

Administration of cannabidiol and imipramine induces antidepressant-like effects in the forced swimming test and increases brain-derived neurotrophic factor levels in the rat amygdala

Réus GZ, Stringari RB, Ribeiro KF, Luft T, Abelaira HM, Fries GR, Aguiar BW, Kapczinski F, Hallak JE, Zuardi AW, Crippa JA, Quevedo J. Administration of cannabidiol and imipramine induces antidepressant-like effects in the forced swimming test and increases brain-derived neurotrophic factor levels in the rat amygdala.

Objective: Cannabidiol is a chemical constituent from *Cannabis sativa* and it has multiple mechanisms of action, including antidepressant effects. The main objective of the present study was to evaluate behavioural and molecular effects induced by administration of cannabidiol and imipramine in rats.

Methods: In the present study, rats were acutely or chronically treated for 14 days once a day with saline, cannabidiol (15, 30 and 60 mg/kg) or imipramine (30 mg/kg) and the animals behaviour was assessed in forced swimming and open-field tests. Afterwards, the prefrontal cortex, hippocampus and amygdala brain-derived neurotrophic factor (BDNF) levels were assessed by enzyme-linked immunosorbent sandwich assay.

Results: We observed that both acute and chronic treatments with imipramine at the dose of 30 mg/kg and cannabidiol at the dose of 30 mg/kg reduced immobility time and increased swimming time; climbing time was increased only with imipramine at the dose of 30 mg/kg, without affecting locomotor activity. In addition, chronic treatment with cannabidiol at the dose of 15 mg/kg and imipramine at the dose of 30 mg/kg increased BDNF levels in the rat amygdala.

Conclusion: In conclusion, our results indicate that cannabidiol has an antidepressant-like profile and could be a new pharmacological target for the treatment of major depression.

Gislaine Z. Réus¹, Roberto B. Stringari¹, Karine F. Ribeiro¹, Tatiana Luft¹, Helena M. Abelaira¹, Gabriel R. Fries², Bianca W. Aguiar², Flávio Kapczinski², Jaime E. Hallak³, Antônio W. Zuardi³, José A. Crippa³, João Quevedo¹

¹Laboratório de Neurociências and Instituto Nacional de Ciência e Tecnologia Translacional em Medicina (INCT-TM), Programa de Pós-Graduação em Ciências da Saúde, Unidade Acadêmica de Ciências da Saúde, Universidade do Extremo Sul Catarinense, Criciúma, SC, Brazil; ²Laboratório de Psiquiatria Molecular and Instituto Nacional de Ciência e Tecnologia Translacional em Medicina (INCT-TM), Centro de Pesquisas, Hospital de Clínicas de Porto Alegre, Porto Alegre, RS, Brazil; and ³Departamento de Neurociências e Ciências do Comportamento and Instituto Nacional de Ciência e Tecnologia Translacional em Medicina (INCT-TM), Faculdade de Medicina de Ribeirão Preto, Universidade de São Paulo, Ribeirão Preto, SP, Brazil

Keywords: brain-derived neurotrophic factor; cannabidiol; depression; forced swimming test; imipramine

Professor João Quevedo, MD, PhD,
Laboratório de Neurociências,
Programa de Pós-Graduação em Ciências da
Saúde,
Unidade Acadêmica de Ciências da Saúde,
Universidade do Extremo Sul Catarinense,
88806-000 Criciúma, SC, Brazil.
Tel: +554834312792;
Fax: +554834312736;
E-mail: quevedo@unesoc.net

Significant outcomes

- Cannabidiol as antidepressant.
- Cannabidiol presented antidepressant-like effects on the forced swimming test.
- Cannabidiol increased BDNF levels in the rat amygdala.

Limitations

The dose of cannabidiol that presented effects on the behaviour test was not the same that presented effect on brain-derived neurotrophic factor (BDNF) levels.

Introduction

Major depression is a serious and disabling psychiatric illness that affects approximately 17% of the population worldwide and has a significant negative impact on public health and productivity (1,2). The current clinically used antidepressants increase the extracellular concentrations of monoamines serotonin or norepinephrine either by inhibiting their reuptake from the synapse or by blocking their degradation by inhibiting monoamine oxidase (3–5). However, the currently used antidepressant drugs show therapeutic efficacies in a maximum of 60–70% of depressive patients, thereby research has been undertaken to find alternative antidepressive treatments (6,7).

The plant *Cannabis sativa* has been used for many centuries. It is known to have therapeutically relevant properties and 400 different identifiable chemical constituents; more than 60 of them are cannabinoids (8). The two main cannabinoids are delta-9-tetrahydrocannabinol and cannabidiol. Cannabidiol is known to antagonise the anxiogenic and psychotomimetic effects of high doses of delta-9-tetrahydrocannabinol (9,10). Cannabinoids exert their effects by interaction with specific endogenous cannabinoid receptors (CB). The CB1 receptor is expressed predominantly in central nervous system, in areas that can mediate most of the effects on cognitive function, pain and short-term memory (hippocampus and cerebral cortex), motor control and coordination (basal ganglia and cerebellum), hypothermia and hyperphagia (hypothalamus) (11), and CB2 expression is restricted to immune cells, T-cells, B-cells, spleen, tonsils and activated microglial cells (12–14).

Cannabidiol is a drug with multiple mechanisms of action (15), including anti-inflammatory effects (16–18), antioxidative and anti-cancer actions (19–21), neuroprotective effects (22–24), regulation of intracellular Ca^{2+} levels (25) and it ameliorates the manifestations of diabetes (26,27). In addition, cannabidiol is known by the action on ischaemia (28), antiepileptic (29,30) and antipsychotic actions (31–34) and anxiolytic effects (35, 36–41), these effects were observed in animal models, as well as in humans. Moreover, it has recently been suggested that the endocannabinoid system may be involved in the pathophysiology of depression (42–45) and that cannabidiol may have agonist

properties at 5-HT_{1A} receptors (46,47), which have been related to the therapeutic effect of antidepressant drugs (48).

Several studies have pointed to the role of BDNF in major depression. In fact, decreased levels of BDNF have been shown in animal models of depression and in patients with depression (49,50). Conversely, administration of antidepressant treatments increases BDNF expression (51) and brain infusion of BDNF produces antidepressant-like actions in rats (50,51).

Thus, the main objective of this study was to evaluate behavioural and molecular effects induced by acute and chronic administration of cannabidiol, imipramine or saline (control group) in rats. The behavioural effects of both drugs were evaluated in the forced swimming test, which is a valid behavioural despair assay widely used for screening antidepressant drugs (52). The BDNF protein levels were measured using an enzyme-linked immunosorbent assay (ELISA) kit in rat prefrontal cortex, hippocampus amygdala acutely and chronically treated with cannabidiol, imipramine or saline.

Materials and methods

Animals

Male adult Wistar rats (60 days old) were obtained from Universidade do Extremo Sul Catarinense (Criciúma, SC, Brazil) breeding colony. They were housed five per cage with food and water available *ad libitum* and were maintained on a 12-h light/dark cycle (lights on at 7:00 h). All experimental procedures involving animals were performed in accordance with the NIH Guide for the Care and Usage of Laboratory Animals and the Brazilian Society for Neuroscience and Behavior (SBNeC) recommendations for animal care and with approval by local Ethics Committee under protocol number 94/2009.

Drugs and treatments

Cannabidiol was obtained from THC-Pharm/STI-Pharm (Frankfurt, Germany) and imipramine, the standard antidepressant, from Novartis Pharmaceutical Industry (São Paulo, Brazil). Different groups of rats ($n = 15$ each) were administered intraperitoneally with saline (control group), imipramine (positive control) (30 mg/kg) or different doses of

cannabidiol (15, 30 and 60 mg/kg) (24) one single time (acute treatment) or over 14 days, once a day (chronic treatment) 60 min before the test sessions. Imipramine was dissolved in saline solution and cannabidiol was suspended in polyoxyethylene-sorbitan monooleate (Tween 80) 2% saline. All treatments were administered in a volume of 1 ml/kg. Rats were tested in the open field and forced swim tests following acute or chronic saline, imipramine and cannabidiol treatments.

Forced swimming test

The forced swimming test was conducted according to previous reports (53–56). The test involves two individual exposures to a cylindrical tank with water in which rats cannot touch the bottom of the tank or escape. The tank is made of transparent Plexiglas, 80 cm tall, 30 cm in diameter and filled with water (22–23 °C) to a depth of 40 cm. In the acute treatment, for the first exposure, rats without drug treatment were placed in the water for 15 min (pretest session). Twenty-four hours later, rats were placed in the water again for a 5-min session (test session). Rats were treated with cannabidiol, imipramine or saline only 60-min before the second exposure to the cylindrical tank of water (test session). During the test session, some behavioural parameters were recorded in seconds, such as immobility time (i.e. no additional activity is observed other than that required to keep the rat's head above the water), climbing time, which is defined as upward-directed movements of the forepaws along the side of the swim chamber, and swimming time (i.e. movement usually horizontal throughout the swim chamber).

In the chronic treatment on the 13th day of chronic treatment, 1 h after drug treatment, rats were individually placed in the cylinder containing water for 15 min (pretest session). On the 14th day, rats received the last intraperitoneal drug treatment and after 1 h they were subjected again to the forced swimming test for a 5-min session (test session) and the immobility, climbing and swimming time of rats were recorded in seconds.

Open-field test

This apparatus consists of a brown plywood arena 45 × 60 cm surrounded by wood 50 cm high walls and containing a frontal glass wall. The floor of the open field was divided into nine rectangles (15 × 20 cm each) by black lines. Animals were gently placed on the left rear quadrant and left to explore the arena. In a separate series of experiments, naïve rats were acutely treated with cannabidiol (15–60 mg/kg), imipramine (30 mg/kg) and saline

60 min before the exposure to the open-field apparatus. In the chronic treatment on 12th day, rats were exposed to the open-field apparatus. The number of horizontal (crossings) and vertical (rearrings) activities performed by each rat during 5-min observation period were counted by an expert observer, in order to assess possible effects of drug treatment on spontaneous locomotor activity.

BDNF analysis

Immediately after the forced swimming test saline, imipramine and cannabidiol-treated rats were killed and the skulls were removed and prefrontal cortex, hippocampus and amygdala were dissected and stored at –70 °C for biochemical analysis. BDNF levels in prefrontal cortex, hippocampus and amygdala were measured by anti-BDNF sandwich-ELISA, according to the manufacturer's instructions (Chemicon, Temecula, CA, USA). Briefly, the rat prefrontal cortex, hippocampus and amygdala were homogenised in phosphate buffer solution with 1 mM phenylmethylsulfonyl fluoride and 1 mM ethylene glycol tetraacetic acid. Microtiter plates (96-well flat bottom) were coated for 24 h with the samples diluted 1:2 in sample diluents and standard curve ranged from 7.8 to 500 pg/ml of BDNF. The plates were then washed four times with sample diluent and a monoclonal anti-BDNF rabbit antibody diluted 1:1000 in sample diluent was added to each well and incubated for 3 h at room temperature. After washing, a peroxidase conjugated anti-rabbit antibody (diluted 1:1000) was added to each well and incubated at room temperature for 1 h. After addition of streptavidin enzyme, substrate and stop solution, the amount of BDNF was determined by absorbance in 450 nm. The standard curve shows a direct relationship between optical density and BDNF concentration. Total protein was measured by Lowry's method using bovine serum albumin as a standard, as previously described by Lowry and et al. (57).

Statistical analysis

All data are presented as mean ± SEM. Differences among experimental groups in the forced swimming and open-field tests and in the assessment of BDNF levels were determined by one-way analysis of variance (ANOVA), followed by Tukey *post hoc* test when ANOVA was significant; *p*-values <0.05 were considered to be statistical significant.

Results

The administration of the standard antidepressant imipramine at the dose of 30 mg/kg and cannabidiol

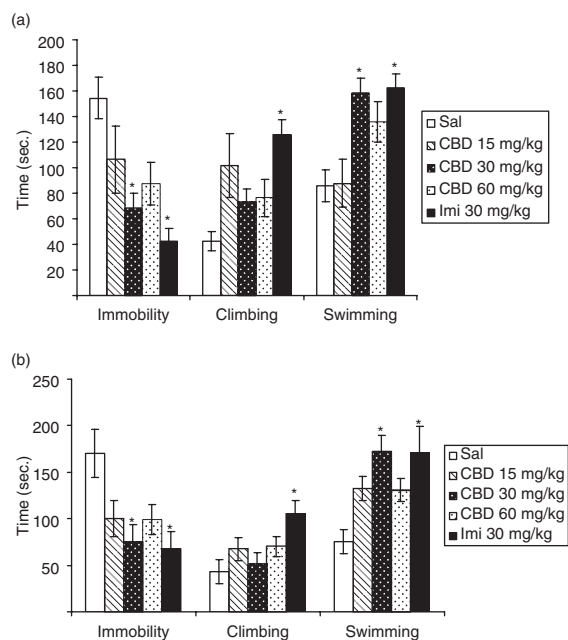


Fig. 1. Effects of acute (a) and chronic (b) administration of cannabidiol (15, 30 and 60 mg/kg), imipramine (30 mg/kg) and saline on the immobility time of rats subjected to the forced swimming test. Bars represent means \pm SEM of 15 rats. * $p < 0.05$ versus saline according to ANOVA followed by Tukey *post hoc* test.

at the dose of 30 mg/kg reduced the immobility time of rats, compared with saline group in both acute (Fig. 1a; $F_{(4-47)} = 4.54$; $p = 0.003$) and chronic (Fig. 1b; $F_{(4-38)} = 3.7$, $p = 0.01$) treatments, and imipramine, but not cannabidiol increased both climbing (Fig. 1a; $F_{(4-43)} = 2.5$, $p = 0.5$ and Fig. 1b; $F_{(4-37)} = 2.89$, $p = 0.03$) and swimming (Fig. 1a; $F_{(4-46)} = 5.5$, $p = 0.001$ and Fig. 1b; $F_{(4-38)} = 5.7$, $p = 0.001$) times. Cannabidiol at the dose of 30 mg/kg increased the swimming time, but not climbing time of rats in both acute and chronic treatments (Fig. 1a; $p = 0.001$ and Fig. 1b; $p = 0.001$).

In the open-field test, acute and chronic treatments with imipramine and cannabidiol at all doses did not modify the number of crossings (acute: $F_{(4-35)} = 0.15$, $p = 0.96$; chronic: $F_{(4-45)} = 0.59$, $p = 0.66$) and rearings (acute: $F_{(4-35)} = 0.76$, $p = 0.55$; chronic: $F_{(4-45)} = 0.44$, $p = 0.77$), compared with saline-treated rats (Table 1).

Figure 2 illustrates the effects of acute and chronic treatments with imipramine (30 mg/kg), cannabidiol (15, 30 and 60 mg/kg) and saline in BDNF protein levels in prefrontal cortex, hippocampus and amygdala. In the acute treatment both imipramine and cannabidiol did not alter BDNF protein levels in prefrontal cortex (Fig. 2a; $F_{(4-32)} = 0.45$, $p = 0.76$), hippocampus (Fig. 2a; $F_{(4-33)} = 0.98$, $p = 0.42$) or amygdala (Fig. 2a; $F_{(4-34)} = 0.33$, $p = 0.85$). In

Table 1. Effects of acute and chronic treatments with cannabidiol (15, 30 and 60 mg/kg), imipramine (30 mg/kg) and saline on rat behaviour in the open-field task

Group	Acute treatment		Chronic treatment	
	Crossing	Rearing	Crossing	Rearing
Saline	45.5 \pm 9.4	22.2 \pm 2.8	29.4 \pm 5.8	10.7 \pm 1.9
CBD 15	39.5 \pm 11.6	21.7 \pm 7.5	27.2 \pm 8.2	12.9 \pm 4.4
CBD 30	40.2 \pm 7.2	12.0 \pm 2.0	24.6 \pm 8.9	7.5 \pm 2.4
CBD 60	42.7 \pm 7.5	20.0 \pm 5.9	26.4 \pm 10.6	11.1 \pm 4.1
IMI 30	48.0 \pm 9.3	17.8 \pm 2.8	40.4 \pm 5.9	12.9 \pm 2.9

Each value shows the mean \pm SEM ($n = 15$). The number of crossings and rearings activity performed by each rat during the 5 min.

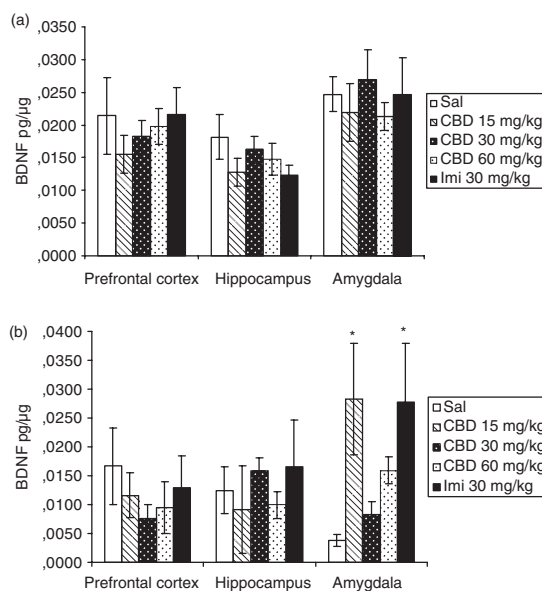


Fig. 2. Effects of acute (a) and chronic (b) administration of cannabidiol (15, 30 and 60 mg/kg), imipramine (30 mg/kg) and saline on the BDNF levels in rat prefrontal cortex, hippocampus and amygdala. Bars represent means \pm SEM of 10 rats. * $p < 0.05$ versus saline according to ANOVA followed by Tukey *post hoc* test.

the chronic treatment, there was an increase in BDNF protein levels in amygdala in rats treated with imipramine at the dose of 30 mg/kg and cannabidiol at the dose of 15 mg/kg, compared with saline group (Fig. 2b; $F_{(4-22)} = 5.38$, $p = 0.004$). In prefrontal cortex (Fig. 2b; $F_{(4-33)} = 0.51$, $p = 0.72$) and in hippocampus (Fig. 2b; $F_{(4-28)} = 0.42$, $p = 0.78$) chronic treatment with imipramine and cannabidiol did not alter BDNF protein levels.

Discussion

It is known that cannabidiol has a wide spectrum of pharmacological actions (17). In this study, we showed that acute and chronic administration with cannabidiol at the dose of 30 mg/kg reduced immobility time and increased swimming time of rats, without affecting locomotor activity.

Other studies have suggested that there are functional interactions between the endogenous cannabinoid system and stress circuitry (43,58), emotional regulation and depression (44,59) and synaptic transmission in the hippocampus (60). In addition, chronic, non-habituating stress resulted in a decrease in functional endogenous cannabinoid signalling within the hippocampus, and the stress-reduced impairment in reversal learning can be reversed by exogenous activation of CB1 receptors (45), suggesting that pharmacological modulation of endogenous cannabinoid signalling could represent a novel approach to the treatment of cognitive deficits that accompany a variety of anxiety-related neuropsychiatric disorders. Moreover, cannabidiol at the dose of 30 mg/kg and imipramine at the dose of 30 mg/kg reduced immobility time in the forced swimming test, without changing exploratory behaviour in mice (45). Furthermore, delta-9-tetrahydrocannabinol and cannabidiol, but not cannabigerol and cannabimol, exhibited a significant antidepressant-like action in mice (42), suggesting these effects may contribute to the overall mood-elevating properties of cannabis.

In this study, we also showed that acute and chronic treatments with imipramine at the dose of 30 mg/kg reduced immobility time and increased both swimming and climbing time in rats, without affecting locomotor activity. Several other studies from our laboratory also have showed that a single injection or chronic administration of imipramine (30 mg/kg) decreased the immobility and increased swimming and climbing times of rats on the forced swimming test, without modifying the locomotor activity (54,55,61,62).

The forced swimming test is a current model and gauges an animal's 'depression-related' responses to acute or chronic inescapable stress (63). In addition, Detke et al. (52) reported that despite the anti-immobility effect antidepressant drugs that enhance noradrenergic neurotransmission increase climbing behaviour, whereas the enhancement of serotonergic neurotransmission increases swimming time in the rat forced swimming test. Our findings indicate that cannabidiol consistently reduced immobility time and significantly increased swimming time, and antidepressant effects observed by cannabidiol may be by serotonergic and noradrenergic neurotransmission actions. In fact, cannabidiol may have properties at 5-HT_{1A} receptors (45–47), which have been related to the therapeutic effect of antidepressant drugs (47). Nevertheless, there are relationships between the serotonin and BDNF (64,65). For example, BDNF stimulates the transcription of genes involved in serotonin function and activation of serotonin receptors by serotonin stimulates expression of the BDNF

factor gene. During brain development this cyclic process promotes outgrowth, synapse formation and survival of serotonin neurons and the eventual innervation of multiple brain regions (66).

BDNF-mediated signalling is involved in neuroplastic responses to stress and antidepressants (63,65). In this data, we investigated the effects of acute and chronic administration of imipramine and cannabidiol in BDNF protein levels in the rat prefrontal cortex, hippocampus and amygdala. These brain areas are implicated in major depression; in addition, the hippocampus has connections with amygdala and prefrontal cortex (67). Our study showed that chronic but not acute treatment increased the BDNF protein levels after administration of imipramine at the dose of 30 mg/kg and cannabidiol at the dose of 15 mg/kg only in the amygdala, suggesting that imipramine and cannabidiol effects are dependent on dose, brain region and treatment regime. In fact, Zanelati et al. (45) showed that both imipramine (30 mg/kg) and cannabidiol (30 mg/kg) did not change hippocampal BDNF levels. Previous studies of our group also showed that acute and chronic administration of imipramine decreased the immobility time of rats in forced swimming test, but did not alter BDNF protein levels in the hippocampus (54,55,61,62); in these studies we did not evaluate the effects of imipramine in amygdala.

Recently, Larsen et al. (68) detailed temporal profiles of the effects of three antidepressants with different pharmacological profiles on the expression of BDNF mRNA and showed a significant increase in BDNF mRNA expression in the granular cell layer after 7 days of treatment with venlafaxine, and after 14 days of treatment with imipramine, but not after 1 day of treatment and a modest decrease in BDNF mRNA expression was observed in the CA3 region after chronic treatment with imipramine. These results indicated that the change in BDNF levels is dependent on treatment time and the region of the hippocampus.

In another study from our group, which investigated the effects of cannabidiol in an animal model of mania induced by D-amphetamine, our group showed that cannabidiol did not have any effect against D-amphetamine-induced hyperactivity, but cannabidiol at the dose of 30 mg/kg reversed the D-amphetamine-induced damage and increased BDNF expression; in addition, cannabidiol (30 or 60 mg/kg) prevented the D-amphetamine-induced formation of carbonyl group in prefrontal cortex (24), suggesting these effects vary depending on the brain regions evaluated and doses of cannabidiol administered.

It is well known that in the hippocampus, as well as prefrontal cortex, increased BDNF results in antidepressant responses (69), the fact that no

differences in these areas reported in this study may be related to differences in the neural circuitry. In fact, stress seems to exert opposite effects in amygdala and hippocampus, for example, stress increases spine synapse formation in amygdala (70), but decreases in hippocampus (71). In addition, in the hippocampus and prefrontal cortex BDNF inhibits depressive symptoms, whereas in the amygdala it facilitates depressive-like symptoms (69). However, acute social stress or repeated restraint exposure reduced amygdala BDNF (72,73), suggesting positive effects of both imipramine and cannabidiol in this study.

In conclusion, given the central role of the amygdala in the modulation of emotional responses (74), the effects of cannabidiol on antidepressant-like behaviour may attribute to changes in amygdalar neuroplasticity or could be because of the combinations of cannabidiol with other effects, e.g. inflammatory effects (18–20), antioxidative action (21,24) and neuroprotective effect (24,25), which are involved in major depression (75,76).

Acknowledgements

This study was supported in part by grants from CNPq-Brazil (J. Q., F. K., J. A. C., A. W. Z. and J. E. H.), FAPESP-Brazil (J. A. C., A. W. Z. and J. E. H.), FAPESC-Brazil (J. Q.), Instituto Cérebro e Mente-Brazil (J. Q. and F. K.) and UNESC-Brazil (J. Q.). J. Q., F. K., J. A. C. and A. W. Z. are recipients of CNPq (Brazil) Productivity Fellowships. G. Z. R. is holder of a CAPES studentship. This study was also sponsored by THC-Pharm (Frankfurt, Germany) and STI-Pharm (UK) who kindly provided cannabidiol.

References

1. BAUNE BT, ADRIAN I, JACOBI F. Medical disorders affect health outcome and general functioning depending on comorbid major depression in the general population. *J Psychosom Res* 2007;**62**:109–118.
2. MATHERS C, LONCAR D. Projections of global mortality and burden of disease from 2002 to 2030. *PLoS Med* 2006;**3**:442.
3. CASTRÉN E. Is mood chemistry? *Nat Rev Neurosci* 2005;**6**:241–246.
4. DUMAN RS, HENINGER GR, NESTLER EJ. A molecular and cellular theory of depression. *Arch Gen Psychiatry* 1997;**54**:597–606.
5. MOREIRA FA, GUIMARÃES FS. Cannabidiol inhibits the hyperlocomotion induced by psychotomimetic drugs in mice. *Eur J Pharmacol* 2005;**512**:199–205.
6. SOUERY D, AMSTERDAM J, DE MONTIGNY C et al. Treatment resistant depression: methodological overview and operational criteria. *Eur Neuropsychopharmacol* 1999;**9**:83–91.
7. TOMBA E, GRANDI S, FAVA GA. Therapeutic factors in depression: new strategies. *Riv Psichiatr* 2009;**44**:95–101.
8. GAONI Y, MECHOULAM RJ. The isolation and structure of delta-1-tetrahydrocannabinol and other neutral cannabinoids from hashish. *Am Chem Soc* 1971;**93**:217–224.
9. ZUARDI AW, SHIRAKAWA I, FINKELFARB E, KARNIOL IG. Action of cannabidiol on the anxiety and other effects produced by Δ^9 -THC in normal subjects. *Psychopharmacology* 1982;**76**:245–250.
10. ZUARDI AW, CRIPPA JAS, HALLAK JEC, MOREIRA FA, GUIMARÃES FS. Cannabidiol, a *Cannabis sativa* constituent, as an antipsychotic drug. *Braz J Med Biol Res* 2006;**39**:421–429.
11. PERTWEE RG. Pharmacology of cannabinoids CB₁ and CB₂ receptors. *Pharmacol Ther* 1997;**74**:129–180.
12. HERKENHAM M, LYNN AB, JOHNSON MR, MELVIN LS, DE COSTA BR, RICE KC. Characterization and localization of cannabinoid receptors in rat brain: a quantitative in vitro autoradiographic study. *J Neurosci* 1991;**11**:563–583.
13. HONÓRIO KM, ARROIO A, SILVA. ABF – therapeutical aspects of compounds of the plant *Cannabis sativa*. *Quím Nova* 2006;**29**:318–325.
14. TSOU K, BROWN S, SANUDO-PENA MC, MACKIE K, WALKER JM. Immunohistochemical distribution of cannabinoid CB1 receptors in the rat central nervous system. *Neuroscience* 1998;**83**:393–411.
15. ZUARDI AW. Cannabidiol: from an inactive cannabinoid to a drug with wide spectrum of action. *Rev Bras Psiquiatr* 2008;**30**:271–280.
16. COSTA B, COLLEONI M, CONTI S et al. Oral anti-inflammatory activity of cannabidiol, a non-psychoactive constituent of cannabis, in acute carrageenan-induced inflammation in the rat paw. *Naunyn Schmiedeberg Arch Pharmacol* 2004;**369**:294–299.
17. SUMARIWALLA PF, GALLILY R, TCHILIBON S, FRIDE E, MECHOULAM R, FELDMANN M. A novel synthetic, nonpsychoactive cannabinoid acid (HU-320) with anti-inflammatory properties in murine collagen-induced arthritis. *Arthritis Rheum* 2004;**50**:985–998.
18. WALTER L, FRANKLIN A, WITTING A et al. Nonpsychoactive cannabinoid receptors regulate microglial cell migration. *J Neurosci* 2003;**23**:1398–1405.
19. HAMPSON AJ, GRIMALDI M, AXELROD J, WINK D. Cannabidiol and delta-9 tetrahydrocannabinol are neuroprotective antioxidants. *Proc Natl Acad Sci U S A* 1998;**95**:8268–8273.
20. JACOBSSON SO, RONGÅRD E, STRIDH M, TIGER G, FOWLER SJ. Serum dependent effects of tamoxifen and cannabinoids upon C6 glioma cell viability. *Biochem Pharmacol* 2000;**60**:1807–1813.
21. MECHOULAM RE, PARKER LA, GALLILY R. Cannabidiol: an overview of some pharmacological aspects. *J Clin Pharmacol* 2002;**42**:11–19.
22. VALVASSORI SS, ELIAS G, DE SOUZA B et al. Effects of cannabidiol on amphetamine-induced oxidative stress generation in an animal model of mania. *J Psychopharmacol* 2011;**25**:274–280.
23. DIRIKOC S, PRIOLA SA, MARELLA M, ZSURGER N, CHABRY J. Nonpsychoactive cannabidiol prevents prion accumulation and protects neurons against prion toxicity. *J Neurosci* 2007;**27**:9537–9544.
24. EL-REMESSY AB, KHALIL IE, MATRAGOON S et al. Neuroprotective effect of delta-9-tetrahydrocannabinol and cannabidiol in N-Methyl-D-Aspartate-induced retinal neurotoxicity. *Am J Pathol* 2003;**163**:1997–2008.
25. RYAN D, DRYSDALE AJ, LAFOURCADE C, PERTWEE RG, PLATT B. Cannabidiol targets mitochondria to regulate intracellular Ca²⁺ levels. *J Neurosci* 2009;**29**:2053–2063.

26. WEISS L, ZEIRA M, REICH S et al. Cannabidiol arrests onset of autoimmune diabetes in NOD mice. *Neuropharmacology* 2008;**54**:244–249.
27. DURST R, DANENBERG H, GALLILY R et al. Cannabidiol, a nonpsychoactive Cannabis constituent, protects against myocardial ischemic reperfusion injury. *Am J Physiol Heart Circ Physiol* 2007;**293**:3602–3607.
28. CARLINI EA, LEITE JR, TANHAUSER M, BERARDI AC. Cannabidiol and *Cannabis sativa* extract protect mice and rats against convulsive agents. *J Pharm Pharmacol* 1973;**25**:664–665.
29. IZQUIERDO I, ORSINGER OA, BERARDI AC. Effect of cannabidiol and other *Cannabis sativa* compounds on hippocampal seizures discharges. *Psychopharmacology* 1973;**28**:95–102.
30. TURKANIS SA, CELY W, OLSEN DM, KARLER R. Anticonvulsant properties of cannabidiol. *Res Commun Chem Pathol Pharmacol* 1974;**8**:231–246.
31. KARNIOL IG, CARLINI EA. Pharmacological interaction between cannabidiol and delta 9-tetrahydrocannabinol. *Psychopharmacology* 1973;**3**:53–70.
32. LONG LE, MALONE DT, TAYLOR DA. Cannabidiol reverses MK-801-induced disruption of prepulse inhibition in mice. *Neuropsychopharmacology* 2006;**31**:795–803.
33. MOREIRA FA, AGUIAR DC, GUIMARÃES FC. Anxiolyticlike effect of cannabidiol in the rat Vogel conflict test. *Prog Neuropsychopharmacol Biol Psychiatry* 2006;**30**:1466–1471.
34. ZUARDI AW, MORAIS SL, GUIMARÃES FS, MECHOUAM R. Antipsychotic effect of cannabidiol. *J Clin Psychiatry* 1995;**56**:485–486.
35. CRIPPA JA, ZUARDI AW, GARRIDO GE et al. Effects of cannabidiol (CBD) on regional cerebral blood flow. *Neuropsychopharmacol* 2004;**29**:417–426.
36. FUSAR-POLI P, CRIPPA JA, BHATTACHARYYA S et al. Distinct effects of delta-9-Tetrahydrocannabinol and cannabidiol on neural activation during emotional processing. *Arch Gen Psychiatry* 2009;**66**:95–105.
37. GUIMARÃES FS, CHIARETTI TM, GRAEFF FG, ZUARDI AW. Antianxiety effect of cannabidiol in the elevated plus-maze. *Psychopharmacology* 1990;**100**:558–559.
38. GUIMARÃES FS, DE AGUIAR JC, MECHOUAM R, BREUER A. Anxiolytic effect of cannabidiol derivatives in the elevated plus-maze. *Gen Pharmacol* 1994;**25**:161–164.
39. ZUARDI AW, COSME RA, GRAEFF FG, GUIMARÃES FS. Effects of ipsapirone and cannabidiol on human experimental anxiety. *J Psychopharmacol* 1993;**7**:82–88.
40. EL-ALFY AT, IVEY K, ROBINSON K et al. Antidepressant-like effect of Delta9-tetrahydrocannabinol and other cannabinoids isolated from *Cannabis sativa* L. *Pharmacol Biochem Behav* 2010;**95**:434–442.
41. HILL MN, GORZALKA BB. Enhancement of the anxiety-like response to the cannabinoid receptor agonist HU-210 following chronic stress. *Eur J Pharmacol* 2004;**24**:291–295.
42. HILL MN, PATEL S, CARRIER EJ et al. Downregulation of endocannabinoid signaling in the hippocampus following chronic unpredictable stress. *Neuropsychopharmacology* 2005;**30**:508–515.
43. ZANELATI TV, BIOJONE C, MOREIRA FA, GUIMARÃES FS, JOCA SR. Antidepressant-like effects of cannabidiol in mice: possible involvement of 5-HT_{1A} receptors. *Br J Pharmacol* 2010;**159**:122–128.
44. RUSSO EB, BURBETT A, HALL B, PARKER KK. Agonistic properties of cannabidiol at 5-HT_{1a} receptors. *Neurochem Res* 2005;**30**:1037–1043.
45. ANDERSON IM. Selective serotonin reuptake inhibitors versus tricyclic antidepressants: a attenuates the effects of antidepressants on the forced swim test in rats. *Brain Res* 1996;**709**:215–220.
46. JOCA SR, PADOVAN CM, GUIMARÃES FS. Stress, depression and the hippocampus. *Rev Bras Psiquiatr* 2003;**2**:46–51.
47. KAREGE F, VAUDAN G, SCHWALD M, PERROUD N, LA HARPE R. Neurotrophin levels in postmortem brains of suicide victims and the effects of antemortem diagnosis and psychotropic drugs. *Brain Res Mol Brain Res* 2005;**136**:29–37.
48. SHIRAYAMA Y, CHEN AC, NAKAGAWA S, RUSSELL DS, DUMAN RS. Brain-derived neurotrophic factor produces antidepressant effects in behavioral models of depression. *J Neurosci* 2002;**22**:3251–3261.
49. NIBUYA M, MORINOBU S, DUMAN RS. Regulation of BDNF and trkB mRNA in rat brain by chronic electroconvulsive seizure and antidepressant drug treatments. *J Neurosci* 1995;**15**:7539–7547.
50. SIUCIAK JA, LEWIS DR, WIEGAND SJ, LINDSAY RM. Antidepressant-like effect of brain-derived neurotrophic factor (BDNF). *Pharmacol Biochem Behav* 1997;**56**:131–137.
51. MCARTHUR R, BORSINI F. Animal models of depression in drug discovery: a historical perspective. *Pharmacol Biochem Behav* 2006;**84**:436–452.
52. DETKE MJ, RICKELS M, LUCKI I. Active behaviors in the rat forced swimming test differentially produced by serotonergic and noradrenergic antidepressants. *Psychopharmacology* 1995;**121**:66–72.
53. GARCIA LB, COMIM CM, VALVASSORI SS et al. Acute administration of ketamine induces antidepressant-like effects in the forced swimming test and increases BDNF levels in the rat hippocampus. *Prog Neuropsychopharmacol Biol Psychiatry* 2008;**32**:140–144.
54. GARCIA LB, COMIM CM, VALVASSORI SS et al. Chronic administration of ketamine elicits antidepressant-like effects in rats without affecting hippocampal brain-derived neurotrophic factor protein levels. *Basic Clin Pharmacol Toxicol* 2008;**103**:502–506.
55. PORSOLT RD, LE PICHON M, JALFRE M. Animal model of depression. *Nature* 1977;**266**:730–732.
56. LOWRY OH, ROSEBOUGH NG, FARR AL, RANDALL RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem* 1951;**193**:265–275.
57. PATEL S, CRAVATT BF, HILLARD CJ. Synergistic interactions between cannabinoids and environmental stress in the activation of the central amygdalae. *Neuropsychopharmacology* 2005;**30**:497–507.
58. MARTIN M, LEDENT C, PARMENTIER M, MALDONADO R, VALVERDE O. Involvement of CB₁ cannabinoid receptors in emotional behavior. *Psychopharmacology* 2002;**159**:379–387.
59. CARLSON G, WANG Y, ALGER BE. Endocannabinoids facilitate the induction of LTP in the hippocampus. *Nat Neurosci* 2002;**5**:723–724.
60. FORTUNATO JJ, RÉUS GZ, KIRSCH TR et al. Acute harmine administration induces antidepressant-like effects and increases BDNF levels in the rat hippocampus. *Prog Neuropsychopharmacol Biol Psychiatry* 2009;**33**:1425–1430.
61. RÉUS GZ, STRINGARI RB, KIRSCH TR et al. Neurochemical and behavioural effects of acute and chronic memantine administration in rats: further support for NMDA as a new pharmacological target for the treatment of depression? *Brain Res Bull* 2010;**81**:585–589.

62. KRISHNAN V, NESTLER EJ. The molecular neurobiology of depression. *Nature* 2008;**455**:894–902.
63. AAN HET ROT M, MATHEW SJ, CHARNEY DS. Neurobiological mechanisms in major depressive disorder. *CMAJ* 2009;**180**:305–313.
64. MARTINOWICH K, LU B. Interaction between BDNF and serotonin: role in mood disorders. *Neuropsychopharmacology* 2008;**33**:73–83.
65. NESTLER EJ, BARROT M, DiLEONE RJ, EISCH AJ, GOLD SJ, MONTEGGIA LM. Neurobiology of depression. *Neuron* 2002;**34**:13–25.
66. DUMAN RS, MONTEGGIA LM. A neurotrophic model for stress-related mood disorders. *Biol Psychiatry* 2006;**59**:1116–1127.
67. LARSEN MH, HAY-SCHMIDT A, RONN LCB, MIKKELSEN JD. Temporal expression of brain-derived neurotrophic factor (BDNF) mRNA in the rat hippocampus after treatment with selective and mixed monoaminergic antidepressants. *Eur J Psychopharmacol* 2008;**578**:114–122.
68. KOVARU H, PAV M, KOVARU F, RABOCH J, FISEROVA A. Cell signalling in CNS and immune system in depression and during antidepressant treatment: focus on glial and natural killer cells. *Neuro Endocrinol Lett* 2009;**30**:421–428.
69. YU H, CHEN Z. The role of BDNF in depression on the basis of its location in the neural circuitry. *Acta Pharmacol Sin* 2011;**32**:3–11.
70. VYAS A, MITRA R, SHANKARANARAYANA RAO BS, CHATTARJI S. Chronic stress induces contrasting patterns of dendritic remodeling in hippocampal and amygdaloid neurons. *J Neurosci* 2002;**22**:6810–6818.
71. PAWLAK R, RAO BS, MELCHOR JP, CHATTARJI S, MCEWEN B, STRICKLAND S. Tissue plasminogen activator and plasminogen mediate stress-induced decline of neuronal and cognitive functions in the mouse hippocampus. *Proc Natl Acad Sci U S A* 2005;**102**:18201–18206.
72. PIZARRO JM, LUMLEY LA, MEDINA W et al. Acute social defeat reduces neurotrophin expression in brain cortical and subcortical areas in mice. *Brain Res* 2004;**1025**:10–20.
73. FANOUS S, HAMMER RP, Jr, Nikulina EM. Short- and long-term effects of intermittent social defeat stress on brain-derived neurotrophic factor expression in mesocorticolimbic brain regions. *Neuroscience* 2010;**167**:598–607.
74. MURRAY EA. The amygdala, reward and emotion. *Trends Cogn Sci* 2007;**11**:489–497.
75. LUCCA G, COMIM CM, VALVASSORI SS et al. Effects of chronic mild stress on the oxidative parameters in the rat brain. *Neurochem Int* 2009a;**54**:358–362.
76. LUCCA G, COMIM CM, VALVASSORI SS et al. Increased oxidative stress in submitochondrial particles into the brain of rats submitted to the chronic mild stress paradigm. *J Psychiatr Res* 2009b;**43**:864–869.