Effect of host diversity and species assemblage composition on bovine tuberculosis (bTB) risk in Ethiopian cattle

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SUMMARY

Current theories on diversity-disease relationships describe host species diversity and species identity as important factors influencing disease risk, either diluting or amplifying disease prevalence in a community. Whereas the simple term 'diversity' embodies a set of animal community characteristics, it is not clear how different measures of species diversity are correlated with disease risk. We therefore tested the effects of species richness, Pielou's evenness and Shannon's diversity on bovine tuberculosis (bTB) risk in cattle in the Afar Region and Awash National Park between November 2013 and April 2015. We also analysed the identity effect of a particular species and the effect of host habitat use overlap on bTB risk. We used the comparative intradermal tuberculin test to assess the number of bTB-infected cattle. Our results suggested a dilution effect through species evenness. We found that the identity effect of greater kudu – a maintenance host – confounded the dilution effect of species diversity on bTB risk. bTB infection was positively correlated with habitat use overlap between greater kudu and cattle. Different diversity indices have to be considered together for assessing diversity–disease relationships, for understanding the underlying causal mechanisms. We posit that unpacking diversity metrics is also relevant for formulating disease control strategies to manage cattle in ecosystems characterized by seasonally limited resources and intense wildlife–livestock interactions.

Key words: evenness, diversity, greater kudu, identity effect, maintenance hosts, multi-host disease ecology, habitat use overlap.

INTRODUCTION

Bovine tuberculosis (bTB), caused by Mycobacter*ium bovis*, is an important zoonotic disease affecting many mammal species, and mainly spreads via aerosol transmission (Skuce et al. 2012). The World Health Organisation (WHO, 2012) identified bTB as one of the eight worldwide neglected zoonoses needing urgent attention, especially in developing countries. The disease is endemic in sub-Saharan African cattle (de Garine-Wichatitsky et al. 2013), and cattle are the main host for M. bovis (Cosivi et al. 1998). A wide range of domestic and wildlife mammals, but also humans can be infected with bTB (Munyeme et al. 2008). Although control programmes have eliminated or nearly eliminated this disease from domestic animals in some developed countries, bTB is still widespread in Great Britain, Ireland, New Zealand and many developing countries, especially in Africa (Renwick et al. 2007; Humblet et al. 2009). In fact, bTB is an important public concern, and can cause economic losses due to livestock deaths, product reduction and trade restrictions (Humblet et al. 2009).

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Africa is recognized as a hotspot for biodiversity, but is suffering from rapid and extensive loss of that diversity (Myers et al. 2000; Olff et al. 2002; Gorenfloa et al. 2012; Di Marco et al. 2014). The continent is also a hotspot for emerging infectious diseases as illustrated by emergence of Ebola, HIV/ AIDS, MERS, among others (Morens et al. 2004). As biodiversity loss is thought to be a major explanatory factor of the increase in emergence of infectious diseases (Keesing et al. 2010; Ostfeld and Keesing, 2012; Huang et al. 2013), it is key to investigate the links between biodiversity, and biodiversity loss on the patterns of infectious diseases in Africa. Recently, several studies have shown that a reduction in biodiversity may increase the prevalence and transmission of diseases (Keesing et al. 2010; Cardinale et al. 2012; Johnson et al. 2013; Myersa et al. 2013; Civitello et al. 2015). The two alternative hypotheses are the dilution and the amplification effect (Keesing et al. 2006; Huang et al. 2013; Hofmeester et al. 2016). The dilution effect predicts that species diversity decreases pathogen prevalence through mechanisms such as decreased host density, reduced encounters between hosts, or reduced host survival (Keesing et al. 2006; Huang et al. 2013; Johnson et al. 2013). In contrast, the amplification effect predicts increased pathogen prevalence with greater species diversity, through increased encounters between hosts, or through the presence of



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secondary hosts (LoGiudice et al. 2003; Keesing et al. 2006). A recent review of the relationships between species diversity and diseases reported dilution effects in up to 80% of the studies examined, and amplification effects in 12% of the studies (Cardinale et al. 2012; Ostfeld and Keesing, 2012). Despite the fact that the dilution effect occurs far more frequently than the amplification effect, our knowledge of which specific systems conform to the dilution effect and the mechanisms underlying the effects of diversity, is incomplete (Ostfeld and Keesing, 2012; Randolph and Dobson, 2012; Huang et al. 2013; Johnson et al. 2013; Miller and Huppert, 2013; Ostfeld, 2013; Hofmeester et al. 2016). Understanding the underlying mechanisms how the risk of disease relates to the level of biodiversity is important, both for predicting disease dynamics in the context of global biodiversity decline, and to provide valuable insights into successful control measures.

Most studies that examine the diversity-disease relationship focus principally on species richness as a measure of biodiversity (Keesing et al. 2006). In fact, biodiversity can be measured in many different ways, as the number of species (species richness), the distribution of individuals over species (species evenness), or a combination of richness and evenness, as represented by diversity indices such as the Shannon index (Magurran, 1988; Tucker and Cadotte, 2013). Many studies have argued that species richness and evenness are two independent indices (Sheldon, 1969; Smith and Wilson, 1996; Gosselin, 2006; Symonds and Johnson, 2008), and suggest treating them separately (Magurran, 1988; Legendre and Legendre, 1998). Ostfeld and Keesing (2000) stated that encounter rate is proportional to the distribution of hosts. Thus, evenness which measure how evenly the individuals are distributed in the community among different species may be most appropriate measure of biodiversity to explain disease risk, because of power to detect the probability of encounter between pathogens and each host species. Thus, despite many studies of the relationship between diversity and diseases, evaluating the effects of different diversity metrics on disease risk has proven to be rare (Chen and Zhou, 2015). Thus, these different metrics of diversity may have different predictive powers for predicting disease risk in the target population. Here we tested for the effect of different diversity metrics on bTB risk in cattle.

Several recent studies suggest that the occurrence of particular species in the animal community may play an important role in disease risk, and in determining whether biodiversity amplifies or dilutes the infectious disease (Fenton and Pedersen, 2005; Keesing *et al.* 2010; Hamer *et al.* 2011; Johnson *et al.* 2013, 2015; Oda *et al.* 2014). This effect of a

particular species on pathogen transmission is known as the identity effect (Hantsch et al. 2013; Huang et al. 2014, 2016). Generally, the identity effect on pathogen transmission can be observed in two different situations (Huang et al. 2016). One is that a key species with particularly high or low reservoir competence may be present in communities when species diversity increases. The other situation is where a species can affect vector abundance (either positively or negatively) (Huang et al. 2016). To our knowledge, the generality of this pattern for directly transmitted or aerosol-borne diseases, such as bTB, has not been established. Thus, understanding the identity effect is an important step in being able to understand the expected impacts of biodiversity loss on disease dynamics. In Africa, buffalo (Syncerus caffer), greater kudu (Tragelaphus strepsiceros) and lechwe (Kobus leche; Cosivi et al. 1995) have been identified as maintenance hosts and implicated in the transmission of M. bovis. Warthog (Phacochoerus africanus) are also thought to be a potential reservoir for this bacteria in Africa (Tschopp, 2015). The presence of species such as the greater kudu and warthog are likely to affect the type of encounters with cattle, which could then alter the relation between biodiversity and disease risk. We thus tested for the existence of an identity effect of greater kudu and warthog. We predict that bTB risk increased with the occurrence of maintenance host species.

Currently, livestock and wild herbivores graze together in many arid and semi-arid rangelands of Africa, with much resource use overlap, as livestock species are ecologically similar, with similar resource requirements as several wild herbivore species (Prins, 2000; Sitters et al. 2009). Overlapping space use can lead to interspecific interactions, and stimulate the spread and prevalence of many diseases (Riley et al. 1998), as most pathogens are able to cross-infect multiple host species. Hence, in areas where wildlife and livestock co-occur, pathogens can emerge and establish in these sympatric host populations (Gortazar et al. 2007). For example, foot and mouth disease, rabies, anthrax, brucellosis and bTB have all been shown to be reciprocally transmissible between livestock and wildlife (Frohlich et al. 2002; Artois, 2003; Ward et al. 2006; Cooper et al. 2010; Proffitt et al. 2011). In this context, resource use overlap between host species can play an important role in pathogen transmission by increasing contact rates and environmental exposure to the agent (Roper et al. 2003; Böhm et al. 2009). How habitat use by hosts affects direct and indirect interactions among hosts is fundamental in understanding multi-host disease transmission (Cooper et al. 2010), and is critical for designing scientifically sound disease control strategies (Hudson et al. 2002). Nevertheless, the role that spatial interactions between livestock and wildlife

host play in disease transmission remains mostly unknown (Böhm et al. 2009; Martin et al. 2011; Tschopp, 2015). For instance, habitat and water resources use overlap may stimulate bTB transmission through increasing wildlife maintenance hostcattle contact, such as observed in and around Awash National Park, Ethiopia, where large numbers of livestock share their habitat with wildlife particularly during the dry season when resources are scarce. We therefore also tested whether habitat use overlap between wildlife maintenance host (greater kudu and warthog) and cattle increased bTB prevalence. Beside the role of host community composition and resource overlap, a positive effect of host (e.g. cattle) densities (Humblet et al. 2009; Dejene et al. 2016) has also been associated with bTB transmission risk. We also tested whether cattle densities were positively correlated with bTB incidence in cattle.

STUDY AREA

We carried out a cross-sectional study in Awash National Park and in the neighbouring Afar Region, Ethiopia. Awash National Park (9°20'N, 40°20'E) is situated in the Ethiopian Rift valley and had an elevation of 960-1050 m above sea level (Fig. 1). It is covered in semi-arid savanna. The Afar region is found in the northeastern part of Ethiopia (between 8°49' to 14°30'N latitude and 39°34' to 42°28'E longitude; Fig. 1) with an area of about 70 000 km^2 (CSA, 2008). It is characterized by an arid and semi-arid climate with low and erratic rainfall, with a mean annual rainfall of 500 mm in the semiarid western escarpments, decreasing to 150 mm in the arid zones to the east. Study sites were included due consideration of variation in wildlife-livestock interactions, concentrations of livestock and wildlife, and the presence of common grazing and water resources (for details see Dejene et al. 2016).

METHODOLOGY

Study design

A cross-sectional multi-stage sampling was used to select study villages with 'sub-region' as the highest level followed by 'district' (n = 17; Fig. 1), and 'sub-district' (n = 34) at the lowest level. Study animals were obtained using a three-stage random sampling procedure. The village within the sub-district was regarded as the primary unit, the herd as secondary unit and individual animal as tertiary unit, following the method of Dejene *et al.* (2016). The desired sample size, which gave us a total of 2550 animals, was calculated following the method of Dejene *et al.* (2016). Tuberculin skin testing was performed using Purified Protein Derivative (supplied by Prionics Lelystad B.V, Lelystad, The Netherlands) to identify bTB-positive animals following the method of Dejene *et al.* (2016).

Dung counts

Plots for dung counts were established using stratified random sampling. First, sub-districts were stratified according to vegetation type. 204 plots (six in each of the 34 sub-districts) of $100 \times 100 \text{ m}^2$ were laid out randomly in these vegetation types and were GPS geo-referenced. In each plot, we surveyed 50 transects of 100 m length and 2 m wide, and counted dung piles. Each pile of dung was attributed to a locally available wildlife species based on the size, shape and form of the pellets by using Stuart and Stuart (2000), and with the help of experienced local trackers. The relative abundances of wild herbivores were estimated based on the frequency of fecal droppings found in the plot transects following Vicente et al. (2004). We divided each 100 m transect into 10 sectors of 10 m length. We defined sign frequency as the average number of 10-m sectors with the presence of wild herbivores droppings. Based on these frequencies, we calculated for each of the species the frequencybased indirect index (FBII):

$$FBII = \frac{1}{n} \sum_{i=1}^{n} s_i$$

where s_i is the number of sign-positive sectors in the *i*th 100 m transect (i.e. S_i varies between 0 and 10), and *n* is the number of 100 m transects considered (i.e. n = 50 for each plot; Vicente *et al.* 2004).

Ethical statements

This study was approved by Haramaya University, Ethiopia (Reference number HUP14/559/15).

Statistical analysis

For each sub-district Pianka's Niche Overlap, mammalian species richness (S), mammalian species diversity (H') and mammalian species evenness (J') were calculated. Habitat use overlap between cattle and greater kudu was calculated according to Pianka's Niche Overlap (Pianka, 1973). This index varies from 0, no overlap, to 1, complete overlap.

$$O_{jk} = \frac{\sum p_{ij} p_{ik}}{\sqrt{\sum p_{ij}^2 \sum p_{ik}^2}}$$

where O_{jk} is the overlapping index between species j and k, and p_{ij} and p_{ik} being the proportions of use of habitat i by the species j and k.

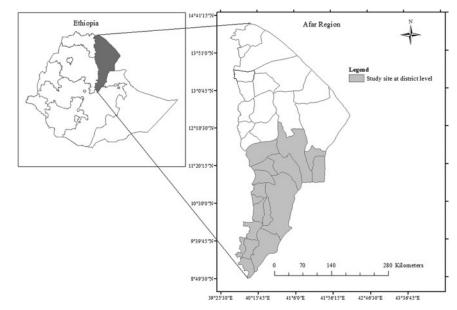


Fig. 1. Map of the study area, the Afar Region in Ethiopia (small inset) and 17 districts (larger map). The location of Awash National Park in the South is indicated by the cross-hatched area.

Shannon's diversity index (H') was used to estimate mammalian species diversity as

$$H' = -\sum_{i=1}^{S} p_i \ln p_i$$

where p_i is the proportion of species *i*, and *S* is the number of species (Hill, 1973).

Pielou's index was used to estimate mammalian species evenness (Hill, 1973), which is most widely used in ecology (Zhang *et al.* 2012).

$$\mathcal{J}' = \frac{H'}{\ln(S)}$$

where H' represents the Shannon diversity index, and S is the total number of species observed. Biodiversity metrics were calculated using package vegan of R v3.2.0 (Oksanen *et al.* 2016).

Generalized Linear Mixed Models (GLMM, family = Poisson) using package lme4 were used to examine the effects of predictors on the sub-district bTB incidence (SI- Table 1). Prior to developing our candidate models, we performed one-by-one univariate analyses to identify potential spatial risk factors, using the number of bTB-infected animals as dependent variable. Predictor variables with P <0.25 recognized as potential spatial risk factors (Huang et al. 2013), and subsequently used to construct multiple regression models. For highly correlated independent variables, only the one causing the largest change in the Log-Likelihood added to the final global model to avoid multi-collinearity, which was assessed by using variance inflation factors. The final variance inflation factor values were all <5 and confirmed the absence of collinearity among variables. From the global model, candidate models constructed using delta AIC (<5), with the best approximating candidate model having the lowest delta AIC, as described in Burnham and Anderson (2002). Model averaging was used to construct the final model based on the lowest Akaike weights of the different candidate models (Anderson *et al.* 2000). In this analysis, we treated district as a random effect to account for repeated sampling. We carried out all analyses in R v3.2.2 (R Core Team, 2015).

RESULTS

Pielou's species evenness (\mathcal{J}') and Shannon's species diversity (H') varied between 0.46–0.90 and 0.72– 2.05, respectively. Habitat use overlap between cattle and kudu varied from 0, no overlap to 0.95, high overlap. The highest Pianka's Niche Overlap index between warthog and cattle was 0.84. Relative abundances of kudu and warthog ranged from 0 to 0.93 and 0 to 0.79, respectively (SI-Table 2).

Univariate analyses

Based on the results of the univariate analyses, we identified seven out of eight variables as potential risk factors, namely, mammalian species richness, Pielou's species evenness (\mathcal{J}'), Shannon's species diversity (H'), habitat use overlap between cattle and greater kudu, habitat use overlap between cattle and warthog, relative density of greater kudu, and relative density of warthog (Table 1). Surprisingly, density of cattle was not associated with the number of bTB-infected cattle in the sub-district (Table 1).

Table 1. Results of the one-by-one GLMM analysis of all variables and summary statistics (regression coefficient *b* with 95% confidence intervals, odds ratio (OR) with 95% confidence intervals, χ^2 and *P*-value) for all predictors against sub-district (n = 34) number of bTB-positive animals from the likelihood ratio test (logLik = log likelihood) and AIC value

Number of bTB-positive animals

Variables	<i>b</i> (95%CI)	OR (95% CI)	logLik	AIC	χ^2	P-value
Habitat use overlap with kudu	1.2(0.6-1.7)	3.3(1.9-5.5)	-66.0	139.5	19.6	<0.001***
Habitat use overlap with warthog	1.4(0.7-1.9)	3.9(2.1-6.9)	-66.8	138.0	20.9	<0.001***
Relative density of kudu	1.3(0.6-1.8)	3.5(1.9-6.0)	-66.1	140.2	18.9	<0.001***
Relative density of warthog	1.4(0.8-2.0)	4.1(2.1-7.5)	-66.6	139.1	19.6	<0.001***
Species diversity (H')	0.7(0.2-1.1)	2.0(1.3-3.1)	-68.5	143.0	14.3	0.002**
Species evenness (\mathcal{Y})	-2.3(-4 to 0.6)	0.9(0.2-1.5)	-70.8	147.5	7.63	0.006**
Species richness	0.1(0.01-0.2)	1.1(1.0-1.2)	-70.1	146.2	9.74	0.001**
Density of cattle	0.2(0.1-0.3)	1.0(0.9-1.1)	-74.1	154.2	0.15	0.693
Presence of Kudu	0.9(0.5-1.2)	2.4(1.6-3.5)	-72.2	147.1	17.21	<0.001***

Kudu = greater kudu; *P < 0.05; **P < 0.01; ***P < 0.001.

Communities that contained greater kudu had a significantly higher bTB incidence than communities without greater kudu (Fig. 2; b = 0.9, 95% CI = 0.5-1.2; OR = 2.4, 95% CI = 1.6-3.5; P < 0.001).

The Spearman's correlation matrix showed that species richness was strongly correlated with Shannon's species diversity index. Habitat use overlap between cattle and warthog, relative density of greater kudu, and relative density of warthog were strongly correlated with habitat use overlap between cattle and greater kudu (r > 0.7; SI-Table 3). Therefore, we only included the latter two variables and species evenness in the multiple variable model to avoid collinearity.

Multiple variable analyses

Variables included in the multiple variable analysis were Pielou's species evenness, Shannon's species diversity and habitat use overlap between cattle and greater kudu (SI-Table 4).

The results of model averaging showed always a negative relationship between Pielou's species evenness and the number of bTB-positive cattle, but we did not find a significant relationship between Shannon's species diversity and the number of bTB-positive cattle, although the effect of species diversity was always positive in the models. In addition, our analysis also identified habitat use overlap between cattle and greater kudu as a significant risk factor for the number bTB-positive cattle in the sub-districts (Table 2; Fig. 3).

DISCUSSION

Our study showed that the bTB infection rate was negatively associated with mammalian species evenness (\mathcal{J}'), in line with our predictions derived from the dilution effect hypothesis. However, contrary

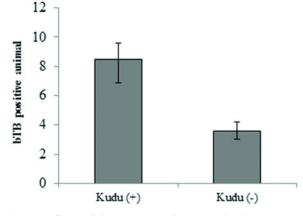


Fig. 2. Effects of the presence of a particular host species, greater kudu, on the number bTB-positive animals at the sub-district level (n = 34). Data shown are means with the 95% confidence intervals

to our expectation we did not find a significant relationship between mammalian species diversity (H')and the number of bTB-infected cattle. There was also a positive effect of habitat use overlap between cattle and greater kudu on bTB incidence in cattle. As proposed by Ostfeld and Keesing (2000), if the encounter rate is proportional to the distribution of the host species, species evenness would seem most appropriate for disease risk, because evenness, not richness, would capture the probability of encounter between pathogens and each host species (Ostfeld and Keesing, 2000; Chen and Zhou, 2015). Our study detected a dilution effect of Pielou's species evenness on the risk of bTB prevalence, an influential aerosol-borne disease. This dilution effect is possibly explained by encounter reduction, in that the addition of alternative hosts may decrease the risk of pathogen transmission by reducing encounter rates between susceptible and infected

Table 2. Summary statistics of the final model, obtained through model averaging, with regression coefficient $(b \pm SE)$, Odds Ratio (OR, 95% confidence interval) and *P*-value from the likelihood ratio test for the effect of species evenness (\mathcal{J}'), species diversity (H') and cattle-greater kudu habitat use overlap on the number of bTB-infected Ethiopian cattle in the sub-districts (n = 34)

Number of bTB-positive animal

Variables	<i>b</i> (95% CI)	OR (95%CI)	<i>P</i> -value	
Pielou's species evenness	-2.01 (-3.9 to 0.1)	0.2 (0.02-0.82)	0.036*	
Shannon's species diversity	0.46 (0.3–1.2)	1.6(0.77 - 3.30)	0.221	
Habitat use overlap with kudu	1.14 (0.3–1.8)	2.8 (1.35-5.94)	0.008**	

Kudu = greater kudu; *P < 0.05; **P < 0.01; ***P < 0.001.

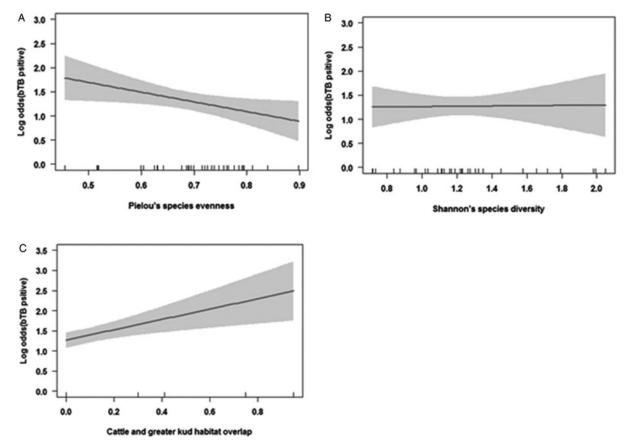


Fig. 3. GLMM results of the effects of explanatory variables on the number of bTB-positive cattle at the sub-district level (log odds scale) in relation to Pielou's species evenness (A), Shannon's species diversity (B) and resource overlap between greater kudu and cattle (C).

hosts (Keesing *et al.* 2006; Chen and Zhou, 2015). In pastoral areas of East Africa, the distribution and abundance of large grazers is negatively associated with the presence of cattle (Voeten and Prins, 1999; de Leeuw *et al.* 2001; Bonnington *et al.* 2007). For instance, de Leeuw *et al.* (2001) observed a significant reduction of species such as Oryx, gerenuk and gazelle in the presence of cattle in Kenya (de Leeuw *et al.* 2001), and Odadi *et al.* (2007) found that the preference of foraging habitat for cattle was lower in the presence of wild grazers (Odadi *et al.* 2007). Many mammal species that can be infected by bTB are spillover or deadend hosts and do not transmit the pathogen efficiently (Corner, 2006; Renwick *et al.* 2007). The presence of these non-competent or spillover mammalian species might act as barriers to cattle herd movement and distribution, and reduce encounter rates among cattle herds by changing the grazing behaviour and habitat preference (e.g. avoidance of sites contaminated by feces or different preferences for feeding patches). Such an 'encounter reduction' (Keesing *et al.* 2006) might lead to decreased probabilities of bTB infection risk, although the exact mechanism behind these correlations needs more attention.

We did not detect significant effects of host species diversity (H') on the bTB infection level. The lack of a significant association between host species diversity and disease risk might occur because the index we chose, the Shannon index, stresses the number of species and presence of rare species (McGarigal and Marks, 1994; Haines-Young and Chopping, 1996; Riitters et al. 2000; Magurran, 2004). Thus, this metric might fail to weigh in the specific importance of particular species that are not rare, which might be addressed better by focusing on the effects of host identity (Hamer et al. 2011). Moreover, studies also criticizing the dilution effect argued that pathogen transmission might increase in high-diversity communities (Randolph and Dobson, 2012; Wood and Lafferty, 2013; Huang et al. 2016) due to the increased chance of including a particular species that has a positive effect on pathogen transmission (Hantsch et al. 2013, 2014). For instance, a recent study on bTB suggested that the presence of buffalo increased disease risk due to its high bTB competence (Huang et al. 2016). Power and Mitchell (2004) also demonstrated how the identity effect of particular host species influence the diversity-disease relationship, and found that more diverse systems had higher rates of infection (i.e., amplification effect), because these species rich assemblages contained highly competent reservoir hosts (Power and Mitchell, 2004). Bouchard et al. (2013) found that the occurrence of white-tailed deer (Odocoileus virginianus), an important host for adult ticks, increase the abundance tick and thus increased the risk of tick-borne diseases (Bouchard et al. 2013). Similarly, we found that the presence of greater kudu and habitat use overlap between cattle and greater kudu was positively associated with the number of bTB infection. In Africa, species habitat use such as of greater kudu is not strongly affected by cattle presence (Prins, 2000), because kudus are almost exclusively browsers and the kudu-cattle dietary niche overlap is relatively small (Fritz et al. 1996). High habitat use overlap between cattle and kudu could increase encounter rates between them and create a positive identity effect of kudu on transmission of bTB, as a known wildlife bTB reservoir host. On the contrary, the presence of opossums created a negative identity effect on tick abundance (Keesing et al. 2009). Thus, high species diversity may amplify or dilute pathogen prevalence depending on the occurrence of a particular species. If the occurrence of the particular species had a negative identity effect, it may enhance the strength of the negative diversitydisease relationship; when the identity effect is positive, it may weaken the negative diversity-disease relationship and lead to a dilution effect (Huang et al. 2016). Another example is the influence of warthogs, which are predominantly grazers and compete with cattle for high-quality food in African savannas (Treydte et al. 2006). The species is also recognized as hosts for ticks, which are vectors of various diseases, including African Swine Fever in eastern Africa (Osofsky et al. 2005). Thus, livestock keepers tend to avoid the areas that are used by warthog for fear of diseases (Maleko et al. 2012). This could decrease the encounter rate between cattle and warthog, and lead to a non-significant identity effect on bTB transmission. This might be the reason for a nonsignificant negative diversity-disease relationship. We recognize that our conclusions are based on correlative studies and that further studies with experimental manipulation, including host behaviour change and contact rates among hosts are required to thoroughly test this hypothesis. However, our results are a necessary first step towards understanding the role of community structure on bTB risk and identifying the underlying mechanisms.

In addition to direct transmission, which requires close contact between host species, indirect transmission via environmental contamination is also possibility for bTB transmission. In the north and northeastern part of Awash National Park, particularly in the northern part of the Park at the hot spring and kudu valley areas, it is common to observe livestock grazing in close proximity with kudu during the dry season. Mycobacterium bovis has been detected in environmental samples in East Africa (Roug et al. 2014), and experimental studies have confirmed that the bacteria can survive for multiple days outside hosts (Fine et al. 2011). Kelly and Collins (1978) suggested that the major factors influencing survival of the bacteria in soil is soil temperature and moisture, as high temperature causes desiccation, and negatively influence survival of the bacteria. Environmental persistence of M. bovis has been proposed to play a role in the transmission of bTB in the UK (Courtenay et al. 2006). Wetlands or humid areas are also potential risk factors, and areas around pounds are generally moister, with greater amounts of shade, which are favourable conditions for *M. bovis* survival (Jackson et al. 1995). In Africa, flooding or soil humidity have also been suggested as propagating factors for M. bovis in the environment, as demonstrated in Tanzania (Cleaveland et al. 2007) and Zambia (Munyeme et al. 2009) by creating favourable conditions for M. bovis survival. The humid marsh-shrub wetland habitat near the hot-spring and kudu valley of Awash National Park and the surrounding water holes may act as potentially high-risk areas for *M. bovis* infection, as these areas are generally moist, with greater amounts of shade. Hence, the correlation of habitat use overlap between greater kudu and cattle with bTB infection in the GLMM analyses might not tell the whole story, as the underlying reasons for this correlation is that it is possible that environmental transmission occurs among African wildlife and livestock. These uncertainty and complex eco-epidemiological scenarios and possible confounding factors require further investigation of the transmission network.

Our results highlight aspects of mammalian species evenness and spatial differences in species assemblage that are likely to affect the risk of disease. Our results support the idea that a greater mammalian species evenness acts as a buffer against disease outbreaks. Our findings also demonstrate that the presence of a particular reservoir hosts can affect the diversity-disease relationship. Hence, it is a prerequisite to understand the identity effect, and predict future outbreaks and minimize the risk of disease transmission. Ecologists, epidemiologists and policy makers need to understand the complex interactions among potential host species to identify risk factors for disease transmission and identify efficient management actions. In order to improve this understanding, further ecological and epidemiological research on disease transmission and contact networks is required.

SUPPLEMENTARY MATERIAL

The supplementary material for this article can be found at https://doi.org/10.1017/S0031182016002511.

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REFERENCES

Anderson, D. R., Burnham, K. P. and Thompson, W. L. (2000). Model hypothesis and testing: problems, prevalence, and an alternative. *Journal of Wildlife Management* 64, 912–923.

Artois, M. (2003). Wildlife infectious disease control in Europe. *Journal Mountain Ecology* 7, 89–97.

Böhm, M., Hutchings, M. and White, P. (2009). Contact networks in a wildlife-livestock host community: identifying high-risk individuals in the transmission of bovine TB among badgers and cattle. *PLoS ONE* **4**, e5016. Bonnington, C., Weaver, D. and Fanning, E. (2007). Livestock and large wild mammals in Kilombero Valley, in southern Tanzania. *African Journal of Ecology* **45**, 658–663.

Bouchard, C., Beauchamp, G., Leighton, P.A., Lindsay, R., Belanger, D. and Ogden, N.H. (2013). Does high biodiversity reduce the risk of Lyme disease invasion? *Parasites and Vectors* **6**, 195.

Burnham, K.P. and Anderson, D.R. (2002). Model Selection and Inference: a Practical Information-Theoretic Approach, 2nd Edn. Springer-Verlag, New York. Cardinale, B. J., Duffy, J. E., Gonzalez, A., Hooper, D. U., Perrings, C., Venail, P., Narwani, A., Mace, G. M., Tilman, D. and Wardle, D. A. (2012). Biodiversity loss and its impact on humanity. *Nature* 486, 59–67.

Central Statistical Agency (CSA) (2008). Agricultural Sample Survey 2006/07, Vol II: Report on Livestock and Livestock Characteristics. Statistical Bulletin 388, Addis Ababa, Ethiopia.

Chen, L. and Zhou, S. (2015). A Combination of species evenness and functional diversity is the best predictor of disease risk in multihost communities. *The American Naturalist* **186**, 755–765.

Civitello, D. J., Cohen, J., Fatima, H., Halstead, N. T., Liriano, J., McMahon, T. A., Ortega, C. N., Sauer, E. L., Sehgal, T., Young, S. and Rohr, J. R. (2015). Biodiversity inhibits parasites: broad evidence for the dilution effect. *Proceedings of the National Academy of Sciences of the United States of America* 112, 8667–8671.

Cleaveland, S., Shaw, D.J., Mfinanga, S.G., Shirima, G., Kazwala, R. R., Eblate, E. and Sharp, M. (2007). *Mycobacterium bovis* in rural Tanzania: risk factors for infection in human and cattle populations. *Tuberculosis* **87**, 30–43.

Cooper, S. M., Scott, H. M., de la Garza, G. R., Deck, A. L. and Cathey, J. C. (2010). Distribution and interspecies contact of feral swine and cattle on rangeland in south Texas: implications for disease transmission. *Journal of Wildlife Disease* **46**, 152–164.

Corner, L. A. L. (2006). The role of wild animal populations in the epidemiology of tuberculosis in domestic animals: how to assess the risk. *Veterinary Microbiology* **112**, 303–312.

Cosivi, O., Meslin, F., Daborn, C. and Grange, J.M. (1995). Epidemiology of *Mycobacterium bovis* infection in animals and humans, with particular reference to Africa. *Scientific and Technical Review of the Office International des Epizooties* **14**, 733–746.

Cosivi, O., Grange, J., Daborn, C., Raviglione, M., Fujikura, T., Cousins, D., Robinson, R., Huchzermeyer, H., Kantor, I. and Meslin, F. (1998). Zoonotic tuberculosis due to *Mycobacterium bovis* in developing countries. *Emerging Infectious Disease* **4**, 59–70.

Courtenay, O., Reilly, L. A., Sweeney, F. P., Hibberd, V., Bryan, S., Ul-Hassan, A., Newman, C., Macdonald, D. W., Delahay, R. J., Wilson, G. J. and Wellington, E. M. H. (2006). Is *Mycobacterium bovis* in the environment important for the persistence of bovine tuberculosis? *Biology Letters* **2**, 460–462.

de Garine-Wichatitsky, M., Caron, A., Kock, R., Tschopp, R., Munyeme, M., Hofmeyr, M. and Michel, A. (2013). Review of bovine tuberculosis at the wildlife-livestock-human interface in sub-Saharan Africa. *Epidemiology and Infection* **147**, 1342–1358.

Dejene, S. W., Heitkönig, I. M. A., Prins, H. H. T., Fitsum, A., Daniel, A., Zelalem, E., Kelkay, Z. T. and de Boer, W. F. (2016). Risk factors for bovine tuberculosis (bTB) in cattle in Ethiopia. *PLoS ONE* **11**, e0159083.

de Leeuw, J., Waweru, M. N., Okello, O. O., Maloba, M., Nguru, P., Said, M.Y., Aligula, H. M., Heitkonig, I. M. A. and Reid, R.S. (2001). Distribution and diversity of wildlife in northern Kenya in relation to livestock and permanent water points. *Biological Conservation* **100**, 297–306.

Di Marco, M., Boitani, L., Mallon, D., Hoffmann, M., Iacucci, A., Meijaard, E., Visconti, P., Schipper, J. and Rondinini, C. (2014). A retrospective evaluation of the global decline of carnivores and ungulates. *Conservation Biology* 28, 1109–1118.

Fenton, A. and Pedersen, A. B. (2005). Community epidemiology framework for classifying disease threats. *Emerging Infectious Diseases* 11, 1815– 1821.

Fine, A. E., Bolin, C. A., Gardiner, J. C. and Kaneene, J. B. (2011). A study of the persistence of *Mycobacterium bovis* in the environment under natural weather conditions in Michigan, USA. *Veterinary Medicine International* **2011**, 1–12.

Fritz, H., De Garine-Wichatitsky, M. and Georges, L. (1996). Habitat use by sympatric wild and domestic herbivores in an African Savanna Woodland: the influence of cattle spatial behaviour. *Journal of Applied Ecology* 33, 589–598.

Frohlich, K., Thiede, S., Kozikowski, T. and Jakob, W. (2002). A review of mutual transmission of important infectious diseases between livestock and wildlife in Europe. *Annals of the New York Academy of Sciences* **969**, 4–13.

Gorenfloa, L. J., Romaineb, S., Russell, A., Mittermeierc, A. R. and Walker-Painemilla, K. (2012). Co-occurrence of linguistic and biological diversity in biodiversity hotspots and high biodiversity wilderness areas. *Proceedings of the National Academy of Sciences of the United States of America* 109, 8032–8037.

Gortazar, C., Ferroglio, E., Hofle, U., Frolich, K. and Vicente, J. (2007). Diseases shared between wildlife and livestock: a European perspective. *European Journal of Wildlife Research* **53**, 241–256.

Gosselin, F. (2006). An assessment of the dependence of evenness indices on species richness. *Journal of Theoretical Biology* 242, 591–597.

Haines-Young, R. and Chopping, M. (1996). Quantifying landscape structure: a review of landscape indices and their application to forested landscapes. *Progress in Physical Geography* **20**, 418–445.

Hamer, G., Chaves, L., Anderson, T., Kitron, U. D., Brawn, J. D., Ruiz, M. O., Loss, S. R., Walker, E. D. and Goldberg, T. L. (2011). Fine-scale variation in vector host use and force of infection drive localized patterns of West Nile Virus transmission. *PLoS ONE* **6**, e23767.

Hantsch, L., Braun, U., Scherer-Lorenzen, M. and Bruelheide, H. (2013). Species richness and species identity effects on occurrence of foliar fungal pathogens in a tree diversity experiment. *Ecosphere* **4**, 1–12. Hill, M. (1973). Diversity and evenness: a unifying notation and its consequences. *Ecology* **54**, 427–432.

Hofmeester, T. R., Coipan, E. C., van Wieren, S. E., Prins, H. H. T., Takken, W. and Sprong, H. (2016). Few vertebrate species dominate the *Borrelia burgdorferi s.l.* life cycle. *Environmental Research Letters* **11**, 1–16. Huang, Z. Y. X., de Boer, W. F., van Langevelde, F., Xu, C., Ben Jebara, K., Berlingieri, F. and Prins, H. H. T. (2013). Dilution effect in bovine tuberculosis: risk factors for regional disease occurrence in Africa. *Proceedings of the Royal Society B – Biological Sciences* **280**, 1–6.

Huang, Z.Y.X., Xu, C., van Langevelde, F., Prins, H. H.T., ben Jebara, K. and de Boer, W.F. (2014). Dilution effect and identity effect by wildlife in the persistence and recurrence of bovine tuberculosis. *Parasitology* **141**, 981–987.

Huang, Z. Y. X., van Langevelde, F., Estrada-Peña, A., Suzán, G. and de Boer, W. F. (2016). The diversity-disease relationship: evidence for and criticisms of the dilution effect. *Parasitology* **143**, 1075–1086.

Hudson, P. J., Rizzoli, A. P., Grenfell, B. T., Heesterbeek, J. P. and Dobson, A. P. (2002). *Ecology of Wildlife Diseases*. Oxford University Press, Oxford, UK.

Humblet, M.F., Boschiroli, M.L. and Saegerman, C. (2009). Classification of worldwide bovine tuberculosis risk factors in cattle: a stratified approach. *Veterinary Research* **40**, 50–63.

Jackson, R., De Lisle, G.W. and Morris, R.S. (1995). A study of the environmental survival of *Mycobacterium bovis* on a farm in New Zealand. *New Zealand Veterinary Journal* **43**, 346–352.

Johnson, P. T., Ostfeld, R. S. and Keesing, F. (2015). Frontiers in research on biodiversity and disease. *Ecology Letters* 18, 1119–1133.

Johnson, P. T. J., Preston, D. L., Hoverman, J. T. and Richgels, K. L. D. (2013). Biodiversity decreases disease through predictable changes in host community competence. *Nature* **494**, 230–233.

Keesing, F., Holt, R.D. and Ostfeld, R.S. (2006). Effects of species diversity on disease risk. *Ecology Letters* 9, 485–498.

Keesing, F., Brunner, J., Duerr, S., Killilea, M., LoGiudice, K., Schmidt, K., Vuong, H. and Ostfeld, R. (2009). Hosts as ecological traps for the vector of Lyme disease. *Proceedings of the Royal Society B* – *Biological Sciences* 276, 3911–3919.

Keesing, F., Belden, L. K., Daszak, P., Dobson, A., Harvell, C. D., Holt, R. D., Hudson, P., Jolles, A., Jones, K. E., Mitchell, C. E., Myers, S. S., Bogich, T. and Ostfeld, R. S. (2010). Impacts of biodiversity on the emergence and transmission of infectious diseases. *Nature* 468, 647–652. Kelly, W. R. and Collins, J. D. (1978). The health significance of some infectious agents present in animal effluents. *Veterinary Science Communications* 2, 95–103.

Legendre, P. and Legendre, L. (1998). Numerical Ecology. Elsevier, Oxford.

LoGiudice, K., Ostfeld, R. S., Schmidt, K. A. and Keesing, F. (2003). The ecology of infectious disease: effects of host diversity and community composition on Lyme disease risk. *Proceedings of the National Academy of Sciences of the United States of America* **100**, 567–571.

Magurran, A.E. (1988). Ecological Diversity and its Measurement. Princeton University Press, Princeton, USA.

Magurran, A.E. (2004). *Measuring Biological Diversity*. Blackwell, Oxford.

Maleko, D.D., Mbassa, G.N., Maanga, W.F. and Sisya, E.S. (2012). Impacts of Wildlife-Livestock Interactions in and around Arusha National Park, Tanzania. *Current Research Journal of Biological Sciences* **4**, 471–476.

Martin, C., Pastoret, P.P., Brochier, B., Humblet, M.F. and Saegerman, C. (2011). A survey of the transmission of infectious diseases/infections between wild and domestic ungulates in Europe. *Veterinary Research* 42, 70–81.

McGarigal, K. and Marks, B. J. (1994). FRAGSTATS: Spatial Analysis Program for Quantifying Landscape Structure. Unpublished report, Oregon State University, Portland, Oregon, USA.

Miller, E. and Huppert, A. (2013). The effects of host diversity on vector borne disease: the conditions under which diversity will amplify or dilute the disease risk. *PLoS ONE* **8**, e80279.

Morens, M. D., Folkers, K. G. and Fauci, S. A. (2004). The challenge of emerging and re-emerging infectious diseases. *Nature* **430**, 242–249.

Munyeme, M., Muma, J. B., Skjerve, E., Nambota, A. M., Phiri, I. G., Samui, K. L., Dorny, P. and Tryland, M. (2008). Risk factors associated with bovine tuberculosis in traditional cattle of the livestock/wildlife interface areas in the Kafue basin of Zambia. *Preventive Veterinary Medicine* **85**, 317–328.

Munyeme, M., Muma, J., Samui, K., Skjerve, E., Nambota, A., Phiri, I., Rigouts, L. and Tryland, M. (2009). Prevalence of bovine tuberculosis and animal level risk factors for indigenous cattle under different grazing strategies in the livestock/wildlife interface areas of Zambia. *Tropical Animal Health and Production* **41**, 345–352.

Myers, N., Mittermeier, R. A., Mittermeier, C. G., Da Fonseca, G. A. and Kent, J. (2000). Biodiversity hotspots for conservation priorities. *Nature* 403, 853–858.

Myersa, S. S., Gaffikin, L., Golden, C. D., Ostfeld, R. S., Redford, K. H., Ricketts, T. H., Turner, W. R. and Osofsky, S. A. (2013). Human health impacts of ecosystem alteration. *Proceedings of the National Academy of Sciences of the United States of America* **110**, 18753–18760.

Oda, E., Solari, A. and Botto-mahan, C. (2014). Effects of mammal host diversity and density on the infection level of *Trypanosoma cruzi* in sylvatic kissing bugs. *Medical and Veterinary Entomology* **28**, 384–390.

Odadi, W. O., Young, T. P. and Okeyo-Owuor, J. B. (2007). Effects of wildlife on cattle diets in Laikipia rangeland, Kenya. *Rangeland Ecology* & *Management* 60, 179–185.

Oksanen, J., Guillaume, F. B., Roeland, K., Pierre, L., Minchin, P. R., O'Hara, R. B., Simpson, L. G., Peter, S., Henry, M., Stevens, H. and Helene, W. (2016). Community Ecology Package. Package 'vegan. http://CRAN.R-project.

Olff, H., Ritchie, M. E. and Prins, H. H. T. (2002). Global environmental controls of diversity in large herbivores. *Nature* **415**, 901–904.

Osofsky, S. A., Cleaveland, S., Karesh, W. B., Kock, M. D., Nyhus, P. J., Starr, L. and Yang, A. (eds) (2005). Conservation and Development Interventions at the Wildlife/Livestock Interface: Implications for Wildlife, Livestock and Human Health. IUCN, Gland, Switzerland and

Ostfeld, R. and Keesing, F. (2000). The function of biodiversity in the ecology of vector-borne zoonotic diseases. *Canadian Journal of Zoology* 78, 2061–2078.

Cambridge, UK.

Ostfeld, R. S. (2013). A Candide response to Panglossian accusations by Randolph and Dobson: biodiversity buffers disease. *Parasitology* **140**(10), 1196–1198.

Ostfeld, R. S. and Keesing, F. (2012). Effects of host diversity on infectious disease. Annual Review of Ecology, Evolution, and Systematics 43, 157–182.

Pianka, E. R. (1973). The structure of lizard communities. *Annual Review* of Ecology and Systematics 4, 53–74.

Power, A. G. and Mitchell, C. E. (2004). Pathogen spillover in disease epidemics. *The American Naturalist* 164, 79–89.

Prins, H. H. T. (2000). Competition between wildlife and livestock in Africa. In *Wildlife Conservation by Sustainable Use* (ed. Prins, H. H. T., Grootenhuis, J. G. and Dolan, T. T.), pp 51–80. Kluwer Academic Publishers. Norwell.

Proffitt, K. M., Gude, J. A., Hamlin, K. L., Garrott, R. A., Cunningham, J. A. and Grigg, J. L. (2011). Elk distribution and spatial overlap with livestock during the brucellosis transmission risk period. *Journal of Applied Ecology* **48**, 471–478.

R Core Team (2015). *R: a Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria. http://R-project.org/.

Randolph, S. E. and Dobson, A. D. M. (2012). Pangloss revisited: a critique of the dilution effect and the biodiversity-buffers-disease paradigm. *Parasitology* **139**, 847–863.

Renwick, A. R., White, P. C. L. and Bengis, R. G. (2007). Bovine tuberculosis in southern Africa wildlife: a multi-species host-pathogen system. *Epidemiology and Infection* **135**, 529–540.

Riitters, K. H., Wickham, J. D., Vogelmann, J. E. and Jones, K. B. (2000). National land-cover pattern data. *Ecology* **81**, 604–612.

Riley, S.P.D., Hadidian, J. and Manski, D.A. (1998). Population density, survival, and rabies in raccoons in an urban national park. *Canadian Journal of Zoology* **76**, 1153–1164.

Roper, T. J., Garnett, B. T. and Delahay, R. J. (2003). Visits to farm buildings and cattle troughs by badgers (*Meles meles*): a potential route for transmission of bovine tuberculosis (*Mycobacterium bovis*) between badgers and cattle. *Cattle Practice* **11**, 9–12.

Roug, A., Clifford, D., Mazet, J., Kazwala, R., John, J., Coppolillo, P. and Smith, W. (2014). Spatial predictors of bovine tuberculosis infection and *Brucella* spp. exposure in pastoralist and agro pastoralist livestock

herds in the Ruaha ecosystem of Tanzania. Tropical Animal Health and Production 46, 837-843.

Sheldon, A. L. (1969). Equitability indices: dependence on the species count. *Ecology* **50**, 466–467.

Sitters, J., Heitkönig, I. M. A., Holmgren, M. and Ojwang, G. S. O. (2009). Herded cattle and wild grazers partition water but share forage resources during dry years in East African savannas. *Biological Conservation* **142**, 738–750.

Skuce, R. A., Allen, A. R. and Stanley, W. (2012). Herd level risk factors for bovine tuberculosis. *Veterinary Medicine International* **2012**, 1–10.

Smith, B. and Wilson, J.B. (1996). A consumer's guide to evenness indices, Oikos 1, 70-82.

Stuart, C. and Stuart, T. (2000). A Field Guide to the Tracks and Signs of Southern and East African Wildlife. Struik Publishers, Cape Town.

Symonds, M.R.E. and Johnson, C.N. (2008). Species richness and evenness in Australian birds. *The American Naturalist* **171**, 480–490.

Treydte, A. C., Bernasconi, S. M., Kreuzer, M. and Edwards, P. J. (2006). Diet of the common warthog (*Phacochoerus africanus*) on former cattle grounds in a Tanzanian savanna. *Journal of Mammalogy* **87**, 889–898.

Tschopp, R. (2015). Bovine tuberculosis at the human–livestock–wildlife interface in sub–Saharan Africa. In *One Health: The Theory and Practice of Integrated Health Approaches* (ed. Zinsstag, J., Schelling, E., Waltner-Toews, D., Whittaker, M. and Tanner, M.), pp. 163–175. CABI, Wallingford.

Tucker, C. M. and Cadotte, M. W. (2013). Unifying measures of biodiversity: understanding when richness and phylogenetic diversity should be congruent. *Diversity and Distributions* **19**, 845–854.

Vicente, J., Segalés, J., Balasch, M., Plana-Durán, J., Domingo, M. and Gortázar, C. (2004). Epidemiological study on porcine circovirus type 2 (PCV2) infection in the European wild boar (*Sus scrofa*). *Veterinary Research* **35**, 243–253.

Voeten, M. M. and Prins, H. T. T. (1999). Resource partitioning between sympatric wild and domestic herbivores in the Tarangire region of Tanzania. *Oecologia* **120**, 287–294.

Ward, A. I., Tolhurst, B. A. and Delahay, R. J. (2006). Farm husbandry and the risks of disease transmission between wild and domestic mammals: a brief review focusing on bovine tuberculosis in badgers and cattle. *Animal Science* **82**, 767–773.

Wood, C. L. and Lafferty, K. D. (2013). Biodiversity and disease: a synthesis of ecological perspectives on Lyme disease transmission. *Trends in Ecology and Evolution* 28, 239–247.

World Health Organization (2012). Global tuberculosis report 2012. http://www.stop.org/wg/news_vaccines/

Zhang, H., John, R., Peng, Z., Yuan, J., Chu, C., Guozhen, D. and Shurong, Z. (2012). The relationship between species richness and evenness in plant communities along a successional gradient: a study from Sub-Alpine Meadows of the Eastern Qinghai-Tibetan Plateau, China. *PLoS ONE* 7, e49024.