell Tissue Research* **345**, 1–19.

Walton NM, Zhou Y, Kogan JH, Shin R, Webster M, Gross AK, Heusner CL, Chen Q, Miyake S, Tajinda K, Tamura K, Miyakawa T, Matsumoto M (2012). Detection of an immature dentate gyrus feature in human schizophrenia/bipolar patients. *Translational Psychiatry* 2, e135.

Wong DF, Kuwabara H, Horti AG, Raymont V, Brasic J, Guevara M, Ye W, Dannals RF, Ravert HT, Nandi A, Rahmim A, Ming JE, Grachev I, Roy C, Cascella N (2010). Quantification of cerebral cannabinoid receptors subtype 1 (CB1) in healthy subjects and schizophrenia by the novel PET radioligand [¹¹C]OMAR. *NeuroImage* 52, 1505–1513.

Zavitsanou K, Garrick T, Huang XF (2004). Selective antagonist [³H]SR141716A binding to cannabinoid CB1 receptors is increased in the anterior cingulate cortex in schizophrenia. *Progress in Neuropsychopharmacology and Biological Psychiatry* 28, 355–360.

Zhang ZJ, Reynolds GP (2002). A selective decrease in the relative density of parvalbumin-immunoreactive neurons in the hippocampus in schizophrenia. *Schizophrenia Research* 55, 1–10.