

Acta Neuropsychiatrica 2017 All rights reserved DOI: 10.1017/neu.2016.56

Review Article

Glial cells as key elements in the pathophysiology and treatment of bipolar disorder

Keshavarz M. Glial cells as key elements in the pathophysiology and treatment of bipolar disorder.

Objective: The exact pathophysiology of bipolar disorder (BD) is not yet fully understood, and there are many questions in this area which should be answered. This review aims to discuss the roles of glial cells in the pathophysiology of BD and their contribution to the mechanism of action of mood-stabilising drugs.

Methods: We critically reviewed the most recent advances regarding glial cell roles in the pathophysiology and treatment of BD and the neuroprotective and neurotrophic effects of these cells.

Results: Postmortem studies revealed a decrease in the glial cell number or density in the specific layers of prefrontal and anterior cingulate cortex in the patients with BD, whereas there was no difference in other brain regions, such as entorhinal cortex, amygdala and hippocampus. Astrocytes and oligodendrocytes were the most important glial types that were responsible for the glial reduction, but microglia activation rather than loss may be implicated in BD. The decreased number or density of glial cells may contribute to the pathological changes observed in neurons in the patients with BD. Alteration of specific neurotrophic factors such as glial cell line-derived neurotrophic factor and S100B may be an important feature of BD. Glial cells mediate the therapeutic effects of mood-stabilising agents in the treatment of BD.

Conclusion: Recent studies provide important evidence on the impairment of glial cells in the pathophysiology and treatment of BD. However, future controlled studies are necessary to elucidate different aspects of glial cells contribution to BD, and the mechanism of action of mood-stabilising drugs.

Mojtaba Keshavarz

Shiraz Neuroscience Research Center, Shiraz University of Medical Sciences, Shiraz, Iran

Keywords: bipolar disorder; glial cells; glial cell line-derived neurotrophic factor mood-stabilising drugs; \$100B

Dr. Mojtaba Keshavarz, Shiraz Neuroscience Research Center, Shiraz University of Medical Sciences, Chamran Hospital, Chamran Boulevard, PO Box: 7194815644, Shiraz, Iran. Tel/Fax: +9871 3623 4508; E-mail: moj.ph60@yahoo.com

Accepted for publication September 24, 2016

First published online October 24, 2016

Significant outcomes

- The review discusses about the potential roles of glial cells in the pathophysiology of bipolar disorder (BD) by emphasising on the neurotrophic and neuroprotective roles of these cells.
- Glial cell loss especially in specific brain regions and glial cells-specific neurotrophic factor deprivation may have important contribution to the pathophysiology of BD.
- Glial cells, at least partly, may mediate the therapeutic effects of mood-stabilising drugs.

Limitations

- In spite of many evidence that addresses glial cell roles in the pathophysiology and treatment of BD, some inconsistencies are present that may be related to the obstacles present in the neuropsychopharmacological studies.
- Establishment of relationship between peripheral and central levels of glial cells-specific neurotrophic factors and assessing glial cell loss in regions revealed by neuroimaging studies seem very necessary.

Introduction

BD is a common, chronic mental disorder with a high rate of relapse, cognitive impairment, psychosocial disability and suboptimal outcome (1). The prevalence of BD is between 1% and 2% around the world (2,3). However, the real prevalence is mainly higher because of under-diagnosed subjects (4,5). BD is a heritable disorder that may disrupt several systems and functioning such as sleep/wake system, autonomic, motoric, cognitive and endocrine systems (6). The exact pathophysiology of this complex disorder remained elusive. However, it has been proposed that multiple layers of body function including behavioural, cellular and molecular systems may be involved in the pathogenesis of this disorder. Recent studies imply that the impaired cellular plasticity of neuronal and glial network or impaired communication of these cells may be responsible for BD (7–9). Several postmortem studies with neuroimaging investigations have implied about the roles of glial cell in the pathophysiology of this disorder. However, there are many questions in this area that should be answered to complete the puzzle of this disorder. This review aims to discuss the roles of glial cells in the pathophysiology of BD and their contribution to the mechanism of action of moodstabilising drugs.

Glial cells

Glial cells have been considered as the passive constitute of central nervous system (CNS) that only have supportive or nutritional roles for the neural network (10,11). However, recent evidence has proposed that high-order CNS functions may be the outcome of glial-neuronal interaction (12).

Astrocytes, oligodendrocytes and microglia are three main types of glial cells in the CNS, which are the most prevalent constitute of the cortex (13). Each glia type can be distinguished by specialised functions and specific morphology. For instance, astrocytes are responsible for the metabolic support of neurons and the development of blood-brain barrier, oligodendrocytes for the myelin production, whereas the function of microglia is important in the cell immunity in the CNS (14). However, dysfunction of any type of glial cells may be involved with the development of neuropsychiatric disorders (15). In general, all the glial types have important physiological roles that can regulate neuronal functions. Regulation of synapses (12), the clearance of extracellular ions and transmitters (16,17), the control of neuronal metabolism and migration (18–20), the regulation of brain energy supplies (15), neurotrophic factor synthesis and release (21), angiogenesis and immune function in the CNS (22), coping with brain insults (23), and myelin formation (24) are the most important functions of the glial cells. Among these functions, several studies have suggested that neurotrophic and neuroprotective roles of the glial cells may be very relevant to the pathophysiology and treatment of mood disorders (25,26).

Neurotrophic and neuroprotective roles of the glial cells

It has been shown that glial cells, especially astrocytes, may possess the potency to exert neuroprotective effects (27). Gliotoxins which abolish astrocyte functions have increased the susceptibility of neurons to the cellular stressors (28). Moreover, *in vivo* ablation of astrocytic function has been increased neuronal cell death and vulnerability to ischaemia-induced injuries (29,30). The roles of the glial cells in the production of neurotrophic factors, homoeostasis of glutamate and production of antioxidants may be the important mechanisms proposed for the neuroprotective actions of these cells.

Glial cells synthesise and release several neurotrophic factors such as brain-derived neurotrophic factor (BDNF), glial derived neurotrophic factor (GDNF) and S100B that regulates neuronal survival and plasticity (14). Accordingly, it was proposed that the deprivation of neurotrophic factors, potentially from the glial cells, may contribute to the neural damage or death (31).

Furthermore, some reports have demonstrated the role of astrocytes in the protection of neurons against neurotoxic agents such as hydrogen peroxide and glutamate (32,33). It has been revealed that antioxidants are present at high concentrations in the astrocytes (34,35), and protect other cells, particularly neurons, from oxidative stress-induced cell death (35). Moreover, neuro-astrocyte co-culture system showed the protective effects of astrocytes against hydrogen peroxide (H₂O₂)- and nitric oxide (NO)-induced neuronal death (32,36). In addition, astrocytes have important roles in the modulation of glutamate level that may be actively involved in the protection of neurons against excitotoxicity. Astrocytes protect neurons against excitotoxicity in the mixed neuro-astrocyte cultures by reducing extracellular level of glutamate (37). Recent findings on the cell loss in the brain of patients with mood disorders have shown that neurotrophic and neuroprotective effects of glial cells may be very relevant to the pathophysiology of these disorders (25).

Glial cell roles in BD

Reduction in the grey matter volume in the patients with BD

Neuroimaging studies have consistently shown a reduction in grey matter volume in the patients with BD. A meta-analysis conducted by Ellison-Wright and Bullmore (38) demonstrated that four regions of the grey matter were decreased in the patients with BD. The decrease in the grey matter volume was occurred in the right insula, perigenual anterior cingulate, left insula and subgenual anterior cingulate (38). The findings of their study were similar to the findings of other studies in which the authors have reported a reduction in the prefrontal lobe volume (39), the left anterior cingulate cortex (ACC) (39,40) and the left subgenual prefrontal cortex (PFC) (40,41). In contrast, a meta-analysis of amygdala volumes did not show any significant change in the patients with BD (42). It has been proposed that the glial cell loss, at least partly, is responsible for a reduction in the grey matter volume in the patients with BD (43). Moreover, it has been revealed that reduction in the neuronal density, which is more subtle than glial alteration, may be relevant to the morphometric changes detected in the patients with BD (44).

Glial cell loss in the brain of patients with BD

In the late 90s, a theory developed that glial cell abnormalities may play an important role in the pathophysiology of mood disorders (13,44). For the first time, Ongur et al. (45) found a selective reduction in the glial cell number and density in the subgenual PFC in the patients with an apparent family history of BD. Afterwards, Rajkowska et al. (46) demonstrated a significant alteration in the density, shape and size of the glial cells in the dorsolateral PFC of the patients with BD. Their results showed that the mean glial cell density was significantly reduced below the control values in the sublayer IIIc of PFC, and was associated with a marginally significant decrease in the sublayer Vb in the brain of patients with BD (46). Moreover, Brauch et al. (47) found a decrease in the glial size of the temporal cortex in the patients with BD may reflect glial dysfunction. In contrast, some studies have reported no glial cell reduction in the dorsolateral PFC of the patients with BD (48,49).

In addition to PFC, some investigators have focused on other brain regions to find glial abnormalities in the brain of patients with BD. In a study conducted in 2011, Gittins and Harrison (50) assessed glial cell density in the ACC and found lower glial density in all layers except Vb in the patients with major depressive disorder (MDD) and

BD. In contrast to the above mentioned investigations, several studies did not find any evidence of glial cell reduction in this region (51,52). It is important to notice that Gittins and Harrison study had the limitation of measuring glial cell density in only two patients with BD.

Some studies measured the density of the glial cells in the amygdala of the patients with BD and controls, but they found no difference in the glial density in the patients with BD (53–55). Interestingly, Bowley et al. (55) found glial cell reduction in the amygdala of un-medicated patients with BD. They proposed that mood-stabilising drugs such as lithium and valproate may attenuate glial reduction in the patients with BD (55). Moreover, Webster et al. (49) studied the glial reduction in the hippocampus and they reported no significant difference in the glial cell density in this region in the patients with BD. Table 1 summarises the studies that evaluated glial cells loss in different brain regions in the patients with BD.

An important limitation in postmortem studies is that many of them have assessed the region that was not match with the information received from the volumetric studies. Neuroimaging studies have shown grey matter volume reduction in the insula and cingulate cortex of patients with BD (38), whereas many postmortem studies have measured glial loss or reduced glial density in the other regions like amygdala or hippocampus. Moreover, some reports showed that glial size has increased in the patients with BD (46). Therefore, it can be suggested that the reduced density of the glial cells in patients with BD may be related to the increased glial size. In addition, in a study conducted by Rajkowska et al. (46), the authors reported that the glial cell alteration may be layer specific. Moreover, the mean age of participants in the majority of studies was above the 40 years, whereas mood disorders are very prevalent among younger adults. Furthermore, another important limitation of almost all of the above mentioned postmortem studies is the small sample size.

Implications of the loss of glial cells in BD

There are some hypotheses about the mechanisms underlying neuronal loss in the patients with mood disorders. However, there are very limited information about the cause and mechanism of glial loss or reduced glial density in the patients with mood disorders. Therefore, the exact mechanism underlying glial loss or reduced density of glial cells in the patients with BD needs to be further elucidated. Recent studies implied that chronic stress, cytokines, excitotoxicity and oxidative stress may be involved

Glial roles in bipolar disorder

Table 1. Glial cells loss in different brain regions in the patients with bipolar disorder

Location	Results	Subjects	Factor measured	Reference	
Prefrontal cortex					
PFC	Layer IV: decreased oligodendrocyte numerical density	15 BD 15 MDD 15 SChZ	Oligodendrocyte density	(137)	
	Decreased one isoform of GFAP	15 controls 24 SChZ 23 BP 19 MDD	Protein	(77)	
dIPFC	Reduced glial density in IIIc	23 controls 10 BD	Glial cell density	(46)	
	and Vb sublayers No difference between BD and controls	11 controls 10 BD 11 MDD 9 SChZ	Glial cell density	(48)	
	No difference between BD and controls	14 controls 15 BD 15 MDD 15 SChZ	Phosphorylated GFAP protein	(49)	
	Decreased oligodendrocyte- related and myelin-related genes mRNA	15 controls 15 BD 15 SChZ 15 controls	Oligodendrocyte-related and myelin-related genes mRNA	(80)	
Subgenual PFC	No change in GFAP mRNA Lower glial number in BD Lower glial number in familial BD	15 BD 14 MDD 15 SChZ 15 controls	GFAP mRNA Glial cells number	(45)	
Entorhinal cortex	No difference in astrocytes density	15 BD 14 MDD 15 SChZ 15 controls	GFAP-positive astrocyte density	(81)	
Anterior cingulate cortex	Lower glial density in all layers but Vb of mood disorders Lower GFAP in white matter of mood disorders	5 MDD 2 BD 9 controls	Glial cell density GFAP protein	(50)	
	No evidence for glial cell reduction	15 BD 15 MDD 15 SChZ 15 controls	Glial cell density	(51)	
	No difference in glial cells	10 BD 11 SChZ 12 controls	Glial cell density	(52)	
Amygdala	No reduction in BD Reduced in non-medicated BD	7 MDD 10 BD 12 controls	Glial cell density	(55)	
	No reduction in BD	8 MDD 9 BD	Glial cell density S100β-positive astrocytes	(53)	
	No difference in neuron or astrocytes in BD	10 controls 15 BD 15 MDD 15 SChZ	GFAP-positive astrocytes	(54)	
Hippocampus	No difference between BD and controls	15 controls 15 BD 15 MDD 15 SChZ 15 control	Phosphorylated GFAP- astrocytes	(49)	

BD, bipolar disorder; dIPFC, dorsolateral PFC; GFAP, glial fibrillary acidic protein; MDD, major depressive disorder; mRNA, messenger RNA; PFC, prefrontal cortex; SChZ, schizophrenia.

The results only show the condition of patients with bipolar disorder compared with control group.

in the glial loss in the patients with BD. It has been proposed that volume change in the patients with mood disorder may be related to the stress-induced reduction in the glial cells (56). Cytokines such as tumour necrosis factor-α (TNF-α) can induce glial apoptosis (57) and reduce glial fibrillary acidic protein (GFAP) content in the primary mouse astrocyte culture and human glioblastoma cells (58). Stimulation of TNF-receptor with enhanced level of TNF- α activates apoptotic routes in the glial cells during mood episodes (57). Glial cells are sensitive to the elevated levels of glutamate (excitotoxicity) and oxidative stress (59-62). These vulnerabilities may contribute to the glial loss in the ischaemia and demyelinating diseases (59,63,64). However, the exact relationship between these factors and glial loss in the patients with BD should be determined (65).

The impact of glial loss or reduced glial cell density in the pathophysiology of BD is not fully elucidated. Moreover, it needs to be fully clarified that the glial reduction is a reaction to the process of underlying mechanism of mood disorders or have a causative role in these disorders (55). However, some evidence shows that decreasing glial number or density may play important roles in BD. Neuronal activity is crucially dependent upon homoeostatic functions of glial cell and glial loss can damage these modulatory roles of the glial cells (55).

Glial cells express a wide range of neurotransmitter receptors such as receptors for serotonin, norepinephrine and dopamine (66). It has been suggested that perturbation of brain monoamine system may be an important mechanism underlying mood disorder (67) and impairment of monoamine neurotransmission may be related, at least partly, to the glial loss in mood disorders (65).

In addition, glutamate activity may contribute to the neuroplasticity impairment detected in the patients with mood disorders (68). In normal condition, glutamate exerts substantial roles in the regulation of neuroplasticity, learning and memory (69). Glutamate also is the most important excitatory neurotransmitter responsible for the neurotoxicity in the CNS, which is known as excitotoxicity (68). Glial cells, particularly astrocytes, have a primary function in the regulation of glutamate level in the brain by glial transporters (70). Reuptake of glutamate by astrocytes decreases synaptic level of this excitatory neurotransmitter and protects neurons from apoptotic insults in the mixed culture (37). Astrocytes convert glutamate to the glutamine via an enzymatic reaction by glutamine synthetase and use glutamine as the source of glutamate and gamma-aminobutyric acid (GABA) for the neurons (71). Therefore, glial loss may give rise to the perturbation of glutamate homoeostasis and enhancement of the risk of excitotoxicity in neurons in the patients with mood disorder. These functions are mainly related to the astrocytes, whereas oligodendrocytes are the other glial type which is involved in the glial loss in the patients with BD (55). Therefore, the effects of oligodendrocyte loss in BD need to be exactly clarified in the future.

Contribution of each glial type to the loss of glial cells in BD

The exact cell type responsible for the glial loss in the patients with BD is not completely clear. Some studies have suggested that astrocytes and oligodendrocytes may be the most relevant cell types in the glial cell loss in patients with BD (14). However, recent reports imply that microglia activation but not loss may contribute to the pathophysiology of BD (53,72).

Contribution of astrocytes to the loss of glial cells in BD

Growing evidence suggests that astrocytes have a potential role in the pathophysiology of several CNS disorders (73). It was demonstrated that the structure and function of astrocytes have been altered in the neuropsychiatric disorders such as neurodegenerative disorders, epilepsy, schizophrenia and mood disorders (74–76). Therefore, many groups have investigated the potential roles of astrocytes in the pathophysiology of mood disorders.

Johnston-Wilson et al. (77) showed that some isoforms of GFAP, specific markers of astrocytes, were reduced in the frontal lobe of patients with BD. Moreover, Gittins and Harrison (50) documented lower level of GFAP in the white matter of patients with mood disorder (both BD and MDD). A study conducted by Gos et al. (78) showed that there was a reduction in S100B-immunocontent of astrocytes in the hippocampus of patients with BD and MDD. Toro et al. (79) also demonstrated that the level of GFAP was reduced in the PFC and orbitofrontal cortex of the patients with BD compared with the healthy controls. On the contrary, some studies have not supported astrocytes role as a main glial type responsible for the glial loss in the patients with BD. In line with this, the finding of a study conducted by Tkachev et al. (80) showed that there was no change in the level of GFAP in the PFC of patients with BD compared with the healthy controls. Similarly, Altshuler et al. (54) found no difference in the density of astrocyte in the amygdala of the patients with BD. Another study in the entorhinal cortex in 13 patients with BD showed that there was no difference in the astrocyte density between the patients with BD

and the healthy controls (81). Bernstein et al. (82) measured the numerical density of glutamine synthetase immunoreactive-astrocytes in eight cortical and two subcortical regions of the patients with BD. However, they found no change in the glutamine synthetase immunoreactive-astrocytes in the patients with BD (82).

It is very hard to make a conclusion about the astrocytic loss in the brain of patients with BD. Several factors can confound the results of the studies on the glial cell loss in the patients with BD. One important factor may be the proliferative effects of the mood-stabilising drugs on the astrocytes that affect the density of these cells in the CNS (83,84). Unfortunately, a majority of these studies have not reported drug effects on the GFAP or astrocyte density.

Contribution of oligodendrocytes to the loss of glial cells in BD

Oligodendrocyte is another cell type that may be responsible for the glial cell loss in the brain of patients with BD. Some studies showed that the gene expression of oligodendrocyte- and myelin-related genes has been greatly reduced and multiple transcription factors with regulatory roles on the myelin gene expression have been altered in the patients with BD (80). Moreover, Uranova et al. (85) showed that the numerical density of oligodendrocytes in the VI layer of the PFC in the patients with BD was reduced compared with the control groups. Because the volume of brain regions was not different between groups, authors concluded that the difference should be related to the reduced oligodendrocytes numbers in the patients with BD. Similarly, follow-up studies have demonstrated a reduction of myelin staining of the dorsolateral PFC in the patients MDD and BD (86). Gos et al. (78) showed that S100B-immunocontent of oligodendrocytes was reduced in the hippocampus of the patients with BD, but it was not reduced in that of the patients with MDD. Therefore, some studies suggested that the glial cells loss in the patients with BD may be related to the oligodendrocytes abnormalities (53,85).

Contribution of microglia to the pathophysiology of BD

Some evidence suggests the involvement of microglia in BD. According to this theory, chronic activation of the immune system mediated by the microglia in the CNS and enhanced production of inflammatory mediators may precipitate mood disturbances (87). Moreover, it has been postulated that the enhanced release of pro-inflammatory cytokines by the activated microglia exerts debilitating effects on the neuroprotective system, thereby interfering with the

pathophysiological changes in BD (65.87). In vivo analysis of peripheral blood showed that the proinflammatory cytokines have increased in various phases of BD (88). Moreover, Söderlund et al. (89) revealed higher level of pro-inflammatory markers in the cerebro-spinal fluids (CSF) of patients with BD. It has been shown that pro-inflammatory cytokines activate microglia and this may result in the enhanced release of glutamate from microglia and excitotoxicity (90). Therefore, microglia activation rather than microglia loss may contribute to the pathophysiology of BD. In this line, Hamidi et al. (53) showed no significant change in the density of microglia in the amygdala of patients with BD. However, there are very limited reports in the literature about the alteration of microglia in the patients with BD.

Roles of glial cells-specific neurotrophic factors in the pathophysiology of BD

Recently, it was hypothesised that the cell loss or the reduced density of glial cells in the patients with BD may be related to the abnormalities of the neurotrophic factor (91). Glial cells loss may be an important feature of BD and it is possible to assume that the abnormalities in the synthesis or release of neurotrophic factors that mainly origin form glial cells may be, at least partly, responsible for the pathologic events in the process of BD. Therefore, we discuss possible roles of some glial cells-specific neurotrophic factors in the pathophysiology and treatment of BD.

GDNF in BD

GDNF is an important neurotrophic factor for the dopaminergic neurons (21). However, this neurotrophic factor is extensively expressed throughout the CNS (92), and produces neuroprotective effects in different neuronal populations (93). It has been demonstrated that GDNF has regulatory effects on the noradrenergic neurons (94) and protects against kainate-induced oxidative stress in the rat hippocampus (95). Furthermore, it has been shown that GDNF can protect both neurons and glial cells against oxidative stress insults (94–96).

Some other studies have shown that GDNF can contribute to the pathophysiology of BD. In 2006, Takebayashi et al. (97) studied the alteration in the level of GDNF in the Japanese patients with BD and reported that total blood level of GDNF was lower in the patients with BD I and II compared with the healthy controls. Moreover, Zhang et al. (98) evaluated a Chinese Han cohort of patients with manic and depressive stages of BD for the serum

GDNF levels and their finding showed a lower level of serum GDNF in both stages of disease and an increment of this neurotrophic factor after 8 weeks of treatment with mood-stabilising drugs (98). On the contrary, Rosa et al. (99) showed that there was a higher level of GDNF in the manic and depressive phases of Brazilian patients with BD. Moreover, Barbosa et al. (26) reported a higher plasma GDNF level in the euthymic patients with BD compared with the manic and healthy controls. In contrast, Otsuki et al. (100) reported no alterations in the expression of GDNF messenger RNA in the peripheral blood cells in the depressive and remissive phases of patients with BD compared with the healthy controls. Tunca et al. (101) studied 92 patients with BD I and II and their finding showed that there was no difference in the serum GDNF level, whereas early onset patients (before 19) had higher serum GDNF level. Furthermore, in a study conducted by Rybakowski et al. (102), the authors reported that there was no difference in the serum level of GDNF in the drug-responder and nonresponder patients with BD. Table 2 summarises the studies which have measured the level of GDNF in the patients with BD.

Findings related to the peripheral measures of GDNF are inconsistent (101). An important limitation of most of these studies is the ignorance of the renal function. It seems that renal impairment may affect plasma level of neurotrophic factors including GDNF. To our knowledge, only one study has reported the renal function (99), whereas others have not reported any information about the renal condition of patients with BD. In addition, most of these studies were limited with the measurement of GDNF

in the peripheral tissues, whereas it is not clear whether these peripheral changes reflect actual changes in the CNS. Previous data indicate that GDNF penetrates very poorly across the brain-blood barrier (103,104). Moreover, small sample size of these studies may be another limitation and future meta-analyses may overcome this limitation. In spite of these controversies, it has been suggested that GDNF may be a non-specific peripheral marker in different phases of BD (98,105).

S100B in BD

Another glial cells-specific neurotrophic factor that may be involved in the pathophysiology of BD is S100B. S100B is an acidic Ca⁺²-binding protein (106) which is primarily found in the CNS, especially in the cytoplasm of astrocytes (107). It regulates cell shape, energy metabolism, contraction, cell-to-cell communication, intracellular signal transduction and cell growth (108). Interestingly, the extracellular S100B can produce both pro- and anti-apoptotic effects depending on its concentration (106). S100B at nano-molar concentrations acts as a growth and/or differentiation factor for neurons and astrocytes, whereas at micro-molar concentrations may activate apoptotic pathways (109).

Several animal and human studies have shown that S100B has a potential role in the pathophysiology of BD. Previous studies have shown that S100B has been changed in both serum (110,111) and CSF (112) of the patients with mood disorder. It was also revealed in an animal model of mania, S100B increased in the CSF of rodents (113). Moreover, Machado-Vieira et al. (114) showed that serum

Table 2.	Glial cell	line-derived	neurotrophic	factor	(GDNF)	level	in the	patients	with r	major	psychiatric	disorder

Subjects	Location	Findings	Drug effect	Reference
17 BD 39 MDD 56 control	Blood	Lower in BD and MDD vs. healthy No difference between BD and MDD	Lithium and antidepressant had no effect	(97)
44 BD I 14 healthy	Plasma	Higher GDNF immunocontent in manic and depressive episodes vs. healthy No difference between euthymic and healthy	NM	(99)
42 BD 60 MDD	Peripheral white blood cells	No difference between BD (depressive or remissive) and healthy controls	Mood stabilisers had no effects	(100)
28 control		Lower in MDD (depressive but not remissive) vs. healthy or BD	Antidepressant or remission in MDD increased GDNF	
40 BD I 50 controls	Serum	Lower in BD (manic or depressive) vs. controls	Antidepressants, antipsychotics and mood stabilisers had no effect	(98)
35 manic 35 euthymic 50 healthy	Plasma	Higher in euthymic BD vs. manic and healthy controls Negatively correlated with YMRS in BD	Drugs had no effect	(26)
92 BD I or II 61 healthy	Serum	Higher in early onset (before 19) than late onset BD No difference between groups	Lithium had no effect	(101)

BD, bipolar disorder; MDD, major depressive disorder; NM, not mentioned; YMRS, Young Mania Rating Scale.

S100B was higher in the 16 drug-naïve patients with acute mania compared with the healthy controls. Similarly, the results of other studies confirmed the elevated level of S100B in the manic and depressive but not euthymic patients (115). Furthermore, a meta-analysis demonstrated that serum S100B is higher in the acute phase of mania in the patients with BD (110). In another meta-analysis, Schroeter et al. (116) studied 174 patients with mood disorder and 102 healthy controls and their findings showed that both young and older patients with BD had higher levels of S100B compared with the healthy control. In addition to the peripheral levels of S100B, a postmortem study which was conducted by Dean et al. (117) revealed that S100B was increased in the area supramarginalis of parietal cortex and decreased in the dorsolateral PFC of patients with BD compared with the healthy controls. As it was mentioned previously, the S100B-immunocontent of astrocytes and oligodendrocytes has decreased in the hippocampus of patients with BD (78). However, it is not fully understood that the altered level of S100B was related to the passive diffusion from destroyed astrocytes or secretion from the activated astrocytes to repair neuronal injury (109,118).Table 3 summarises the studies which have measured S100B level in the patients with BD.

In addition, genetic studies provide new evidence for the association of S100B with BD. The gene encoding S100B in humans is located on chromosome 21q22.3 (119). This region shows a genetic linkage to BD (120, 121). Whole genome scan revealed that variants within S100B are related to BD (121). Moreover, Dagdan et al. (122) found that single nucleotide polymorphisms within the promoter of S100B gene are related to BD. Furthermore, a powerful whole genome gene expression studies with mega-analytic evidence has demonstrated an increase in the S100B gene expression in the hippocampus of patients with BD, but not in that of the patients with MDD (123).

Consequently, it has been suggested that S100B may be an important factor involved in the pathophysiology of BD. However, the exact role of this neurotrophic factor in the pathophysiology of BD remains to be elucidated.

Glial cell roles in the mechanism of action of mood stabiliser drugs

Regarding the roles of the glial cells in the pathophysiology of BD, it is possible to assume that these cells may be involved in the mechanism of action of moodstabilising drugs. Several studies have demonstrated that mood-stabilising drugs such as lithium increase the

Table 3. S100B level in the patients with mood disorders

Subjects	Location	Finding in BD	Reference
Part I	Serum	Higher in acute and remission phase vs. control	(110)
10 MDD		No difference between acute and remission phase	
10 control			
Part II	meta-analysis	Effect size:	
86 depressive		MDD: 2.57 ± 0.70	
63 manic		Manic: 1.53 ± 0.13	
44 euthymic		Euthymic: 2.54 ± 2.48	
		Higher in the acute phases (MDD and manic)	
8 BD I	Brain	Decreased in dorsolateral PFC compared with control and increased in area	(117)
20 control		supramarginalis of parietal cortex in BD compared with control	
20 BD	Serum	Higher in BD	(114)
20 control			
7 MDD	Serum	Higher in MDD and mania in admission and discharge	(107)
11 manic			
8 control			
57 BD	Brain, Letter to editor,	Increase in gene expression in hippocampus	(111)
60 control	mega-analysis		
9 MDD	Hippocampus	Decreased S100-immunocontent astrocytes in BD and MDD vs. control	(78)
6 BD		Decreased S100-oligodendrocytes in BD only	
13 control			
21 MDD	Serum	Increased in depressive and manic but not euthymic	(115)
32 manic			
31 euthymic BD			
32 healthy			
Rat	CSF in animal model of mania	30% increase in manic model animals	(113)

BA, Brodmann area; BD, bipolar disorder; CSF, cerebro-spinal fluid; MDD, major depressive disorder.

volume of brain grev matter (124). An magnetic resonance imaging (MRI) study has reported that chronic lithium therapy increased cortical grey matter volume (125) and N-acetyl aspartate which is a marker of cell viability in the CNS (126). Accordingly, it was suggested that lithium may act as a neurotrophic and neuroprotective agent (127). Some have proposed that this increase in grey matter volume may be related to the anti-apoptotic or proliferative effects of these agents on the glial cells. It has been demonstrated an increase in the GFAP (83) and astrocytosis (84) in the hippocampus of rats after 4 weeks treatment with lithium. Moreover, Rajkowska et al. (128) showed that 4 weeks treatment with lithium increases the total number of glial cells and the density of astrocytes in the hippocampus, but not PFC of mice. In our previous study, we showed that chronic lithium therapy increased bcl-2, an important anti-apoptotic agent, only in the astrocytes but not in the neuronal culture (129). It is likely that lithium may exert glioprotective effects by promoting bcl-2 up-regulation (55) that can be possibly relevant to the therapeutic mechanism of action of this agent (126,130). Human studies have supported glio-protective effects of mood-stabilising agents. Bowley et al. (55) showed that the two patients with BD who were not treated with lithium or valproic acid had low glial numbers that were significantly lower than either control or treated patients. They suggested that the mood-stabilising drugs may be effective in the modulation of the glial cell changes in the patients with mood disorders (55). They proposed that mood-stabilising drugs effect on the glial cells may be responsible, at least partly, for the therapeutic effects of these agents. However, their suggestion was based on only two patients and future studies may be warranted to further investigate this idea.

Regarding the glial cells-specific neurotrophic factors, especially GDNF and S100B roles in the pathophysiology of BD, it seems logical to assume that these neurotrophic factors may have important functions in the mechanism of action of moodstabilising drugs. Our study showed that chronic lithium therapy in the therapeutically relevant concentration increased the level of GDNF in the astrocytes culture (131). Moreover, lithium treatment has increased GDNF concentration in some cortical regions of the Flinders resistant rats (132). It has been reported that up-regulation of neurotrophic factors like GDNF, originated from astrocytes, may be responsible for the neurotrophic and neuroprotective effects of valproic acid (25). In contrast, it was shown that acute or chronic lithium treatments did not change GDNF protein expression in the brain of rat (133), spinal cord-derived progenitor cells (134) and neural precursor cells (135). Moreover, human clinical studies have shown no relationship between the serum level of GDNF and mood-stabilising drugs (97,100).

As mentioned above, there are some inconsistencies in the effects of mood-stabilising drugs on the GDNF. However, regarding the *in vitro* and animal studies, these differences may be related to different setting of experiments (cell culture vs. animal model or various animal models that were employed). Moreover, to our knowledge, there is no controlled clinical trial in the literature that specifically assessed the effects of mood-stabilising drugs on the level of GDNF and there is no established relationship between CNS and the serum level of GDNF.

There are few investigations on the lithium effects on the S100B. Chronic lithium therapy does not increase S100B in the rat primary astrocytes culture (136). Human studies reported that there is no relationship between mood-stabilising drugs and the serum level of S100B (107). Currently, it is impossible to draw any conclusive comment about the relationship of the glial cells-specific neurotrophic factors and the mechanism of action of the mood-stabilising drugs. However, future studies may be warranted to further investigate the possible roles of these neurotrophic factors in the mechanism of action of mood-stabilising drugs.

Consequently, it seems that glial cells may be important mediators for the action of mood-stabilising drugs. However, future controlled trials may shed more light on the contribution of these cells in the mechanism of action of these agents.

Future direction and concluding remarks

Some postmortem studies, but not all, have shown that there is a loss of glial cells or decreased in the glial density in some layers of PFC and ACC, but there is no difference in other regions of brain including entorhinal cortex, amygdala and hippocampus in the patients with BD. Astrocytes and oligodendrocytes may be the most important glial types which are responsible for the glial reduction in the brain of patients with BD. However, it seems necessary to assess the loss of glial cell in the regions that were established with neuroimaging studies. Furthermore, some studies have shown the alteration of the glial cells-specific neurotrophic factors (GDNF and S100B) in the different phases of BD. However, it is necessary to evaluate relationship between plasma and central levels of these neurotrophic factors. In addition, glial cells, at least partly, may mediate therapeutic effects of mood-stabilising agents in the treatment of BD.

Acknowledgements

The author would like to sincerely acknowledge Dr. Masoumeh Emamghoreishi for her guidance. Moreover, the author also thanks Mrs. Gholami for revising the manuscript.

Conflicts of interest

None.

References

- Belmaker R. Bipolar disorder. New Engl J Med 2004;351: 476–486.
- OSWALD P, SOUERY D, KASPER S et al. Current issues in bipolar disorder: a critical review. Eur Neuropsychopharmacol 2007;17:687–695.
- PINI S, DE QUEIROZ V, PAGNIN D et al. Prevalence and burden of bipolar disorders in European countries. Eur Neuropsychopharmacol 2005;15:425–434.
- Angst J, Azorin J-M, Bowden CL et al. Prevalence and characteristics of undiagnosed bipolar disorders in patients with a major depressive episode: the BRIDGE study. Arch Gen Psychiat 2011;68:791–799.
- GHAEMI SN, BOIMAN EE, GOODWIN FK. Diagnosing bipolar disorder and the effect of antidepressants: a naturalistic study. J Clin Psychiat 2000;61:804–808.
- Schloesser RJ, Huang J, Klein PS, Manji HK. Cellular plasticity cascades in the pathophysiology and treatment of bipolar disorder. Neuropsychopharmacol 2008;33:110–133.
- KAPCZINSKI F, FREY BN, KAUER-SANT'ANNA M, GRASSI-OLIVEIRA R. Brain-derived neurotrophic factor and neuroplasticity in bipolar disorder. Expert Rev Neurother 2008;8:1101–1113.
- CARLSON PJ, SINGH JB, ZARATE CA, DREVETS WC, MANJI HK. Neural circuitry and neuroplasticity in mood disorders: insights for novel therapeutic targets. NeuroRx 2006;3:22–41.
- CZEH B, FUCHS E, FLUGGE G. Altered glial plasticity in animal models for mood disorders. Curr Drug Targets 2013;14: 1249–1261.
- ABBOTT NJ, RÖNNBÄCK L, HANSSON E. Astrocyte-endothelial interactions at the blood-brain barrier. Nat Rev Neurosci 2006;7:41-53.
- 11. Acosta MT, Giola GA, Silva AJ. Neurofibromatosis type 1: new insights into neurocognitive issues. Curr Neurol Neurosci Rep 2006;6:136–143.
- ARAQUE A, PARPURA V, SANZGIRI RP, HAYDON PG. Tripartite synapses: glia, the unacknowledged partner. Trends Neurosci 1999;22:208–215.
- COTTER DR, PARIANTE CM, EVERALL IP. Glial cell abnormalities in major psychiatric disorders: the evidence and implications. Brain Res Bull 2001;55:585–595.
- RAJKOWSKA G, MIGUEL-HIDALGO J. Gliogenesis and glial pathology in depression. CNS Neurol Disord Drug Targets 2007;6:219.
- Eroglu C, Barres BA. Regulation of synaptic connectivity by glia. Nature 2010;468:223–231.
- VERKHRATSKY A, ORKAND RK, KETTENMANN H. Glial calcium: homeostasis and signaling function. Physiol Rev 1998;78: 99–141.
- Mennerick S, Zorumski CF. Glial contributions to excitatory neurotransmission in cultured hippocampal cells. Nature 1994;368:59–62.

- 18. TSACOPOULOS M, MAGISTRETTI PJ. Metabolic coupling between glia and neurons. J Neurosci 1996;16:877–885.
- MAGISTRETTI PJ. Role of glutamate in neuron-glia metabolic coupling. Am J Clin Nutr 2009;90:875S–880S.
- Anton ES, Cameron RS, Rakic P. Role of neuron-glial junctional domain proteins in the maintenance and termination of neuronal migration across the embryonic cerebral wall. J Neurosci 1996:16:2283–2293.
- LIN LF, DOHERTY DH, LILE JD, BEKTESH S, COLLINS F. GDNF: a glial cell line-derived neurotrophic factor for midbrain dopaminergic neurons. Science 1993;260: 1130–1132.
- Pont-Lezica L, Béchade C, Belarif-Cantaut Y, Pascual O, Bessis A. Physiological roles of microglia during development. J Neurochem 2011;119:901–908.
- Tremblay M-È, Stevens B, Sierra A, Wake H, Bessis A, Nimmerjahn A. The role of microglia in the healthy brain. J Neurosci 2011;31:16064–16069.
- BAUMANN N, PHAM-DINH D. Biology of oligodendrocyte and myelin in the mammalian central nervous system. Physiol Rev 2001:81:871–927.
- 25. CHEN PS, PENG G, LI G et al. Valproate protects dopaminergic neurons in midbrain neuron/glia cultures by stimulating the release of neurotrophic factors from astrocytes. Mol Psychiatry 2006;11:1116–1125.
- BARBOSA IG, HUGUET RB, SOUSA LP et al. Circulating levels of GDNF in bipolar disorder. Neurosci Lett 2011;502: 103–106.
- DHANDAPANI KM, HADMAN M, DE SEVILLA L, WADE MF, MAHESH VB, BRANN DW. Astrocyte protection of neurons: role of transforming growth factor-β signaling via Ac-Jun-AP-1 protective pathway. J Biol Chem 2003;278: 43329–43339.
- LARGO C, CUEVAS P, HERRERAS O. Is glia disfunction the initial cause of neuronal death in ischemic penumbra? Neurol Res 1996;18:445–448.
- NAWASHIRO H, BRENNER M, FUKUI S, SHIMA K, HALLENBECK JM. High susceptibility to cerebral ischemia in GFAPnull mice. J Cereb Blood Flow Metab 2000;20:1040–1044.
- Cui W, Allen ND, Skynner M, Gusterson B, Clark AJ. Inducible ablation of astrocytes shows that these cells are required for neuronal survival in the adult brain. Glia 2001;34:272–282.
- 31. Connor B, Dragunow M. The role of neuronal growth factors in neurodegenerative disorders of the human brain. Brain Res Rev 1998:27:1–39.
- 32. Desagher S, Glowinski J, Premont J. Astrocytes protect neurons from hydrogen peroxide toxicity. J Neurosci 1996;16:2553–2562.
- COTTER D, MACKAY D, LANDAU S, KERWIN R, EVERALL I. Reduced glial cell density and neuronal size in the anterior cingulate cortex in major depressive disorder. Arch Gen Psychiatry 2001;58:545–553.
- 34. PEUCHEN S, BOLAÑOS JP, HEALES SJ, ALMEIDA A, DUCHEN MR, CLARK JB. Interrelationships between astrocyte function, oxidative stress and antioxidant status within the central nervous system. Prog Neurobiol 1997;52:261–281.
- 35. Dringen R, Gutterer JM, Hirrlinger J. Glutathione metabolism in brain. Eur J Biochem 2000;**267**:4912–4916.
- TANAKA J, TOKU K, ZHANG B, ISHIHARA K, SAKANAKA M, MAEDA N. Astrocytes prevent neuronal death induced by reactive oxygen and nitrogen species. Glia 1999;28:85–96.

- 37. Rothstein JD, Dykes-Hoberg M, Pardo CA et al. Knockout of glutamate transporters reveals a major role for astroglial transport in excitotoxicity and clearance of glutamate. Neuron 1996;16:675–686.
- Ellison-Wright I, Bullmore E. Anatomy of bipolar disorder and schizophrenia: a meta-analysis. Schizophr Res 2010;117:1–12.
- ARNONE D, CAVANAGH J, GERBER D, LAWRIE S, EBMEIER K, McIntosh A. Magnetic resonance imaging studies in bipolar disorder and schizophrenia: meta-analysis. Br J Psychiatry 2009;195:194–201.
- Kempton MJ, Geddes JR, Ettinger U, Williams SC, Grasby PM. Meta-analysis, database, and meta-regression of 98 structural imaging studies in bipolar disorder. Arch Gen Psychiatry 2008:65:1017–1032.
- McDonald C, Zanelli J, Rabe-Hesketh S et al. Meta-analysis of magnetic resonance imaging brain morphometry studies in bipolar disorder. Biol Psychiatry 2004;56:411–417.
- HAJEK T, KOPECEK M, KOZENY J, GUNDE E, ALDA M, HÖSCHL C. Amygdala volumes in mood disorders – metaanalysis of magnetic resonance volumetry studies. J Affect Disord 2009;115:395–410.
- ÖNGÜR D, BECHTHOLT AJ, CARLEZON WA JR, COHEN BM. Glial abnormalities in mood disorders. Harv Rev Psychiatry 2013;22:334–337.
- RAJKOWSKA G. Cell pathology in bipolar disorder. Bipolar Disord 2002;4:105–116.
- ÖNGÜR D, DREVETS WC, PRICE JL. Glial reduction in the subgenual prefrontal cortex in mood disorders. Proc Natl Acad Sci USA 1998;95:13290–13295.
- RAJKOWSKA G, HALARIS A, SELEMON LD. Reductions in neuronal and glial density characterize the dorsolateral prefrontal cortex in bipolar disorder. Biol Psychiatry 2001;49:741–752.
- 47. Brauch RA, Adnan El-Masri M, Parker JC Jr, El-Mallakh RS. Glial cell number and neuron/glial cell ratios in postmortem brains of bipolar individuals. J Affect Disord 2006;91:87–90.
- 48. COTTER D, MACKAY D, CHANA G, BEASLEY C, LANDAU S, EVERALL IP. Reduced neuronal size and glial cell density in area 9 of the dorsolateral prefrontal cortex in subjects with major depressive disorder. Cereb Cortex 2002;12:386–394.
- Webster M, Knable M, Johnston-Wilson N, Nagata K, Inagaki M, Yolken R. Immunohistochemical localization of phosphorylated glial fibrillary acidic protein in the prefrontal cortex and hippocampus from patients with schizophrenia, bipolar disorder, and depression. Brain Behav Immun 2001;15:388–400.
- GITTINS RA, HARRISON PJ. A morphometric study of glia and neurons in the anterior cingulate cortex in mood disorder. J Affect Disord 2011;133:328–332.
- 51. CHANA G, LANDAU S, BEASLEY C, EVERALL IP, COTTER D. Two-dimensional assessment of cytoarchitecture in the anterior cingulate cortex in major depressive disorder, bipolar disorder, and schizophrenia: evidence for decreased neuronal somal size and increased neuronal density. Biol Psychiatry 2003;53:1086–1098.
- Benes FM, Vincent SL, Todtenkopf M. The density of pyramidal and nonpyramidal neurons in anterior cingulate cortex of schizophrenic and bipolar subjects. Biol Psychiatry 2001;50:395–406.

- Hamidi M, Drevets WC, Price JL. Glial reduction in amygdala in major depressive disorder is due to oligodendrocytes. Biol Psychiatry 2004;55:563–569.
- ALTSHULER LL, ABULSEOUD OA, FOLAND-ROSS L et al. Amygdala astrocyte reduction in subjects with major depressive disorder but not bipolar disorder. Bipolar Disord 2010;12:541–549.
- Bowley MP, Drevets WC, Öngür D, Price JL. Low glial numbers in the amygdala in major depressive disorder. Biol Psychiatry 2002;52:404–412.
- CZÉH B, MÜLLER-KEUKER JI, RYGULA R et al. Chronic social stress inhibits cell proliferation in the adult medial prefrontal cortex: hemispheric asymmetry and reversal by fluoxetine treatment. Neuropsychopharmacology 2007;32:1490–1503.
- BRIETZKE E, KAPCZINSKI F. TNF-α as a molecular target in bipolar disorder. Prog Neuropsychopharmacol Biol Psychiatry 2008;32:1355–1361.
- 58. Murphy GM, Lee YL, Jia XC et al. Tumor necrosis factor-α and basic fibroblast growth factor decrease glial fibrillary acidic protein and its encoding mRNA in astrocyte cultures and glioblastoma cells. J Neurochem 1995;65:2716–2724.
- DEWAR D, UNDERHILL SM, GOLDBERG MP. Oligodendrocytes and ischemic brain injury. J Cereb Blood Flow Metab 2003;23:263–274.
- 60. McDonald JW, Levine JM, Qu Y. Multiple classes of the oligodendrocyte lineage are highly vulnerable to excitotoxicity. Neuroreport 1998;9:2757–2762.
- 61. JUURLINK BH, THORBURNE SK, HERTZ L. Peroxide-scavenging deficit underlies oligodendrocyte susceptibility to oxidative stress. Glia 1998;22:371–378.
- 62. DANILOV CA, CHANDRASEKARAN K, RACZ J, SOANE L, ZIELKE C, FISKUM G. Sulforaphane protects astrocytes against oxidative stress and delayed death caused by oxygen and glucose deprivation. Glia 2009;57:645–656.
- 63. SUGAWARA T, LEWÉN A, NOSHITA N, GASCHE Y, CHAN PH. Effects of global ischemia duration on neuronal, astroglial, oligodendroglial, and microglial reactions in the vulnerable hippocampal CA1 subregion in rats. J Neurotrauma 2002;19: 85–98.
- 64. MATUTE C, ALBERDI E, DOMERCQ MA, PÉREZ-CERDÁ F, PÉREZ-SAMARTÍN A, SÁNCHEZ-GÓMEZ MAV. The link between excitotoxic oligodendroglial death and demyelinating diseases. Trends Neurosci 2001;24:224–230.
- RAJKOWSKA G. Postmortem studies in mood disorders indicate altered numbers of neurons and glial cells. Biol Psychiatry 2000;48:766–777.
- 66. Henn F. Neurotransmitters and astroglia lead to neuromodulation. Prog Brain Res 1982;55:241–252.
- 67. MILLAN MJ. The role of monoamines in the actions of established and 'novel' antidepressant agents: a critical review. Eur J Pharmacol 2004;500:371–384.
- 68. MAENG S, ZARATE CA. The role of glutamate in mood disorders: results from the ketamine in major depression study and the presumed cellular mechanism underlying its antidepressant effects. Curr Psychiatry Rep 2007;9: 467–474.
- RIEDEL G, PLATT B, MICHEAU J. Glutamate receptor function in learning and memory. Behav Brain Res 2003;140:1–47.
- 70. Anderson CM, Swanson RA. Astrocyte glutamate transport: review of properties, regulation, and physiological functions. Glia 2000;32:1–14.
- Pfrieger FW, Barres BA. Synaptic efficacy enhanced by glial cells in vitro. Science 1997;277:1684–1687.

- WATKINS C, SAWA A, POMPER M. Glia and immune cell signaling in bipolar disorder: insights from neuropharmacology and molecular imaging to clinical application. Transl Psychiatry 2014;4:e350.
- 73. Sofroniew MV, Vinters HV. Astrocytes: biology and pathology. Acta Neuropathol 2010;**119**:7–35.
- Volterra A, Meldolesi J. Astrocytes, from brain glue to communication elements: the revolution continues. Nat Rev Neurosci 2005;6:626–640.
- 75. MILLER G. The dark side of glia. Science 2005;308:778–781.
- Seifert G, Schilling K, Steinhäuser C. Astrocyte dysfunction in neurological disorders: a molecular perspective. Nat Rev Neurosci 2006;7:194–206.
- JOHNSTON-WILSON N, SIMS C, HOFMANN J et al. Diseasespecific alterations in frontal cortex brain proteins in schizophrenia, bipolar disorder, and major depressive disorder. Mol Psychiatry 2000;5:142–149.
- Gos T, Schroeter ML, Lessel W et al. S100Bimmunopositive astrocytes and oligodendrocytes in the hippocampus are differentially afflicted in unipolar and bipolar depression: a postmortem study. J Psychiat Res 2013;47:1694–1699.
- TORO CT, HALLAK JE, DUNHAM JS, DEAKIN JF. Glial fibrillary acidic protein and glutamine synthetase in subregions of prefrontal cortex in schizophrenia and mood disorder. Neurosci Lett 2006;404:276–281.
- TKACHEV D, MIMMACK ML, RYAN MM et al. Oligodendrocyte dysfunction in schizophrenia and bipolar disorder. Lancet 2003;362:798–805.
- DAMADZIC R, BIGELOW LB, KRIMER LS et al. A quantitative immunohistochemical study of astrocytes in the entorhinal cortex in schizophrenia, bipolar disorder and major depression: absence of significant astrocytosis. Brain Res Bull 2001;55:611–618.
- 82. Bernstein H-G, Meyer-Lotz G, Dobrowolny H et al. Reduced density of glutamine synthetase immunoreactive astrocytes in different cortical areas in major depression but not in bipolar I disorder. Front Cell Neurosci 2015:9:273.
- ROCHA E, RODNIGHT R. Chronic administration of lithium chloride increases immunodetectable glial fibrillary acidic protein in the rat hippocampus. J Neurochem 1994;63: 1582–1584.
- 84. ROCHA E, ACHAVAL M, SANTOS P, RODNIGHT R. Lithium treatment causes gliosis and modifies the morphology of hippocampal astrocytes in rats. Neuroreport 1998;9: 3971–3974.
- 85. Uranova N, Orlovskaya D, Vikhreva O et al. Electron microscopy of oligodendroglia in severe mental illness. Brain Res Bull 2001;55:597–610.
- REGENOLD WT, PHATAK P, MARANO CM, GEARHART L, VIENS CH, HISLEY KC. Myelin staining of deep white matter in the dorsolateral prefrontal cortex in schizophrenia, bipolar disorder, and unipolar major depression. Psychiatry Res 2007;151:179–188.
- MUNKHOLM K, BRAÜNER JV, KESSING LV, VINBERG M.
 Cytokines in bipolar disorder vs. healthy control subjects:

 a systematic review and meta-analysis. J Psychiatr Res 2013;47:1119–1133.
- BRIETZKE E, STERTZ L, FERNANDES BS et al. Comparison of cytokine levels in depressed, manic and euthymic patients with bipolar disorder. J Affect Disord 2009;116:214–217.

- SÖDERLUND J, OLSSON SK, SAMUELSSON M et al. Elevation of cerebrospinal fluid interleukin-1β in bipolar disorder. J Psychiatry Neurosci 2011:36:114.
- TAKEUCHI H, JIN S, WANG J et al. Tumor necrosis factor-α induces neurotoxicity via glutamate release from hemichannels of activated microglia in an autocrine manner. J Biol Chem 2006;281:21362–21368.
- BERK M, KAPCZINSKI F, ANDREAZZA A et al. Pathways underlying neuroprogression in bipolar disorder: focus on inflammation, oxidative stress and neurotrophic factors. Neurosci Biobehay Rev 2011;35:804–817.
- POCHON NM, MENOUD A, TSENG J, ZURN A, AEBISCHER P. Neuronal GDNF expression in the adult rat nervous system identified by in situ hybridization. Eur J Neurosci 1997;9: 463–471.
- AIRAKSINEN MS, SAARMA M. The GDNF family: signalling, biological functions and therapeutic value. Nat Rev Neurosci 2002;3:383–394.
- QUINTERO E, WILLIS L, ZAMAN V et al. Glial cell line-derived neurotrophic factor is essential for neuronal survival in the locus coeruleus-hippocampal noradrenergic pathway. Neuroscience 2004;124:137–146.
- CHENG H, Fu YS, Guo JW. Ability of GDNF to diminish free radical production leads to protection against kainate-induced excitotoxicity in hippocampus. Hippocampus 2004;14:77– 86.
- GRATACÒS E, PÉREZ-NAVARRO E, TOLOSA E, ARENAS E, ALBERCH J. Neuroprotection of striatal neurons against kainate excitotoxicity by neurotrophins and GDNF family members. J Neurochem 2001;78:1287–1296.
- 97. Takebayashi M, Hisaoka K, Nishida A et al. Decreased levels of whole blood glial cell line-derived neurotrophic factor (GDNF) in remitted patients with mood disorders. Int J Neuropsychopharmacol 2006;9:607–612.
- ZHANG X, ZHANG Z, SHA W et al. Effect of treatment on serum glial cell line-derived neurotrophic factor in bipolar patients. J Affect Disord 2010;126:326–329.
- Rosa AR, Frey BN, Andreazza AC et al. Increased serum glial cell line-derived neurotrophic factor immunocontent during manic and depressive episodes in individuals with bipolar disorder. Neurosci Lett 2006;407:146–150.
- OTSUKI K, UCHIDA S, WATANUKI T et al. Altered expression of neurotrophic factors in patients with major depression. J Psychiatr Res 2008;42:1145–1153.
- 101. Tunca Z, Ozerdem A, Ceylan D et al. Alterations in BDNF (brain derived neurotrophic factor) and GDNF (glial cell linederived neurotrophic factor) serum levels in bipolar disorder: the role of lithium. J Affect Disord 2014;166:193–200.
- 102. Rybakowski JK, Permoda-Osip A, Skibinska M, Adamski R, Bartkowska-Sniatkowska A. Single ketamine infusion in bipolar depression resistant to antidepressants: are neurotrophins involved? Hum Psychopharmacol Clin Exp 2013:28:87–90.
- Albeck DS, Hoffer BJ, Quissell D, Sanders LA, Zerbe G, Granholm A-CE. A non-invasive transport system for GDNF across the blood-brain barrier. Neuroreport 1997;8: 2293–2298.
- Kastin AJ, Akerstrom V, Pan W. Glial cell line-derived neurotrophic factor does not enter normal mouse brain. Neurosci Lett 2003;340:239–241.
- SCOLA G, ANDREAZZA AC. The role of neurotrophins in bipolar disorder. Prog Neuropsychopharmacol Biol Psychiatry 2015;56:122–128.

- SCHÄFER BW, HEIZMANN CW. The S100 family of EF-hand calcium-binding proteins: functions and pathology. Trends Biochem Sci 1996;21:134–140.
- Schroeter ML, Abdul-Khaliq H, Diefenbacher A, Blasig IE. S100B is increased in mood disorders and may be reduced by antidepressive treatment. Neuroreport 2002;13:1675–1678.
- ZIMMER D, CHAPLIN J, BALDWIN A, RAST M. S100-mediated signal transduction in the nervous system and neurological diseases. Cell Mol Biol 2005;51:201–214.
- VAN ELDIK LJ, WAINWRIGHT MS. The Janus face of glialderived S100B: beneficial and detrimental functions in the brain. Restor Neurol Neurosci 2003:21:97–108.
- SCHROETER ML, ABDUL-KHALIQ H, KREBS M, DIEFENBACHER A, BLASIG IE. Serum markers support disease-specific glial pathology in major depression. J Affect Disord 2008;111: 271–280.
- SCHROETER ML, STEINER J. Elevated serum levels of the glial marker protein S100B are not specific for schizophrenia or mood disorders. Mol Psychiatry 2009;14:235–237.
- Grabe HJ, Ahrens N, Rose H-J, Kessler C, Freyberger HJ. Neurotrophic factor S100beta in major depression. Neuropsychobiology 2001;44:88–90.
- MACHADO-VIEIRA R, SCHMIDT AP, ÁVILA TT et al. Increased cerebrospinal fluid levels of S100B protein in rat model of mania induced by ouabain. Life Sci 2004;76:805–811.
- 114. Machado-Vieira R, Lara D, Portela L et al. Elevated serum S100B protein in drug-free bipolar patients during first manic episode: a pilot study. Eur Neuropsychopharmacol 2002;12:269–272.
- ANDREAZZA AC, CASSINI C, ROSA AR et al. Serum S100B and antioxidant enzymes in bipolar patients. J Psychiatr Res 2007;41:523–529.
- SCHROETER ML, STEINER J, MUELLER K. Glial pathology is modified by age in mood disorders – a systematic metaanalysis of serum S100B in vivo studies. J Affect Disord 2011;134:32–38.
- 117. Dean B, Gray L, Scarr E. Regionally specific changes in levels of cortical S100β in bipolar 1 disorder but not schizophrenia. Aust NZ J Psychiatry 2006;40:217–224.
- 118. Hetzel G, Moeller O, Evers S et al. The astroglial protein S100B and visually evoked event-related potentials before and after antidepressant treatment. Psychopharmacology 2005;178:161–166.
- ALLORE R, O'HANLON D, PRICE R et al. Gene encoding the beta subunit of S100 protein is on chromosome 21: implications for Down syndrome. Science 1988;239:1311–1313.
- 120. McQuillin A, Bass N, Kalsi G et al. Fine mapping of a susceptibility locus for bipolar and genetically related unipolar affective disorders, to a region containing the C21ORF29 and TRPM2 genes on chromosome 21q22. 3. Mol Psychiatry 2006;11:134–142.
- 121. ROCHE S, CASSIDY F, ZHAO C et al. Candidate gene analysis of 21q22: support for S100B as a susceptibility gene for bipolar affective disorder with psychosis. Am J Med Genet B Neuropsychiatr Genet 2007;144:1094–1096.
- 122. Dagdan E, Morris DW, Campbell M et al. Functional assessment of a promoter polymorphism in S100B, a putative risk variant for bipolar disorder. Am J Med Genet B Neuropsychiatr Genet 2011;156:691–699.

- 123. SCHROETER ML, STEINER J, SCHÖNKNECHT P, MUELLER K. Further evidence for a role of S100B in mood disorders: a human gene expression mega-analysis. J Psychiatr Res 2014;53:84–86.
- 124. Bora E, Fornito A, Yücel M, Pantelis C. Voxelwise meta-analysis of gray matter abnormalities in bipolar disorder. Biol Psychiatry 2010;67:1097–1105.
- 125. Moore GJ, Вевсник JM, Wilds IB, Chen G, Manji HK. Lithium-induced increase in human brain grey matter. Lancet 2000;**356**:1241–1242.
- 126. Moore GJ, Bebchuk JM, Hasanat K et al. Lithium increases N-acetyl-aspartate in the human brain: *in vivo* evidence in support of bcl-2's neurotrophic effects? Biol Psychiatry 2000;**48**:1–8.
- 127. Machado-Vieira R, Manji HK, Zarate CA Jr. The role of lithium in the treatment of bipolar disorder: convergent evidence for neurotrophic effects as a unifying hypothesis. Bipolar Disord 2009;11(Suppl. 2):92–109.
- 128. RAJKOWSKA G, CLARKE G, MAHAJAN G et al. Differential effect of lithium on cell number in the hippocampus and prefrontal cortex in adult mice: a stereological study. Bipolar Disord 2016;18:41–51.
- 129. Keshavarz M, Emamghoreishi M, Nekooeian AA, Warsh JJ, Zare HR. Increased bcl-2 protein levels in rat primary astrocyte culture following chronic lithium treatment. Iran J Med Sci 2013;38:255.
- 130. Manji HK, Moore GJ, Chen G. Lithium up-regulates the cytoprotective protein Bcl-2 in the CNS in vivo: a role for neurotrophic and neuroprotective effects in manic depressive illness. J Clin Psychiatry 2000;61(Suppl. 9): 1478–1496.
- 131. EMAMGHOREISHI M, KESHAVARZ M, NEKOOEIAN AA. Acute and chronic effects of lithium on BDNF and GDNF mRNA and protein levels in rat primary neuronal, astroglial and neuroastroglia cultures. Iran J Basic Med Sci 2015;18:240.
- 132. ANGELUCCI F, ALOE L, JIMÉNEZ-VASQUEZ P, MATHÉ AA. Lithium treatment alters brain concentrations of nerve growth factor, brain-derived neurotrophic factor and glial cell line-derived neurotrophic factor in a rat model of depression. Int J Neuropsychopharmacol 2003:6:225–231.
- 133. Fukumoto T, Morinobu S, Okamoto Y, Kagaya A, Yamawaki S. Chronic lithium treatment increases the expression of brain-derived neurotrophic factor in the rat brain. Psychopharmacology 2001;158:100–106.
- 134. Su H, Chu T-H, Wu W. Lithium enhances proliferation and neuronal differentiation of neural progenitor cells *in vitro* and after transplantation into the adult rat spinal cord. Exp Neurol 2007;**206**:296–307.
- 135. Qu Z, Sun D, Young W. Lithium promotes neural precursor cell proliferation: evidence for the involvement of the non-canonical GSK-3β-NF-AT signaling. Cell Biosci 2011;1:1.
- EMAMGHOREISHI M, KESHAVARZ M, NEKOOEIAN AA. Chronic lithium treatment increased intracellular s100ß levels in rat primary neuronal culture. Acta Med Iranica 2015;53:89–96.
- 137. Uranova NA, Vostrikov VM, Orlovskaya DD, Rachmanova VI. Oligodendroglial density in the prefrontal cortex in schizophrenia and mood disorders: a study from the Stanley Neuropathology Consortium. Schizophr Res 2004;67:269–275.