

Review

Prevalence of extended-spectrum β -lactamase-producing *Escherichia coli* and residual antimicrobials in the environment in Vietnam

Shinji Yamasaki^{1*}, Tuyen Danh Le², Mai Quang Vien³, Chinh Van Dang⁴ and Yoshimasa Yamamoto⁵

¹ Graduate School of Life and Environmental Sciences, Osaka Prefecture University, 1-58, Rinkuourai-kita, Izumisano, Osaka 598-8531, Japan

² National Institute of Nutrition, 48B, Tang Bat Ho, Hai Ba Trung, Ha Noi, Vietnam

³ Pasteur Institute in Nha Trang, 8-10 Tran Phu, Xuong Huan, Nha Trang, Khanh Hoa, Vietnam

⁴ Institute of Public Health in Ho Chi Minh City, 159 Hung Phu, ward, district 8, Ho Chi Minh city, Vietnam

⁵ Graduate School of Pharmaceutical Sciences, Osaka University, 1-6, Yamadaoka, Suita, Osaka 565-0871, Japan

Received 2 October 2017; Accepted 3 November 2017

Abstract

Emergence and spread of antimicrobial-resistant bacteria, including extended-spectrum β -lactamase (ESBL)-producing *Escherichia coli*, have become serious problems worldwide. Recent studies conducted in Vietnam revealed that ESBL-producing *E. coli* are widely distributed in food animals and people. CTX-M-9 and CTX-M-1 are the most prevalent β -lactamases among the identified ESBLs. Furthermore, most of the ESBL-producing *E. coli* isolates were multi-drug resistant. Residual antimicrobials such as sulfamethoxazole, trimethoprim, sulfadimidine, cephalexin, and sulfadiazine were also detected at a high level in both animal meats and environmental water collected from several cities, including Ho Chi Minh city and Can Tho city. These recent studies indicated that improper use of antimicrobials in animal-originated food production might contribute to the emergence and high prevalence of ESBL-producing *E. coli* in Vietnam. Although clonal ESBL-producing *E. coli* was not identified, CTX-M-55 gene-carrying plasmids with similar sizes (105–139 kb) have been commonly detected in the ESBL-producing *E. coli* strains isolated from various food animals and human beings. This finding strongly suggests that horizontal transfer of the CTX-M plasmid among various *E. coli* strains played a critical role in the emergence and high prevalence of ESBL-producing *E. coli* in Vietnam.

Keywords: ESBL, CTX-M type, *E. coli*, antimicrobial residue.

Introduction

Emergence of antimicrobial-resistant bacteria has become a crucial issue in the world. In particular, emergence and rapid spread of extended-spectrum β -lactamase (ESBL)-producing bacteria have posed threat to the people not only in developing countries but also in developed countries, because ESBL can degrade not

only penicillin but also other classes of β -lactams. Furthermore, most of the ESBL-producing bacteria are multi-drug resistant. Thus, it is a serious problem in clinical settings.

ESBL-producing Enterobacteriaceae were first reported in the early 1980s (Knothe *et al.*, 1983), soon after the broad-spectrum cephalosporins were introduced into clinical settings. ESBLs can be generally divided into three groups, the TEM, SHV, and CTX-M types. In the 1980s and 1990s, the TEM and SHV types were most prevalent and were involved in hospital-associated infections caused by ESBL-producing

*Corresponding author. E-mail: shinji@vet.osakafu-u.ac.jp

Enterobacteriaceae. However, since the late 1990s, when third generation of β -lactams were widely introduced into the clinical settings, CTX-M types have become predominant and replaced TEM and SHV types very rapidly. Recent studies have revealed that widespread ESBL-producing bacteria, including *Escherichia coli*, were observed not only in the nosocomial settings but also in the community. It is of interest to note that CTX-M-type ESBL-producing Enterobacteriaceae were mostly *E. coli*.

Although at least 168 subtypes were reported for CTX-M types (as of 16 July 2017; <http://www.lahey.org/studies/other.asp>), the CTX-M type is divided into five subgroups, CTX-M-1, -2, -8, -9, and -25, based on the homology of amino acid sequences (D'Andrea *et al.*, 2015). In the early 2000s, carriage of CTX-M-type ESBL-producing *E. coli* in healthy people was still <10% of the community. However, in 2008, the carriage rate suddenly increased up to 60% in Thailand (Sasaki *et al.*, 2010). After 2008, the highest prevalence of ESBL-producing *E. coli* has been reported from Southeast Asia, followed by the Eastern Mediterranean and Western Pacific (Woerther *et al.*, 2013). Furthermore, the carriage rate of ESBL-producing *E. coli* in healthy individuals has been increasing year by year all over the world. The prevalence of ESBL-producing *E. coli* in Vietnam was not clear until recent initiation of a collaborative project between Vietnam and Japan (Woerther *et al.*, 2013; Le *et al.*, 2015; Nakayama *et al.*, 2015).

It has been suggested that the spread of ESBL-producing *E. coli* from animals to people is strongly associated with animal-originated food (Ewers *et al.*, 2012). Therefore, a two-country research collaborative project, titled 'Determining the outbreak mechanisms and development of a surveillance model for multi-drug resistant bacteria', was conducted in Vietnam with the support of the Japan International Cooperation Agency (JICA)/Japan Agency for Medical Research and Development (AMED) under the program of Science and Technology Research Partnership for Sustainable Developments (SATREPS). The SATREPS project focused on the prevalence of ESBL-producing *E. coli* from livestock to human beings, including animal-originated foods and residual antimicrobials in foods and environmental water. This review mostly summarizes the situation of the prevalence of ESBL-producing *E. coli* and antimicrobial residues in the environment, based on the results of the SATREPS project conducted in Vietnam between 2012 and 2017. The authors also reviewed recent Vietnamese studies of antimicrobial residues in meat and the environment.

Prevalence of ESBL-producing bacteria in livestock and fish in Vietnam

Chicken and pig production are among the most important agricultural industries in Vietnam. It has been suggested that overuse and improper use of antimicrobials in animal-originated food production could be one of the possible reasons why antimicrobial-resistant bacteria, including ESBL-producing *E. coli*, have emerged and spread. However, little is known about the prevalence of ESBL-producing *E. coli* in the livestock in Vietnam. Recently, Hinenoya *et al.* (2017) investigated the prevalence of

ESBL-producing *E. coli* in apparently healthy broiler chickens and pigs and their environment in Can Tho, the largest city in the southern part of Vietnam. ESBL-producing *E. coli* was isolated from 86.7% of cloacal swabs of chickens (13/15), 55.0% of rectal swabs of pigs (11/20), and 100% of their surrounding environments (4/4). All these isolates were multi-drug resistant, which was defined as resistant against more than three different classes of antimicrobials. Most of the chicken isolates carried both CTX-M-1- and TEM-type genes, while most pig isolates carried both CTX-M-9- and TEM-type genes. Furthermore, some of the ESBL-producing *E. coli* harbored virulence genes for diarrheagenic *E. coli*, such as *eae* and *astA*, suggesting that these *E. coli* isolates have a human pathogenic potential. It is of interest to note that 94.7 and 97.4% of chicken isolates were resistant to ciprofloxacin and were *mcr-1* gene-positive, which is responsible for colistin resistance, while only 13% of pig isolates were resistant to ciprofloxacin and none were *mcr-1* positive.

Aquaculture is also one of the most important agricultural industries in Vietnam. Therefore, the prevalence of ESBL-producing *E. coli* was also investigated for 16 wild fish and 32 cultured fish (16 each of *Pangasianodon hypophthalmus* and *Oreochromis* spp.). Hon *et al.* (2016) isolated ESBL-producing *E. coli* from 44.4 and 36.1% of wild (8/18) and cultured (13/36) fish, respectively. In total, 42 ESBL-producing *E. coli* were isolated from 54 fish and most of them were multi-drug resistant. Among 12 antimicrobials tested, 95.1, 92.9, 88.3, 88.1, 88.1, 78.6, 78.6, 76.2, 69.0, and 38.1% of the ESBL-producing *E. coli* were resistant to trimethoprim-sulfamethoxazole, nalidixic acid, gentamicin, tetracycline, streptomycin, chloramphenicol, ciprofloxacin, kanamycin, ceftazime, and ceftoxitin, respectively, while 100% were resistant to ampicillin and cefotaxime. It should be noted that the ESBL-producing *E. coli* obtained from cultured fish were more resistant to antimicrobials than those from wild fish.

Prevalence of ESBL-producing bacteria in various retail meats in Vietnam

Because it was demonstrated that chickens and pigs were highly contaminated with ESBL-producing *E. coli* (Hinenoya *et al.*, 2017), the prevalence of ESBL-producing *E. coli* in not only those meats but also in shrimp was investigated. Le *et al.* (2015) reported that the prevalence of ESBL-producing *E. coli* in retail poultry, pork, and shrimp was 58.7 (84/143), 32.0 (47/147), and 18.3% (11/60), respectively. Among 142 ESBL-producing *E. coli* isolated, 131, 85, and four isolates carried genes for CTX-M (92.3%), TEM (59.9%), and SHV (2.82%), respectively. The prevalence of CTX-M type in the isolates was 55.0 and 45.0% in CTX-M-1 and CTX-M-9, respectively, but no other CTX-M type was detected.

Prevalence and characteristics of ESBL-producing bacteria in healthy people in Vietnam

Carriage of CTX-M-type ESBL-producing *E. coli* was examined in healthy individuals in the northern part of Vietnam (Bui *et al.*,

2015). A total of 597 stool specimens were collected from 199 healthy individuals every 6 months, and the prevalence of ESBL-producing *E. coli* was determined to be 46.7, 52.8, and 46.3% at each sampling time, respectively. The CTX-M-9 type was predominant throughout the experimental period, such as 71.0, 86.7, and 85.9% in June 2013, November 2013, and June 2014, respectively. Furthermore, ESBL-producing *E. coli* were detected at least once in 175 individuals during the experimental periods at the individual level, regardless of CTX-M type. PFGE analysis revealed that none of the identical PFGE patterns of CTX-M-type ESBL-producing *E. coli* were observed in the same person, indicating a short carriage period of CTX-M-type ESBL-producing *E. coli* in healthy Vietnamese people (Bui *et al.*, 2015). These data suggested that CTX-M-type ESBL-producing *E. coli* was carried no more than 6 months in healthy individuals in a rural area in Vietnam. Hoang *et al.* (2017b) also reported isolation and characterization of CTX-M-type ESBL-producing *E. coli* from healthy adults in Ho Chi Minh city. Among ESBL-producing *E. coli* isolates from healthy adults, CTX-M-9 was the most prevalent, followed by CTX-M-1, and some of them carried virulence genes for diarrheagenic *E. coli* such as *astA*, EAF, *eae*, *elt*, and *eagg*, suggesting that healthy adults carried an *E. coli* potentially pathogenic to human beings.

The high positive rate of healthy carriage of ESBL-producing *E. coli* in Vietnam was comparable to that in other Southeast Asian countries such as Thailand and Laos (Table 1). Luvsansharav *et al.* (2012) reported that of 417 healthy residents in the community in Thailand, 69.3% carried ESBL-producing Enterobacteriaceae. Similarly, Nakayama *et al.* (2015) reported that among 198 and 57 healthy residents in Vietnam and Laos, 51.0 and 71.9% were positive for ESBL-producing Enterobacteriaceae, respectively, and all of them were identified to be ESBL-producing *E. coli*. Similar surveillance was also conducted in Japan and the results indicated, however, that out of 218 individuals, only 6.4% of healthy residents were positive for ESBL-producing Enterobacteriaceae (Luvsansharav *et al.*, 2011). Among those identified with ESBL-producing Enterobacteriaceae, 92.1, 94.8, 97.6, and 92.9% were CTX-M gene-positive in Vietnam, Thailand, Laos, and Japan, respectively (Table 1). Interestingly, among CTX-M types, in Southeast Asian countries, CTX-M-1 and CTX-M-9 were predominant and CTX-M-2 and CTX-M-9 were moderately high, while CTX-M-1 and CTX-M-8 were relatively high in Japan. These data indicate that the prevalence of CTX-M-1 and CTX-M-9 types in Southeast Asian countries may be common, and most probably CTX-M-1 could have originated from chicken, while CTX-M-9 could have originated from pigs.

Possible transmission route of ESBL-producing *E. coli* between animals, meats, and people

It has been suggested that the transmission of ESBL-producing *E. coli* occurs between people and animals, including poultry (Kluytmans *et al.*, 2013). Ueda *et al.* (2015) examined if transmission of ESBL-producing *E. coli* between chickens and human

beings occurred by comparing phylogeny, sequence typing of various house-keeping genes, antimicrobial susceptibility profile, and plasmid replicon type of ESBL-producing *E. coli* isolated from people and chickens. However, it was found that none of the *E. coli* isolated from people and chickens showed same characteristics, indicating that the transmission of CTX-M-9-type ESBL-producing *E. coli* between people and poultry may be limited.

On the other hand, Hoang *et al.* (2017a) conducted a study to compare the association between *E. coli* isolates carrying CTX-M-55-type plasmid from pork meats, wholesale market workers, and patients with urinary tract infection, because increasing prevalence of CTX-M-55 in human patient isolates has been reported (Zhang *et al.*, 2014). It was shown that *E. coli* harboring plasmid carrying *bla*_{CTX-M-55} genes found among pork, food seller, and patient isolates seemed to be clonally different, but these strains contained almost the same size of plasmid carrying *bla*_{CTX-M-55} genes with 104–139 kb. There is a report stating that the *bla*_{CTX-M-55} gene was transferred from *Sbigella sonnei*, isolated from a hospitalized patient showing abdominal pain, diarrhea, and fever, to an *E. coli* J53 recipient by conjugation (Lee *et al.*, 2013). Taken together, these observations suggest that clonal ESBL-producing *E. coli* might not be spread from meats to people, but that horizontal transfer of ESBL plasmid carrying *bla*_{CTX-M-55} gene could also be associated with the dissemination of ESBL-producing *E. coli* from meat to people.

Residual antimicrobial in various foods and environmental water in Vietnam

Residual antimicrobials in various food samples were examined by LC-MS/MS. Yamaguchi *et al.* (2015) reported that among 28 different antimicrobials tested, six (sulfaclozine, sulfamonomethoxine, enrofloxacin, difloxacin, norfloxacin, tilmicosin), one (sulfamethazine) and two (sulfamethazine, difloxacin) antimicrobials were detected in chicken, pork, and beef, respectively. As shown in Table 2, 17.3, 8.77, and 11.9% of chicken, pork, and beef, respectively, were contaminated with one of the antimicrobials tested. The contamination rate and tendency were different in geographic locations in Vietnam. In general, the contamination rate was higher in Ho Chi Minh city than that in Nha Trang. For example, 11.9% of chicken was contaminated with sulfaclozine in Ho Chi Minh city, while none in Nha Trang. On the other hand, 3.57% of chicken in Nha Trang was contaminated with norfloxacin but none in Ho Chi Minh city.

Similarly, among 32 antimicrobials tested, eight antimicrobial agents (ciprofloxacin, enrofloxacin, norfloxacin, ofloxacin, oxolinic acid, sulfamethazine, sulfamethoxazole, trimethoprim) were detected in fish and shrimp in Ho Chi Minh city, Nha Trang, and Thai Binh as shown in Table 2 (Uchida *et al.*, 2016). Fish was more contaminated with antimicrobials than shrimp in Ho Chi Minh city. Enrofloxacin was most prevalent antimicrobial, which was detected in 13.4 and 7.5% of fish and shrimp in Ho Chi Minh city, respectively. Fish was not contaminated with oxolinic acid, while shrimp was not contaminated with

Table 1. Prevalence of extended-spectrum β -lactamase-producing Enterobacteriaceae in residents of Japan and Southeast Asia

	Vietnam ^a	Thailand ^b	Laos ^a	Japan ^c
Sampling period	June 2013	November 2010	November 2012	July 2009 to June 2010
No. of participants	198	417	57	218
Age, mean	31	47	47	41.9
Age, range	1–90	20–85	17–75	20–70
Sex, male (%)	52	38	18	50.9
No. of ESBL producer	101 (51.0% ^d)	289 (69.3% ^d)	41 (71.9% ^d)	14 (6.4% ^d)
No. of CTX-M gene positive	93 (92.1% ^e)	274 (94.8% ^e)	40 (97.6% ^e)	13 (92.9% ^e)
CTX-M genotype groups				
M-1 group	34 (33.0% ^e)	106 (38.7% ^e)	19 (47.5% ^e)	2 (15.4% ^e)
M-2 group	1 (1.0% ^e)	0 (0% ^e)	0 (0% ^e)	4 (30.8% ^e)
M-8 group	2 (1.9% ^e)	2 (0.7% ^e)	2 (5.0% ^e)	2 (15.4% ^e)
M-9 group	66 (64.1% ^e)	166 (60.6% ^e)	19 (47.5% ^e)	5 (38.5% ^e)

^aNakayama *et al.* (2015).^bLuvsansharav *et al.* (2012).^cLuvsansharav *et al.* (2011).^dThe value was calculated by No. of participants in each country as a denominator.^eThe value was calculated by No. of ESBL producers in each country as a denominator.**Table 2.** Analysis of antimicrobial residues in food animals in Vietnam

Type of food animal	Compound (range $\mu\text{g kg}^{-1}$)	HCM (%)	NT (%)	TB (%)	Total (%)
Chicken ^a	Sulfaclozine (11–2710)	11.9 (10 ^b /84 ^c)	0	NT	6.41 (10/156)
	Sulfamonomethoxine (45)	0	1.19 (1/84)	NT	0.641 (1/156)
	Enrofloxacin (12–121)	3.57 (3/84)	1.19 (1/84)	NT	2.56 (4/156)
	Difloxacin (15)	1.19 (1/84)	0	NT	0.641 (1/156)
	Norfloxacin (21–123)	0	3.57 (3/84)	NT	1.92 (3/156)
	Tilmicosin (153–449)	5.95 (5/84)	5.95 (5/84)	NA	6.41 (10/156)
	Subtotal	20.2 (17/84)	11.9 (10/84)	NT	17.3 (27/156)
Pork ^a	Sulfamethazine (10–3,560)	14.0 (12/86)	3.53 (3/85)	NT	8.77 (15/171)
Beef ^a	Sulfamethazine (13–25)	5.88 (4 ^d /68)	NT	NT	5.88 (4*/68)
	Difloxacin (11–24)	2.94 (2 ^d /68)	NT	NT	2.94 (2*/68)
	Subtotal	7.35 (5/68)	NT	NT	7.35 (5/68)
Total	15.0 (34/226)	7.69 (13/169)	NA	11.9 (47/395)	
Fish/shrimp ^e	Ciprofloxacin	7.43 (15/202)/2.50 (4/160)	NT/0 (0/31)	0 (0/62)/0 (0/56)	5.68 (15/264)/1.62 (4/247)
	Enrofloxacin (6–4,002)	13.4 (27/202)/7.50 (12/160)	NT/0	0/0	10.2 (27/264)/4.86 (12/247)
	Norfloxacin (146)	0.495 (1/202)/0	NT/0	0/0	0.378 (1/264)/0
	Ofloxacin (11–28)	2.97 (6/202)/0	NT/0	0/0	2.27 (6/264)/0
	Oxolinic acid (6)	0/0.625 (1/160)	NT/0	0/0	0/0.405 (1/247)
	Sulfamethazine (6–42)	1.98 (4/202)/1.25 (2/160)	NT/3.23 (1/31)	0/0	1.52 (4/264)/0.810 (2/247)
	Sulfamethoxazole (12–5,760)	0.990 (2/202)/0	NT/0	0/1.79 (1/56)	0.758 (2/264)/0.810 (2/247)
	Trimethoprim (11–1,010)	2.97 (6/202)/0	NT/3.23 (1/31)	0/1.79 (1/56)	2.27 (6/264)/0.405 (1/247)
	Total	19.3 (39/202)/8.75 (14/160)	NA	0/1.79 (1/56)	14.8 (39/264)/6.48 (16/247)

^aModified from the data described in Yamaguchi *et al.* (2015).^bPositive number.^cTotal number tested.^dBoth sulfamethazine and difloxacin were detected in one sample.^eModified by the data described in Uchida *et al.* (2016). Chicken, pork, and beef include meat and internal organs such as heart, liver, etc.

HCM, Ho Chi Minh city; NT, Nha Trang; TB, Thai Binh; NT, not tested; NA, not applicable.

norfloxacin and ofloxacin, regardless of origin. Although β -lactams are often used in Vietnam, and the highest penicillin resistance was reported in Vietnam among Asian countries, β -lactams were not detected in their study. It cannot be excluded, however, that β -lactams might be degraded in the environment.

Although ampicillin is a commonly used antibiotic, it is rarely detected in the environment as described above, because it is easily degraded in water containing Ca^{2+} and Mg^{2+} . Sy *et al.* (2017), therefore, attempted to detect 2-hydroxy-3 phenylpyrazine (HPP), one of the degraded products of ampicillin and β -lactam antibiotics, from 98 environmental water samples including rivers, household ponds, and aquaculture ponds in Ha Noi, Thai Binh, and Can Tho. HPP residues were detected in 60 samples (11 from 26 rivers, 49 from 62 household pond, none from 10 aquaculture pond samples). The range of HPP residue concentrations was 1.3–410 ng l^{-1} . These data suggest that β -lactams used in medical settings or the veterinary field might contaminate environmental water.

Similarly, research was also conducted with other food such as eggs. Various antibiotics were detected in chicken eggs sold in supermarkets in Ho Chi Minh city (Yamaguchi *et al.*, 2017). A total of 111 egg packages from 11 different companies were examined for 28 different antimicrobials, and 16 samples were positive for any of eight compounds (enrofloxacin, ciprofloxacin, norfloxacin, sulfadimethoxine, sulfamethazine, sulfamonomethoxine, tilmicosin, and triphthoprim). Enrofloxacin was most prevalent and was detected in eight samples, in which two showed more than 1000 $\mu\text{g kg}^{-1}$ concentrations. Among 11 companies, five showed positive for any of eight antimicrobials, while six were not. In one company, more than 50% were positive for any of these antimicrobials.

Nakayama *et al.* (2017) investigated antimicrobial residues, antimicrobial resistance genes (ARGs), and microbiota in 12 freshwater sites from three different aquaculture systems, such as traditional backyard-based (V-A, V-B, V-C, and V-D), river-based (Ri-H), and land-based (Aq-F, Aq-I, Aq-J, and Aq-K) aquacultures and rivers (Ri-E, Ri-G, and Ri-L), in Can Tho city (shown in Fig. 1). Among 45 antimicrobials tested, only sulfamethoxazole, trimethoprim, sulfadimidine, cephalixin, and sulfadiazine were detected in 10, nine, seven, six, and five sites, respectively (Table 3). Only sulfadiazine and sulfamethoxazole were detected in V-A and V-B, respectively. However, four antimicrobials were detected in V-C and V-D. It should be noted that all five antimicrobials were detected in Ri-E, although two or three antimicrobials were detected in Ri-G, Ri-H, and Ri-L. However, three to four antimicrobials were detected in land-based aquaculture places. It should be noted that sulfamethoxazole, trimethoprim, and sulfadiazine were detected in all land-based aquaculture places and that highest concentrations of sulfadimidine, cephalixin, and sulfadiazine were detected in V-D.

On the other hand, *sul1*, *bla*_{CTX-M-1}, *bla*_{CTX-M-9}, *bla*_{SHV}, *bla*_{TEM}, *tetC*, *sul2*, and *tetM* genes were detected in 11, 10, nine, nine, nine, nine, and eight sites, respectively (Table 3). It is striking that all eight ARGs examined were detected in V-A, V-C, Ri-E, Aq-Ri-H, Ri-L, Aq-F, Aq-J, and Aq-K sites.

It should be emphasized that none of the genes were detected in a V-B site, and only one and two genes were detected in Aq-I and V-D sites, respectively. It is also specially mentioned that very high copy number of *bla*_{CTX-M-1}, *bla*_{CTX-M-9}, *bla*_{SHV}, *bla*_{TEM}, and *tetM* were detected in V-C sites. Remarkably high copy numbers of *tetC* and *sul1* genes were detected in both Ri-L and Aq-K sites. It should be emphasized that V-B was not contaminated with antimicrobials (only SMX was detected) and those genes, while V-C, Ri-E, Aq-J, and Aq-K places were highly contaminated with both antimicrobials and those genes. Although it is not known why V-B was not contaminated with antimicrobials and related genes, it is most likely that the use of antimicrobials might be well controlled. One of the most possible reasons could be that V-B had the least fish density at 0.79 fish/m³ (range 0.79–69.4 fish m⁻³). Alternatively, V-B is a little bit further from a residential area in comparison to other places, indicating that the contamination of antimicrobials from people could be limited. It is of interest to note that the V-A site was highly contaminated with antimicrobial genes, but only sulfadiazine was detected, while only a small copy of *sul1* was detected with a high concentration of sulfamethoxazole and low concentrations of trimethoprim, sulfadimidine, and cephalixin in the Aq-I site. Nevertheless, these data indicated that the improper use of antimicrobials in aquaculture systems might occur in Can Tho, resulting in the contamination of residual antimicrobials, which could be used for the veterinary field and most probably for medical settings, in environmental water in Can Tho. Furthermore, Managaki *et al.* (2007) detected sulfamethazine in the water of the region of Mekong Delta, and sulfamethazine was also detected in a pig farm pond in northern Vietnam (Hoa *et al.*, 2011), indicating that this compound could also be often used in Vietnam.

Although β -lactam antibiotics were not always detected in animal-originated food and environmental water, related genes such as *bla*_{CTX-M}, *bla*_{SHV}, *bla*_{TEM} were often detected with even high copy number (Table 3). Taken together, these results indicated that β -lactam antibiotics might be widely used in the agricultural field and might contribute to the emergence and spread of ESBL-producing *E. coli*, most probably through horizontal gene transfer of ESBL plasmid carrying *bla*_{CTX-M}. Alternatively, co-selection of ESBL-producing *E. coli* is also possible by other antimicrobials, such as sulfa drugs, tetracycline, etc., because most of the ESBL-producing *E. coli* isolated in Vietnam are multi-drug resistant.

Consumption of antimicrobials in Vietnam

Although exact statistical data regarding the consumption of antimicrobials used in medical and veterinary settings in Vietnam are not available, some literature or reports described the use of antimicrobials in Vietnam. For example, a report from the Ministry of Health of the Socialist Republic of Vietnam in collaboration with the global antibiotic resistance partnership and the Oxford University clinical research unit described that most commonly used antimicrobial classes in 15 representative hospitals in Vietnam were cephalosporins,

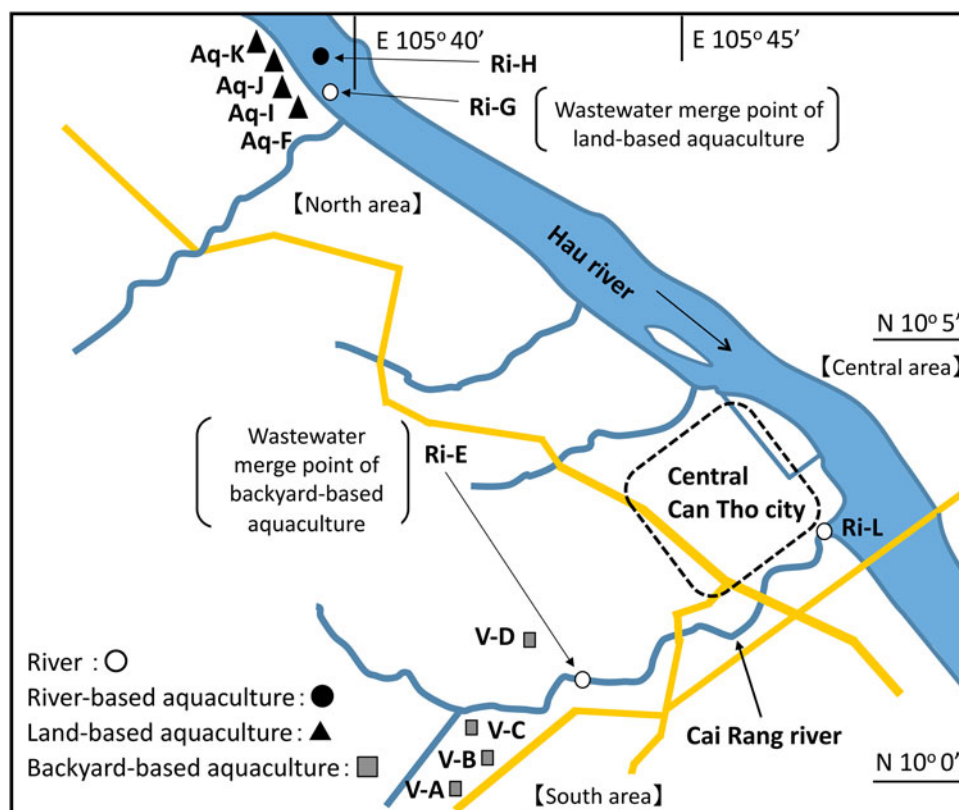


Fig. 1. Sampling sites in Can Tho city and its suburbs. Four backyard-based aquaculture sites (V-A, V-B, V-C, and V-D), four land-based aquaculture sites (Aq-F, Aq-I, Aq-J, and Aq-K), and four river sites (Ri-E, Ri-G, Ri-H, and Ri-L) including one river-based aquaculture site (Ri-H). Modified by Fig. 1 described in Nakayama *et al.* (2017).

Table 3. Detection of antimicrobial residue and antimicrobial resistance genes in 12 freshwater sites in Can Tho

Sample site	Residual antimicrobials (ng l ⁻¹)					Residual antimicrobial genes (relative copy number/1 ng DNA)							
	SMX	TMP	SDD	CEX	SDZ	Sul1	CTX-M-1	CTX-M-9	SHV	TEM	TetC	Sul2	TetM
1 V-A	ND	ND	ND	ND	43	120	85	3.4	99	7.3	100	4.4	59
2 V-B	11	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
3 V-C	47	ND	19	14	6.6	210	1200	1100	320	8800	440	33	360
4 V-D	ND	2.8	260	130	110	360	8.2	ND	ND	ND	ND	ND	ND
5 Ri-E	93	2.8	7.9	D	0.3	530	2900	680	330	3900	990	29	120
6 Ri-G	60	1.4	ND	ND	ND	25	460	60	21	70	24	6.8	ND
7 *Ri-H	42	1.0	ND	ND	ND	5.3	570	52	5.6	240	1.4	35	1.8
8 Ri-L	73	3.2	ND	ND	2.8	2400	310	50	290	66	43,000	10	28
9 Aq-F	87	1.5	18	ND	ND	110	51	2.4	32	320	640	2.7	21
10 Aq-I	140	2.9	16	D	ND	8.1	ND	ND	ND	ND	ND	ND	ND
11 Aq-J	68	2.6	19	D	ND	140	75	3.0	42	890	1300	3.9	12
12 Aq-K	130	3.3	14	6.0	ND	1900	910	150	38	350	57,000	42	40
Total positive (No.)	10	9	7	6	5	11	10	9	9	9	9	9	8

Modified from the data described in Nakayama *et al.* (2017).

V, household backyard-based aquaculture; Ri, river; *Ri, river-based aquaculture; Aq, land-based aquaculture; ND, not detected; D, detected but exact value is unknown; SMX, sulfamethoxazole; TMP, trimethoprim; SDD, sulfadimidine; CEX, cephalixin; SDZ, sulfadiazine.

followed by penicillins, macrolides, and quinolones. Although the sum of each antimicrobial consumption is not clear, β -lactam antibiotics are most frequently used in clinical settings in Vietnam, indicating that the emergence of ESBL-producing *E. coli* might occur.

To estimate the consumption of antimicrobials in chicken and pig production (Nguyen *et al.*, 2016), a total of 1462 commercial feed formulations were examined. Overall, it was estimated that 77 and 290 mg of in-feed antimicrobials were used to raise 1 kg of live chickens and pigs, respectively. It was calculated that

bacitracin (16%), chlortetracycline (11%), and enramycin (11%) were the most common antimicrobials used in chicken feed formulations, while bacitracin (25%), chlortetracycline (24%), and florfenicol (17%) were the most common antimicrobials used in pig feed formulations. The overall estimated amount of antimicrobials used for chicken and pig production in Vietnam was 42 and 980 tons, respectively. The most used antimicrobials were florfenicol (220 tons), chlortetracycline (210 tons), colistin (190 tons), bacitracin (150 tons), and tylosin (86 tons). However, about 57% of total usage of antimicrobials such as amoxicillin, colistin, chlortetracycline, neomycin, oxytetracycline, and bacitracin are considered as of critical importance or of high importance for human medicine. Therefore, it is important to reduce the use of antimicrobials, or at least these six antimicrobials, in the veterinary field in Vietnam.

Conclusions

Recent studies conducted in Vietnam indicated that the CTX-M plasmid might play a critical role on the dissemination of ESBL-producing *E. coli* among communities as well as animal-originated foods, rather than an association with particular clones of ESBL-producing *E. coli*. Use of antimicrobials contributes to not only multiplication of AMR bacteria, but also multiplication of AMR genes, which are relatively stable in the environment and could be utilized for horizontal gene transfer. Indeed, high levels of residual antimicrobials were detected in foods, and antimicrobials and ARGs were detected in the environment, including animal-originated foods in Vietnam. In Vietnam, farmers can purchase antibiotics easily without a prescription, and many farmers have insufficient information about appropriate use of the drugs. Nevertheless, risk factors for the dissemination of ESBL-producing *E. coli* could be animal-originated food and food animals, pets and healthy people, as well as patients, in particular those with UTI. Even among human beings, aging populations may also be particularly at risk. Poor access to drinking water, poverty, and a high population density are heavily associated with the dissemination of ESBL-producing bacteria. Therefore, it is important to limit the use of antimicrobials, not only to the agricultural sector but also to medical settings. Improved hygiene, such as encouragement of hand washing, also warrants more attention.

Acknowledgments

The authors would like to deeply thank Drs Kazuo Harada (Osaka University, Japan), Tatsuya Nakayama (Osaka University), Kazuto Uchida (Osaka Institute of Public Health), and Takahiro Yamaguchi (Osaka Institute of Public Health) for providing useful information and comments on this review article. The works described in this review article were supported in part by the Japan Agency for Medical Research and Development (AMED)/Japan International Cooperation Agency (JICA) as part of the Science and Technology Research Partnership for Sustainable Development (SATREPS).

References

- Bui TM, Hirai I, Ueda S, Bui TK, Hamamoto K, Toyosato T, Le DT and Yamamoto Y (2015). Carriage of *Escherichia coli* producing CTX-M-type extended-spectrum β -lactamase in healthy Vietnamese individuals. *Antimicrobial Agents and Chemotherapy* **59**: 6611–6614.
- D'Andrea MM, Arena F, Pallecchi L and Rossolini GM (2015). CTX-M-type β -lactamases: a successful story of antibiotic resistance. *International Journal of Medical Microbiology: IJMM* **303**: 305–317.
- Ewers C, Bethe A, Semmler T, Guenther S and Wieler LH (2012). Extended-spectrum β -lactamase-producing and AmpC-producing *Escherichia coli* from livestock and companion animals, and their putative impact on public health: a global perspective. *Clinical Microbiology and Infection* **18**: 646–655.
- Hinenoya A, Tran STT, Nguyen NT, Nguyen HC, Nguyen DDL, Hoang PH, Awasthi SP, Hassan J, Sumimura Y, Yamamoto Y and Yamasaki S (2017). Isolation and molecular characterization of extended-spectrum β -lactamase (ESBL)-producing *Escherichia coli* from industrial food animals in Mekong Delta, Vietnam. *Japanese Journal of Veterinary Research*, in press.
- Hoa PT, Managaki S, Nakada N, Takada H, Shimizu A, Anh DH, Viet PH and Suzuki S (2011). Antibiotic contamination and occurrence of antibiotic-resistant bacteria in aquatic environments of northern Vietnam. *Science of the Total Environment* **409**: 2894–2901.
- Hoang TA, Nguyen TN, Ueda S, Le QP, Tran TT, Nguyen TN, Dao TV, Tran MT, Le TT, Le TL, Nakayama T, Hirai I, Do TH, Vien QM and Yamamoto Y (2017a). Common findings of bla CTX-M-55-encoding 104-139 kbp plasmids harbored by extended-spectrum β -lactamase-producing *Escherichia coli* in pork meat, wholesale market workers, and patients with urinary tract infection in Vietnam. *Current Microbiology* **74**: 203–211.
- Hoang PH, Awasthi SP, Nguyen PD, Nguyen NLH, Nguyen DTA, Le NH, Dang CV, Hinenoya A and Yamasaki S (2017b). Antimicrobial resistance profiles and molecular characterization of *Escherichia coli* strains isolated from healthy adults in Ho Chi Minh city, Vietnam. *Journal of Veterinary Medical Science* **79**: 479–485.
- Hon NTN, Hoa TTT, Thinh NQ, Hinenoya A, Nakayama T, Harada K, Asayama M, Warisaya M, Hirata K, Phuong NT and Yamamoto Y (2016). Spread of antibiotic and antimicrobial susceptibility of ESBL-producing *Escherichia coli* isolated from wild and cultured fish in the Mekong Delta, Vietnam. *Fish Pathology* **51**: S75–S82.
- Kluytmans JA, Overdeest IT, Willemsen I, Kluytmans-van den Bergh MF, van der Zwaluw K, Heck M, Rijnsburger M, Vandembroucke-Grauls CM, Savelkoul PH, Johnston BD, Gordon D and Johnson JR (2013). Extended-spectrum β -lactamase-producing *Escherichia coli* from retail chicken meat and humans: comparison of strains, plasmids, resistance genes, and virulence factors. *Clinical Infectious Diseases* **56**: 478–487.
- Knothe H, Shah P, Krcmery V, Antal M and Mitsuhashi S (1983). Transferable resistance to cefotaxime, cefoxitin, cefamandole and cefuroxime in clinical isolates of *Klebsiella pneumoniae* and *Serratia marcescens*. *Infection* **11**: 315–317.
- Le QP, Ueda S, Nguyen TNH, Dao TVK, Hoang TAV, Tran TTN, Hirai I, Nakayama T, Kawahara R, Do TH, Vien QM and Yamamoto Y (2015). Characteristics of extended-spectrum β -lactamase-producing *Escherichia coli* in retail meats and shrimp at a local market in Vietnam. *Foodborne Pathogens and Disease* **12**: 719–725.
- Lee W, Chung HS, Lee H, Yum JH, Yong D, Jeong SH, Lee K and Chong Y (2013). CTX-M-55-type extended-spectrum β -lactamase-producing *Shigella sonnei* from a Korean patient who had travelled to China. *Annals of Laboratory Medicine Annals of Laboratory Medicine* **33**: 141–144.
- Luvansharav UO, Hirai I, Niki M, Arisa N, Aya Yoshinaga A, Moriyama T and Yamamoto Y (2011). Prevalence of fecal carriage of extended-spectrum β -lactamase-producing *Enterobacteriaceae*

- among healthy adult people in Japan. *Journal of Infection and Chemotherapy* **17**: 722–725.
- Luvansharav UO, Hirai I, Nakata A, Imura K, Yamauchi K, Niki M, Komalamisra C, Kusolsuk T and Yamamoto Y (2012). Prevalence of and risk factors associated with faecal carriage of CTX-M β -lactamase-producing Enterobacteriaceae in rural Thai communities. *Journal of Antimicrobial Chemotherapy* **67**: 1769–1774.
- Managaki S, Murata A, Takada H, Tuyen BC and Chiem NH (2007). Distribution of macrolides, sulfonamides and trimethoprim in tropical waters: ubiquitous occurrence of veterinary antibiotics in the Mekong Delta. *Environmental Science & Technology* **41**: 8004–8010.
- Nakayama T, Ueda S, Huong BT, Tuyen le D, Komalamisra C, Kusolsuk T, Hirai I and Yamamoto Y (2015). Wide dissemination of extended-spectrum β -lactamase-producing *Escherichia coli* in community residents in the Indochinese peninsula. *Infection and Drug Resistance* **8**: 1–5.
- Nakayama T, Hoa TTT, Harada K, Warisaya M, Asayama M, Hinenoya A, Lee JW, Phu TM, Ueda S, Sumimura Y, Hirata K, Phuong NT and Yamamoto Y (2017). Water metagenomics analysis reveals low bacterial diversity and the presence of antimicrobial residues and resistance genes in a river containing wastewater from backyard aquacultures in the Mekong Delta Vietnam. *Environmental Pollution* **222**: 294–306.
- Nguyen VC, Nguyen NT, Nguyen HN, Nguyen TMH, Nguyen VT, Thwaites G and Carrique-Mas J (2016). Antimicrobial consumption in medicated feeds in Vietnamese pig and poultry production. *Ecobhealth* **13**: 490–498.
- Sasaki T, Hirai I, Niki M, Nakamura T, Komalamisra C, Maipanich W, Kusolsuk T, Sa-Nguankiat S, Pubampen S and Yamamoto Y (2010). High prevalence of CTX-M β -lactamase-producing Enterobacteriaceae in stool specimens obtained from healthy individuals in Thailand. *Journal of Antimicrobial Chemotherapy* **65**: 666–668.
- Sy NV, Harada K, Asayama M, Warisaya M, Dung LH, Sumimura Y, Diep KT, Ha LV, Thang NN, Hoa TTT, Phu TM, Khai PN, Phuong NT, Tuyen LD, Yamamoto Y and Hirata K (2017). Residues of 2-hydroxy-3-phenylpyrazine, a degradation product of some β -lactam antibiotics, in environmental water in Vietnam. *Chemosphere* **172**: 355–362.
- Uchida K, Konishi Y, Harada K, Okihashi M, Yamaguchi T, Do MHN, Bui LT, Nguyen TD, Nguyen PD, Khong DT, Tran HT, Nguyen TN, Chau HVV, Dao KTV, Nguyen HTN, Kajimura K, Kumeda Y, Pham KT, Pham KN, Bui CT, Vien MQ, Le NH, Dang CV, Hirata K and Yamamoto Y. (2016) Monitoring of antibiotic residues in aquatic products in urban and rural areas of Vietnam. *Journal of Agricultural and Food Chemistry* **64**: 6133–6138.
- Ueda S, Ngan BTK, Huong BTM, Hirai I, Tuyen LD and Yamamoto Y (2015). Limited transmission of *bla*_{CTX-M-9}-type-positive *Escherichia coli* between humans and poultry in Vietnam. *Antimicrobial Agents and Chemotherapy* **59**: 3574–3577.
- Woerther PL, Burdet C, Chachaty E and Andremont A (2013). Trends in human fecal carriage of extended-spectrum β -lactamases in the community: toward the globalization of CTX-M. *Clinical Microbiology Reviews* **26**: 744–758.
- Yamaguchi T, Okihashi M, Harada K, Konishi Y, Uchida K, Do MH, Bui HD, Nguyen TD, Nguyen PD, Chau VV, Dao KT, Nguyen HT, Kajimura K, Kumeda Y, Bui CT, Vien MQ, Le NH, Hirata K and Yamamoto Y (2015). Antibiotic residue monitoring results for pork, chicken, and beef samples in Vietnam in 2012–2013. *Journal of Agricultural and Food Chemistry* **63**: 5141–5145.
- Yamaguchi T, Okihashi M, Harada K, Konishi Y, Uchida K, Hoang Ngoc Do M, Thi Bui L, Nguyen TD, Phan HB, Dang Thien Bui H, Nguyen PD, Kajimura K, Kumeda Y, Van Dang C, Hirata K and Yamamoto Y (2017). Detection of antibiotic in chicken eggs obtained from supermarkets in Ho Chi Minh city, Vietnam. *Journal of Environmental Science and Health. Part. B, Pesticides, Food Contaminants, and Agricultural Wastes* **52**: 430–433.
- Zhang J, Zheng B, Zhao L, Wei Z, Ji J, Li L and Xiao Y (2014). Nationwide high prevalence of CTX-M and an increase of CTX-M-55 in *Escherichia coli* isolated from patients with community-onset infections in Chinese county hospitals. *BMC Infectious Diseases* **14**: 659.