Effect of bovine age on the proliferative activity, global DNA methylation, relative telomere length and telomerase activity of granulosa cells

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Summary

Granulosa cells influence the growth and acquisition of the developmental competence of oocytes. We investigated the effects of ageing on the proliferative activity, global genomic DNA methylation, relative telomere length and telomerase activity of bovine granulosa cells. The proliferative activity of cells was examined by bromodeoxyuridine (BrdU) assay, genomic DNA methylation was examined by enzyme-linked immunosorbent assay (ELISA), and relative telomere length and telomerase activity were examined by real-time polymerase chain reaction. We first compared the proliferative activity of the granulosa cells of the medium follicles between in dominant phase ovaries and growth phase ovaries. We observed that the proliferative activity of the granulosa cells of dominant phase ovaries was significantly lower than those of growth phase ovaries. In addition, the proliferative activity of granulosa cells was inversely associated with follicular size. Based on the results, we used granulosa cells harvested from the medium follicles (3–5 mm in diameter) on the surfaces of the dominant phase ovaries collected from cows at a slaughterhouse. The proliferative activity of the granulosa cells harvested from the ovaries of old cows (N = 8; average age 165.1 months) was lower than that of the cells from young cows (N = 8; average age 30.9 months). Global loss of cytosine methylation was detected in the granulosa cells of old cows (N = 12; average age 141.0 months) compared with young cows (N = 15; average age 27.4 months). Although the relative telomere lengths of cumulus cells were similar in the two age groups, the relative telomere lengths and telomerase activity of the granulosa cells from old cows (N = 17 and 9; average age, 164.6 and 151.3 months, respectively) tended to be shorter than those of the cells from young cows (N = 17 and 10; average age 30.6 and 28.1 months, respectively); however, this difference was not significant p = 0.09 and 0.053, respectively). In conclusion, the proliferative activity and genomic global DNA methylation significantly decreased, and the relative telomere lengths and telomerase activity of granulosa cells tended to be shorter with the age of donor cows.

Keywords: Ageing, DNA methylation, Granulosa cells, Proliferative activity, Telomere length

Introduction

During the female mammalian reproductive life, follicles grow from primordial follicles surrounded by a small number of flat granulosa cells to large antral follicles containing a net of 21 doublings of granulosa cell numbers (Wezel & Rodgers, 1996; Rodgers & Irving-Rodgers, 2010). In addition to their proliferation, granulosa cells differentiate into cumulus cells and mural granulosa cells. During each round of the somatic cell cycle, telomeric DNA replication results in telomere shortening due to the end replication problem. Once the telomeric length shortens to a critical level, the cell exits the cell cycle and undergoes cell senescence (Olovnikov, 1996). However, granulosa cells reportedly maintain their telomerase activity during follicle development (Lavranos *et al.*, 1999; Russo *et al.*, 2006; Liu & Li,

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2010), which may enable their numerous replications. In addition, the telomerase activity of granulosa cells differed according to follicle size and their localization in the follicles such that granulosa cells either from small follicles or in the middle layers had higher telomerase activity than those from large follicles or in the basal layers (Lavranos *et al.*, 1999).

Age-associated decline in fertility is a concern in modern society, and vast literature regarding the relationship between maternal ageing and oocyte quality in humans exists (Ottolenghi et al., 2004; Baird et al., 2005; Broekmans et al., 2007). However ethical constraints have limited the sample numbers and hindered thorough investigations. A bovine model of human reproduction has been introduced by Malhi et al. to study the age-associated problems of human fertility, and there is increasing evidence demonstrating age-associated decline in bovine fertility (Malhi et al., 2005, 2006, 2008; Yamamoto et al., 2010; Iwata et al., 2011). In our previous study, oocytes were harvested from old cows (age, >120 months) and young cows (age, 25-35 months); the progression of nucleic meiotic maturation of oocytes from old cows was faster than that of oocytes from young cows. Furthermore, when oocytes were denuded of surrounding cells, the differences between the two age groups disappeared (Yamamoto et al., 2010). On the basis of these observations, we speculate that ageing also affects the characteristics of the granulosa cells in cows.

It is widely accepted that the quality of cumulus and granulosa cells is closely associated with oocyte development through their mutual interactions (Orisaka *et al.*, 2009; Downs, 2010; Pandey *et al.*, 2010). In humans, it has been demonstrated that ageing influences the quality of granulosa cells (Tatone *et al.*, 2006; Ito *et al.*, 2010). However, few reports regarding the effects of ageing on bovine granulosa cells exists.

Proliferative activity is a primary characteristic reflecting the quality of cells and the cell character depends on the genomic epigenetic information which closely associated with cytosine methylation; thus, the aim of the present study was to examine the effect of ageing on the proliferative activity and global DNA methylation of granulosa cells, and relative telomere lengths and telomerase activity of the granulosa cells harvested from the ovaries of old and young cows. Although the characteristics of granulosa cells differ considerately among follicular developmental phases and individual cows, to the best of our knowledge, this is the first report regarding age-associated changes in the characteristics of bovine granulosa cells. We observed that the proliferative activity and global cytosine methylation of the granulosa cells was profoundly affected by donor age. However, no significant difference was

observed between the relative telomere lengths of the cumulus cells derived from old and young cows. In addition, the relative telomere length and telomerase activity of the granulosa cells derived from old cows were shorter than those from young cows, though the difference was not significant.

Materials and methods

Unless otherwise stated, all chemicals were purchased from Nacalai Tesque (Kyoto, Japan). Granulosa cells were cultured in a TCM-199 medium (Gibco, Paisley, UK) supplemented with 0.5 mM pyruvic acid and 5% fetal calf serum (FCS).

Ovary and oocyte collection

Ovaries with a functional luteum were harvested from cows in a local slaughterhouse and stored at 25 °C in phosphate-buffered saline (PBS) containing 10 mM glucose, 10 mM sucrose and antibiotics; these were transported to our laboratory within 4 h. Granulosa cells and cumulus–oocyte complexes (COCs) were harvested from the follicles on the surfaces of the ovaries using a syringe with an 18-gauge needle.

Preparation of granulosa and cumulus cells

The telomerase activity of granulosa cells differs in accordance with the localization in the follicle. Thus, we collected granulosa cells from two distinct positions: granulosa cells distal to the oocyte, which were aspirated from the follicle and granulosa cells proximal to the oocyte, which are defined as cumulus cells in the present study. The contents aspirated from the follicles were suspended in a wash solution (PBS with 1 mg/ml BSA (bovine serum albumin)) and washed by centrifugation. Next, the granulosa cells were carefully collected from the pellet and re-suspended in a wash solution containing 0.2% hyaluronidase. The cells were separated by vortexing for 3 min and then washed by centrifugation. The resulting cell pellet was subjected to DNA extraction or used for in vitro culture. For culture, the cells were suspended in a culture medium at a final concentration of 5×10^5 viable cells/ml. The proportion of viable cells was assessed by trypan blue staining. This cell suspension was used for proliferation assays.

For cumulus cell preparation, the COCs harvested from each cow were vortexed in the wash solution containing 0.25% hyaluronidase. After removing the oocytes, the cell suspension was centrifuged, and the resulting cell pellet was subjected to DNA extraction immediately after collection or after passage culture. For passage culture, the cumulus cells were cultured in a 4-well plate (Nunc) for 5–7 days and sub-cultured four times.

Proliferation assay

The suspension of the granulosa cells derived from each cow was dispensed into three wells of a 96-well plate (100 μ l/well; Falcon) and incubated for 24 h at 38.5 °C with maximum humidity in an atmosphere of 5% CO₂ in air. The cell proliferative activity was assessed by the cell proliferation ELISA BrdU assay kit (Roche), according to the manufacturer's protocol. The average of triplicate determinations was used for each cow.

DNA extraction and telomere length assessment

Granulosa and cumulus cells were prepared as described above. These cells were diluted in 100 µl of the extraction buffer (20 mM of Tris, 0.4 mg/ml of pronase K, 0.9% of Nonidet-40 and 0.9% of Tween 20), and DNA was extracted at 55 °C for 30 min followed by 95 °C for 5 min. Real-time quantification of relative telomere length was performed based on the protocol described by Mastromonaco et al. (2006). Briefly, reactions were performed in triplicate in 20-µl reaction volumes (6 µl of sample per reaction) for a standard DNA sample and all experimental samples. The PCR (polymerase chain reaction) mixture (20 µl) contained 6 µl of extracted DNA, 0.5 µM of each primer set, and 10 µl of MESA Blue (qPCR TM-Mastermix Plus for SYBR Assay; Eurogentec, Belgium). This assay included amplification of the telomere and a single-copy gene (BTF3) for normalization. The teh telomere and single-copy gene amplifications were performed in series. The telomere primer sequences were as follows: Tel 1b: 5'-CGGTTTGTTTGGGTTTGGGTTTGGGTTTGGGTTT-GGGTT-3' and Tel 2b: 5'-GGCTTGCCTTACCCTTAC-CCTTACCCTTACCCT-3'. The primer sequences for the single-copy gene were as follows: BTF3u: 5'-AGGAACTGCTCGCAGAAAGA-3' and BTF3d: 5'-GCCCGTAATGGTGAAAGTGT-3'.

The telomere length assay consisted of a 10-min incubation at 95 °C for enzyme activation followed by 20 cycles at 95 °C for 5 s, 56 °C for 10 s, and 72 °C for 60 s and fluorescent signal acquisition. The single-copy gene assay consisted of a 10 min incubation at 95 °C followed by 35 cycles at 95 °C for 5 s, 60 °C for 5 s and 72 °C for 20 s and fluorescence signal acquisition. Real-time PCR was performed using the Rotor-Gene 6500 real-time rotary analyser (Corbett Research). The samples that contained the highest amounts of DNA were selected as external standards and three serial dilutions were produced using this DNA. DNA extracted from the cumulus cells of a cow aged 27 months and DNA from the granulosa cells of a

cow aged 147 months were used as external standards. The concentrations of the telomere and single-copy gene were defined as 1. In this experiment, relative telomere concentration (T) and relative concentration of a single-copy gene (S) were calculated using an external standard that was amplified in all assays and was used for calculating the sample T/S. The sample T/S was compared with the standard sample to obtain the relative T/S ratio [(sample T/S)/(standard T/S)]. The relative T/S ratio of each sample indicates its telomere length relative to the standard sample.

Quantitative telomerase assay

Granulosa cells were aspirated from the medium follicles on the surface of dominant phase ovaries. Telomerase detection was conducted according to the manufacture's protocol (Cat No. MT3010, Allied Biotech Inc.). In brief, granulosa cells were washed and suspended in lysis buffer followed by centrifugation. The cell extract was subjected to real-time PCR to determine the telomerase activity through its ability to synthesize telomeric repeats onto an oligonucleotide substrate *in vitro*.

Global DNA methylation sift assay

The global methylation shift assay was conducted according to the manufacture's protocol (Imprint Methylated DNA Quantification Kit; Sigma-Aldrich). Granulosa cells were aspirated from the medium follicles on the surface of dominant phase ovaries. Purified DNA was prepared from 4×10^6 granulosa cells of each cow using NucleoSpin Tissue XS (Takara), and the concentration of the DNA was measured. One hundred nanograms each of the extracted DNA and control methylated DNA were subjected to the measurement and the relative methylation of the each sample compared with the methylated DNA control was calculated.

Experimental protocols

Experiment 1

Follicle development occurs in a wave-like pattern during the oestrous cycle, and the ovaries were categorized into two distinct follicular development phases; the dominant phase and growth phase (Hagemann, 1999). We then selected ovaries in the two distinct follicular development phases and examined the effect of the follicular development phase on the proliferation of granulosa cells. To establish the two different follicular development phases, an ovary with a functional luteum and at least one large follicle (>9 mm in diameter) was defined as a dominant phase ovary, and an ovary with a functional luteum but without large follicles (>7 mm in diameter) was defined as a growth phase ovary. Dominant and growth phase ovaries were harvested from 14 cows aged 27–29 months (7 cows for each phase). Granulosa cells were collected from teh medium follicles (3– 5 mm in diameter) on the surfaces of the ovaries. Cell proliferative activity was examined as described above.

Experiment 2

Here we examined the effect of follicular size on the proliferative activity of granulosa cells. Dominant phase ovaries with functional luteum were harvested from eight cows aged 26–29 months and granulosa cells were harvested from the small (1–3 mm in diameter), medium (3–5 mm in diameter) and large follicles (>5 mm in diameter) on the surfaces of individual ovaries. The proliferative activity of the granulosa cells was examined.

Experiment 3

Here we examined the effect of donor age on the proliferative activity of granulosa cells. Dominant phase ovaries with a functional luteum were harvested from eight young cows aged 26–35 months (average age, 30.9 months) and old cows aged 144–185 (average age, 165.1 months). The proliferative activity of the granulosa cells harvested from medium follicles was examined.

Experiment 4

We examined the effect of donor age on the global cytosine methylation of granulosa cells. Granulosa cells were harvested from the medium follicles on the surface of dominant phase ovaries with a functional luteum of 15 young cows aged 20–32 (average 27.4) months and 12 old cows aged 120–170 (average 141.0) months. The methylation of genomic DNA was assessed as described above.

Experiment 5

Here we examined the validity of the relative telomere length assay. We harvested dominant phase ovaries with a functional luteum from five young cows aged 26–27 months. Cumulus cells were collected from the COCs derived from the medium follicles of each ovary. DNA was extracted from the cumulus cells immediately after collection and after four passages in culture (1 month). Relative telomere lengths of the primary and continuously cultured cells were compared.

Experiment 6

The relative telomere length of the cumulus and granulosa cells collected from the medium follicles on the surface of dominant phase ovaries with functional luteum were compared by the age of donors. Ovaries of 18 young cows aged 25–30 months (average age 27.7 months) and those of 15 older cows aged 137–198 months (average age 160.9 months) were used to compare the relative telomere lengths of cumulus cells. Ovaries of 16 young cows aged 29–47 months (average age 30.6 months) and 17 older cows aged 129–223 months (average age, 164.6 months) were used to compare relative telomere lengths of granulosa cells. We compared the relative telomere lengths of the cumulus and granulosa cells between the two age groups.

Experiment 7

Here we compared the relative telomerase activity in the granulosa cells of nine old cows aged 120– 190 (average age, 151.3) months and 10 young cows aged 26–38 (average age, 28.1) months. The telomerase activity of the cells was measured as described above.

Statistical analysis

The proliferative activity among different follicle sizes was compared by analysis of variance (ANOVA) followed by Tukey's test. The proliferative activity, telomere length, global cytosine methylation, and telomerase activity of the two groups were compared using Student's *t*-test. A *p*-value of less than 0.05 was considered significant.

Results

Figure 1 shows the proliferative activities of the granulosa cells harvested from the two distinct follicular development phases. The proliferative activity of the granulosa cells of the medium follicles from dominant phase ovaries was defined as follows: the proliferative activity was 2.4-fold and significantly higher in the granulosa cells from the medium follicles of the growth phase ovaries than those of the dominant phase ovaries.

Figure 2 shows that the proliferative activity of granulosa cells was inversely associated with follicular size. The activity shown by the rate to the cells from large follicles was 3.2 and 4.6 from medium and small follicles, respectively. These values were significantly different among the three follicle sizes (p < 0.05).

In Experiment 3, granulosa cells were harvested from the medium follicles on the surfaces of dominant phase ovaries from old and young cows. A comparison of the proliferative activity of the granulosa cells between old and young cows is shown in Fig. 3. The proliferative activity of cells from young cows was shown by the rate to that from old cows. The activity from young cows was 1.5-fold higher than that of cells from old cows (p < 0.05).

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Figure 1 Proliferative activity of granulosa cells harvested from growth and dominant phase ovaries. Dominant: dominant phase ovaries with a functional luteum and a large follicle (>9 mm). Growth: growth phase ovaries with a functional luteum but not large follicles (<7 mm). The proliferative activity of granulosa cells from dominant phase ovaries was defined as 1. a, b: Values between columns with different letters were significantly different (p < 0.05).



Figure 2 Proliferative activity of granulosa cells harvested from small (1–3 mm in diameter), medium (3–5 mm) and large follicles (>9 mm) on the surface of the dominant phase ovaries. The proliferative activity of granulosa cells derived from large follicles was defined as 1. (a–c) Values between columns with different letters were significant different (p < 0.05).

In Experiment 4, the percentage of the overall genomic methylation of the granulosa cells derived from old cows was 31.2% which was significantly lower (p = 0.04) than that of the cells derived from young cows (38.9%).

In Experiment 5, we examined the validity of the relative telomerase length assay by checking



Figure 3 Proliferative activity of granulosa cells harvested from medium follicles on the surface of dominant phase ovaries from young and old cows. The proliferative activity of granulosa cells from old cows was defined as 1. (a, b) Values between columns with different letters were significantly different (p < 0.05).

for telomere length following passage in culture. When cumulus cells were sub-cultured four times, a decrease in telomere lengths was observed (Fig. 4; p < 0.05).

In Experiment 6, the relative telomere length in the granulosa and cumulus cells of young and old cows was compared. A comparison of the relative telomere length of the granulosa and cumulus cells is shown in Figs. 5 and 6, respectively. The relative telomere length of the granulosa cells derived from an old cow (142 months) was defined as follows: although the mean value of the relative telomere length was 5.4 and 2.3 in the granulosa cells derived from young (average age, 30.6 months) and old cows (average age, 164.6 months) respectively, these values were not significant different (p = 0.09).

In Fig. 6, the relative telomere length of the cumulus cells harvested from a young cow (27 months) was defined as follows: the relative telomere length of the cumulus cells derived from old cows was similar to that derived from young cows and there was no significant difference between the two groups (p = 0.28).

In Experiment 7, the telomerase activity of the granulosa cells collected from the medium size follicles of young cows was defined as 1 (means \pm S.E,



Figure 4 Telomere lengths of cumulus cells during culture. Cumulus cells were harvested from medium follicles on the surface of dominant phase ovaries of young cows. Telomere length of cumulus cells just after harvest (primary) was compared with those after fourth passage. (a, b) Values between columns with different letters were significantly different (p < 0.05).



Figure 5 Relative telomere lengths in granulosa cells from young and old cows. Granulosa cells were harvested from medium follicles on the surface of dominant phase ovaries. Difference between young and old cows was not significant (p = 0.09).

 1.0 ± 0.28), and the relative telomerase activity of the granulosa cells from old cows was 0.32 ± 0.15 . The difference was not significant (p = 0.053).



Figure 6 Relative telomere lengths in cumulus cells from young and old cows. Cumulus cells were harvested from cumulus oocytes complexes collected from medium follicles on the surface of dominant phase ovaries. Difference between young and old cows was not significant (p = 0.28).

Discussion

This study demonstrated that the proliferative activity and global genomic methylation of granulosa cells was profoundly affected by the age of donor cow. Although the relative telomere lengths were similar in the cumulus cells of young and old cows, the relative telomere length and telomerase activity of the granulosa cells from old cows tended to be shorter and less than those from young cows (p = 0.09 and 0.053, respectively).

In cows, once small antral follicles are recruited for development, one or two of the largest follicles are selected to become dominant follicles, while the remaining small follicles regress. (Aerts & Bols, 2010). Thus, antral follicle development is divided into the following two phases: growth phase ovaries and dominant phase ovaries (Fortune *et al.*, 2004). Oocytes derived from growth phase ovaries have higher developmental competence than those from dominant phase ovaries, and the frequency of apoptosis among the follicular cells derived from dominant phase is also higher than that from growth phase ovaries (Hagemann *et al.*, 1999).

In Experiment 1, the proliferative activity was higher for the granulosa cells originating from the medium follicles on the surface of growth phase ovaries as compared with those of dominant phase ovaries; this result is consistent with previous reports. In addition, the proliferative activity of granulosa cells was inversely associated with follicular size, with the highest proliferative activity observed for the granulosa cells derived from small follicles. Lavranos *et al.* (1999) reported that telomerase activity is high in small antral follicles and is reduced in large follicles in cows. Furthermore, the amount of estradiol secretion, the number of receptors for this hormone and the frequency of apoptosis varies among follicles of different sizes (Gutiérrez *et al.*, 1997; Lavranos *et al.*, 1999; Schams & Berisha, 2002). On the basis of these findings, the difference in proliferative activity depending on follicular size may be reflected by different degrees of differentiation, apoptosis and/or telomere lengths of the cells in the present study.

In general, the characteristics of granulosa and cumulus cells differ considerably among follicles. It is important to collect uniform samples for studying the properties of granulosa cells. Hence we attempt to minimize the effect of follicle characteristics such as the follicular development phase of ovaries and size by using the medium follicles on the surface of dominant phase ovaries. It is worth noting that the characteristics of the granulosa cells used in this study did not precisely reflect the properties of granulosa cells *in vivo*, as a follicle destined for ovulation is the largest follicle or dominant follicle on the surface of the ovary at the end of a follicular wave.

Both granulosa and cumulus cells support oocyte development through energy metabolism, amino acid uptake, hormone secretion and by providing suitable microenvironments. In contrast, the oocytes themselves regulate the granulosa and cumulus cell functions and differentiation through growth/differentiation factor 9, bone morphogenetic protein 15 and others (Hunter, 2000; Hamel *et al.*, 2008; Su *et al.*, 2009; Huang & Wells, 2010). Moreover, the quality of oocytes is closely related to the apoptosis rate among cumulus cells as well as cumulus and granulosa cell gene expressions (Hamel *et al.*, 2008; Huang & Wells, 2010).

Many reports have demonstrated the effects of ageing on the quality of oocytes and age-associated changes in the characteristics of the granulosa cells in humans. Ito *et al.* (2010) reported that p38 MAPK activity was high in the granulosa cells of aged women and suggested that this enhanced activity was caused by oxidative stress. In addition, mRNA levels of the glutathione *S*-transferase family are reduced in the granulosa cells of aged females (Ito *et al.*, 2008), and the mRNA and protein levels of SOD1, SOD2 and catalase are also decreased in the granulosa cells of aged women.

To the best of our knowledge, the present study is the first to demonstrate that the proliferative activity of granulosa cells harvested from old cows differ significantly from that of younger cows. Furthermore, it has been previously demonstrated that bovine oocyte quality reduces as donor age increases (Malhi *et al.*, 2007; Yamamoto *et al.*, 2010). Taken together, the decline in the proliferative activity of the granulosa cells observed in the present study may have influenced the quality of oocytes derived from old cows.

During ageing, the DNA methylation pattern of somatic cells changed, and DNA methyltransferase activity decreased (Fuke *et al.*, 2004; Kim *et al.*, 2009; Murgatroyd *et al.*, 2010). Furthermore, we detected greater loss of global cytosine methylation in the granulosa cells of old cows compared with those of young cows. It is plausible that this change may influence telomere length because telomere length is regulated by neighbouring histone modifications (Kim *et al.*, 2009). Although many factors are possible candidates for the age-related down-regulation in the proliferative activity of granulosa cells, we focused on the relative telomere lengths of the granulosa cells.

In primordial follicles, flattened granulosa cells surrounding oocytes remain quiescent for many years, and once follicle development starts, massive proliferation of the granulosa cells supports follicle development from primordial to antral follicles. It is generally accepted that passage in culture shortens the telomere lengths of somatic cells. However, cumulus and granulosa cells have high telomerase activity, (Lavranos et al., 1999; Russo et al., 2006; Liu & Li. 2010), which may enable their numerous replications. In the present study, the relative telomere length of cumulus cells decreased significantly following four serial passages in culture, which demonstrated the validity of the relative telomere length assay. When the relative telomere length of cumulus cells was compared between the two age groups, similar relative telomere length was observed. However, large deviations in telomere length of the granulosa cells were observed with an approximately 44-fold difference between the maximum and minimum values. In addition, the relative telomere length and telomerase activity of the granulosa cells from old cows was shorter and lower than those from young cows, although the difference was not significant (p = 0.09 and 0.053, respectively). The results suggest that the cumulus cells collected from COCs were less affected by ageing than the granulosa cells collected by aspiration of follicles, and the granulosa cells are more heterogeneous than the cumulus cells and more likely to be affected by the age of donor cows. The telomerase activity of granulosa cells differed between middle and basal layers in the follicles (Lavranos et al., 1999). In addition, Russo et al. (2006) used a quantitative FISH technique and demonstrated that pig granulosa cells have high telomerase activity from preantral to antral stage follicles, and only cumulus cells and antral granulosa layer cells, not basal granulosa cells, harbour elongated telomeres.

On the basis of these observations, we hypothesize that the causes of the low proliferative activity of the granulosa cells from old cows may be as follows: (1) the changes in global cytosine methylation affect the physiological conditions of the granulosa cells of old cows; (2) the frequency of atretic follicles in the ovaries of old cows is greater than that in the ovaries of young cows; and (3) the granulosa cells of old cows lose telomerase activity faster than those of young cows. However, additional studies are required.

It is worth noting that granulosa cells immediately differentiate during *in vitro* culture (Luck *et al.*, 1990; Wathes *et al.*, 1995; Zheng *et al.*, 2008), and culture conditions, such as serum supplementation, presence of occytes and the maturation stage of the occytes, profoundly affect the function and differentiation of granulosa cells (Maillet *et al.*, 2002; Lucidi *et al.*, 2003; Gilchrist *et al.*, 2004; Parborell *et al.*, 2005). In the present study, we used the TCM-199 medium supplemented with 0.5 mM pyruvate and 5% FCS, and experiments were conducted within 29 h. During this period, our culture conditions may have affected the degree of differentiation and apoptosis of the cells, and this will require further studies.

In conclusion, the relative telomere lengths and telomerase activity of granulosa cells tend to differ in young and old cows, and the proliferative activity and global genomic methylation of the granulosa cells of old cows were significantly lower than those of young cows.

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References

- Aerts, J.M. & Bols, P.E. (2010). Ovarian follicular dynamics. A review with emphasis on the bovine species. Part II: Antral development, exogenous influence and future prospects. *Reprod. Domest. Anim.* 45, 180–7.
- Baird, D.T., Collins, J., Egozcue, J., Evers, L.H., Gianaroli, L., Leridon, H., Sunde, A., Templeton, A., Van, Steirteghem, A., Cohen, J., Crosignani, P.G., Devroey, P., Diedrich, K., Fauser, B.C., Fraser, L., Glasier, A., Liebaers, I., Mautone, G., Penney, G. & Tarlatzis, B; ESHRE, Capri Workshop Group. (2005). Fertility and ageing. *Hum. Reprod. Update* **11**, 261–76.
- Broekmans, F.J., Knauff, E.A., Te, Velde, E.R., Macklon, N.S. & Fauser, B.C. (2007). Female reproductive ageing: current knowledge and future trends. *Trends Endocrinol. Metab.* 18, 58–65.
- Downs, S.M. Regulation of the G2/M transition in rodent oocytes. *Mol. Reprod. Dev.* (2010). 77, 566–85.
- Fortune, J.E., Rivera, G.M. & Yang, M.Y. (2004). Follicular development: the role of the follicular microenvironment in selection of the dominant follicle. *Anim. Reprod. Sci.* 82– 83:109–26.

- Fuke, C., Shimabukuro, M., Petronis, A., Sugimoto, J., Oda, T., Miura, K., Miyazaki, T., Ogura, C., Okazaki, Y. & Jinno, Y. (2004). Age related changes in 5-methylcytosine content in human peripheral leukocytes and placentas: an HPLCbased study. *Ann. Hum. Genet.* 68, 196–204.
- Gilchrist, R.B., Ritter, L.J. & Armstrong, D.T. (2004). Oocytesomatic cell interactions during follicle development in mammals. *Anim. Reprod. Sci.* 82–83, 431–46.
- Gutiérrez, C.G., Campbell, B.K. & Webb, R. (1997). Development of a long-term bovine granulosa cell culture system: induction and maintenance of estradiol production, response to follicle-stimulating hormone, and morphological characteristics. *Biol. Reprod.* 56, 608–16.
- Hagemann, L.J. (1999). Influence of the dominant follicle on oocytes from subordinate follicles. *Theriogenology* 51, 449– 59.
- Hagemann, L.J., Beaumont, S.E., Berg, M., Donnison, M.J., Ledgard, A., Peterson, A.J., Schurmann, A. & Tervit, H.R. (1999). Development during single IVP of bovine oocytes from dissected follicles: interactive effects of estrous cycle stage, follicle size and atresia. *Mol. Reprod. Dev.* 53, 451–8.
- Hamel, M., Dufort, I., Robert, C., Gravel, C., Leveille, M.C., Leader, A. & Sirard, M.A. (2008). Identification of differentially expressed markers in human follicular cells associated with competent oocytes. *Hum. Reprod.* 23, 1118– 27.
- Huang, Z. & Wells, D. (2010). The human oocyte and cumulus cells relationship: new insights from the cumulus cell transcriptome. *Mol. Hum. Reprod.* 16, 715–25.
- Hunter, M.G. (2000). Oocyte maturation and ovum quality in pigs. *Rev. Reprod.* **5**, 122–30.
- Ito, M., Muraki, M., Takahashi, Y., Imai, M., Tsukui, T., Yamakawa, N., Nakagawa, K., Ohgi, S., Horikawa, T., Iwasaki, W., Iida, A., Nishi, Y., Yanase, T., Nawata, H., Miyado, K., Kono, T., Hosoi, Y. & Saito, H. (2008). Glutathione S-transferase theta 1 expressed in granulosa cells as a biomarker for oocyte quality in age-related infertility. *Fertil. Steril.* **90**, 1026–35.
- Ito, M., Miyado, K., Nakagawa, K., Muraki, M., Imai, M., Yamakawa, N., Qin, J., Hosoi, Y., Saito, H. & Takahashi, Y. (2010). Age-associated changes in the subcellular localization of phosphorylated p38 MAPK in human granulosa cells. *Mol. Hum. Reprod.* 16, 928–37.
- Iwata, H., Goto, H., Tanaka, H., Sakaguchi, Y., Kimura, K., Kuwayama, T. & Monji, Y. (2011). Effect of maternal age on mitochondrial DNA copy number, ATP content and IVF outcome of bovine oocytes. *Reprod. Fertil. Dev.* 23, 424– 32.
- Kim, K.C., Friso, S. & Choi, S.W. (2009). DNA methylation, an epigenetic mechanism connecting folate to healthy embryonic development and aging. J. Nutr Biochem. 20, 917–26.
- Lavranos, T.C., Mathis, J.M., Latham, S.E., Kalionis, B., Shay, J.W. & Rodgers, R.J. (1999). Evidence for ovarian granulosa stem cells: telomerase activity and localization of the telomerase ribonucleic acid component in bovine ovarian follicles. *Biol. Reprod.* **61**, 358–66.
- Liu, J.P. & Li, H. (2010). Telomerase in the ovary. *Reproduction* **140**, 215–22.
- Lucidi, P., Bernabø, N., Turriani, M., Barboni, B. & Mattioli, M. (2003). Cumulus cells steroidogenesis is influenced by

the degree of oocyte maturation. *Reprod. Biol. Endocrinol.* **28**, 45.

- Luck, M.R., Rodgers, R.J. & Findlay, J.K. (1990). Secretion and gene expression of inhibin, oxytocin and steroid hormones during the in vitro differentiation of bovine granulosa cells. *Reprod. Fertil. Dev.* **2**, 11–25.
- Maillet, G., Bréard, E., Benhaïm, A., Leymarie, P. & Féral, C. (2002). Hormonal regulation of apoptosis in rabbit granulosa cells in vitro: evaluation by flow cytometric detection of plasma membrane phosphatidylserine externalization. *Reproduction* **123**, 243–51.
- Malhi, P.S., Adams, G.P. & Singh, J. (2005). Bovine model for the study of reproductive aging in women: follicular, luteal, and endocrine characteristics. *Biol. Reprod.* **73**, 45– 53.
- Malhi, P.S., Adams, G.P., Pierson, R.A. & Singh, J. (2006). Bovine model of reproductive aging: response to ovarian synchronization and superstimulation. *Theriogenology* **66**, 1257–66.
- Malhi, P.S., Adams, G.P., Mapletoft, R.J. & Singh, J. (2007). *Reproduction* **134**, 233–9.
- Malhi, P.S., Adams, G.P., Mapletoft, R.J. & Singh, J. (2008). Superovulatory response in a bovine model of reproductive aging. *Anim. Reprod. Sci.* **109**, 100–9.
- Mastromonaco, G.F., Perrault, S.D., Betts, D.H. & King, W.A. (2006) Role of chromosome stability and telomere length in the production of viable cell lines for somatic cell nuclear transfer. *BMC Dev. Biol.* **6**, 41.
- Murgatroyd, C., Wu, Y., Bockmühl, Y. & Spengler, D. (2010). The Janus face of DNA methylation in aging. *Aging* **20**, 107–10.
- Olovnikov, A.M. (1996). Telomeres, telomerase, and aging: origin of the theory. *Exp. Gerontol.* **31**, 443–8.
- Orisaka, M., Tajima, K., Tsang, BK. & Kotsuji, F. (2009). Oocyte-granulosa-theca cell interactions during preantral follicular development. *J. Ovarian Res.* **2**, 9.
- Ottolenghi, C., Uda, M., Hamatani, T., Crisponi, L., Garcia, J.E., Ko, M., Pilia, G., Sforza, C., Schlessinger, D. & Forabosco, A. (2004). Aging of oocyte, ovary, and human reproduction. *Ann. NY Acad. Sci.* 1034, 117– 31.
- Pandey, A.N., Tripathi, A., Premkumar, K.V., Shrivastav, T.G. & Chaube, S.K. (2010). Reactive oxygen and nitrogen

species during meiotic resumption from diplotene arrest in mammalian oocytes. *J. Cell Biochem.* **111**, 521–8.

- Parborell, F., Irusta, G., Vitale, A., Gonzalez, O., Pecci, A. & Tesone, M. (2005). Gonadotropin-releasing hormone antagonist antide inhibits apoptosis of preovulatory follicle cells in rat ovary. *Biol. Reprod.* 72, 659–66.
- Rodgers, R.J. & Irving-Rodgers, H.F. (2010). Morphological classification of bovine ovarian follicles. *Reproduction* **139**, 309–18.
- Russo, V., Berardinelli, P., Martelli, A., Di, Giacinto, O., Nardinocchi, D., Fantasia, D. & Barboni, B. (2006). Expression of telomerase reverse transcriptase subunit (TERT) and telomere sizing in pig ovarian follicles. *J. Histochem. Cytochem.* 54, 443–55.
- Schams, D. & Berisha, B. (2002). Steroids as local regulators of ovarian activity in domestic animals. *Domest. Anim. Endocrinol.* **23**, 53–65.
- Su, Y.Q., Sugiura, K. & Eppig, J.J. (2009). Mouse oocyte control of granulosa cell development and function: paracrine regulation of cumulus cell metabolism. *Semin. Reprod. Med.* 27, 32–42.
- Tatone, C., Carbone, M.C., Falone, S., Aimola, P., Giardinelli,
 A., Caserta, D., Marci, R., Pandolfi, A., Ragnelli, A.M.
 & Amicarelli, F. (2006). Age-dependent changes in the expression of superoxide dismutases and catalase are associated with ultrastructural modifications in human granulosa cells. *Mol. Hum. Reprod.* 12, 655–60.
- Wezel, I.L. & Rodgers, R.J. (1996). Morphological characterization of bovine primordial follicles and their environment *in vivo*. *Biol. Reprod.* **55**, 1003–11.
- Wathes, D.C., Perks, C.M., Davis, A.J. & Denning-Kendall, P.A. (1995). Regulation of insulin-like growth factor-I and progesterone synthesis by insulin and growth hormone in the ovine ovary. *Biol. Reprod.* **53**, 882–9.
- Yamamoto, T., Iwata, H., Goto, H., Shiratuki, S., Tanaka, H., Monji, Y. & Kuwayama, T. (2010). Effect of maternal age on the developmental competence and progression of nuclear maturation in bovine oocytes. *Mol. Reprod. Dev.* 77, 595– 604.
- Zheng, X., Price, C.A., Tremblay, Y., Lussier, J.G. & Carrière, P.D. (2008). Role of transforming growth factorbeta1 in gene expression and activity of estradiol and progesterone-generating enzymes in FSH-stimulated bovine granulosa cells. *Reproduction* **136**, 447–57.