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Author for correspondence:

S. R. Singh, E-mail: shivramsingh22@gmail. com

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Effect of balanced fertilizers on soil quality and lentil yield in Gangetic alluvial soils of India

S. R. Singh¹, D. K. Kundu¹, P. Dey², Pushpa Singh³ and B. S. Mahapatra¹

¹ICAR-Central Research Institute for Jute and Allied Fibres, Barrackpore, Kolkata 700120, India; ²ICAR-Indian Institute of Soil Science, Nabibagh, Berasia Road, Bhopal 462038, India and ³ICAR-Indian Institute of Sugarcane Research, Raibareilly Road, Lucknow 226002, India

Abstract

Declining pulse production has caused wide concern in recent years. A field experiment was conducted to investigate effects of balance fertilizers based on soil test values and targeted yield equations on soil biological activities, soil quality, nutrient acquisition and grain yield of lentil. Treatments included the use of farmyard manure (FYM), bio-inoculants and inorganic fertilizers at different rates and combinations. The results revealed significant improvement in nodulation, microbial counts, microbial biomass carbon (MBC), soil respiration, soil enzymes and soil organic carbon (SOC) with integrated approaches (i.e. fertilizer plus FYM or bio-inoculants); these improvements led to achievement of the specific target yield of 2.0 t/ha. Although the highest yield was achieved with fertilizers applied for a target yield of 2.0 t/ha, there was significant decline in nodulation, microbial counts, MBC, soil respiration, soil enzymes, SOC and soil quality. Correlation between soil quality index (SQI) and grain yield suggested a significant influence of balanced fertilization based on soil tests and target yield. Principal component analysis revealed the average contribution of soil quality indicators towards SQI was in descending order of SOC > acid phosphatase activity > total culturable fungi > available phosphorus > BMC, which are crucial for sustainable lentil production in alluvial soils.

Introduction

Lentil is a major international pulse crop grown during the winter season in many countries where warm winters and hot summers are predominant. It is an important food legume with various uses as food and feed owing to its protein-rich grains and straw (Abbeddou *et al.* 2011). It is cultivated globally as a rain-fed crop on 5.48 m ha with 6.32 metric tonnes (mt) grain production and average productivity of 1.15 t/ha. In India, lentil occupies about 1.42 m ha with a production of 1.13 mt and average productivity of 0.70 t/ha (FAOSTAT 2016). The lower than average productivity is due mainly to the prevalence of moisture deficit, unbalanced fertilization, lack of improved varieties, and biotic and abiotic stresses. Improved varieties have shown yields of up to 5 t/ha in research fields and up to 3 t/ha in farmers' fields in Ethiopia (Schneider & Anderson 2010). In lentil, yield gaps of 30–105% with an average of 42% have been reported from different production zones in India (Ali & Gupta 2012). A major portion of this yield gap may be filled with improved nutrient management.

Lentil is a high-protein pulse crop that obtains its nitrogen (N) requirements for protein synthesis primarily from biological N_2 fixation. Bremer et al. (1989) and Matus et al. (1997) reported that lentil yield derives about 0.35 of its seed N from soil, <0.10 from applied N fertilizer and up to 0.70 from N_2 fixation. *Rhizobium* symbiosis has significant potential for N_2 fixation but depends upon the effectiveness of bacterial strain colonization. Hence, crop yield declines due to low N-fertilizer application: starter doses (basal) are insufficient and cannot fulfil the whole N requirement. In India, application of N as a starter dose (20 kg/ha) along with phosphorus (P, 40 kg/ha) and potassium (K, 20 kg/ha) and inoculation of seed with Rhizobium is a commonly recommended fertilizer regime for lentil. The prescription of N is based on the hypothesis that most of the N requirement is fulfilled by biological N fixation but sometimes this is negligible due to biotic and abiotic stress, leading to a decline in productivity of the crop. Moreover, despite decreased fertility levels and the introduction of new, high-yielding varieties, the recommended doses of mineral fertilizers have not been refined and in practice still remain old and outdated. Hence, lentil yield cannot attain its potential yield due to imbalances in fertilization. Gan et al. (2009) reported that application of higher doses of N fertilizer improved yield and timely maturity of chickpea. Application of 56 kg N/ha apparently increased yield of dry bean (Phaseolus vulgaris) when compared to N received only from N₂ fixation (Nleya et al. 2001). Nitrogen application could therefore be a more controllable means of increasing the productivity of lentil in India.

Alluvial soil of the Indo-Gangetic plains (IGP) is rich in K due to the presence of illite in its clay lattice. Therefore, in general farmers avoid K application as they perceive that the soils

already contain sufficient K for crop requirements. However, in practice, K uptake is consistently more than that applied in most of the crops grown in this soil; therefore, K reserves are being depleted gradually (Sarkar *et al.* 2014). Potassium is an essential component for enzyme activation, stomatal dynamics and water regulation but is applied in very low amounts, which affects the bio-chemical properties of plants adversely. Overexhausting organic matter, lower water retention capacity and low fertility status are other concerns in the Gangetic alluvial soils of India that have negative impacts on crop yield and soil quality (Singh *et al.* 1998). Therefore, crop yield declines due to sub-optimal fertilizer doses applied for legume cultivation.

Nutrient management is considered to be one of the most important factors for narrowing yield gap in lentil. Optimizing N, P and K doses for achieving the highest productivity may be based on soil testing, soil nutrient content in the rhizosphere (Chen et al. 2010), combination of organic resources and mineral fertilizer (Chivenge et al. 2011), site-specific nutrient management, and fertilizer recommendation based on soil testing, yield targets and crop responses (He et al. 2009; Singh et al. 2015). However, in order to improve nutrient use efficiency and achieve the highest crop yields of lentil, integrated nutrient management practices require urgent attention because unbalanced fertilization has become a common issue in India over recent years, particularly for macronutrients the N, P and K (He et al. 2009; Cui et al. 2010). Sub-optimal doses of inorganic fertilizer not only decrease crop yield but also affect nutrient status and soil quality (Singh et al. 2015, 2017). According to Hussain et al. (1999), soil quality assessment may be considered as a tool for evaluating sustainability of soil and crop management. Doran & Parkin (1994) defined soil quality as the capacity of soil to function within ecosystem boundaries to sustain biological productivity, maintain environmental quality and promote plant and soil health. Soil function capacity is normally reflected by its physical, chemical and biological properties, which have been noted to pertain to soil quality in agricultural land, grassland and forest soils (Schoenholtz et al. 2000). It influences basic soil functions including soil water movement and supply to plants, nutrient cycling and resistibility to organic and inorganic pollutants. Therefore, modification of fertilizer recommendations is essential to meet actual nutrient requirements through balanced fertilization in lentil.

A soil test and targeted yield equation (ST-TYE) approach was developed considering the three components, i.e. nutritional requirement of the crop, and per cent contribution of the available nutrient both in soil and the applied fertilizer (Ramamoorthy & Velayutham 1971). The ST-TYE approach is based on targeted yield response, agronomic efficiency and soil-test-based sitespecific nutrient management principles employed with the 'four R's method' (right source, right dose, right time and right place). In the present study, the continual use of fertilizer recommendations have been assessed through ST-TYE: (1) to determine whether the targeted yield of lentil is achieved with application of balanced fertilizers on the basis of initial soil test values and TYEs; (2) to compare the effect of balanced fertilization based on initial soil test values and targeted yields with the conventionally recommended dose of fertilizer and farmers' practice in respect of grain yield, soil quality and nutrient status of soil; and (3) to examine whether inoculation (Rhizobium + phosphate solubilizing bacteria) or application of farmyard manure (FYM) can reduce the required doses of N, P and K fertilizer without apparent reduction in soil quality and crop yield.

Materials and methods

Site description

A field experiment was carried out over three consecutive crop seasons (last week of November to first week of March) during 2010-2011, 2011-2012 and 2012-2013 at the Central Research Institute for Jute and Allied Fibres, Kolkata, West Bengal, India (88°26'E, 22°45'N, 9 m a.s.l., mean annual precipitation 1550 mm, Table 1). The experimental site is located on New Gangetic alluvial soils (Typic Ustocrept) according to the USDA classification system (Soil Survey Staff 1998), developed by deposition of alluvium carried out through the Ganga river and its tributaries in the IGP. Non-expanding illite minerals are dominant in these soils, followed by kaolinite and chlorite. The experimental site was non-saline and alkaline, moderately deep, well drained and composed of sandy clay loam (Table 2). Mean minimum and maximum air temperature during the crop season were 21.0 and 31.3 °C, respectively. Mean relative humidity in the morning and at noon was 92.7 and 62.2%, respectively.

Experimental design

A field experiment comprising nine treatments (Table 3) was laid out in a randomized block design (RBD) with three replicates. The treatments were: T1: Control (no mineral fertilizers, FYM or inoculants), T₂: FYM at the rate of 5 t/ha, T₃: Bio-inoculants (mixture of *Rhizobium* + *Bacillus megaterium*), T₄: Farmers' practice (15:30:20 kg N: phosphorus pentoxide (P_2O_5): potassium oxide (K₂O)/ha), T₅: Recommended dose of mineral fertilizers (30:40:20 kg N:P₂O₅:K₂O/ha), T₆: Balanced application of mineral fertilizers (47:50:107 kg N:P2O5:K2O/ha) based on initial soil test values and targeted yield equations (STV-TYE) for achieving target grain yield of 1.50 t/ha, T₇: Integrated approached (43:45:97 kg N:P₂O₅:K₂O/ha) based on initial STV-TYE for achieving target grain yield of 1.50 t/ha + FYM @ 5 t/ha and T_8 : Integrated approached (43:45:97 kg N:P2O5:K2O/ha) based on initial STV-TYE for achieving target yield of 1.50 t/ha lentil grain + bio-inoculants and T9: Balanced application of mineral fertilizers (73:72:153 kg N:P₂O₅:K₂O/ha) based on initial STV-TYE for achieving target grain yield of 2.0 t/ha. Each treatment was imposed on a fixed plot size $(5 \times 5 \text{ m}^2)$ in each year. Well-decomposed FYM @ 5 t/ha was incorporated into respective plots before preparation of the seedbed. It contained 40% moisture, 6.1 g N/kg, 3.0 g P/kg, 7.0 g K/kg, 0.18 g sulphur (S)/kg, 1.9 g iron (Fe)/kg, 0.039 g zinc (Zn)/kg, 0.16 g manganese (Mn)/kg and 0.0037 g copper (Cu)/kg on dry weight basis and had a C/N ratio of 39.1. Each plot was tilled separately with a power tiller to a depth of 20 cm, followed by levelling and planking with wooden planks. Indigenous symbiotic, free-living N-fixing bacterium Rhizobium leguminosarum and phosphatesolubilizing bacterial (PSB) strain B. megaterium var. phosphaticum were procured from the Survey, Selection and Mass Production Unit, Bidhan Chandra Krishi Vishva Vidyalaya, District, Nadia, West Bengal (India). These strains were selected for their effective N fixation, phosphate-solubilizing ability, growth-promoting capacity, their potential to suppress root pathogens and their capacity to interact positively with each other. In each year of the experiment, lentil seeds were inoculated with R. leguminosarum and B. megaterium var. phosphaticum at the rate of 500 g culture/ha seed before sowing: 10⁶-10⁷ colony forming units (cfu) per seed were applied via seed encapsulation. Seeds were air-dried for 30 min after inoculation and then sown. Apart from seed inoculation, charcoal-based bio-inoculant culture was also broadcast at the rate of 2.0 kg/ha

	Ma	ximum tem	iperature (°	°C)	Mir	nimum tem	perature (°	c)	Η	ımidity at r	norning (%	(-	Humidity at	1000 (%)			Rainfall	(mm)	
	2010	2011	2012	2013	2010	2011	2012	2013	2010	2011	2012	2013	2010	2011	2012	2013	2010	2011	2012	2013
Jan	25.3	23.7	23.9	27.6	9.8	13.3	10.5	10.5	92.8	96.0	95.7	95.7	51.4	61.9	46.4	46.4	0.00	40.0	0.00	0.00
Feb	23.3	28.5	27.6	34.6	14.9	14.3	14.0	14.0	96.7	92.5	93.3	93.3	45.6	42.9	46.9	46.9	15.0	30.8	8.80	8.80
Mar	28.8	33.0	34.6	35.3	22.1	19.6	19.9	19.9	91.3	90.5	91.1	91.1	42.3	49.8	40.7	40.7	1.40	29.1	0.00	0.00
Apr	35.2	34.4	35.3	36.7	26.7	23.0	23.6	23.6	87.3	91.0	87.9	87.9	54.1	54.6	49.2	49.2	19.4	68.1	78.6	78.6
May	37.4	34.3	36.7	35.2	26.0	24.7	26.2	26.2	89.0	91.5	87.3	87.3	64.0	68.5	55.9	55.9	165.1	178.8	58.5	58.5
Jun	35.3	33.5	35.2	32.1	26.7	26.3	26.7	26.7	93.3	93.8	89.4	89.4	70.1	72.9	68.5	68.5	193.2	387.3	150.1	150.1
Jul	34.8	32.4	32.1	31.8	26.4	26.0	25.6	25.6	92.9	93.0	93.4	93.4	75.5	75.5	83.1	83.1	204.6	357.5	420.1	420.1
Aug	32.9	31.2	31.8	32.0	26.1	25.9	25.4	25.4	93.2	94.7	94.0	94.0	77.1	83.7	17.6	79.6	240.3	587.5	232.5	232.5
Sep	32.6	31.4	32.0	31.5	25.0	25.5	25.1	25.1	93.7	96.3	93.6	93.6	77.6	78.8	76.6	76.6	148.5	545.8	302	302
Oct	32.6	32.8	31.5	28.5	22.6	23.8	21.6	21.6	94.8	92.8	91.6	91.9	68.9	60.0	62.1	62.1	59.2	57.6	129.8	129.8
Nov	32.2	30.3	28.5	24.8	18.7	18.2	17.6	17.8	94.9	93.6	92.7	92.7	57.5	50.6	52.8	52.8	0.00	0.00	44.8	44.8
Dec	31.0	25.3	24.8	23.9	1.11	13.3	12.2	12.2	92.9	96.1	96.4	96.4	50.5	55.9	56.4	56.4	0.00	0.00	9.40	9.40

Table 1. Temperature, humidity and rainfall during the experimental period (2010-2013)

before preparation of the seedbed, to ensure microbial inoculation. Lentil (var. B 256) was sown at the rate of 35 kg/ha in the rabi season (last week of November to the first week of March) in each year at a planting distance of 25×5 cm². A TYE (FN = 4.94 T-0.09 SN-0.07 ON, FP₂O₅ = 4.43 T-0.24 SP-0.16 OP and FK₂O = 9.03 T-0.24 SN-0.17 OK) was developed for lentil during 2008-2009 for alluvial soil of the IGP of India (i.e. the experimental site) considering three basic components, i.e. nutrient requirement to achieve grain yield of 100 kg and utilization efficiencies of nutrients available both in soil and those applied through inorganic fertilizers and organic sources (Ramamoorthy & Velayutham 1971). Based on initial soil test values and achievable fixed target yield of lentil, requirements of mineral fertilizer for treatments T₆, T₇, T₈ and T₉ were calculated using the target yield equations. A full dose of N, P and K fertilizers were applied as a basal dressing of urea, DAP (diammonium phosphate) and MOP (muriate of potash), respectively, at sowing.

Experimental methods

In order to count nodule numbers and record the fresh and dry weight of nodules, ten healthy plants were gently removed randomly from each plot, without disturbing the roots, at early bloom stage (Erskine et al. 1990). The freshly removed plants were washed thoroughly with tap water, surface-dried with filter paper to enable the recording of nodule number and fresh weight before drying in a hot air oven to obtain the dry weight, firstly at 70 °C for 48 h followed by 80 °C for 72 h, until a constant weight of nodules was obtained. Data on grain and straw yield were taken at maturity by difference methods. The total dry weights of biological and grain yields were measured separately. Straw yield was obtained by deducting grain yield from biological yield (straw yield = biological yield – grain yield). Three representative surface (0-15 cm) soil samples were collected from each plot using an auger before sowing and at harvest in each year. Composite soil samples were prepared from the three representative soil samples after proper mixing and homogenization and each soil sample was divided into two parts. One half of the fresh moist soil samples were packed in airtight plastic bags and stored at -20 °C for analysis of microbial biomass carbon (MBC), microbial counts and extra-cellular enzymatic activities. The remaining half was air-dried, ground and passed through a 2-mm sieve and used for analysis of physical and chemical properties. Soil pH was measured with a compound electrode using soil-to-water ratio of 1: 2.5. Soil organic carbon was determined according to Walkley & Black (1934) method. Total N was determined by the Kjeldahl method and available N (N_a) by the alkaline permanganate (KMnO₄-N) method using an auto N analyser as suggested by Subbiah & Asija (1956). Available phosphorus (P_a) was determined by the Olsen method (Olsen et al. 1954) and available potassium (K_a) was analysed by the 1 N ammonium acetate (NH₄OAc) method given by Jackson (1973). Representative straw and grain samples were collected from each plot at harvest for determination of nutrient content in the straw and grain and were dried in a hot air oven at 70 °C for 5 days. The dried samples were ground in a stainless steel Wiley mill and wet-digested in concentrated sulphuric acid (H₂SO₄) for determination of N and in di-acid mixture (nitric acid, HNO3 and perchloric acid, HClO4: 4:1) for determination of total P and K. Nitrogen content was determined by the Kjeldahl method, P in plant extracts by the vanadomolybadate yellow colour method using a UV-spectrophotometer and K in plant extracts by flame photometer. The nutrient uptake (kg/ha) in straw and grains was measured by multiplying the dry

Table 2. Initial soil properties recorded at the start of the study

Parameters	Values (0–15 cm soil layer)	Methods employed
pH (soil: water: 1:2.5)	6.77	Jackson (1973)
EC (dS/m)	0.18	Jackson (1973)
SOC (g/kg)	7.17	Walkley & Black (1934)
Total N (g/kg)	0.62	Wet digestion method
Available N (mg/kg)	146.0	Alkaline KMnO ₄ method (Subbiah & Asija, 1956)
Available P (mg/kg)	34.0	0.5 M NaHCO ₃ extractable P (Olsen et al. 1954)
Extractable K (mg/kg)	59.0	1 N NH ₄ OAc (Jackson 1973)
CEC (c mol ⁺ /kg)	20.9	Jackson (1973)
Sand (%)	54.5	Hydrometer method
Silt (%)	27.0	
Clay (%)	18.5	
Texture	Sandy clay loam	

EC, Electrical conductivity; SOC, soil organic carbon; N, nitrogen; P, phosphorus; K, potassium; CEC, cation exchange capacity; KMnO₄, potassium permanganate; NaHCO₃, sodium bicarbonate; NH₄OAc, ammonium acetate.

		Fertilizer doses (kg/ha)			
Symbol	Description of treatment	Ν	P ₂ O ₅	K ₂ 0	
T ₁	Control (no mineral fertilizers, farmyard manure or inoculation)	-	-	-	
T ₂	Farmyard manure at the rate of 5 t/ha ^a	30.5	15	35	
T ₃	Bio-inoculants (mixture of Rhizobium + Bacillus megaterium)	-	-	-	
T ₄	Farmers practice	15	30	20	
T ₅	Recommended dose of mineral fertilizers	30	40	20	
T ₆	Balance application of mineral fertilizers based on initial soil test values and targeted yield equations (STV–TYE) for achieving target yield of 1.50 t/ha lentil grain	47	50	107	
T ₇	Integrated approached based on initial STV–TYE for achieving target yield of 1.50 t/ha lentil grain + FYM @ 5 t/ha	43	45	97	
T ₈	Integrated approached based on initial STV-TYE for achieving target yield of 1.50 t/ha lentil grain + Bio-inoculants	43	45	97	
T ₉	Balance application of mineral fertilizers based on initial STV-TYE for achieving targeted yield of 2.0 t/ha lentil grain	73	72	153	

Initial soil test values: available nitrogen (N_a) = 292.2 kg N/ha, available phosphorus (P_a) = 68.8 kg P/ha, available potassium (K_a) = 118.2 kg K/ha. Targeted yield equation for grain yield of lentil (var. B 256):

FN = 4.94 T-0.09 SN-0.07 ON.

FP₂O₅ = 4.43 T-0.24 SP-0.16 OP

FK₂O = 9.03 T-0.24 SN-0.17 OK.

where FN, FP₂O₅ and FK₂O are mineral fertilizers N, phosphorus pentoxide (P₂O₅) and potassium oxide (K₂O) (kg/ha); SN, SP and SK are initial soil test values estimated before sowing of crop; ON, OP and OK are the N, P and K in their elemental form (kg/ha) mineralized from farmyard manure (FYM). To make symmetry, P₂O₅ and K₂O were converted into P and K by applying factor 2.29 and 1.20, respectively. T-targeted grain yield of lentil (t/ha). N, P₂O₅ and K₂O fertilizers were calculated on the basis of above equations for the T₅, T₆ and T₇ treatments. ^aFYM contained 0.61% N, 0.30% P and 0.70% K.

biomass (kg/ha) of straw and grain with its nutrient content (%). Crude protein (CP; dry matter basis) was calculated by applying N content in lentil grain \times 6.25, specific for pulses (AOAC 1999). The apparent nutrient recovery of fertilizer use was calculated by comparing nutrient uptake in above-ground biomass between treatments and the control, expressed in percentage as follows:

$$ANR_{NPK} = \frac{NU_{\rm T} - NU_{\rm C}}{F_{\rm N}} \times 100 \tag{1}$$

where ANR_{NPK} is the apparent nutrient recovery of N, P and K (%); NU_T and NUc is the nutrient uptake by treated and control plot

(kg/ha), respectively and $F_{\rm N}$ is the amount of fertilizer applied (kg/ha).

The serial dilution plate technique was used for enumeration of cultivable microbial counts using selective media for specific groups of micro-organisms. Nutrient agar containing 50 mg/l cyclohexamide (Parkinson *et al.* 1971), Rose Bengal chloramphenicol agar media containing 100 mg/l chloramphenicol (Martin 1950) and Ken knight agar medium were used for the counting of total cultivable bacteria (TCB), fungi (TCF) and actinomycetes (TCA), respectively. Ashby's N-free mannitol agar and Pikovskaya's agar media (Pikovskaya 1948) were used for enumeration of *Azotobacter* (AZO) and phosphate solubilizing microorganisms (PSM). The chloroform fumigation-extraction method (Vance et al. 1987) was used for measurement of MBC. Soil respiration was determined by incubating moist soils in airtight sealed flasks along with a small flask containing 10 ml 1 N sodium hydroxide (NaOH) for 24 h at 25 °C. Evolved carbon dioxide-carbon (CO₂-C) trapped in NaOH was measured by titrating with barium chloride as prescribed by Anderson (1982). Dehydrogenase activity in soil was determined by the method of Tabatabai (1982), whereas fluorescein diacetate hydrolytic activity (FDHA) was measured by quantifying fluorescein content released with the help of a spectrophotometer at 490 nm (Schnurer & Rosswall 1982). Urease activity (UA) was determined by measuring NH⁺₄ released during soil incubation with urea for 120 min at 37 °C. Acid phosphatase (ACP) and alkaline phosphatase (ALP) activities were measured based on detection of p-nitrophenol (PNP) released after incubation of soil (37 °C, 60 min) at pH 6.5 with p-nitrophenyl phosphate disodium (Tabatabai & Bremner 1969).

Soil quality index

The soil quality index (SQI) was calculated by using crop productivity as a management goal. The minimum dataset (MDS) or soil quality indicators include the most significant variables that represent soil functions associated with selected goal. After normalization of MDS indicators, all the indicators and scores were integrated into an overall index of soil quality (Andrews et al. 2002a). Soil properties that had significant treatment differences were used to determine MDS through principal component analysis (PCA) employed as a data reduction technique (Andrews et al. 2002b). Principal components having high eigenvalues (>1) and variables that explained at least 5% of variation in the data with high factor loading within 10% were assumed to be the variables that best represented attributes (Wander & Bollero 1999). Correlation analysis was performed to determine whether some of highest weighted variables were redundant and further reduction could be operated (Andrews et al. 2002a).

A non-linear scoring method was used for transforming each observation in every MDS indicator. Indicators were arranged in order depending on whether a higher value was considered beneficial for, or detrimental to, soil fertility (Andrews et al. 2002b). In the present study, all the indicators identified under MDS were considered beneficial and scored as 'more is better', as increasing levels of indicators related directly to increases in soil quality. Under the 'more is better' approach, each observed value was divided by the highest value in such a way that the highest observed value received a maximum score of 1. The PCA results were employed to weight each observed value of MDS variables after transformation. A certain amount (%) of variation in the total dataset was explained by each PC, and the percentage divided by total percentage of variance explained by all PCs (eigenvectors >1) provided the weighted factor for each variant chosen under a given PC. After that, the weighted MDS variable scores were summed up for each observation using the following equation:

$$SQI = \sum_{i=1}^{n} W_i S_i$$
 (2)

where W_i denotes the assigned weight of each indicator deduced from PCA, S_i is the score for the subscripted variable and n is the number of indicators in the final MDS.

Better soil quality or greater performance of soil functions were expected if a higher index score was recorded. The calculated

SQI values showed the contribution (%) of each final key indicator tested for their level of significance at P < 0.05.

Statistical analysis

Statistical analysis of data was carried out using SPSS version 16.0. The homogeneity of error variance was tested using Bartlett's χ^2 test. As error variance was homogeneous, pooled analysis was performed according to Cochran & Cox (1957). A two-factor analysis of variance (ANOVA) was carried out to determine the effects of fertilizer treatment, season and their interactions on grain yield (Table 4). The mean data of three crop seasons were used to analyse the treatment variance (ANOVA) by employing RBD at P < 0.05 prior to using experimental error rate and presented with standard errors of means (s.E.M. $\pm = \sqrt{MSE/r}$). The data were normalized before performing PCA to assess the impact of various fertilizer treatments on soil quality (Wold *et al.* 1987). Correlations among the variables were assessed by determining Pearson correlation coefficients and probability of P < 0.05 and P < 0.01 levels of significance.

Results

Nodulation and yield of lentil

Application of mineral fertilizers through different approaches significantly (P < 0.05) affected nodule counts as well as fresh and dry weight of nodules over the control (T₁). Application of FYM (T₂) and bio-inoculants (T₃) increased nodulation significantly (P < 0.05) over the control but were comparable with T₅ and T₉ (Fig. 1). However, the highest nodule counts (37.6 nodules/plant), fresh weight (37.8 mg/plant) and dry weight (22.3 mg/plant) of nodules were measured in T₈, closely comparable with T₇. Application of balanced fertilizers to achieve a higher grain yield (T₉) reduced nodule counts, fresh and dry weight of nodules by 26.6, 18.5 and 18.4%, respectively, over T₈.

Grain and straw yield were significantly (P < 0.05) enhanced in the fertilizer treatments compared with the unfertilized control. Application of balanced fertilizers calculated to achieve specific target yields of 1.50 and 2.0 t/ha returned only 90.7% (1.36 t/ ha) and 82.5% (1.65 t/ha), respectively, of the target yield (Table 5). However, integration of FYM with balanced fertilizers (T₇) achieved the specific target yield of 1.50 t/ha, and that of balanced fertilizers with bio-inoculants (T₈) increased yield to 109.3% (1.64 t/ha) of the target. Also, the T₈ treatment decreased N, P₂O₅ and K₂O requirement by 30, 27 and 56 kg/ha, respectively, over balanced fertilizers alone (T₉) without reduction of grain yield. Application of FYM (T₂) and bio-inoculants (T₃) resulted in grain yields comparable with T₅ and T₆. Treatments T₈ and T₉ recorded significantly (P < 0.05) higher grain and

 Table
 4.
 ANOVA tables showing interaction effects of year, treatments, replication and their interactions

Source	DF	MS	F-value	<i>P</i> -value
Year	2	0.534	20.08	<0.001
Treatment	8	0.453	18.93	<0.001
Replication	2	0.001	0.063	NS
Year × treatment	16	0.024	2.725	<0.01

DF, degrees of freedom; MS, mean squares.



T₁: Control (no mineral fertilizers, farmyard manure (FYM) or bio-inoculants), T₂: FYM @ 5 t/ha, T₃: Bio-inoculants (mixture of *Rhizobium* + *Bacillus megaterium*), T₄: Farmers practice, T₅: Recommended dose of mineral fertilizers, T₆: Precise application of mineral fertilizers based on initial soil test values and targeted yield equations(STV-TYE) for achieving target yield of 1.5 t/ha lentil grain, T₇: Integrated approached based on initial STV-TYE for achieving target yield of 1.5 t/ha lentil grain + FYM @ 5 t/ ha, T₈: Integrated approached based on initial STV-TYE for achieving target yield of 2.0 t/ha lentil grain + Bio-inoculants and T₉: Precise application of mineral fertilizers based on initial STV-TYE for achieving target yield of 2.0 t/ha lentil grain. ± represents standard errors of mean (s.E.M.±).

 68 ± 2.6

77 ± 1.9

76 ± 2.2

6.01

 7.3 ± 0.31

 8.6 ± 0.20

 8.2 ± 0.16

0.81

 36 ± 1.0

 41 ± 0.8

 39 ± 0.2

2.92

 262 ± 2.5

 268 ± 3.4

 265 ± 3.1

7.29

63.6

86.2

50.4

17.0

23.1

14.2

16.7

23.6

13.9

 1.84 ± 0.037

 2.14 ± 0.014

 2.03 ± 0.056

0.18

 1.56 ± 0.182

 1.81 ± 0.091

 1.65 ± 0.094

0.16

 1.57 ± 0.067

 1.62 ± 0.081

 1.79 ± 0.044

0.14

 1.35 ± 0.074

 1.50 ± 0.096

 1.52 ± 0.081

0.16

 1.50 ± 0.035

 1.64 ± 0.042

 1.65 ± 0.035

0.11

 T_7

 T_8

T₉

CD (P < 0.05)

0.05 (Table 5). Nutrient uptake (N, P and K) varied significantly (P < 0.05) between the treatments. The highest N (76.6 kg/ha), P (8.60 kg/ha) and K (40.7 kg/ha) uptake was recorded in T₈ and lowest in T₁ treatment (41.6, 3.74 and 23.1 kg/ha). The T₈ treatment improved N, P and K uptake by 9.30, 1.29 and 5.10 kg, respectively, over T₇. Similarly, crude protein was also highest in T₈ (268 g/kg), followed by T₉ (265 g/kg) and T₇ (262 g/kg).

Nutrient recovery exhibited a direct relationship with crop yield and nutrient utilization. Nutrient recovery varied with different fertilizer doses applied in different treatments (Table 5). The highest apparent N and P recovery were recorded in T₈ followed by T₇ whereas K recovery was highest in T₄ followed by T₈. Application of bio-inoculants with balanced fertilizers (T₈) increased N and P use efficiency by 39.3 and 60.4%, respectively, over the T₅ treatment, and by 35.5 and 35.9% over the T₇ treatment. Application of higher doses of mineral fertilizers for a higher target yield (T₉) also achieved 50.4, 14.2 and 13.9% N, P and K recovery, respectively.

Microbial biomass carbon, microbial counts and soil enzymes

Application of mineral fertilizers significantly altered TCB, TCA, TCF, AZO and PSM counts over the control (P < 0.05; Table 6). Treatment T₇ (FYM with balanced fertilizers) showed significantly (P < 0.05) higher TCB counts (9.67×10^5 cfu/g soil), TCF (4.01×10^4 cfu/g soil) and AZO counts (24.3×10^4 cfu/g soil) over rest of the treatments, whereas TCA count (5.52×10^5 cfu/g soil) was highest in T₉ and PSM counts (16.4×10^4 cfu/g soil) was highest in T₈. Unbalanced application of mineral fertilizers (T₄ and T₅) could not improve TCF and AZO counts compared to T₁. Application of higher doses of mineral fertilizers for a higher target yield (T₉) decreased TCB, TCF, AZO and PSM significantly (P < 0.05) over T₇ and T₈. *Azotobacter* and PSM counts were highly susceptible to large doses of N and P fertilizer compared with TCB, TCA and TCF. The TCB/TCA ratio was highest (24.1) in T₈ whereas the TCA/TCF ratio was highest (16.4) in T₉.

Soil MBC varied significantly (P < 0.05) between the treatments and was highest in T_7 (379.2 µg c/g) and lowest in the control (131.4 μ g c/g). Treatment T₇ increased MBC by 35.4, 25.9 and 80.5% over that of T₉, T₈ and T₆, respectively (Table 7). As with MBC, the highest (9.14 μ g CO₂/g soil/h) and lowest (5.82 μ g CO₂/ g soil/h) soil respiration rates were found in T_7 and T_1 , respectively. Application of fertilizers through different approaches significantly (P < 0.05) affected DHA, FDHA, urease, ACP and ALP activities over the control (T_1) . The highest DHA (2.01 µg triphenyl formazan (TPF)/g/h), FDHA (106.1 µg fluorescein/g/ h), urease (58.7 μ g NH₄⁺/g/h) and ACP (351.1 μ g PNP/g/h) were measured in the T_7 treatment, while the highest ALP (240.6 µg PNP/g/h) was recorded in T₈. The lowest enzymatic activities were recorded in T₁. Treatment T₉ reduced enzymatic activities in soil but levels were still generally higher than T₄ and T₅ in most cases. Treatment T7 increased DHA, FDHA and urease activities by 10.4, 7.44 and 14.5%, respectively, over T₈. However, T₈ showed 12.5% higher ALP over T₇.

Nutrient status

Significant differences (P < 0.05) were seen in soil organic carbon (SOC), available nitrogen (N_a), phosphorus (P_a) and potash (K_a) over the control (T₁) due to fertilization through different approaches (Table 8). Similarly, all treatments apparently increased N_a, P_a and K_a over initial levels, with the exception of T₁. Balanced fertilization (T₆, T₇, T₈ and T₉) had pronounced effects on improving SOC, N_a, P_a and K_a over unbalanced fertilization (T₄ and T₅). The SOC contents ranged from 7.23 g/kg in T₁ to 7.97 g/kg in T₇, significantly (P < 0.05) higher than the rest of the treatments. Treatments T₇, T₈ and T₉ increased SOC contents by 11.2, 8.50 and 9.21%, respectively, over the initial SOC level of 7.17 g/kg. The highest N_a (164 mg/kg) and K_a (71 mg/kg) were recorded in T₉, improvements of 18 and 12 mg/kg over initial N_a (146 mg/kg) and K_a (59 mg/kg) status, respectively. Available *P* was significantly (P < 0.05) higher in T₈ than in the

Table 6. Effect of different treatments on microbial activity of lentil were presented with standard error of mean (s.E.M.±)

Treatment	TCB (× 10 ⁵ cfu/g)	TCA (× 10 ⁵ cfu/g)	TCF (×10 ⁴ cfu/g)	<i>Azotobacter</i> (×10 ⁴ cfu/g)	PSM (×10 ⁴ cfu/g)	TCB/TCF ratio	TCA/TCF ratio
T ₁	5.0 ± 0.15	4.0 ± 0.09	2.8 ± 0.11	17.6 ± 0.81	12.7 ± 0.50	17.7	14.1
T ₂	7.3 ± 0.38	4.8 ± 0.30	3.5 ± 0.10	21.1 ± 0.87	14.4 ± 0.68	21.1	14.0
T ₃	6.6 ± 0.55	4.6 ± 0.24	2.8 ± 0.14	18.9 ± 0.96	13.6 ± 0.72	23.3	16.2
T ₄	5.4 ± 0.16	4.5 ± 0.10	3.0 ± 0.17	17.7 ± 0.99	13.6 ± 0.53	18.3	15.2
T ₅	6.0 ± 0.18	4.8 ± 0.17	3.1 ± 0.12	18.8 ± 0.87	14.6 ± 0.57	19.5	15.6
T ₆	6.7 ± 0.20	4.6 ± 0.11	3.4 ± 0.14	20.3 ± 0.94	14.1 ± 0.55	19.7	13.3
T ₇	9.7 ± 0.29	5.5 ± 0.13	4.0 ± 0.16	24 ± 1.4	16.2 ± 0.63	24.1	13.6
T ₈	8.3 ± 0.25	4.6 ± 0.16	3.6 ± 0.14	24 ± 1.2	16.4 ± 0.64	23.3	12.7
T ₉	6.0 ± 0.18	5.5 ± 0.13	3.4 ± 0.19	17.9 ± 0.97	13.3 ± 0.52	17.9	16.4
CD (P < 0.05)	0.86	0.41	0.39	2.53	1.83	-	-

TCB, total culturable bacteria; TCA, total culturable actinomycetes, TCF, total culturable fungi; PSM, total culturable phosphate solubilizing microorganisms, TCB/TCF, ratio of total culturable bacteria and total culturable fungi, TCB/TCA, ratio of total culturable bacteria and total culturable actinomycetes; cfu/g, colony forming unit per gram soil.

 T_1 : Control (no mineral fertilizers, farmyard manure(FYM) or bio-inoculants), T_2 : FYM @ 5 t/ha, T_3 : Bio-inoculants, T_4 : Farmers practice, T_5 : Recommended dose of mineral fertilizers, T_6 : Precise application of mineral fertilizers based on initial soil test values and targeted yield equations for achieving target yield of 1.5 t/ha lentil grain, T_7 : Integrated approached based on initial soil test values and targeted yield equations for achieving target yield of 1.5 t/ha lentil grain, T_7 : Integrated approached based on initial soil test values and targeted yield equations for achieving target yield of 1.5 t/ha lentil grain + FYM @ 5 t/ha, T_8 : Integrated approached based on initial soil test values and targeted yield equations for achieving target yield of 1.5 t/ha lentil grain + Bio-inoculants and T_9 : Precise application of mineral fertilizers based on initial soil test values and targeted yield equations for achieving target yield of 1.5 t/ha lentil grain + Bio-inoculants and T_9 : Precise application of mineral fertilizers based on initial soil test values and targeted yield equations for achieving target yield of 2.0 t/ha lentil grain. \pm represents standard errors of mean (s.e.m. \pm).

Treatment	Microbial biomass carbon (μg c/g soil)	Soil respiration (µg CO ₂ /g soil/h)	Dehydrogenase (µg TPF/g/h)	Fluorescein diacetate hydrolytic activity (µg fluorescein/g soil/h)	Urease (µg NH₄+/g soil/h)	Acid phosphatase (μg PNP/g soil/h)	Alkaline phosphatase (μg PNP/g soil/h)
T ₁	131 ± 4.3	5.7 ± 0.44	0.9 ± 0.03	83 ± 2.4	41 ± 1.2	291 ± 5.5	146 ± 4.1
T ₂	239 ± 5.1	6.3 ± 0.41	1.4 ± 0.15	94 ± 3.1	43 ± 1.9	325 ± 6.0	190 ± 5.61
T ₃	217 ± 4.6	5.9 ± 0.37	1.2 ± 0.10	87 ± 2.6	41 ± 1.6	304 ± 4.1	171 ± 4.4
T ₄	141 ± 4.6	5.8 ± 0.22	1.2 ± 0.03	87 ± 2.5	39 ± 1.1	301 ± 5.8	169 ± 4.7
T ₅	197 ± 6.4	5.8 ± 0.39	1.4 ± 0.05	93 ± 2.7	40 ± 1.2	309 ± 5.4	165 ± 4.6
T ₆	210 ± 6.9	5.9 ± 0.51	1.4 ± 0.04	96 ± 2.8	46±1.3	318 ± 4.5	178 ± 5.0
T ₇	379 ± 6.6	9.1 ± 0.34	2.0 ± 0.06	106 ± 3.1	59 ± 1.7	351 ± 3.6	211 ± 5.9
T ₈	301 ± 5.6	8.7 ± 0.40	1.8 ± 0.05	98 ± 2.9	50 ± 1.5	346 ± 5.8	241 ± 6.7
T ₉	280 ± 6.5	7.7 ± 0.29	1.7 ± 0.05	97 ± 2.8	41 ± 1.2	326 ± 4.1	207 ± 5.8
CD (<i>P</i> < 0.05)	16.2	0.62	0.13	4.79	4.22	15.7	13.8

Table 7. Effect of different treatments on microbial biomass carbon and enzymatic activity

CO₂, carbon dioxide; TPF, triphenyl formazan; NH⁺₄, ammonium; PNP, p-nitrophenol.

 T_1 : Control (no mineral fertilizers, farmyard manure(FYM) or bio-inoculants), T_2 : Farmyard manure @ 5 t/ha, T_3 : Bio-inoculants (mixture of *Rhizobium* + *Bacillus megaterium*), T_4 : Farmers practice, T_5 : Recommended dose of mineral fertilizers, T_6 : Precise application of mineral fertilizers based on initial soil test values and targeted yield equations (STV–TYE) for achieving target yield of 1.5 t/ha lentil grain, T_7 : Integrated approached based on initial STV–TYE for achieving target yield of 1.5 t/ha lentil grain, T_7 : Integrated approached based on initial STV–TYE for achieving target yield of 1.5 t/ha lentil grain + FYM @ 5 t/ha, T_8 : Integrated approached based on initial STV–TYE for achieving target yield of 2.0 t/ha lentil grain + Bio-inoculants and T_9 : Precise application of mineral fertilizers based on initial STV–TYE for achieving target yield of 2.0 t/ha lentil grain + Bio-inoculants and T_9 : Precise application of mineral fertilizers based on initial STV–TYE for achieving target yield of 2.0 t/ha lentil grain + Bio-inoculants and T_9 : Precise application of mineral fertilizers based on initial STV–TYE for achieving targeted yield of 2.0 t/ha lentil grain + Bio-inoculants and T_9 : Precise application of mineral fertilizers based on initial STV–TYE for achieving targeted yield of 2.0 t/ha lentil grain + Bio-inoculants and T_9 : Precise application of mineral fertilizers based on initial STV–TYE for achieving targeted yield of 2.0 t/ha lentil grain + Bio-inoculants and T_9 : Precise application of mineral fertilizers based on initial STV–TYE for achieving target yield of 2.0 t/ha lentil grain + Bio-inoculants and T_9 : Precise application of mineral fertilizers based on initial STV–TYE for achieving targeted yield of 2.0 t/ha lentil grain + Bio-inoculants and T_9 : Precise application of mineral fertilizers based on initial STV–TYE for achieving targeted yield of 2.0 t/ha lentil grain + Bio-inoculants and T_9 : Precise application of mineral fertilizers based on i

other treatments; it was 8.80 mg/kg higher than initial P_a levels (34 mg/kg). After completing three cycles of lentil cultivation, SOC had improved slightly in all treatments; however, N_a was greatly reduced in T_2 (-3.0 mg/kg) compared with initial levels, followed by T_3 (-2.0 mg/kg). However, P_a (-1.20 mg/kg) and K_a (-3.0 mg/kg) were reduced slightly in the control plot only.

Correlation among the variables

Correlation matrix amongst various variable is given in Table 9. Lentil grain yield had strong significant positive correlation with straw yield ($R^2 = 0.937$, P < 0.001), crude protein ($R^2 = 0.944$, P < 0.001), P_a ($R^2 = 0.895$, P < 0.001), K_a ($R^2 = 0.832$, P < 0.005), SR ($R^2 = 0.872$, P < 0.01), ALP ($R^2 = 0.876$, P < 0.002), SOC ($R^2 = 0.876$), R = 0.002), SOC ($R^2 = 0.002$), SOC ($R^2 = 0.0$

Table 8. Effect of different treatments on soil organic carbon (SOC) and nutrient status (nitrogen (N), phosphorus (P) and potassium (K)) after harvest of third cycle of lentil

					Nutrient b	alance over initia cycle of len	l levels after com til cultivation	pleting third
Treatment	SOC (g/kg)	N (mg/kg)	P (mg/kg)	K (mg/kg)	SOC (g/kg)	N (mg/kg)	P (mg/kg)	K (mg/kg)
Initial	7.17	146.0	34.0	59.0	-	-	-	-
T ₁	7.2 ± 0.11	146 ± 3.8	32.8 ± 0.56	56 ± 2.8	0.06	0.00	-1.20	-3.00
T ₂	7.7 ± 0.22	143 ± 4.4	36.2 ± 0.58	62 ± 3.1	0.51	-3.10	2.20	2.80
T ₃	7.4 ± 0.15	144 ± 3.7	37.9 ± 0.62	63 ± 3.2	0.20	-2.10	3.90	4.10
T ₄	7.3 ± 0.16	151 ± 3.9	38.3 ± 0.70	64 ± 3.0	0.13	5.00	4.30	5.00
T ₅	7.4 ± 0.08	153 ± 2.7	37.0 ± 0.67	66 ± 2.5	0.23	7.00	3.00	7.00
T ₆	7.5 ± 0.07	155 ± 4.2	36.4 ± 0.64	64 ± 2.6	0.30	9.00	2.40	5.00
T ₇	8.0 ± 0.13	162 ± 4.5	40.2 ± 0.68	67 ± 3.6	0.80	16.00	6.20	8.00
T ₈	7.8 ± 0.09	158 ± 4.1	42.8 ± 0.74	64 ± 2.7	0.61	12.00	8.80	5.00
T ₉	7.8 ± 0.11	164 ± 3.5	40.4 ± 0.69	71 ± 2.9	0.66	18.00	6.40	12.00
CD (<i>P</i> < 0.05)	0.29	12.2	3.05	6.39	-	-	-	-

 T_1 : Control (no mineral fertilizers, farmyard manure(FYM) or bio-inoculants), T_2 : Farmyard manure @ 5 t/ha, T_3 : Bio-inoculants (mixture of *Rhizobium* + *Bacillus megaterium*), T_4 : Farmers practice, T_5 : Recommended dose of mineral fertilizers, T_6 : Precise application of mineral fertilizers based on initial soil test values and targeted yield equations (STV–TYE) for achieving target yield of 1.5 t/ha lentil grain, T_7 : Integrated approached based on initial STV–TYE for achieving target yield of 1.5 t/ha lentil grain + FYM @ 5 t/ha, T_8 : Integrated approached based on initial STV–TYE for achieving target yield of 1.5 t/ha lentil grain + FYM @ 5 t/ha, T_8 : Integrated approached based on initial STV–TYE for achieving target yield of 2.0 t/ha lentil grain + Bio-inoculants and T_9 : Precise application of mineral fertilizers based on initial STV–TYE for achieving targeted yield of 2.0 t/ha lentil grain, ± represents standard errors of mean (s.t.m.±).

0.840, P < 0.005), SR ($R^2 = 0.799$, P < 0.01) than TCA ($R^2 = 0.704$, P< 0.034), TCF ($R^2 = 0.671$, P < 0.048), FDHA ($R^2 = 0.727$, P < 0.027) and N₂ ($R^2 = 0.673$, P < 0.047). However, CP had strong positive relationship with P_a ($R^2 = 0.936$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$, P < 0.001), DHA ($R^2 = 0.837$), $R^2 = 0.837$, $R^2 = 0.001$), DHA ($R^2 = 0.837$, $R^2 = 0.001$), DHA ($R^2 = 0.837$), $R^2 = 0.001$), DHA ($R^2 = 0.837$, $R^2 = 0.001$), DHA ($R^2 = 0.837$), $R^2 = 0.001$), DHA ($R^2 = 0.837$, $R^2 = 0.001$), DHA ($R^2 = 0.837$), $R^2 = 0.001$), DHA ($R^2 = 0.837$), $R^2 = 0.001$), DHA ($R^2 = 0.837$), $R^2 = 0.001$), $R^2 = 0.001$, $R^2 = 0.001$), $R^2 = 0.001$, $R^2 = 0.001$, $R^2 = 0.001$), $R^2 = 0.001$, $R^2 = 0.001$), $R^2 = 0.001$, $R^2 = 0$ 0.006), K_a ($R^2 = 0.822$, P < 0.006), ALP ($R^2 = 0.813$, P < 0.008) and SOC ($R^2 = 0.804$, P < 0.009). The total culturable bacteria counts, the backbone of biochemical reactions in soil, correlated strongly with urease $(R^2 = 0.934, P < 0.001)$, AZO $(R^2 = 0.930, P < 0.001)$, ACP ($R^2 = 0.917$, P < 0.001), MBC ($R^2 = 0.907$, P < 0.001), PSM $(R^2 = 0.897, P < 0.001), \text{ TCF} (R^2 = 0.886, P < 0.001), \text{ DHA} (R^2 = 0.001), \text{ DHA$ 0.849, P < 0.004), SR ($R^2 = 0.816$, P < 0.007), FDHA ($R^2 = 0.819$, P< 0.007) and SOC ($R^2 = 0.806$, P < 0.009). The TCF counts exhibited significant strong positive relationship with ACP ($R^2 = 0.953$, P <0.001), FDHA ($R^2 = 0.935$, P < 0.001), DHA ($R^2 = 0.930$, P < 0.001), DHA ($R^2 = 0.930$, P < 0.001), P($R^2 = 0.930$, P < 0.001), DHA ($R^2 = 0.930$, P < 0.001), DHA ($R^2 = 0.930$, P < 0.001), DHA ($R^2 = 0.930$, P < 0.001), DHA ($R^2 = 0.930$, P < 0.001), DHA ($R^2 = 0.930$, P < 0.001), DHA ($R^2 = 0.930$, P < 0.001), DHA ($R^2 = 0.930$, P < 0.001), DHA ($R^2 = 0.930$, P < 0.001), DHA ($R^2 = 0.930$, P < 0.001), DHA ($R^2 = 0.930$, P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), R = 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), P < 0.001), DHA ($R^2 = 0.930$), R = 0.001), DHA ($R^2 = 0.930$), R = 0.001), DHA ($R^2 = 0.930$), R = 0.001), DHA ($R^2 = 0.930$), R = 0.001), DHA ($R^2 = 0.930$), R = 0.001), DHA (R = 0.001), DHA ($R^2 = 0.930$), R = 0.001), DHA (R = 0.000.001), MBC ($R^2 = 0.904$, P < 0.001), SOC ($R^2 = 0.901$, P < 0.001), UA ($R^2 = 0.874$, P < 0.002) and SR ($R^2 = 0.831$, P < 0.005). PSM had strong positive correlation with ACP ($R^2 = 0.867$, P < 0.002), UA $(R^2 = 0.827, P < 0.003)$ and DHA $(R^2 = 0.814, P < 0.008)$. MBC also had significant correlation with ACP ($R^2 = 0.945$, P <0.001), SOC ($R^2 = 0.923$, P < 0.001), DHA ($R^2 = 0.915$, P < 0.001), SR ($R^2 = 0.865$, P < 0.003), FDHA ($R^2 = 0.831$, P < 0.005) and ALP $(R^2 = 0.824, P < 0.006)$. SOC showed positive correlation with K_a $(R^2 = 0.755, P < 0.019)$ and P_a $(R^2 = 0.751, P < 0.020)$.

Soil quality index

The impact of mineral fertilizer application through different approaches was assessed by computing a SQI. Considering the 16 microbial, enzymatic and chemical attributes of soil, three PCs were identified with eigenvalues >1 (Table 10). The total variance explained by these PCs was 86%. PC1, PC2 and PC3 contributed 56.9, 18.7 and 10.3% of the variance. The first PC had five highly weighted variables (SOC, TCF, SR, TCB and MBC) within 10% of the highest factor loading, which explained 56.9% of variation (Table 9). Among these five variables, SOC, TCF and MBC were identified for the MDS. In PC2, ACP was selected whereas in PC3 only P_a was retained in the MDS. Therefore, the final MDS included SOC, TCF, MBC, ACP and Pa. After deciding the shape of the anticipated response (more is better), the threshold values were assigned taking into account site-specific characteristics and management goals (Table 11). All MDS indicators scored in the range of 0.1-1.0. Weights were assigned to MDS indicators using the PCA outcomes (Table 11), which were equal to percentage of variance explained by the PC. The SQI was measured by employing data of estimated factors to soil quality indicators using integrated quality index equations (Eqn (2)).

$$SQI = \frac{0.19S_{SOC} + 0.19S_{TCF} + 0.19S_{MBC} + 0.19S_{ACP} + 0.19S_{P_a}}{0.86}$$
(3)

$$SQI = 0.22S_{SOC} + 0.22S_{TCF} + 0.22S_{MBC} + 0.22S_{ACP} + 0.12S_{P_a}$$

where *S* is the score for the subscribed variable and coefficient are the weighting factors.

Among the treatments, SQI ranged from 0.66 in the control treatment (T₁) to 0.92 in T₇. Treatment T₇ measured the highest SQI (0.92, P < 0.05) followed by T₈ (0.85) and T₉ (0.82). The lowest SQI (0.66, P < 0.05) was recorded in T₁. The average contributions of each MDS indicator were 20.7, 19.5, 18.3, 10.2 and 9.31%

towards SQI through SOC, ACP, TCF, P_a and MBC, respectively (Fig. 2). Correlation analysis results showed that SQI had significant correlation with CP ($y = 7.42 \times +20.05$; $R^2 = 0.735$) (Fig. 3 (*a*)) and grain yield ($y = 2.395 \times -0.518$; $R^2 = 0.701$) (Fig. 3(*b*)).

Discussion

Nodulation, yield, nutrient acquisition and nutrient use efficiency

In the present study, significant variations were observed in nodule number, and fresh and dry weight of nodules in response to fertilizer regime. Integration of bio-inoculants and FYM with balanced fertilization showed higher nodulation than unbalanced fertilization. This might be due to balanced fertilization providing adequate nutrient supply at critical stages of plant development, promoting proliferation and development of the root system. The negative impact of low mineral N on nodulation under unbalanced fertilizer treatments could be related to nutrient stress for the host plant and the high energetic cost of the process (Robertson & Groffman 2015). Inoculation of *B. megaterium* as a P-solubilizer enhanced nodulation by promoting early root and lateral fibre formation resulting in improved P uptake, as clearly depicted in the current results. Shahzad et al. (2008) suggested that P played a vital role in energy transformation and translocation, which helps nodule formation and their activation. Further, co-inoculation was more effective than FYM integrated with balanced fertilizers, owing to the synergistic effect of co-inoculated Rhizobium and B. megaterium. Together, these enhance nodulation by reducing endogenous ethylene production by plants, producing antibiotics against pathogenic organisms and siderophores that chelate insoluble cations and colonizing root surfaces, thereby out-competing pathogens (Contesto et al. 2008). Application of FYM ensured a balanced supply of macro- and micro-nutrients, besides improving the physical, chemical and microbiological properties of soil, and is also used as a carbon substrate by native as well as inoculated bio-inoculants, resulting in better nodulation. Moreover, the inoculated Rhizobium strain is more efficient than native populations, again resulting in better nodulation (Zafar et al. 2012; Iqbal et al. 2016). Robertson & Groffman (2015) revealed that the higher mineral N concentration in the soil solution common in high fertilizer regimes would limit nodulation and biological N fixation. The current results also showed that higher doses of mineral fertilizers, applied for higher yield targets (T_9) , reduced nodulation notably compared with that of balanced fertilizer treatments due to increasing nitrate (NO_3^-) levels, leading to greater NO_3^- uptake. The decrease in nodulation in most legume specie sowing to substantial nitrous oxide (NO) uptake by the legumes is usually associated with decreasing nodule weight. This decrease was reported by Andrews et al. (1992) to be greater in lentil (92%) than field bean (60%).

Grain and straw yield of lentil in the present study were altered significantly in the fertilizer treatments compared with the unfertilized control. Similar results have been reported in mung bean by Choudhary *et al.* (2011). Several researchers have reported that application of conventionally recommended doses of inorganic fertilizers had poor impact on yield, nutrient uptake, crop quality and fertility status (Ramamurthy *et al.* 2009; Singh *et al.* 2015). This failure could be due to unbalanced application of nutrients through old and out-dated general recommendations of fertilizers under intensive cropping systems. Mono-cropped lentil has been reported to remove 52–67 kg N/ha from the soil, which cannot be met through unbalanced fertilization. Also, lentil GY

SY

																		-	-
GY	1																		
SY	0.937**	1																	
СР	0.944**	0.949**	1																
тсв	0.576	0.452	0.614	1															
TCA	0.704*	0.574	0.679*	0.523	1														
TCF	0.671*	0.479	0.584	0.886**	0.673*	1													
AZO	0.480	0.451	0.526	0.930**	0.374	0.797*	1												
PSM	0.588	0.592	0.646	0.897**	0.365	0.793*	0.960**	1											
MBC	0.711*	0.55	0.684*	0.907**	0.767*	0.904**	0.813**	0.751*	1										
SR	0.799**	0.690*	0.727*	0.816**	0.631	0.831**	0.773*	0.766*	0.865**	1									
DHA	0.872**	0.755*	0.827**	0.849**	0.782*	0.930**	0.774*	0.814**	0.915**	0.899**	1								
FDHA	0.727*	0.540	0.665	0.819**	0.711*	0.935**	0.665	0.716*	0.831**	0.828**	0.907**	1							
UA	0.501	0.342	0.480	0.934**	0.424	0.874**	0.858**	0.827**	0.800**	0.823**	0.790*	0.815**	1						
ACP	0.793*	0.660	0.741*	0.917**	0.662	0.953**	0.860**	0.867**	0.945**	0.915**	0.967**	0.901**	0.848**	1					
ALP	0.876**	0.794*	0.813**	0.757*	0.551	0.782*	0.714*	0.765*	0.824**	0.891**	0.876**	0.793*	0.656	0.918**	1				
SOC	0.840**	0.685*	0.804**	0.806**	0.839**	0.901**	0.669*	0.701*	0.923**	0.795*	0.953**	0.904**	0.682*	0.928**	0.852**	1			
Na	0.673*	0.651	0.549	0.217	0.531	0.468	0.212	0.337	0.308	0.589	0.609	0.532	0.367	0.452	0.429	0.442	1		
Pa	0.895**	0.924**	0.936**	0.624	0.611	0.573	0.590	0.688*	0.686*	0.824**	0.801**	0.678*	0.506	0.761*	0.880**	0.751*	0.534	1	
Ka	0.832**	0.792*	0.822**	0.313	0.877**	0.490	0.183	0.296	0.542	0.513	0.711*	0.601	0.203	0.539	0.560	0.755*	0.660	0.712*	1

MBC

SR

DHA

FDHA

UA

ACP

ALP

SOC

N₂

Р

Grain yield (GY), straw yield (SY), crude protein (CP), total cultivable bacteria (TCB), total culturable actinomycetes (TCA), total culturable fungi (TCF), Azotobacter (AZO), phosphate solubilizing microorganism (PSM), microbial biomass carbon (MBC), soil respiration (SR) and soil enzyme activities viz., dehydrogenase (DHA), fluorescence diacetatehydrolytic (FDHA), urease (UA), acid phosphatase (ACP), alkaline phosphatase (ALP), Soil organic carbon (SOC), available nitrogen (Na), available phosphorus (P_a), available potassium (K_a).

n=9, **Pearson correlation is significant at P < 0.01, *Correlation is significant at P < 0.05.

тсв

CP

TCA

TCF

AZO

PSM

 $\ensuremath{\textbf{Table 10}}$. Principal component analysis of soil quality indicator for the first four PCs

Principal components	PC1	PC2	PC3	PC4
Eigenvalue	9.11	2.99	1.65	0.93
Variability (%)	56.9	18.7	10.3	5.80
Cumulative %	56.9	75.6	85.9	91.7
Eigenvectors				
ТСВ	0.294	-0.209	-0.152	0.074
TCA	0.264	0.317	0.020	-0.133
TCF	0.302	0.006	-0.176	0.285
AZO	0.265	-0.311	-0.109	0.195
PSM	0.265	-0.311	-0.109	0.195
MBC	<u>0.283</u>	-0.211	-0.228	-0.166
SR	0.295	-0.145	0.138	-0.209
DHA	-0.126	-0.271	0.470	0.359
FDHA	0.107	0.340	-0.053	0.721
UA	0.252	0.215	-0.279	-0.044
ACP	0.234	0.358	-0.087	-0.193
ALP	0.222	-0.291	0.179	-0.045
SOC	0.316	0.011	0.058	-0.046
N _a	0.227	0.186	0.382	0.150
Pa	0.237	-0.064	0.479	-0.196
K _a	0.206	0.332	0.351	-0.039

Variables viz., Total culturable bacteria (TCB), total culturable actinomycetes (TCA), total culturable fungi (TCF), *Azotobacter* (AZO), total culturable phosphate solubilizing microorganisms (PSM), microbial biomass carbon (MBC), soil respiration (SR), dehydrogenase activity (DHA), fluorescence diacetate hydrolytic activity (FDHA), urease activity (UA) acid phosphatase activity (ACP), alkaline phosphatase activity (ALP), soil organic carbon (SOC), available N (N_a), available P (P_a) and available K (K_a). Eigenvalues in bold correspond to the PCs examined for the index.

Component loadings in bold are considered highly weighted, Bold-underlined component loadings correspond to indicators included in MDS.

crops do not acquire much mineralizable-N from soil layers deeper than 0.6 m due to their shallow root system and higher risk of N losses through de-nitrification, which minimize N availability and cause adverse effects on crop yield (Kutschera 1960). However, in the present study the highest yield (straw and grain) was observed in balanced treatments (T_6 , T_7 , T_8 and T_9)

 $\label{eq:table_$

Indicators	SF	La	Ва	Ua	Weighting
SOC (g/kg)	More is better	2.0	4.0	8.0	0.22
TCF (×10 ⁴ cfu/g soil)	More is better	0	2.0	4.0	0.22
MBC (µg c/g soil)	More is better	75	225	550	0.22
ACP (µg PNP/g/h)	More is better	90	180	360	0.22
P _a (mg/kg)	More is better	11.2	27.9	44.6	0.12

SOC, soil organic carbon; TCF, total culturable fungi; MBC, microbial biomass carbon; ACP, acid phosphatase; P_a, available phosphorus; La, lower threshold, at which or below score is 0; Ba, baseline, at which score is 0.5; Ua, upper threshold, at which or above score is 1.

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due to improvements in nodulation, dry matter yield and nutrient uptake. The lower target yield (1.5 t/ha) was achieved by the application of balanced fertilizers because the gap between applied nutrients and their availability was smaller. However, the higher target yield (2.0 t/ha) could not be achieved, which might be attributed to a mismatch between nutrient availability and removal. Adequate supply of nutrients to the crop under balanced fertilizer treatments, based on soil tests and target yields, significantly enhanced plant growth, nodulation and dry matter, which are directly proportional to the grain yield of lentil (Lavanya & Toms 2009; Kundu et al. 2013). Integration of FYM and bio-inoculants with balanced fertilizer treatment improved grain yield by 14.0 and 21.3% over balanced fertilizers alone, in spite of lower doses of mineral fertilizers being applied. This could be attributed to increased MBC, soil enzymes and nutrient status, promoting nutrient supply to the plant. The current results clearly revealed that grain yields had a positive relationship with straw yield, crude protein, TCA, TCF, MBC, SR, DHA, FDHA, ACP, ALP, SOC, Na, Pa and Ka. These results corroborate those of Elfstrand et al. (2007). Rhizobium-Bacillus association promoted nodulation, dry matter and nutrient concentrations to produce higher levels of yield despite some extent of subsidy from mineral fertilizers. Beside N₂ fixation and P solubilizing efficiency, these bacteria produce various biologically active compounds, viz. auxins, enzymes, phytohormones and siderophores, which play a crucial role for yield improvement (Egamberdiyeva 2005).

Nutrient acquisition varied significantly due to application of fertilizer through different approaches. The highest nutrient removal was found in the bio-inoculant plus balanced fertilizer treatment, despite application of 41.1, 37.5 and 36.6% lower doses of N, P₂O₅ and K₂O than in the higher fertilized treatment. This might be attributed to increased nutrient content in plants and grains resulting in higher biomass yield owing to balance supply of nutrient at active stages of crop growth. The effect of bio-inoculation plus balanced fertilizers on nutrient acquisition was more pronounced than FYM application because co-inoculation had a direct impact on the nutrient supply system. Although FYM application increased SOC, SR, microbial counts and soil enzymes, thereby improving the physical condition of soil (Stark et al. 2007), this had only indirect effects on nutrient uptake. Rhizobium is a symbiotic N fixer which has high potential to fix N₂ into NH₃, thereby improving N concentration in plants and grains. Similar results were also reported by Basu & Bansyopadhyay (1990). Bacillus megaterium solubilizes insoluble P into available P by production of organic acids, which enhances P concentration in the rhizosphere of plants. Rhizobium and Bacillus, having synergistic effects, played a vital role in promoting their activity and nutrient content in plant biomass (Gull et al. 2004). Crude protein was found to be highest in balanced fertilizer treatments. It seems that the higher nodule dry weight and shoot N contents supplied sufficient N for synthesis of amino acids, which translated into higher seed protein content. Moreover, a balanced nutrient supply system in the soil and symbiotic N fixation carried out by Rhizobium may enhance N availability in soil, resulting in higher N content in grains (Hoque & Haq 1994). The increase in seed protein percentage can be linked with better N supply to the plant.

Apparent nutrient recovery is an indicator for assessing the impacts of different fertilizer treatments. The maximal N and P recovery in an integrated, balanced fertilizer treatment is due to it having the highest N and P uptake and biomass yield. The

Fig. 2. Average effect of different treatments on soil quality index (SQI) and individual contribution of minimum data set (MDS) indicator (scored and weighted) to overall index value. T1: Control (no mineral fertilizers, farmyard manure (FYM) or Bio-inoculants), T₂: FYM @ 5 t/ha, T₃: Bio-inoculants, T₄: Farmers practice, T₅: Recommended dose of mineral fertilizers, T₆: Precise application of mineral fertilizers based on initial soil test values and targeted yield equations for achieving target yield of 1.5 t/ha lentil grain, T₇: Integrated approached based on initial soil test values and targeted yield equations for achieving target yield of 1.5 t/ha lentil grain + FYM @ 5 t/ha, T₈: Integrated approached based on initial soil test values and targeted yield equations for achieving target yield of 1.5 t/ha lentil grain + Bio-inoculants and T₉: Precise application of mineral fertilizers based on initial soil test values and targeted yield equations for achieving targeted yield of 2.0 t/ha lentil grain. P_a, available phosphorus; ACP, acid phosphatase; MBC, microbial biomass carbon; TCF, total culturable fungi; SOC, soil organic carbon.



higher N recovery seen in balanced fertilizer treatments compared with other treatments might be due to symbiotic N_2 fixation improving N availability and its translocation into the plants and grains of lentil, resulting in higher biomass yield. However, enhancing P recovery is the result of improving ACP and ALP, which promoted P dynamics in the soil. In the present study, P_a was positively correlated with ACP, which indicated that a balanced supply of P during the growth period of lentil accelerated P recovery. However, a lower dose of balanced fertilization recorded the highest K recovery rate. This may have been due to effective uptake of applied nutrients (Bera *et al.* 2006). The lowest P recovery, seen under farmers' practice, might be due to



Fig. 3. (a) Correlation between soil quality index (SQI) values and crude protein (CP) of lentil; (b) Correlation between soil quality index (SQI) values and grain yield of lentil.

imbalances in microbial (bacterial, fungal, actinomycetes, *Azotobacter* and PSM) and enzymatic dynamics in soil that adversely affected the conversion and translocation of plant nutrients.

Microbial biomass carbon, microbial counts and enzymatic activity

Balanced fertilization based on soil tests and target yield had a beneficial effect on MBC, SR, microbial counts and enzymatic activities. The current results revealed that application of FYM or bio-inoculants alone or in combination with inorganic fertilizers significantly increased bacterial, fungal, actinomycetes, Azotobacter and PSM counts, whereas unbalanced application of mineral fertilizers caused lower MBC and SR. Kumar et al. (2013) reported that organic manure containing vast quantities of readily utilizable energy sources and balanced fertilizers being rich in available nutrients that stimulate growth of microorganisms (TCB, TCA, TCF, AZO and PSM), positively correlated with MBC and SR. In addition to this, organic manure improved build-up of SOC in the soil and had a significant positive correlation with MBC, thus helping to increasing MBC and SR. Moreover, FYM supplies organic C substrates that help microorganisms not only for maintenance but also for production of new biomass. The current findings revealed that SOC had a positive relationship with TCB and exhibited an active role for enhancing the microbial count, resulting in higher MBC and SR.

Integration of FYM or bio-inoculants with balanced inorganic fertilizers significantly enhanced TCB, TCA, TCF, AZO and PSM in the present study. These results support those of Kanazawa *et al.* (1988), who reported that application of FYM or bio-inoculation increased SOC and MBC, which had significant positive effects on microbial counts. However, application of higher doses of mineral fertilizers had a positive effect on TCA but a negative effect on *Azotobacter* counts, as it is more sensitive to nitrogenous fertilizers. Although ammonium is the preferred N source for most bacteria and fungi (Marzluf 1997), when applied at high rates through urea and ammonium fertilizers it may inhibit soil microorganisms due to ammonia toxicity, increase in soil pH and ionic strength (Omar & Ismail 1999).

Chang et al. (2007) and Ai et al. (2012) reported that application of organic manure and inorganic fertilizers not only affect microbial counts but also soil enzymes. Enzymatic activity is widely used as an indicator of soil quality and has a close relationship to nutrient transformation of soil (Yang et al. 2008). Li et al. (2008) reported that soil microbial biomass was positively correlated with soil enzymatic activities. Higher enzymatic activities were observed in FYM plus balanced fertilizer treatments in the present study, based on soil test values and target yield equations. These results are coherent with the findings reported by Singh et al. (2017). Geisseler & Horwath (2009) found an increased availability of mineral N due to production of extracellular enzymes involved in the C, N and P cycle. These results are in line with the present study, which showed a marked increase in DHA, FDHA, ACP and ALP activities when available N increased. Yang et al. (2007) explained that UA was affected slightly by mineral fertilizers applied alone, even though urea is a common N fertilizer, but improved significantly due to integration of FYM or bio-inoculants with balanced fertilizers. Bio-inoculation improved enzymatic activity due to increasing microbial counts, MBC, SOC and availability of N and P, which had a direct impact on soil enzymes. These results, attributed to N₂ fixation, P solubilization, and production of plant growthpromoting and bio-controlling substances, might promote bacterial, actinomycetes, fungal, PSM and Azotobacter counts at adequate supply of nutrient. The increasing microbial counts result in enhanced enzymatic activity because they have a direct correlation with microbial counts, as depicted in the current results.

Fertility status and soil quality

Application of balanced fertilizer had pronounced effects on improving SOC, N_a, P_a and K_a over the unbalanced fertilization and initial nutrient status. In the present study, balanced mineral fertilization with FYM increased SOC by 10.2% compared with the control. These findings are corroborated with results of Körschens et al. (2013). With increasing lentil productivity over the years, the major portion of plant residues (rhizo-deposition, root, shoot and branch biomass) is returned to the soil during crop growth and at harvest has a positive impact on SOC (Ladha et al. 2011), microbial counts (TCB, TCA, TCF, AZO and PSM), MBC, SR and enzymatic activity (DHA, FDHA, urease, ACP and ALP). Zhong & Cai (2007) illustrated that soil available nutrients (N_a, P_a and K_a) were notably altered by the application of mineral fertilizers in comparison to a control, which is in accordance with the current results. The increase in SOC seen in the present study for FYM plus balanced mineral fertilization might be attributed to increasing microbial activities (bacteria, actinomycetes, fungal counts), MBC, grain and straw yield by the addition of residue biomass and stabilization in to SOC that had significant positive correlation with TCB. Moreover, the current findings also indicate that microbial biomass is more important than plant biomass for enhancing SOC as the FYM plus balanced fertilizer treatment had higher SOC than balanced fertilizers alone, where higher amounts of plant biomass were added. These results are accordance with the findings of Gregorich et al. (2001) and Srinivasrao et al. (2012). Application of higher doses of mineral fertilizers in an attempt to achieve higher target yields based on soil test values and target yield equations showed the highest N_a and K_a. This indicates that balanced fertilization improved the build-up of N_a and K_a status in soil, despite having the highest nutrient acquisition. The N fixed in plant nodules is partly used by the plant as indicated by improvement in N contents of straw and grains. The remaining N is added to the soil and can be utilized by the subsequent crop. Similar observations reported by Daterao *et al.* (1990) also revealed increased soil N after the harvest of green gram. However, inoculation of bio-inoculants, especially P-solubilizers, solubilized unavailable forms of inorganic P into available inorganic P through production of organic acids.

In the present study, SOC (0.316) and TCF (0.302) were retained in the MDS owing to higher factor loading values, whereas TCB counts and SR were redundant owing to low factor loading values and significant correlation with SOC (Rezaei et al. 2006). Microbial biomass carbon was selected as a soil quality indicator despite a lower factor loading value than TCB and SR in PC1 because of strong significant correlations with SOC, grain yield, crude protein, TCB, TCA, TCF, AZO and PSM which promote grain yield and soil quality. Soil organic carbon, an agent of labile nutrients, is a determinant for establishment of soil quality in management systems (Qi et al. 2009). Moreover, SOC was not only significantly correlated with grain yield, straw yield and crude protein but also significantly correlated with TCB, TCA, TCF, SR, DHA, FDHA, ACP and ALP. Therefore, SOC was retained in PC1 as a soil quality indicator, although it was significantly correlated with MBC. Total cultivable fungi was selected as a soil quality indicator with the next highest factor loading value after SOC because it plays a vital role in decomposition of crop residues, mobilization/solubilization of nutrients, producing plant growth promoting substances and addition of fungal biomass. Also, the fungal mycelium helps to stabilize soil aggregates (Ellouze et al. 2014). Acid phosphatase, FDHA and K_a were the only high-loading factor variables in PC2. Acid phosphatase was retained in the MDS in PC2 since it had a higher loading factor value (0.358) than FDHA (0.340) or K_a (0.332). Fluorescein diacetate hydrolytic activity had a strong, significant correlation with ACP and low factor loading value; therefore, it could not be retained in PC2. Phosphorous is an important factor for growth and development, especially in leguminous plants that have very low recovery rates. Hence, this clearly indicates the preference of available P to lentil. In the present study, P_a was a primary factor in PC3 having high factor loading (0.479). Urease activity and N_a could not be retained in the MDS due to lower loading values. The choice among wellcorrelated variables could also be based on the practicability of the variables; hence, one could use the final MDS considering the logical utility and interpretability (Andrews et al. 2002a). Considering these objectives, options were utilized to retain or eliminate the variables from the MDS. Hence, the final MDS consisted of SOC, TCF, MBC, ACP and Pa. Among these five indicators, SOC was the most dominant factor.

Integration of FYM or bio-inoculants with balanced fertilizer treatments enhanced SQI due to improvement in MBC, microbial counts, enzymatic activity, SOC and nutrient content of soil. Soil organic carbon is well known for improving soil quality owing to significant correlations with MBC, microbial counts, enzymatic activity and soil fertility. Application of higher doses of mineral fertilizer to achieve higher yield targets also enhances soil quality due to increases MBC, SOC and available nutrient in soil, despite slightly lower microbial counts and enzymatic activity than integration of FYM and bio-inoculants (Singh *et al.* 2015). Coming to the role of MBC, microbial biomass consists mostly of bacteria and fungi, which decompose crop residue and organic matter in soil and release nutrients into the soil that are then available for plant uptake. Several studies have suggested that MBC is a useful and sensitive measure of changes in soil organic matter by

microbial activities that are the driving force behind soil organic matter transformation through mineralization and immobilization. These transformations are the basis of plant decomposition, nutrient availability, soil pH, soil aggregation and soil tilth (Dhull *et al.* 2004). Chinnadurai *et al.* (2014) also reported that MBC is influenced strongly by management practices and that it provides an indication of a soil's ability to store and recycle nutrients and energy. In lentil, being a leguminous crop, the contribution of ACP to yield and SQI is important as ACP is the most important enzyme for P dynamics. Hence, P_a is not only important in energy transformation and storage for metabolic processes but also plays a very crucial role in root establishment, proliferation and N_2 fixation.

Conclusions

The results implied that application of balanced fertilization based on initial soil test values and target yields achieved the lower of two target grain yields of lentil with slight deviation (-9.33%)but increased soil microbial counts, MBC, soil respiration and enzymatic activities, rendering significant improvement in SQI through maintenance of soil N, P and K status. The highest increase in microbial counts, enzymatic activities, nutrient status and SQI were seen in balanced fertilizers applied as per initial soil test value and target yield equations with organic manure (FYM) and co-inoculation of *Rhizobium* + *B. megaterium*. However, unbalanced fertilization such as farmers' practice and recommended doses of fertilizers led to a deterioration of the soil quality, which impacted grain and straw yields of lentil adversely. Overall, the present study therefore clearly reflected that the balanced application of mineral fertilizers based on sitespecific soil test values and target grain yields in combination with organic manures and bio-fertilizers is the best approach for accelerating nutrient acquisition, fertility status and improving SQI for sustaining lentil productivity in the New Gangetic alluvial soils of India.

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Conflict of Interest. None.

References

- Abbeddou S, et al. (2011) Nutritional composition of lentil straw, vetch hay, olive leaves and saltbush leaves and their digestibility as measured in fattailed sheep. Small Ruminant Research 96, 126–135.
- Ai C, et al. (2012) Responses of extracellular enzyme activities and microbial community in both the rhizosphere and bulk soil to long term fertilization practices in a fluvo-aquic soil. Geoderma 173–174, 330–338.
- Ali M and Gupta S (2012) Carrying capacity of Indian agriculture: pulse crops. *Current Science* **102**, 874–881.
- Anderson JPE (1982) Soil respiration. In Page AL, Miller RH and Keeney DR (eds). *Methods of Soil Analysis, Part 2*. Madison, WI: ASA, SSSA, pp. 831– 871.

- Andrews M, et al. (1992) Nitrate effects on leaf growth of grain legumes prior to nodulation: species differences relate to nitrate uptake. In Ramsay G and Middlefell-Williams JE (eds). Proceedings of the 1st European Conference on Grain Legumes. Paris, France: L'Union Nationale Interprofessionnelle des Plantes Riches en Proteins, pp. 139–140.
- Andrews SS, et al. (2002a) On farm assessment of soil quality in California's central valley. Agronomy Journal 94, 12–23.
- Andrews SS, Karlen DL and Mitchell JP (2002b) A comparison of soil quality indexing methods for vegetable production systems in Northern California. *Agriculture, Ecosystems and Environment* **90**, 25–45.
- AOAC (Association of Official Analytical Chemists) (1999) Official Methods of Analysis, 16th edn. Gaithersburg, Maryland: Association of Official Analytical Chemists.
- Basu TK and Bansyopadhyay S (1990) Effect of *Rhizobium* inoculation and nitrogen application on some yields attributes of mung. *Environment and Ecology* 8, 650–654.
- Bera R, et al. (2006) Targeted yield concept and a framework of fertilizers recommendation in irrigated rice domains of subtropical India. Journal of Zhejiang University Science B7, 963–968.
- Bremer E, van Kessel C and Karamanos R (1989) Inoculant, phosphorus and nitrogen responses of lentil. *Canadian Journal of Soil Science* 69, 691–701.
- Chang E-H, Chung RS and Tsai YW (2007) Effect of different application rates of organic fertilizer on soil enzyme activity and microbial population. *Soil Science & Plant Nutrition* 53, 132–140.
- Chen XP, et al. (2010) Optimizing soil nitrogen supply in the root zone to improve maize management. Soil Science Society of America Journal 74, 1367–1373.
- Chinnadurai C, Gopalaswamy G and Balachandar D (2014) Long term effects of nutrient management regimes on abundance of bacterial genes and soil biochemical processes for fertility sustainability in a semi-arid tropical Alfisol. *Geoderma* 232-243, 563-752.
- Chivenge P, Vanlauwe B and Six J (2011) Does the combined application of organic and mineral nutrient sources influence maize productivity? A meta-analysis. *Plant and Soil* 342, 1–30.
- Choudhary HR, et al. (2011) Effect of organic sources and chemical fertilizers on productivity of mungbean. Journal of Food Legumes 24, 324–326.
- **Cochran WG and Cox GM** (1957) *Experimental Designs*, 2nd edn. New York: John Wiley and Sons.
- **Contesto C, et al.** (2008) Effects of rhizobacterial ACC deaminase activity on *Arabidopsis* indicate that ethylene mediates local root responses to plant growth-promoting rhizobacteria. *Plant Science* **175**, 178–189.
- Cui ZL, et al. (2010) In-season nitrogen management strategy for winter wheat: maximizing yields, minimizing environment impact in an overfertilization context. Field Crops Research 116, 140–146.
- Daterao SH, et al. (1990) Effect of *Rhizobium* seed inoculation of green gram with and without molybdenum on grain yield and nitrogen status of soil. *PKV Research Journal* 14, 75–77.
- Dhull S, et al. (2004) Microbial biomass carbon and microbial activities of soils receiving chemical fertilizers and organic amendments. Archive of Agronomy and Soil Science 50, 641–647.
- **Doran JW and Parkin TB** (1994) Defining and assessing soil quality. In Doran JW, Coleman DC, Bezdicek DF and Stewart BA (eds). *Defining Soil Quality for A Sustainable Environment*. Special Publication No. 35. Madison, WI: SSSA, pp. 3–21.
- Egamberdiyeva D (2005) Plant-growth-promoting rhizobacte-ria isolated from a Calcisol in a semi-arid region of Uzbekistan: biochemical characterization and effectiveness. *Journal of Plant Nutrition and Soil Science* 168, 94–99.
- Elfstrand S, Hedlund K and Martensson A (2007) Soil enzyme activities, microbial community composition and function after 47 years of continuous green manuring. *Applied Soil Ecology* **35**, 610–621.
- Ellouze W, et al. (2014) Soil fungal resources in annual cropping systems and their potential for management. *BioMed Research International* 2014, 1–15, Article ID 531824. doi: 10.1155/2014/531824.
- Erskine W, Muehlbaure FJ and Short RW (1990) Stages of development in lentil. *Experimental Agriculture* 26, 297–302.
- **FAOSTAT** (2016) *Agricultural Data: Agriculture and Food Trade.* Rome, Italy: FAO. Available at http://faostat.fao.org (Accessed 20 February 2018).

- Gan Y, et al. (2009) Adaptability of chickpea in northern high latitude areas maturity responses. Agricultural and Forest Meteorology 149, 711–720.
- Geisseler D and Horwath WR (2009) Relationship between carbon and nitrogen availability and extracellular enzyme activities in soil. *Pedobiologia* 53, 87–98.
- Gregorich EG, Drury CF and Baldock JA (2001) Changes in soil carbon under long-term maize in monoculture and legume-based rotation. *Canadian Journal of Soil Science* 81, 21–31.
- **Gull M, et al.** (2004) Phosphorus uptake and growth promotion of chickpea by co-inoculation of mineral phosphate solubilization bacteria and a mixed culture. *Australian Journal of Experimental Agriculture* **44**, 623–628.
- He P, et al. (2009) Performance of an optimized nutrient management system for double-cropped wheat-maize rotations in north-central China. *Agronomy Journal* 101, 1489–1496.
- Hoque MM and Haq MF (1994) *Rhizobium* inoculation and fertilization of lentil in Bangladesh. *LENS Newsletter* **21**, 29–30.
- Hussain I, et al. (1999) Adaptation of soil quality indices and application to three tillage systems in southern Illinois. Soil and Tillage Research 50, 237–249.
- Iqbal MA, et al. (2016) Integrated use of *Rhizobium leguminosarum*, plant growth promoting rhizobacteria and enriched compost for improving growth, nodulation and yield of lentil (*Lens culinaris* medik.). *Chilean Journal of Agricultural Research* 72, 104–110.
- Jackson ML (1973) Soil Chemical Analysis. New Delhi, India: Prentice Hall of India Pvt., Ltd.
- Kanazawa S, Asakawa S and Takai Y (1988) Effect of fertilizer and manure application on microbial numbers, biomass and enzymes activities in volcanic ash soils. *Soil Science and Plant Nutrition* **34**, 429–439.
- Körschens M, et al. (2013) Effect of mineral and organic fertilization on crop yield, nitrogen uptake, carbon and nitrogen balances, as well as soil organic carbon content and dynamics: results from 20 European long-term field experiments of the twenty-first century. Archives of Agronomy and Soil Science 59, 1017–1040.
- Kumar S, et al. (2013) Balanced fertilization along with farmyard manures enhances abundance of microbial groups and their resistance and resilience against heat stress in a semi-arid inceptisol. *Communications in Soil Science* and Plant Analysis 44, 2299–2313.
- Kundu R, Mandal J and Majumder A (2013) Growth and production potential of green gram (*Vigna radiata*). influenced by *Rhizobium* inoculation with different nutrient sources. *International Journal of Agriculture*, *Environment and Biotechnology* 6, 344–350.
- Kutschera L (1960) Wurzelatlasmitteleuropaischer Ackerunkrauter und Kulturpflanzen. Frankfurt, Germany: DLG-Verlags-GmbH.
- Ladha JK, et al. (2011) Role of nitrogen fertilization in sustaining organic matter in cultivated soils. *Journal of Environmental Quality* 40, 1756– 1766.
- Lavanya GR and Toms B (2009) Association and interrelationship among yield contributing characters in mungbean. *Journal of Food Legumes* 22, 65–67.
- Li J, et al. (2008) Effects of long-term combined application of organic and mineral fertilizers on microbial biomass, soil enzyme activities and soil fertility. March 2008. Agricultural Sciences in China 7, 336–343.
- Martin JP (1950) Use of acid, rose bengal and streptomycin in the plate method for estimating soil fungi. *Soil Science* **69**, 215–232.
- Marzluf GA (1997) Genetic regulation of nitrogen metabolism in the fungi. Microbiology and Molecular Biology Reviews 61, 17–32.
- Matus A, et al. (1997) The influence of tillage and crop rotation on nitrogen fixation in lentil and pea. Canadian Journal of Plant Science 77, 197–200.
- Nleya T, Walley F and Vandenberg A (2001) Response of four common bean cultivars to granular inoculant in a short season dryland production system. *Canadian Journal of Plant Science* 81, 385–390.
- **Olsen SR, et al.** (1954) Estimation of Available Phosphorus in Soils by Extraction with Sodium Bicarbonate. United States Department of Agriculture Circular no. 939. Washington DC: USDA.
- **Omar SA and Ismail MA** (1999) Microbial populations, ammonification and nitrification in soil treated with urea and inorganic salts. *Folia Microbiologica* **44**, 205–212.

- Parkinson D, Gray TRG and Williams ST (1971) Methods for Studying the Ecology of Soil Microorganisms. International Biological Programme Handbook 19. Oxford, UK: Blackwell Scientist Publications.
- Pikovskaya AI (1948) Mobilization of phosphorus in soil in connection with vital activity of some microbial species. *Microbiology* 17, 362–370.
- Qi YB, et al. (2009) Evaluating soil quality indices in an agricultural region of Jiangsu Province, China. *Geoderma* 149, 325–334.
- Ramamoorthy B and Velayutham M (1971) Soil Test Crop Response Correlation Work in India. World Soil Resources Report No. 41: 96-105, Rome, Italy: FAO.
- Ramamurthy V, et al. (2009) Soil-based fertilizer recommendations for precision farming. Current Science 97, 641–647.
- Rezaei SA, Gilkes RJ and Andrews SS (2006) A minimum data set for assessing soil quality in rangelands. *Geoderma* 136, 229–234.
- Robertson GP and Groffman PM (2015) Nitrogen transformations. In Paul EA (ed.). *Soil Microbiology, Ecology and Biochemistry*, 4th edn. Amsterdam, the Netherlands: Elsevier, pp. 421–446.
- Sarkar GK, et al. (2014) Depletion of soil potassium under exhaustive cropping in Inceptisols and Alfisols. Communications in Soil Science and Plant Analysis 45, 61–72.
- Schneider K and Anderson L (2010) Yield Gap and Productivity Potential in Ethiopian Agriculture: Staple Grains & Pulses. EPAR Brief No. 98. Seattle, WA: Evans School Policy Analysis and Research (EPAR), University of Washington.
- Schnurer J and Rosswall T (1982) Fluorescein diacetate hydrolysis as a measure of total microbial activity in soil and litter. *Applied and Environmental Microbiology* 43, 1256–1261.
- Schoenholtz SH, Van Miegroet H and Burger JA (2000) A review of chemical and physical properties as indicators of forest soil quality: challenges and opportunities. *Forest Ecology and Management* 138, 335–356.
- Shahzad SM, et al. (2008) Integrated use of plant growth promoting bacteria and p-enriched compost for improving growth, yield and nodulation of chickpea. Pakistan Journal of Botany 40, 1735–1441.
- Singh HP, et al. (1998) Prospects of Indian agriculture with special reference to nutrient management under rainfed systems. In Swarup A, Damodar Reddy D and Prasad RN (eds). Long Term Soil Fertility Management Through Integrated Plant Nutrient Supply. Bhopal, India: Indian Institute of Soil Science, pp. 34–54.
- Singh SR, et al. (2015) Impact of balanced fertilization on nutrient acquisition, fibre yield of jute and soil quality in New Gangetic alluvial soils of India. Applied Soil Ecology 92, 24–34.
- Singh SR, et al. (2017) Identification of minimum data set under balanced fertilization for sustainable rice production and maintaining soil quality in alluvial soils of Eastern India. Communications in Soil Science and Plant Analysis 48, 2170–2192.
- Soil Survey Staff (1998) Keys to Soil Taxonomy, 8th edn. Washington DC: USDA National Conservation Service.
- Srinivasrao C, et al. (2012) Long-term effects of soil fertility management on carbon sequestration in a rice-lentil cropping system of the Indo-Gangetic Plains. Soil Science Society of America Journal 76, 168–178.
- Stark C, et al. (2007) Influence of organic and mineral amendments on microbial soil properties and processes. Applied Soil Ecology 35, 79–93.
- Subbiah BV and Asija GL (1956) A rapid procedure for estimation of available nitrogen in soils. Current Science 25, 259–260.
- Tabatabai MA (1982) Soil enzymes. In Page AL, Miller RH and Keeney DR (eds). Methods of Soil Analysis Part 2: Chemical and Microbiological Properties. New York: Academic Press, pp. 903–947.
- Tabatabai MA and Bremner JM (1969) Use of p-nitrophenyl phosphate for assay of soil phosphatase activity. Soil Biology and Biochemistry 1, 301–307.
- Vance ED, Brookes PC and Jenkinson DS (1987) An extraction method for measuring soil microbial biomass C. Soil Biology and Biochemistry 19, 703– 707.
- Walkley A and Black CA (1934) An examination of Degtjareff method for determining soil organic matter and a proposed modification of the chromic acid titration method. *Soil Science* 37, 29–38.
- Wander MM and Bollero GA (1999) Soil quality assessment of tillage impacts in Illinois. Soil Science Society of America Journal 63, 961–971.

Wold S, Esbensen K and Geladi P (1987) Principal component analysis. Chemometrics and Intelligent Laboratory Systems 2, 37–52.

 Yang JY, et al. (2007) Residual soil nitrogen in soil landscapes of Canada as affected by land use practices and agricultural policy scenarios. Land Use Policy 24, 89–99.
 Yang L, et al. (2008) Fertilization regulates soil enzymatic activity and fertility

dynamics in a cucumber field. Scientia Horticulturae 116, 21–26.

- Zafar M, et al. (2012) Effect of plant growth-promoting rhizobacteria on growth, nodulation and nutrient accumulation of lentil under controlled conditions. *Pedosphere* 22, 848–859.
- Zhong WH and Cai ZC (2007) Long-term effects of inorganic fertilizers on microbial biomass and community functional diversity in a paddy soil derived from quaternary red clay. *Applied Soil Ecology* **36**, 84–91.