Ketogenic diets and the nervous system: a scoping review of neurological outcomes from nutritional ketosis in animal studies

Rowena Field¹* ^(b), Tara Field², Fereshteh Pourkazemi¹ and Kieron Rooney¹ ¹*The University of Sydney, Faculty of Medicine and Health, Sydney, Australia* ²*The New South Wales Ministry of Health (NSW Health), Sydney, Australia*

Abstract

Objectives: Ketogenic diets have reported efficacy for neurological dysfunctions; however, there are limited published human clinical trials elucidating the mechanisms by which nutritional ketosis produces therapeutic effects. The purpose of this present study was to investigate animal models that report variations in nervous system function by changing from a standard animal diet to a ketogenic diet, synthesise these into broad themes, and compare these with mechanisms reported as targets in pain neuroscience to inform human chronic pain trials.

Methods: An electronic search of seven databases was conducted in July 2020. Two independent reviewers screened studies for eligibility, and descriptive outcomes relating to nervous system function were extracted for a thematic analysis, then synthesised into broad themes.

Results: In total, 170 studies from eighteen different disease models were identified and grouped into fourteen broad themes: alterations in cellular energetics and metabolism, biochemical, cortical excitability, epigenetic regulation, mitochondrial function, neuroinflammation, neuroplasticity, neuroprotection, neurotransmitter function, nociception, redox balance, signalling pathways, synaptic transmission and vascular supply.

Discussion: The mechanisms presented centred around the reduction of inflammation and oxidative stress as well as a reduction in nervous system excitability. Given the multiple potential mechanisms presented, it is likely that many of these are involved synergistically and undergo adaptive processes within the human body, and controlled animal models that limit the investigation to a particular pathway in isolation may reach differing conclusions. Attention is required when translating this information to human chronic pain populations owing to the limitations outlined from the animal research.

Keywords: Ketogenic diet: Animal: Neurological: Beta-hydroxybutyrate: Inflammation

(Received 11 February 2021; revised 17 May 2021; accepted 7 June 2021; accepted manuscript published online 28 June 2021)

Introduction

Nutrition Research Reviews

Nutrition is directly responsible for the delivery of energy for cellular metabolism, as well as providing the diverse array of substrates required for normal physiological function. It also plays a central role in the modulation of inflammatory and disease processes^(1–3) and, thus, can be utilised as a therapeutic intervention. Nutritional therapies that limit substrate availability and produce ketosis (such as fasting, calorie restriction and ketogenic diets) directly impact metabolism and cellular energetics^(2,4). Ketosis has been reported to be effective in neurological conditions characterised by neurodegeneration^(5–9), psychological disorders⁽¹⁰⁾, brain injury^(6,11) and nervous system excitability^(12–15). More recently, the presence of ketones has been suggested to influence pain mechanisms^(16–18). Given this, ketosis produced through a ketogenic diet may be an appropriate treatment strategy for persistent pain, a dysfunction within the nervous system involving changes in both cortical structure and function^(19,20). Neuroplastic remodelling facilitates increased connectivity and amplification of pain perception and is required to shift into a persistent pain state⁽²¹⁾. Broadly, nutritional interventions have been shown to improve pain outcomes^(22–24). Directly targeting neurobiology through a ketogenic diet could potentially modulate maladaptive change and become an additional strategy to add to comprehensive chronic pain management⁽²⁵⁾.

The concept of nutritional neurobiology for chronic pain management is starting to appear in the literature, where dietary intake can be both a trigger for upregulated pain mechanisms but also potentially provide therapeutic options^(25,26). There have been three systematic reviews published to date^(22–24) that report outcomes on human participants with chronic pain from dietary interventions, all published in the last 2 years. These reviews report the effectiveness of improved nutrition generally as a pain management option, particularly when considering nutrientdense whole-food diets and the removal of discretionary ultraprocessed foods high in sugar and fat. They are unable to clearly point to any specific diet as the best treatment, however. A recent review suggested both the Mediterranean diet and a carbohydrate-restricted diet were promising diets for reducing the impact of chronic pain by either a reduction in inflammation or a reduction in oxidative stress⁽²⁶⁾. The authors note, however, that only two studies reviewed were specifically assessing the context of chronic pain (knee osteoarthritis) and the rest were examining participants with metabolic dysregulation (such as elevated cardiovascular risk or obesity).

Nutritional ketosis is achieved through a ketogenic diet by restricting dietary carbohydrates sufficiently to shift cellular energetics from glucose to fat oxidation as the main fuel source⁽²⁷⁾. Ketone bodies (β-hydroxybutyrate and acetoacetate) are produced in the liver (ketosis) and delivered via the bloodstream as part of this alternate fuel pathway, providing both a fuel source and a signalling molecule that can modulate many physiological processes⁽⁶⁾. As a signalling molecule, β -hydroxybutyrate is a metabolic intermediary that can act as an endogenous class I and II histone deacetylase inhibitor involved in the regulation of longevity and antioxidant defences, diseases of aging, and also diabetes and cancer^(10,28,29). It acts as a ligand for G-protein-coupled receptors (hydroxycarboxylic acid receptor 2) and free fatty acid receptor 3, which bind short-chain fatty acids, regulate metabolism and play a role in the development of metabolic disease states⁽²⁸⁾. Ketone signalling via a ketogenic diet has been reported to beneficially effect physiological processes involved in many disease conditions, including obesity, cancer, diabetes, epilepsy, Parkinson's disease, Alzheimer's disease, multiple sclerosis, peripheral neuropathy, liver disease, inherited metabolic disorders, muscle degeneration, polycystic ovarian syndrome, irritable bowel syndrome, migraine and fibromyalgia^(12,30).

Whilst ketogenic diets have reported efficacy clinically for humans in a variety of neurological conditions, the evidence for plausible physiological mechanisms by which the nervous system may be modulated relies heavily on animal models (both *in vivo* and *in vitro*) to explore the mechanistic pathways. The mechanisms suggested are neuroprotective and neuromodulatory, whereby decreasing glycolytic metabolism and shifting to fat oxidation raises ATP and adenosine levels and improves cellular energetics. It also activates multiple signalling pathways involved in the reduction of reactive oxygen species in neurological tissues, increased mitochondrial number and function, synaptic regulation, and inhibition of pro-inflammatory cytokine mediators^(18,31-35). The overall effect would seem to be restoring homeostatic synaptic function and excitability.

To date, there is limited published literature on human clinical trials that examine a ketogenic diet as a treatment for chronic pain. The purpose of this scoping review was to investigate animal models that report outcomes related to the nervous system by changing from a standard animal diet to a ketogenic diet. It includes multiple models of nervous system dysfunction and synthesises the outcomes presented into broader themes by which a ketogenic diet may plausibly modulate biological pathways associated with human chronic pain perception. It also discusses the potential issues with clinical translation from animal models to human models of dietary interventions.

Methods

Protocol

The framework for this review was based on relevant items of the scoping review protocol and PRISMA-ScR checklist from the Joanna Briggs Institute^(36,37) to answer the research question: 'How does a ketogenic diet in animal models influence the nervous system?'

Eligibility criteria

Studies were included if they met the following criteria:

- 1. Mammal models that report an *ad libitum* high-fat, lowcarbohydrate ketogenic diet that is ≥ 7 d (% energy from fat ≥ 69 % or 3:1 ratio of fat:protein + carbohydrate + fibre + extras) as the intervention. The minimum diet length of 1 week was used to ensure all studies were captured, and was based on similar systematic reviews reporting studies where the minimum reported length of diet was 2 weeks in rat and mouse models assessing both metabolic and nervous system outcomes^(38,39).
- Studies that report objective outcomes related to nervous system function including neuroinflammation.
- Experimental study designs: longitudinal pre-post intervention trials including randomised controlled trials.

Studies were excluded if:

- 1. The diets were both high in fat and carbohydrate, carbohydrate levels exceeded 10 % or where the chow was not described, and the ketogenic status could not be confirmed.
- 2. The diet was not *ad libitum* or provided in the form of whole food, including oral gavage, intraperitoneal models, food extracts or exogenous ketones.
- 3. The subjects were human or in vitro cultures.
- 4. The model used represented cancer or genetic syndromes.
- 5. The paper was not in English.

Information sources and search strategy

An electronic database search including Medline, EMBASE, Cochrane Library for controlled trials, AMED via OVID, CINAHL via Ebsco, Web of Science and PubMed was carried out on 5 July 2020 and included dates from database inception to the search date. A preliminary search refined the search strategy, with the key terms outlined in Supplementary Table 1. Additional searches included a Google Scholar search to check identified articles 'cited by' and 'related articles' links, and reference checks on identified articles with subsequent hand search for these and inclusion if they met the criteria. Retrieved references were downloaded into EndNote reference management software (Endnote X7.7.1, Thomson Reuters 2016) and then imported using Covidence systematic review software (Veritas Health Innovation, Melbourne, Australia). 270

Study selection and screening

Duplicates were removed, then titles and abstracts were assessed in Covidence by two reviewers independently (R.F. and T.F) against the eligibility criteria. Full texts of identified studies were then screened by two reviewers independently (R.F. and T.F) for final eligibility, with any disagreements resolved by a third reviewer.

Data items

The primary outcomes of interest were changes in nervous system function (such as excitability) or energetics (such as altered substrate measured by blood glucose or ketones levels) that report a plausible biological mechanism by which a ketogenic diet may influence the nervous system. Additional data extracted included: author, year of study, animal, animal variant, chow ratios, disease model and intervention diet length. Critical appraisal of the literature to assess risk of bias was not carried out due to the frequently poor-quality methods employed in animal research. This includes lack of randomisation, lack of blinding and incorrect statistical methods^(40,41). Nervous system outcomes were taken as presented by the study authors.

Data charting process and synthesis of results

Data items were extracted and compiled in an excel spreadsheet. Primary outcomes were reviewed, and a subjective thematic analysis was carried out by R.F. The process of thematic analysis involved building a list of categories that best fit the outcome description given by the study author. These were then further synthesised into broad themes. Studies with more than one relevant theme could be allocated into more than one theme. A random sample of thirty-five studies (20 %) was independently reviewed by T.F to ensure consistency of theme allocation.

Results

A systematic search of the databases retrieved 7045 studies screened for eligibility after duplicates were removed. A total of 341 full-text articles were assessed with a total of 170 meeting the inclusion criteria and included in the scoping review (Fig. 1). Of the ninety-nine studies excluded for being either >10 % carbohydrate or ≤ 69 % fat, only three studies were described as lower in ketogenic (but still included 32 % carbohydrate, 20 % carbohydrate or 30 % fat). The remaining studies were captured by the search term 'high fat', which retrieved studies high in both fat and carbohydrate designed to produce obesity or metabolic dysfunction.

Characteristics of included studies

The studies comprised 103 rat studies, 63 mouse studies, 2 that included rats and mice, and 1 that included rats and gerbils as well as a canine case study. There was a range of nervous system dys-function models, including five age-related degeneration⁽⁴²⁻⁴⁶⁾, four Alzheimer's disease^(47–50), seven autism^(51–57), four cerebral ischaemia^(58–61), two pain perception^(16,62), twenty-four general central nervous system^(63–86), two diabetes^(87,88), ninety-one epilepsy^(89–179), two metabolic syndrome^(180,181), one mild cognitive

impairment⁽¹⁸²⁾, two multiple sclerosis^(183,184), one nerve toxin⁽¹⁸⁵⁾, four optic nerve dysfunction^(186–189), two Parkinson's disease^(190,191), three peripheral nerve dysfunction^(192–194), four spinal cord injury^(195–198), three stroke^(199–201) and nine traumatic brain injury^(202–210). The length of the dietary intervention ranged from 1 week to 6 months.

Fourteen broad themes involving nervous system function were identified. These themes, the disease models used, and further details are presented in Table 1. Detailed information on individual study characteristics and reported outcomes is compiled in Supplementary Table 2, which references all 170 included studies.

- 1. Alterations in cellular energetics and metabolism (reported in twenty-eight studies across nine disease models^(42,43,46,49,63,68,70,72,74-76,83,85,87,91,95,97,108,125,136,142,156, 187,188,192,197,202,207)). The reduced glucose consumption of a ketogenic diet resulted in lower glucose availability within the nervous system and a shift to fat-based metabolism with up-regulation of processes required to deliver this alternate energy substrate. Fat-based metabolism was reported to improve energy availability, utilisation and efficiency. It was also reported to reduce low-grade inflammation driven by a low energy state.
- 2. *Biochemical (reported in three studies in epilepsy models*⁽¹⁰²⁻¹⁰⁴⁾). Elemental changes (P, S, K, Ca, Fe, Cu, Zn and Se) within the hippocampus were assessed via X-ray fluorescence microscopy with significant changes, with a significant decrease in P, K and Zn, and a significant increase in Ca and Se as a result of the ketogenic diet. As hippocampal levels of Ca increase with seizures, these changes did not provide evidence supporting a mechanism for seizure reduction. Additionally, the ratio of absorbance for specific biological macromolecules (such as ketones and lipids) was increased with the possibility of these molecules being involved in anti-seizure mechanism rather than elemental changes.
- 3. Cortical/neuronal excitability (reported in fifty-three studies of which forty-nine were epilepsy models^(53,57,67,89–94,104,107,109–111,113–116,118,124,125,127,129,131,135,140–142,144,150–152,154–156,158–164,166,167, 170–173,175,177–179,209)). The ketogenic diet was broadly reported to restore the balance of nervous system excitability toward homeostatic levels; however, some studies reported neutral or negative findings^(91,93,140,150,151,154,166,175,178). This category was largely composed of epilepsy models that described reductions in frequency, threshold, duration, latency and spread of seizures. Restoration of circadian rhythms within the brain was also reported.
- 4. *Epigenetic regulation (reported in thirty studies across nine disease models*^{(44,55,61,69,72,76,95,97,99,100,105,106,108,122,127,132,135, 145,146,149,157,165,172,174,176,177,181,182,192,208)). The genes reportedly altered by the ketogenic diet generally pertained to the disease model being investigated. Overall, they tended to up-regulate beneficial genetic expression regarding neuroinflammation, neurodegeneration and neuroprotection.}
- 5. *Mitochondrial function (reported in eighteen studies across ten disease models*^(45,51,52,76,78,97,117,120,134,173,180,184,187,192, 199,203–205)). The overall reported benefit to the mitochondria

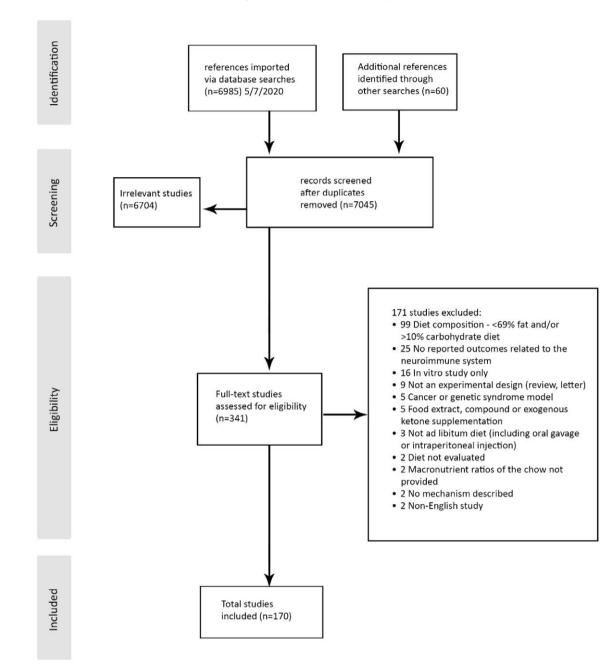


Fig. 1. Inclusion flowchart.

within the nervous system was positive, with increases in number, and improvements in structure and function including energy production and redox balance.

- 6. Neuroinflammation (reported in seven studies across six disease models^(123,183,188,191,199,204,206)). Ketones were reported to inhibit the NOD-, LRR- and pyrin domain-containing protein 3 (NLRP3) inflammasome expressed in the nervous system and subsequent reduction of the downstream inflammatory signalling pathways it generates. Neuroinflammation was also reportedly reduced through a reduction in reactive oxygen species. Neuroinflammation was frequently reported in terms of signalling pathways, so many of the relevant studies were reported in theme 12.
- Neuroplasticity and structural integrity (reported in twentysix studies across ten disease models^(45,47,50,54,73,77,79,80,124, 130,133,134,141,143,144,147,183,184,186,189,193,194,197,199-201)). Improved

synaptic plasticity (long-term potentiation) and a reduction of maladaptive plasticity (such as mossy fibre sprouting in epilepsy models) was reported on a ketogenic diet. Other structural changes reported across a range of disease models included: improved myelin formation, reduced axonal degeneration, improved white matter development, reduction in β -amyloid, increased neuronal progenitor cells following seizure, prevention of neuronal loss in the ipsilateral hippocampus, reversal of hippocampal atrophy and lesions, improved neuronal recovery following insult 272

when the diet commenced pre-injury, and reduction in retinal ganglion cell loss.

- 8. *Neuroprotection (reported in six studies across four disease models*^(58,59,88,185,190,191)). A reduction of neuronal apoptosis and neuronal death was reported as a result of the diet in a variety of disease models. Protection against seizure was commonly reported in the epilepsy models and reported in theme 3.
- 9. Neurotransmitter function (reported in ten studies across four disease models^(45,64,70,74,85,96,98,101,153,190)). Various mechanisms around neurotransmitter production and clearance were reported with proposed benefit from the diet being an improved GABA levels or GABA-to-glutamate ratio. Parkinson's disease models reported improvements around dopamine levels.
- Nociception (reported in three studies, two for pain and one for peripheral nerve dysfunction^(16,62,193)). A reduction in both allodynia and thermal pain sensitivity was reported that was not dependent on lowered glucose levels.
- 11. *Redox balance (reported in fifteen studies across eight disease models*^(63,71,78,86,92,120,180,181,186,190,192,195,196,203,210)). Several studies found an improvement in redox balance through either a reduction in nervous system reactive oxygen species or an increase in antioxidant defence.
- 12. Signalling pathways (reported in thirty-six studies across eight disease models ^{(48,54,56,60,61,63,65,66,69,71,82,84,112,119,121,123, 125,126,131,135,137–139,143,148,149,161,164,168,169,187,196,198–201)). A variety of signalling pathways were reported depending on the disease model being used. These centred around other key mechanisms such as reduced neuroinflammation, reduced oxidative stress, altered neuronal energy metabolism, reduced cortical excitability and reduced neurodegeneration.}
- 13. Synaptic transmission (reported in seven studies across three disease models^(42,44,67,95,97,128,178)). Improved clearance and levels of protein transporters for neurotransmitters was reported to improve synaptic transmission. Cortical excitability was described as improved due to a reduction in long-term potentiation, without any change in baseline excitability or impact on normal brain activity. Not all studies noted reduced long-term potentiation⁽⁶⁷⁾.
- 14. Vascular supply (reported in three studies across three disease models^(48,61,81)). The size of cerebral infarct and oedema was reduced with a ketogenic diet. Alzheimer's models reported increased blood flow providing positive outcomes. In epilepsy, positive outcomes due to a decrease in capillarisation associated with seizures were also reported.

Discussion

The aim of this scoping review was to investigate animal models that report outcomes related to the nervous system by changing from a standard animal diet to a ketogenic diet. We identified fourteen broad themes of biological mechanisms from eighteen different disease models by which a ketogenic diet is reported to influence the nervous system in animal models (Table 1). Multiple themes may be present within a single study, with many of the different mechanisms and pathways reported resulting in similar overarching effects, including reduction of inflammation and oxidative stress, normalisation of neuronal excitability and improved cell viability. The themes outlined are consistent with other broader reviews that included *in vitro* and hypothetical models^(211,212). The purpose of describing these themes was to provide insight into how altering dietary macronutrients to produce ketosis in humans could also plausibly exert influence on the nervous system in a chronic pain model. The ketogenic diet appears to utilise metabolic modulation to engage the reported mechanisms in animal studies, and thus could also potentially facilitate positive changes within a human nervous system that has undergone aberrant neuroplasticity leading to a persistent pain state.

There are many mechanisms presented that fit with current priorities in pain neuroscience research, such as targeting inflammation. An increase in pro-inflammatory cytokines is often seen in chronic lifestyle disease⁽²¹³⁾, but also frequently occurs with chronic pain^(214,215). The failure of the inflammatory response to resolve perpetuates the development of metabolic diseases, but also potentially contributes to persisting pain by shifting the nervous system towards a pathologically maladapted state⁽²¹⁶⁾. Neuroinflammation is a common finding in many neurological conditions and was frequently reported in the outcomes from the extracted studies. Modulation of neuroinflammation across various models from the ketogenic diet was attributed to as many as nine mechanisms (Supplementary Table 2): (a) suppression of nuclear factor (NF)-kß resulting in reduced expression of proinflammatory cytokines tumour necrosis factor α (TNF- α), interleukin (IL)-1 β and interferon (IFN)- $\gamma^{(196)}$; (b) a decrease in hippocampal mRNA levels of IL-16(106); (c) reduced pro-inflammatory cytokine hippocampal TNF- α levels with reduced NF-k β dependant cyclooxygenase (COX)-2 (enzyme for prostaglandin synthesis) signalling pathway⁽¹²³⁾; (d) activation of the peroxisome proliferator-activated receptor (PPAR)-y⁽¹⁶¹⁾ (a nuclear transcription factor involved in detecting and metabolising lipids) which also suppresses the COX-2 dependant pathway⁽¹²³⁾ and regulates catalyse expression⁽¹²⁶⁾; (e) central and peripheral suppression of inflammatory cytokines/chemokines coupled with a reduction in reactive oxygen species (ROS)⁽¹⁸³⁾; (f) meeting the cellular energy demand which inhibits AMP-activated protein kinase (AMPK) (which senses and regulates cellular energy levels) and reduces low-energy facilitated inflammation⁽¹⁸⁸⁾; (g) inhibition of the NOD-, LRR- and pyrin domain-containing protein 3 (NLRP3) inflammasome through ketone action on hydroxycarboxylic acid receptor 1 (HCAR1)^(188,196); (h) altered NAD+/ NADH ratio (which is coupled to glycolysis) and regulates inflammation⁽⁶³⁾; and (i) reduced mitochondrial ROS production⁽¹⁹⁹⁾. The use of a ketogenic diet for chronic pain management could be theoretically targeting any of these mechanisms to lower inflammation and reduce pain perception(4,18,217), and is supported mechanistically by the outcomes from animal research.

Mitochondrial pathology is another theme presented that has been implicated in central sensitisation seen in chronic pain, with dysfunctional mitochondria observed in the muscle cells of fibromyalgia patients⁽²¹⁸⁾, and two recent studies reporting between 67 % and 91 % of patients with mitochondrial diseases also reporting chronic pain^(219,220). Given this, strategies to restore or optimise mitochondrial function would be an

Table 1. Overall themes presented for beneficial ketogenic diet outcomes

Models	Mechanistic theme	Detail
ARD, ALZ, CNS, D, EP, ON, PND, SCI, TBI	Cellular energetics and metabolism	 Reduction in glucose availability with concomitant increase in alternate fuel substrates (ketones, lactate, glutamate) Adaptive regulation of energy transporters reflecting change in fuels (reduction in glucose transporters and increase in ketone transporters) Improved cerebral energy metabolism, utilisation and efficiency
		Increased energy reserves/ATP
		Increased seizure resistance when combined with calorie restriction
50		Reduction in maladaptation driven by low-energy-facilitated inflammation
	Biochemical	Increased calcium selenium, decreased phosphorus, potassium and zinc areas of hippocampus
AUT, CNS, EP, TBI	Cortical/neuronal excitability	Balance of excitation and inhibition restored towards more normal levels
		 Reduction in seizure events, threshold, duration, intensity, latency and spreading. Delayed progression of seizure stage
		Restoration of circadian rhythms
ARD, AUT, CI, CNS, EP, MetS, MCI, PND,	Epigenetic regulation	Modulation of genes for neurotransmitter production and function, synaptic transmission and neuroplasticity
TBI		Altered mitochondrial gene expression favouring biogenesis, improved function and efficiency
		Up-regulation of genes for brain adaptation following ischaemia
		Up-regulation of differentially regulated transcripts encoding energy metabolism enzymes
		Up-regulation of intracellular signal transduction pathways
		Modulation of genes for inflammatory signalling pathways favouring anti-inflammation
		Down-regulation of genes related to apoptosis and neuronal death
		 Altered gene expression for various factors related to seizure production, ameliorated seizure-induced DNA methylation
		Down-regulation of genes related to neurodegeneration
ARD, AUT, CNS, EP, MetS, MS, ON, PND,	Mitochondria	Increased biogenesis and mass
ST, TBI		 Improved bioenergetic profile, oxygen consumption and maximal respiration rates
		Improved mitochondrial antioxidant defence
		Improved mitochondrial autophagy
		 Decreased percentage of damaged mitochondria post-seizure with increased expression of autophagy proteins and decreased apoptosis
EP, MS, ON, PKD, ST, TBI	Neuroinflammation	Suppression of inflammatory cytokines/chemokines and reactive oxygen species
		Inhibition of the NLRP3 inflammasome
ADD ALZ AUT ONO ED MO ON DND		Reduced oedema
ARD, ALZ, AUT, CNS, EP, MS, ON, PND,	Neuroplasticity/structural integrity	Improved hippocampal synaptic plasticity
SCI, ST		Improved myelin formation and white matter development
		 Neuroanatomical differences with prenatal exposure Prevention of neuronal loss
		Increased neuronal recovery post-seizure
		Reduced supragranular mossy fibre sprouting in epilepsy models
CI, D, NT, PKD,	Neuroprotection	Reduced likelihood of seizure.
0, 0, 1, 11, 110,	Neuroprotection	Elimination of post-ischaemia hippocampal neurodegeneration
		Reduced neuronal death
		Attenuated toxicity from a neurotoxin
ARD, CNS, EP, PKD	Neurotransmitter function	Improved GABA-to-glutamate ratio
		Increased dopamine activity in the motor and somatosensory cortex
		Altered gut biome resulting in systemic GABA and elevated hippocampal GABA/glutamate levels
		Inhibited decrease of striatal dopamine and metabolites
CP, PND	Nociception	Decreased thermal pain sensitivity
		Protection from allodynia
CNS, EP, MetS, ON, PKD, PND, SCI, TBI	Redox balance	Improved redox state
		Improved neurochemical metabolite ratios

Ketogenic diets and the nervous system

Reviews
Research
Nutrition
\$

ŝ

Table 1. (Continued)		
Models	Mechanistic theme	Detail
ALZ, AUT, CI, CNS, EP, ON, SCI, ST	Signalling pathways	 Increase in hippocampal mitochondrial glutathione Improved brain oxidative stress responses Improved neurovascular function via mechanistic target of rapamycin (mTOR) inhibition Increased effector substrates for glutamate, serotonin, dopamine and tryptophan Activation of multiple pathways providing neuroprotection Activation of detoxification pathways tross Activation of pathways impacting attives Activation of pathways impacting circadian timing Activation of pathways involved in cortical excitability and seizure regulation Activation of pathways supressing neuroinflammation
ARD, CNS, EP	Synaptic transmission	 Activation of molecules involved in neuronal growth and survival Restoration of vesicular transporters levels for GABA and glutamate Transmission resistant to low glucose levels and metabolic stress
ALZ, CI, CNS	Vascular supply	 Reduced long-term potentiation that preferentially limits excess excitability whilst preserving normal brain activity Increased cerebral blood flow in ALZ model Decreased capillarisation associated with tumour growth and seizure prevention Reduced infarct size
ARD, age-related degeneration; ALZ, Alzheimer's c MS, multiple sclerosis; NT, nerve toxin; ON, opl	lisease; AUT , autism; CI, cerebral ischaernia; C tic nerve; PKD, Parkinson's disease; PND, pe	ARD, age-related degeneration; ALZ, Alzheimer's disease; AUT, autism; CI, cerebral ischaemia; CP, chronic pain; CNS, central nervous system generally; D, diabetes; EP, epilepsy; MetS, metabolic syndrome; MCI, mild cognitive impairment; MS, multiple sclerosis; NT, nerve toxin; ON, optic nerve; PKD, Parkinson's disease; PND, peripheral nerve dysfunction; SCI, spinal cord injury; ST, stroke; TBI, traumatic brain injury.

appropriate pain management strategy⁽²²¹⁾. Beneficial outcomes on mitochondria were frequently reported in the extracted studies (Table 1); however, the result is less clear when examining the outcomes of individual studies (Supplementary Table 2). Kephart et al.⁽¹⁸⁰⁾ reported no benefit to mitochondrial quality in brain tissue sampled following a long-term ketogenic diet. A study by Lauritzen et al.⁽⁴⁵⁾ was one of the few to report negative outcomes. This study was designed specifically to examine a mouse model of mitochondrial dysfunction bred to express a mutated mitochondrial DNA repair gene (mutUNG1) designed to represent DNA damage that occurs in neurological disorders. They reported an increase in mitochondrial mass in the hippocampus and upregulated mitochondrial antioxidant defences, which would appear positive; however, this did not correlate with their overall observation of accelerated neurodegeneration from impaired mitochondrial dynamics and function. The context of their experiment becomes important, where the ketogenic diet increased mitochondrial biogenesis, but this increase was of dysfunctional mitochondria, compounding the neurodegeneration and energy demands. This study highlights the difficulty in extrapolating these results to human application. Their research does not necessarily apply to a ketogenic diet applied in the absence of this specific mitochondrial gene mutation, but as the authors⁽⁴⁵⁾ conclude, the diet also cannot be considered always beneficial for every type of mitochondrial pathology. Theoretically, the ketogenic diet appears to have potential for pain management through the improvement of mitochondrial function with subsequent reduction of oxidative stress and inflammation. Variability in clinical efficacy is likely to exist due to nuance in the mechanism of mitochondrial pathology.

Difficulty in extrapolating results also exists where an animal is fed the diet, but the analysis occurs in a dissected animal which is no longer a part of a complex adaptive system. One of the inclusion criteria for the current review was that the experiment had to have fed a ketogenic diet to the animal; cell culture and *in vitro* studies were excluded. The lack of an intact noradrenergic system may limit the effect of the ketogenic diet and produce disparate results⁽¹⁶⁴⁾ and may also account for the differences seen between animal and human trials involving ketogenic diets.

Chronic pain involves an increase in neuronal excitability^(222,223), with links suggested between these mechanisms and those involved in seizures, and the use of anticonvulsant medications to treat neuropathic pain⁽¹⁸⁾. A ketogenic diet has been widely used clinically as a treatment for epilepsy with several trials in adults⁽¹⁵⁾ as well as children⁽²²⁴⁾. A similar interpretive difficulty lies in the animal research for epilepsy where clinical human trials report generally favourable outcomes, but the animal research results can range between anticonvulsant to pro-convulsant outcomes^(91,166,175) (Supplementary Table 2). Again, experiment design becomes important, with the eightynine epilepsy studies including: different animal models (species, strain and age), multiple different seizure induction models (using different chemicals with different target receptors, and some using electrical shock), inconsistent levels of ketosis achieved, different chow content and quality, different chow quantity (with some diets employing calorie restriction in conjunction with the ketogenic diet), different lengths of dietary intervention, mismatched animal weight between groups resulting from different diets⁽¹⁵⁰⁾, and different

Neurotransmitter function was frequently reported in the included studies as a change within the nervous system favouring a reduction or restoration of normal levels of neuronal excitability. The mechanism reported was improved GABA-to-glutamate ratios usually via increased GABA (inhibitory) and/or decreased glutamate (excitatory) levels, with outcomes being a reduction in various seizure metrics in the animals tested. The research exploring the relationship between chronic pain and neurotransmitter levels is inconsistent. There is evidence supporting motor cortex disinhibition that is more pronounced in neuropathic pain⁽²²²⁾; however, whether this is due to a loss of GABAergic inhibition, as has been suggested, is still unclear. A recent systematic review reported altered neurotransmitter levels demonstrated in a small number of human chronic pain trials. There were increased levels of Glx (glutamate and glutamine combined) reported, but no corresponding reduction in GABA as might be expected⁽²²⁵⁾. The authors reported that different pain conditions may present with unique neurometabolite signatures, but the research was limited by inadequate reporting and standardisation of magnetic resonance spectroscopy techniques used.

A further variable that may contribute to the inconsistencies reported is that of the chow. Problems exist where the control diets are not matched appropriately to the ketogenic chow. Differences in vitamins, minerals and fibre exist between the diets as well as the macronutrient properties, limiting the ability to assess the ketogenic component of the diet. A number of issues also exist with the commercial rodent ketogenic diet formulations, including restriction of protein, choline deficiency⁽²²⁶⁾ and poor-quality fats (such as hydrogenated vegetable oils) rather than fats with a more beneficial inflammatory profile (such as omega-3)⁽²²⁷⁾.

The evidence presented in animal models supporting positive changes from a ketogenic diet, such as seen with anti-inflammatory mechanisms, appears compelling. However, the reported outcomes overall are often inconsistent and ambiguous⁽⁶⁷⁾, and there are many difficulties when extrapolating from animal models to human models of chronic pain⁽²²⁸⁾. The use of specific animal strains and sex may reduce the heterogeneity and increase the likelihood of detecting an effect, but may be poor representations of the diversity in target human pain populations⁽²²⁹⁾. These translational issues could be explored by also including natural animal models (such as using the ketogenic diet on naturally occurring pain presentations in domestic animals)⁽²²⁹⁾ as well as more consistency in experimental design, and reporting which more clearly acknowledges the limitations of the research. These strategies may allow the data to better inform human clinical trials of chronic pain.

Conclusion

Fourteen broad themes were identified from the literature outlining how a ketogenic diet influences nervous system function from animal models. The mechanisms presented centred around the reduction of inflammation and oxidative stress as well as a reduction in nervous system excitability. These mechanisms are potential drivers of chronic pain, and treatment strategies which target these have implications for chronic pain management. Given the multiple potential mechanisms presented, it is likely that many of these are involved synergistically and undergo adaptive processes within the human body, and controlled animal models that limit the investigation to a particular pathway in isolation may reach differing conclusions. Attention is required when translating this information to human chronic pain populations owing to the limitations outlined from the animal research.

Financial support

This research did not receive any specific grant from funding agencies in the public, commercial or not-for-profit sector.

Conflicts of interest

The authors declare that there is no conflict of interest regarding the publication of this article.

Supplementary material

To view supplementary material for this article, please visit https://doi.org/10.1017/S0954422421000214

References

- 1. Bosma-den Boer M, van Wetten M & Pruimboom L (20) Chronic inflammatory diseases are stimulated by current lifestyle: how diet, stress levels and medication prevent our body from recovering. *Nutr Metab* 2012, **9**, 1–14.
- Shen Y, Kapfhamer D, Minnella A, *et al.* (2017) Bioenergetic state regulates innate inflammatory responses through the transcriptional co-repressor CtBP. *Nat Commun* 8, 624.
- Kopp W (2019) How Western diet and lifestyle drive the pandemic of obesity and civilization diseases. *Diabetes Metab Syndr Obes* 12, 2221–2236.
- Ruskin D (2016) Metabolic therapy and pain. In: Masino S, ed. Ketogenic Diet and Metabolic Therapies: Expanded Roles in Health and Disease. New York: Oxford University Press, 196–208.
- Davis J, Fournakis N & Ellison J (2021) Ketogenic diet for the treatment and prevention of dementia: a review. J Geriatr Psychiatry Neurol 34, 3–10.
- Camberos-Luna L & Massieu L (2020) Therapeutic strategies for ketosis induction and their potential efficacy for the treatment of acute brain injury and neurodegenerative diseases. *Neurochem Int* 133, 104614.
- Phillips MCL, Murtagh DKJ, Gilbertson LJ, Asztely FJS & Lynch CDP (2018) Low-fat versus ketogenic diet in Parkinson's disease: a pilot randomized controlled trial. *Mov Disord* 33, 1306–1314.
- 8. Norwitz N, Hu M & Clarke K (2019) The mechanisms by which the ketone body D-beta-hydroxybutyrate may improve the multiple cellular pathologies of Parkinson's disease. *Front Nutr* **6**, 63.

R. Field et al.

- 276
 - Taylor M, Sullivan D, Mahnken J, Burns J & Swerdlow R (2018) Feasibility and efficacy data from a ketogenic diet intervention in Alzheimer's disease. *Alzbeimers Dement* 4, 28–36.
 - Morris G, Puri B, Carvalho A, *et al.* (2020) Induced ketosis as a treatment for neuroprogressive disorders: food for thought? *Int J Neuropsychopbarmacol* 23, 366–384.
 - 11. McDougall A, Bayley M & Munce S (2018) The ketogenic diet as a treatment for traumatic brain injury: a scoping review. *Brain Inj* **32**, 416–422.
 - Li R, Liu Y, Liu H & Li J (2020) Ketogenic diets and protective mechanisms in epilepsy, metabolic disorders, cancer, neuronal loss, and muscle and nerve degeneration. *J Food Biochem* 44, e13140.
 - 13. Sadeghifar F & Penry V (2019) Mechanisms and uses of dietary therapy as a treatment for epilepsy: a review. *Glob Adv Health Med* **8**, 2164956119874784.
 - Masino S & Rho J (2019) Metabolism and epilepsy: ketogenic diets as a homeostatic link. *Brain Res* 1703, 26–30.
 - 15. Mahmoud S, Ho-Huang E & Buhler J (2020) Systematic review of ketogenic diet use in adult patients with status epilepticus. *Epilepsia Open* **5**, 10–21.
 - Ruskin DN, Kawamura M & Masino SA (2009) Reduced pain and inflammation in juvenile and adult rats fed a ketogenic diet. *PLoS One* 4, e8349.
 - Rho J & Stafstrom C (2012) The ketogenic diet as a treatment paradigm for diverse neurological disorders. *Front Pharmacol* 3, 59.
 - Masino S & Ruskin D (2013) Ketogenic diets and pain. J Child Neurol 28, 993–1001.
 - Schabrun S, Elgueta-Cancino E & Hodges P (2017) Smudging of the motor cortex is related to the severity of low back pain. *Spine* 42, 1172–1178.
 - Schabrun S, Christensen S, Mrachacz-Kersting N & Graven-Nielsen T (2016) Motor cortex reorganization and impaired function in the transition to sustained muscle pain. *Cereb Cortex* 26, 1878–1890.
 - Kuner R & Flor H (2017) Structural plasticity and reorganisation in chronic pain. *Nat Rev Neurosci* 18, 20–30.
 - Elma Ö, Yilmaz S, Deliens T, *et al.* (2020) Do nutritional factors interact with chronic musculoskeletal pain? A systematic review. *J Clin Med* 9, 702.
 - Brain K, Burrows T, Rollo M, *et al.* (2018) A systematic review and meta-analysis of nutrition interventions for chronic noncancer pain. *J Hum Nutr Diet* **32**, 198–225.
 - Field R, Pourkazemi F, Turton J & Rooney K (2020) Dietary interventions are beneficial for patients with chronic pain: a systematic review with meta-analysis. *Pain Med* doi: 10. 1093/pm/pnaa1378
 - Nijs J, Elma Ö, Yilmaz S, *et al.* (2019) Nutritional neurobiology and central nervous system sensitisation: missing link in a comprehensive treatment for chronic pain? *Br J Anaesth* 123, 539–543.
 - Kaushik AS, Strath LJ & Sorge RE (2020) Dietary interventions for treatment of chronic pain: oxidative stress and inflammation. *Pain Ther* 9, 487–498.
 - 27. Hite A, Cavan D, Cywes R, *et al.* Clinical guidelines for the prescription of carbohydrate restriction as a therapeutic intervention V1.1. Low Carb USA. https://www.lowcarbusa.org/ clinical-guidelines/
 - 28. Newman JC & Verdin E (2014) Ketone bodies as signaling metabolites. *Trends Endocrinol Metab* **25**, 42–52.
 - 29. Shimazu T, Hirschey MD, Newman J, *et al.* (2013) Suppression of oxidative stress by β -hydroxybutyrate, an endogenous histone deacetylase inhibitor. *Science* **339**, 211–214.

- Eendfeldt A & Scher B The science of low carb and keto. DietDoctor.com. https://www.dietdoctor.com/low-carb/ science Accessed 29/10/2020.
- Paoli A, Rubini A, Volek J & Grimaldi K (2013) Beyond weight loss: a review of the therapeutic uses of very-low-carbohydrate (ketogenic) diets. *Eur J Clin Nutr* 67, 789–796.
- 32. Youm Y, Nguyen K, Grant R, *et al.* (2015) The ketone metabolite [beta]-hydroxybutyrate blocks NLRP3 inflammasomemediated inflammatory disease. *Nat Med* **21**, 263–269.
- Rho J (2017) How does the ketogenic diet induce anti-seizure effects? *Neurosci Lett* 637(Suppl. C), 4–10.
- 34. Miller V, Villamena F & Volek J (2018) Nutritional ketosis and mitohormesis: potential implications for mitochondrial function and human health. *J Nutr Metab* **2018**, 1–27.
- 35. Ruskin D & Masino S (2012) The nervous system and metabolic dysregulation: emerging evidence converges on ketogenic diet therapy. *Front Neurosci* **6**, 33.
- 36. Peters M, Godfrey C, McInerney P, et al. (2020) Chapter 11: scoping reviews. In: Aromataris E & Munn Z, eds. JBI Reviewers Manual 2020. The Joanna Briggs Institute. https://reviewersmanual.joannabriggs.org/
- Tricco A, Lillie E, Zarin W, *et al.* (2018) PRISMA extension for scoping reviews (PRISMA-ScR): checklist and explanation. *Ann Intern Med* 169, 467–473.
- 38. Campbell G, Senior A & Bell-Anderson K (2017) Metabolic effects of high glycaemic index diets: a systematic review and meta-analysis of feeding studies in mice and rats. *Nutrients* **9**, 646.
- Auvinen H, Romijn J, Biermasz N, *et al.* (2011) Effects of high fat diet on the basal activity of the hypothalamus-pituitaryadrenal axis in mice: a systematic review. *Horm Metab Res* 43, 899–906.
- Hooijmans CR & Ritskes-Hoitinga M (2013) Progress in using systematic reviews of animal studies to improve translational research. *PLoS Med* **10**, e1001482.
- Pound P & Ritskes-Hoitinga M (2020) Can prospective systematic reviews of animal studies improve clinical translation? *J Transl Med* 18, 15.
- Hernandez AR, Hernandez CM, Campos KT, *et al.* (2017) The antiepileptic ketogenic diet alters hippocampal transporter levels and reduces adiposity in aged rats. *J Gerontol A Biol Sci Med Sci* 73, 450–458.
- 43. Hernandez A, Hernandez C, Campos K, *et al.* (2018) A ketogenic diet improves cognition and has biochemical effects in prefrontal cortex that are dissociable from hippocampus. *Front Aging Neurosci* **10**, 391.
- 44. Hernandez A, Hernandez C, Truckenbrod L, *et al.* (2019) Age and ketogenic diet have dissociable effects on synapse-related gene expression between hippocampal subregions. *Front Aging Neurosci* **11**, 239.
- 45. Lauritzen KH, Hasan-Olive MM, Regnell CE, *et al.* (2016) A ketogenic diet accelerates neurodegeneration in mice with induced mitochondrial DNA toxicity in the forebrain. *Neurobiol Aging* **48**, 34–47.
- 46. Zhang Y, Xu K, Kerwin T, LaManna J & Puchowicz M (2018) Impact of aging on metabolic changes in the ketotic rat brain: glucose, oxidative and 4-HNE metabolism. Vol 1072. TypeOxygen Transport to Tissue XL. Advances in Experimental Medicine and Biology. Thews O, LaManna J, Harrison D: Springer International Publishing.
- 47. Beckett T, Studzinski C, Keller J, Paul Murphy M & Niedowicz D (2013) A ketogenic diet improves motor performance but does not affect beta-amyloid levels in a mouse model of Alzheimer's disease. *Brain Res* **1505**, 61–67.

2

Nutrition Research Reviews

- Ma D, Wang AC, Parikh I, *et al.* (2018) Ketogenic diet enhances neurovascular function with altered gut microbiome in young healthy mice. *Sci Rep* 8, 6670.
- Roy M, Nugent S, Tremblay-Mercier J, *et al.* (2012) The ketogenic diet increases brain glucose and ketone uptake in aged rats: a dual tracer PET and volumetric MRI study. *Brain Res* 1488, 14–23.
- Van der Auwera I, Wera S, Van Leuven F & Henderson ST (2005) A ketogenic diet reduces amyloid beta 40 and 42 in a mouse model of Alzheimer's disease. *Nutr Metab (Lond)* 2, 28.
- Ahn Y, Narous M, Tobias R, Rho J & Mychasiuk R (2014) The ketogenic diet modifies social and metabolic alterations identified in the prenatal valproic acid model of autism spectrum disorder. *Dev Neurosci* 36, 371–380.
- 52. Ahn Y, Sabouny R, Villa B, *et al.* (2020) Aberrant mitochondrial morphology and function in the BTBR mouse model of autism is improved by two weeks of ketogenic diet. *Int J Mol Cell Med* **21**, 3266.
- 53. Dai Y, Zhao Y, Tomi M, *et al.* (2017) Sex-specific life course changes in the neuro-metabolic phenotype of Glut3 null heterozygous mice: ketogenic diet ameliorates electroencephalographic seizures and improves sociability. *Endocrinology* **158**, 936–949.
- Mychasiuk R & Rho J (2017) Genetic modifications associated with ketogenic diet treatment in the BTBR T+Tf/j mouse model of autism spectrum disorder. *Autism Res* 10, 456–471.
- Newell C, Shutt T, Ahn Y, *et al.* (2016) Tissue specific impacts of a ketogenic diet on mitochondrial dynamics in the BTBRT+tf/j mouse. *Front Physiol* 7, 654.
- Newell C, Johnsen V, Yee N, *et al.* (2017) Ketogenic diet leads to O-G1cNAc modification in the BTBRT + tf/j mouse model of autism. *Biochim Biophys Acta Mol Basis Dis* 1863, 2274–2281.
- Smith J, Rho J & Teskey G (2016) Ketogenic diet restores aberrant cortical motor maps and excitation-to-inhibition imbalance in the BTBR mouse model of autism spectrum disorder. *Behav Brain Res* **304**, 67–70.
- Tai KK & Truong DD (2007) Ketogenic diet prevents seizure and reduces myoclonic jerks in rats with cardiac arrestinduced cerebral hypoxia. *Neurosci Lett* **425**, 34–38.
- Tai K, Nguyen N, Pham L & Truong D (2008) Ketogenic diet prevents cardiac arrest-induced cerebral ischemic neurodegeneration. *J Neural Transm* 115, 1011–1017.
- 60. Tai K, Pham L & Truong D (2009) Intracisternal administration of glibenclamide or 5-hydroxydecanoate does not reverse the neuroprotective effect of ketogenic diet against ischemic brain injury-induced neurodegeneration. *Brain Inj* 23, 1081–1088.
- Yang Q, Guo M, Wang X, *et al.* (2017) Ischemic preconditioning with a ketogenic diet improves brain ischemic tolerance through increased extracellular adenosine levels and hypoxia-inducible factors. *Brain Res* 1667, 11–18.
- Ruskin DN, Suter TACS, Ross JL & Masino SA (2013) Ketogenic diets and thermal pain: dissociation of hypoalgesia, elevated ketones, and lowered glucose in rats. *J Pain* 14, 467–474.
- Elamin M, Ruskin D, Masino S & Sacchetti P (2018) Ketogenic diet modulates NAD+- dependent enzymes and reduces DNA damage in hippocampus. *Front Cell Neurosci* 12, 263.
- Fukushima A, Ogura Y, Furuta M, *et al.* (2015) Ketogenic diet does not impair spatial ability controlled by the hippocampus in male rats. *Brain Res* 1622, 36–42.
- Genzer Y, Dadon M, Burg C, Chapnik N & Froy O (2016) Effect of dietary fat and the circadian clock on the expression of brain-derived neurotrophic factor (BDNF). *Mol Cell Endocrinol* **430**, 49–55.

- Heischmann S, Gano L, Quinn K, *et al.* (2018) Regulation of kynurenine metabolism by a ketogenic diet. *J Lipid Res* 59, 958–966.
- 67. Huang J, Li Y, Wu C, *et al.* (2019) The effect of ketogenic diet on behaviors and synaptic functions of naive mice. *Brain Behav* **9**, e01246.
- Leino RL, Gerhart DZ, Duelli R, Enerson BE & Drewes LR (2001) Diet-induced ketosis increases monocarboxylate transporter (MCT1) levels in rat brain. *Neurochem Int* 38, 519–527.
- 69. Ling Y, Wang D, Sun Y, Zhao D & Ni H (20) Neuro-behavioral status and the hippocampal expression of metabolic associated genes in wild-type rat following a ketogenic diet. *Front Neurol* 10, 65.
- Melo TM, Nehlig A & Sonnewald U (2006) Neuronal-glial interactions in rats fed a ketogenic diet. *Neurochem Int* 48, 498–507.
- 71. Milder JB, Liang L-P & Patel M (2010) Acute oxidative stress and systemic Nrf2 activation by the ketogenic diet. *Neurobiol Dis* **40**, 238–244.
- 72. Pifferi F, Tremblay S, Croteau E, *et al.* (2011) Mild experimental ketosis increases brain uptake of 11C-acetoacetate and 18Ffluorodeoxyglucose: a dual-tracer PET imaging study in rats. *Nutr Neurosci* **142**, 51–58.
- Rho J, Sarnat H, Sullivan P, Robbins C & Kim D (2004) Lack of long-term histopathologic changes in brain and skeletal muscle of mice treated with a ketogenic diet. *J Child Neurol* 19, 555–557.
- Roy M, Beauvieux M, Naulin J, *et al.* (2015) Rapid adaptation of rat brain and liver metabolism to a ketogenic diet: an integrated study using H-1- and C-13-NMR spectroscopy. *J Cereb Blood Flow Metab* 35, 1154–1162.
- Samala R, Klein J & Borges K (2011) The ketogenic diet changes metabolite levels in hippocampal extracellular fluid. *Neurochem Int* 58, 5–8.
- 76. Selfridge J, Wilkins H, Lezi E, *et al.* (2015) Effect of one month duration ketogenic and non-ketogenic high fat diets on mouse brain bioenergetic infrastructure. *J Bioenerg Biomembr* **47**, 1–11.
- Strandberg J, Kondziella D, Thorlin T & Asztely F (2008) Ketogenic diet does not disturb neurogenesis in the dentate gyrus in rats. *Neuroreport* 19, 1235–1237.
- Sullivan PG, Rippy NA, Dorenbos K, *et al.* (2004) The ketogenic diet increases mitochondrial uncoupling protein levels and activity. *Ann Neurol* 55, 576–580.
- Sussman D, Germann J & Henkelman M (2015) Gestational ketogenic diet programs brain structure and susceptibility to depression & anxiety in the adult mouse offspring. *Brain Behav* 5, e00300.
- Thio L, Rensing N, Maloney S, *et al.* (2010) A ketogenic diet does not impair rat behavior or long-term potentiation. *Epilepsia* 51, 1619–1623.
- Viggiano A, Meccariello R, Santoro A, *et al.* (2019) A calorierestricted ketogenic diet reduces cerebral cortex vascularization in prepubertal rats. *Nutrients* 11, 2681.
- 82. Vizuete AF, de Souza DF, Guerra MC, *et al.* (2013) Brain changes in BDNF and S100B induced by ketogenic diets in Wistar rats. *Life Sci* **92**, 923–928.
- Wang X, Liu Q, Zhou J, Wu X & Zhu Q (2017) Beta hydroxybutyrate levels in serum and cerebrospinal fluid under ketone body metabolism in rats. *Exp Anim* 66, 177–182.
- Zarnowski T, Choragiewicz T, Tulidowicz-Bielak M, et al. (2012) Ketogenic diet increases concentrations of kynurenic acid in discrete brain structures of young and adult rats. J Neural Transm 119, 679–684.
- 85. Zhang Y, Zhang S, Marin-Valencia I & Puchowicz M (2015) Decreased carbon shunting from glucose toward oxidative

N Nutrition Research Reviews

metabolism in diet-induced ketotic rat brain. J Neurochem 132, 301-312.

- 86. Ziegler DR, Ribeiro LC, Hagenn M, et al. (2003) Ketogenic diet increases glutathione peroxidase activity in rat hippocampus. Neurochem Res 28, 1793-1797.
- 87. Morrison C, Hill C, DuVall M, et al. (2020) Consuming a ketogenic diet leads to altered hypoglycemic counter-regulation in mice. [Diabetes Complications 34, 107557.
- 88 Yamada KA, Rensing N & Thio LL (2005) Ketogenic diet reduces hypoglycemia-induced neuronal death in young rats. Neurosci Lett 385, 210-214.
- 89. Bough K & Eagles D (1999) A ketogenic diet increases the resistance to pentylenetetrazole-induced seizures in the rat. Epilepsia 40, 138–143.
- Bough K, Valiyil R, Han FT & Eagles D (1999) Seizure resis-90. tance is dependent upon age and calorie restriction in rats fed a ketogenic diet. Epilepsy Res 35, 21-28.
- 91. Bough K, Matthews P & Eagles D (2000) A ketogenic diet has different effects upon seizures induced by maximal electroshock and by pentylenetetrazole infusion. Epilepsy Res 38, 105-114.
- 92. Bough K, Yao S & Eagles D (2000) Higher ketogenic diet ratios confer protection from seizures without neurotoxicity. Epilepsy Res 38, 15-25.
- 93. Bough K, Gudi K, Han F, Rathod A & Eagles D (2002) An anticonvulsant profile of the ketogenic diet in the rat. Epilepsy Res 50. 313-325
- 94. Bough K, Schwartzkroin P & Rho J (2003) Calorie restriction and ketogenic diet diminish neuronal excitability in rat dentate gyrus in vivo. Epilepsia 44, 752-760.
- 95 Bough K, Wetherington J, Hassel B, et al. (2006) Mitochondrial biogenesis in the anticonvulsant mechanism of the ketogenic diet. Ann Neurol 60, 223-235.
- 96. Bough K, Paquet M, Pare J, et al. (2007) Evidence against enhanced glutamate transport in the anticonvulsant mechanism of the ketogenic diet. Epilepsy Res 74, 232-236.
- 97. Bough K (2008) Energy metabolism as part of the anticonvulsant mechanism of the ketogenic diet. Epilepsia 49, 91-93.
- 98. Calderón N, Betancourt L, Hernández L & Rada P (2017) A ketogenic diet modifies glutamate, gamma-aminobutyric acid and agmatine levels in the hippocampus of rats: a microdialysis study. Neurosci Lett 642, 158-162.
- 99. Cheng C, Kelley B, Wang J, et al. (2003) A ketogenic diet increases brain insulin-like growth factor receptor and glucose transporter gene expression. Endocrinology 144, 2676-2682.
- 100. Cheng C, Hicks K, Wang J, Eagles D & Bondy C (2004) Caloric restriction augments brain glutamic acid decarboxylase-65 and -67 expression. J Neurosci Res 77, 270-276.
- 101 Church WH, Adams RE & Wyss LS (2014) Ketogenic diet alters dopaminergic activity in the mouse cortex. Neurosci Lett 571, 1-4.
- 102. Chwiej J, Patulska A, Skoczen A, et al. (2015) Elemental changes in the hippocampal formation following two different formulas of ketogenic diet: an X-ray fluorescence microscopy study. J Biol Inorg Chem 20, 1277-1286.
- 103. Chwiej J, Skoczen A, Matusiak K, et al. (2015) The influence of the ketogenic diet on the elemental and biochemical compositions of the hippocampal formation. Epilepsy Behav 49, 40-46.
- 104. Chwiej J, Patulska A, Skoczen A, et al. (2017) Various ketogenic diets can differently support brain resistance against experimentally evoked seizures and seizure-induced elemental anomalies of hippocampal formation. J Trace Elem Med Biol 42, 50-58.
- 105. Cullingford T, Eagles D & Sato H (2002) The ketogenic diet upregulates expression of the gene encoding the key

ketogenic enzyme mitochondrial 3-hydroxy-3-methylglutaryl-CoA synthase in rat brain. Epilepsy Res 49, 99-107.

- 106. Dupuis N, Curatolo N, Benoist J & Auvin S (2015) Ketogenic diet exhibits anti-inflammatory properties. Epilepsia 56, e95-98
- 107. Dutton S, Sawyer N, Kalume F, et al. (2011) Protective effect of the ketogenic diet in Scn1a mutant mice. Epilepsia 52, 2050-2056
- 108. Forero-Quintero LS, Deitmer JW & Becker HM (2017) Reduction of epileptiform activity in ketogenic mice: the role of monocarboxylate transporters. Sci Rep 7, 4900.
- 109. Gama I, Trindade-Filho E, Oliveira S, et al. (2015) Effects of ketogenic diets on the occurrence of pilocarpine-induced status epilepticus of rats. Metab Brain Dis 30, 93-98.
- 110. Gietzen DW, Lindstrom SH, Sharp JW, Teh PS & Donovan MJ (2018) Indispensable amino acid-deficient diets induce seizures in ketogenic diet-fed rodents, demonstrating a role for amino acid balance in dietary treatments for epilepsy. J Nutr **148** 480–489
- 111. Godlevskii LS, Polyasny VO, Ovchinnikova OG, et al. (2012) Modulation of the state of the antiepileptic cerebral system by the influence of a ketogenic diet under conditions of the resistant epileptic syndrome. Neurophysiology 43, 503-506.
- 112. Gomez-Lira G, Mendoza-Torreblanca J & Granados-Rojas L (2011) Ketogenic diet does not change NKCC1 and KCC2 expression in rat hippocampus. Epilepsy Res 96, 166-171.
- 113. Hansen SL, Nielsen AH, Knudsen KE, et al. (2009) Ketogenic diet is antiepileptogenic in pentylenetetrazole kindled mice and decrease levels of N-acylethanolamines in hippocampus. Neurochem Int 54, 199-204.
- 114 Harney J, Madara J, Madara J & I'Anson H (2002) Effects of acute inhibition of fatty acid oxidation on latency to seizure and concentrations of beta hydroxybutyrate in plasma of rats maintained on calorie restriction and/or the ketogenic diet. Epilepsy Res 49, 239-246.
- 115. Hartman A, Lyle M, Rogawski M & Gasior M (2008) Efficacy of the ketogenic diet in the 6-Hz seizure test. *Epilepsia* **49**, 334-339.
- 116. Hartman A, Zheng X, Bergbower E, Kennedy M & Hardwick J (2010) Seizure tests distinguish intermittent fasting from the ketogenic diet. Epilepsia 51, 1395-1402.
- 117. Hasan-Olive MM, Lauritzen KH, Ali M, et al. (2019) A ketogenic diet improves mitochondrial biogenesis and bioenergetics via the PGC1alpha-SIRT3-UCP2 axis. Neurochem Res 44, 22 - 37.
- 118. Hori A, Tandon P, Holmes G & Stafstrom C (1997) Ketogenic diet: effects on expression of kindled seizures and behavior in adult rats. Epilepsia 38, 750-758.
- 119 Hu X, Cheng X, Fei J & Xiong Z (2011) Neuron-restrictive silencer factor is not required for the antiepileptic effect of the ketogenic diet. Epilepsia 52, 1609-1616.
- 120. Jarrett SG, Milder JB, Liang LP & Patel M (2008) The ketogenic diet increases mitochondrial glutathione levels. J Neurochem 106, 1044-1051.
- 121. Jeon BT, Lee DH, Kim KH, et al. (2009) Ketogenic diet attenuates kainic acid-induced hippocampal cell death by decreasing AMPK/ACC pathway activity and HSP70. Neurosci Lett 453, 49-53.
- 122. Jeong H, Kim H, Kim Y, et al. (2010) The ketogenic diet suppresses the cathepsin E expression induced by kainic acid in the rat brain. Yonsei Med J 51, 653-660.
- 123. Jeong E, Jeon B, Shin H, et al. (2011) Ketogenic diet-induced peroxisome proliferator-activated receptor-y activation decreases neuroinflammation in the mouse hippocampus after kainic acid-induced seizures. Exp Neurol 232, 195-202.

- Jiang Y, Yang Y, Wang S, *et al.* (2012) Ketogenic diet protects against epileptogenesis as well as neuronal loss in amygdaloid-kindling seizures. *Neurosci Lett* **508**, 22–26.
- Kawamura M, Ruskin D, Geiger J, Boison D & Masino S (2014) Ketogenic diet sensitizes glucose control of hippocampal excitability. *J Lipid Res* 55, 2254–2260.
- Knowles S, Budney S, Deodhar M, et al. (2018) Ketogenic diet regulates the antioxidant catalase via the transcription factor PPARgamma2. *Epilepsy Res* 147, 71–74.
- 127. Kobow K, Kaspi A, Harikrishnan K, et al. (2013) Deep sequencing reveals increased DNA methylation in chronic rat epilepsy. Acta Neuropathol **126**, 741–756.
- Koranda JL, Ruskin DN, Masino SA & Blaise JH (2011) A ketogenic diet reduces long-term potentiation in the dentate gyrus of freely behaving rats. *J Neurophysiol* **106**, 662–666.
- Kresyun V, Polyasny V, Godovan V & Godlevsky L (2013) Changes in brain cortex sensitivity to epileptogens under conditions of ketogenic diet. *Bull Exp Biol Med* **154**, 457–459.
- Kwon Y, Jeong S, Kim D, Choi E & Son B (2008) Effects of the ketogenic diet on neurogenesis after kainic acid-induced seizures in mice. *Epilepsy Res* 78, 186–194.
- 131. Likhodii S, Musa K, Mendonca A, *et al.* (2000) Dietary fat, ketosis, and seizure resistance in rats on the ketogenic diet. *Epilepsia* **41**, 1400–1410.
- 132. Lin GW, Lu P, Zeng T, *et al.* (2017) GAPDH-mediated posttranscriptional regulations of sodium channel Scn1a and Scn3a genes under seizure and ketogenic diet conditions. *Neuropharmacology* **113**, 480–489.
- 133. Linard B, Ferrandon A, Koning E, Nehlig A & Raffo E (2010) Ketogenic diet exhibits neuroprotective effects in hippocampus but fails to prevent epileptogenesis in the lithium-pilocarpine model of mesial temporal lobe epilepsy in adult rats. *Epilepsia* **51**, 1829–1836.
- 134. Luan G, Zhao Y, Zhai F, Chen Y & Li T (2012) Ketogenic diet reduces Smac/Diablo and cytochrome c release and attenuates neuronal death in a mouse model of limbic epilepsy. *Brain Res Bull* 89, 79–85.
- Lusardi T, Akula K, Coffman S, *et al.* (2015) Ketogenic diet prevents epileptogenesis and disease progression in adult mice and rats. *Neuropharmacology* **99**, 500–509.
- 136. Mantis J, Meidenbauer J, Zimick N, Centeno N & Seyfried T (2014) Glucose reduces the anticonvulsant effects of the ketogenic diet in EL mice. *Epilepsy Res* **108**, 1137–1144.
- 137. Martillotti J, Weinshenker D, Liles L & Eagles D (2006) A ketogenic diet and knockout of the norepinephrine transporter both reduce seizure severity in mice. *Epilepsy Res* 68, 207–211.
- Masino S, Li T, Theofilas P, *et al.* (2011) A ketogenic diet suppresses seizures in mice through adenosine A 1 receptors. *J Clin Invest* **121**, 2679–2683.
- 139. McDaniel S, Rensing N, Thio L, Yamada K & Wong M (2011) The ketogenic diet inhibits the mammalian target of rapamycin (mTOR) pathway. *Epilepsia* 52, e7–e11.
- Melo IT, Rego EM, Bueno NB, *et al.* (2018) Ketogenic diet based on extra virgin coconut oil has no effects in young Wistar rats with pilocarpine-induced epilepsy. *Lipids* 53, 251–254.
- Muller-Schwarze AB, Tandon P, Liu Z, *et al.* (1999) Ketogenic diet reduces spontaneous seizures and mossy fiber sprouting in the kainic acid model. *Neuroreport* 10, 1517–1522.
- Nakazawa M, Kodama S & Matsuo T (1983) Effects of ketogenic diet on electroconvulsive threshold and brain contents of adenosine nucleotides. *Brain Dev* 5, 375–380.
- 143. Ni H, Zhao D & Tian T (2016) Ketogenic diet change cPLA2/ clusterin and autophagy related gene expression and correlate with cognitive deficits and hippocampal MFs sprouting following neonatal seizures. *Epilepsy Res* **120**, 13–18.

- 144. Noh H, Kim Y, Lee H, *et al.* (2003) The protective effect of a ketogenic diet on kainic acid-induced hippocampal cell death in the male ICR mice. *Epilepsy Res* **53**, 119–128.
- Noh HS, Lee HP, Kim DW, et al. (2004) A cDNA microarray analysis of gene expression profiles in rat hippocampus following a ketogenic diet. Mol Brain Res 129, 80–87.
- 146. Noh H, Kang S, Kim D, *et al.* (2005) Ketogenic diet increases calbindin-D28k in the hippocampi of male ICR mice with kainic acid seizures. *Epilepsy Res* **65**, 153–159.
- 147. Noh H, Kim D, Kang S, Cho G & Choi W (2005) Ketogenic diet prevents clusterin accumulation induced by kainic acid in the hippocampus of male ICR mice. *Brain Res* 1042, 114–118.
- 148. Noh H, Kim D, Cho G, Choi W & Kang S (2006) Increased nitric oxide caused by the ketogenic diet reduces the onset time of kainic acid-induced seizures in ICR mice. *Brain Res* 1075, 193–200.
- Noh HS, Kim DW, Kang SS, *et al.* (2006) Ketogenic diet decreases the level of proenkephalin mRNA induced by kainic acid in the mouse hippocampus. *Neurosci Lett* **395**, 87–92.
- 150. Nylen K, Likhodii S, Abdelmalik P, Clarke J & Burnham W (2005) A comparison of the ability of a 4:1 ketogenic diet and a 6.3:1 ketogenic diet to elevate seizure thresholds in adult and young rats. *Epilepsia* **46**, 1198–1204.
- 151. Nylen K, Likhodii S, Hum K & Burnham W (2006) A ketogenic diet and diallyl sulfide do not elevate afterdischarge thresholds in adult kindled rats. *Epilepsy Res* **71**, 23–31.
- 152. de Almeida Rabello Oliveira M, da Rocha Ataíde T, de Oliveira SL, *et al.* (2008) Effects of short-term and long-term treatment with medium- and long-chain triglycerides ketogenic diet on cortical spreading depression in young rats. *Neurosci Lett* **434**, 66–70.
- Olson C, Vuong H, Yano J, *et al.* (2018) The gut microbiota mediates the anti-seizure effects of the ketogenic diet. *Cell* 173, 1728–1741.
- 154. Raffo E, Francois J, Ferrandon A, Koning E & Nehlig A (2008) Calorie-restricted ketogenic diet increases thresholds to all patterns of pentylenetetrazol-induced seizures: critical importance of electroclinical assessment. *Epilepsia* **49**, 320–328.
- 155. Rho J, Kim D, Robbins C, Anderson G & Schwartzkroin P (1999) Age-dependent differences in flurothyl seizure sensitivity in mice treated with a ketogenic diet. *Epilepsy Res* **37**, 233–240.
- Samala R, Willis S, Borges K (2008) Anticonvulsant profile of a balanced ketogenic diet in acute mouse seizure models. *Epilepsy Res* 81, 119–127.
- 157. Silva MC, Rocha J, Pires CS, *et al.* (2005) Transitory gliosis in the CA3 hippocampal region in rats fed on a ketogenic diet. *Nutr Neurosci* **8**, 259–264.
- 158. Simeone K, Wilke J, Milligan H, *et al.* (2009) Ketogenic diet treatment abolishes seizure periodicity and improves diurnal rhythmicity in epileptic Kcna1-null mice. *Epilepsia* **50**, 2027–2034.
- 159. Simeone K, Matthews S, Rho J & Simeone T (2016) Ketogenic diet treatment increases longevity in Kcna1-null mice, a model of sudden unexpected death in epilepsy. *Epilepsia* **57**, e178–e182.
- 160. Simeone T, Samson K, Matthews S & Simeone K (2014) In vivo ketogenic diet treatment attenuates pathologic sharp waves and high frequency oscillations in in vitro hippocampal slices from epileptic K(v)1.1 alpha knockout mice. *Epilepsia* 55, E44–E49.
- Simeone T, Matthews S, Samson K & Simeone K (2017) Regulation of brain PPARgamma2 contributes to ketogenic diet anti-seizure efficacy. *Exp Neurol* 287, 54–64.

R. Field et al.

- 162. Stafstrom C, Wang C & Jensen F (1999) Electrophysiological observations in hippocampal slices from rats treated with the ketogenic diet. *Dev Neurosci* **21**, 393–399.
- Su S, Cilio M, Sogawa Y, *et al.* (2000) Timing of ketogenic diet initiation in an experimental epilepsy model. *Brain Res* 125, 131–138.
- Szot P, Weinshenker D, Rho J, Storey T & Schwartzkroin P (2001) Norepinephrine is required for the anticonvulsant effect of the ketogenic diet. *Brain Res Dev Brain Res* 129, 211–214.
- Tabb K, Szot P, White S, Liles L & Weinshenker D (2004) The ketogenic diet does not alter brain expression of orexigenic neuropeptides. *Epilepsy Res* 62, 35–39.
- Thavendiranathan P, Mendonca A, Dell C, et al. (2000) The MCT ketogenic diet: effects on animal seizure models. Exp Neurol 161, 696–703.
- Thavendiranathan P, Chow C, Cunnane S & Burnham W (2003) The effect of the 'classic' ketogenic diet on animal seizure models. *Brain Res* **959**, 206–213.
- 168. Tian T, Ni H & Sun B (2015) Neurobehavioral deficits in a rat model of recurrent neonatal seizures are prevented by a ketogenic diet and correlate with hippocampal zinc/lipid transporter signals. *Biol Trace Elem Res* 167, 251–258.
- 169. Tian T, Li LL, Zhang S & Ni H (2016) Long-term effects of ketogenic diet on subsequent seizure-induced brain injury during early adulthood: relationship of seizure thresholds to zinc transporter-related gene expressions. *Biol Trace Elem Res* 174, 369–376.
- Todorova M, Tandon P, Madore R, Stafstrom C & Seyfried T (2000) The ketogenic diet inhibits epileptogenesis in EL mice: a genetic model for idiopathic epilepsy. *Epilepsia* 41, 933–940.
- 171. Viggiano A, Stoddard M, Pisano S, *et al.* (2016) Ketogenic diet prevents neuronal firing increase within the substantia nigra during pentylenetetrazole-induced seizure in rats. *Brain Res Bull* **125**, 168–172.
- 172. Wang S, Ding Y, Ding X-Y, *et al.* (2016) Effectiveness of ketogenic diet in pentylenetetrazol-induced and kindling rats as well as its potential mechanisms. *Neurosci Lett* **614**, 1–6.
- 173. Wang B, Hou Q, Lu Y, *et al.* (2018) Ketogenic diet attenuates neuronal injury via autophagy and mitochondrial pathways in pentylenetetrazol-kindled seizures. *Brain Res* **1678**, 106–115.
- 174. Xu X, Sun R & Jin R (2008) The effect of the ketogenic diet on hippocampal GluR(5) and GluR(6) mRNA expression and Q/R site editing in the kainate-induced epilepsy model. *Epilepsy Behav* **13**, 445–448.
- Zarnowska I, Luszczki JJ, Zarnowski T, et al. (2017) Proconvulsant effects of the ketogenic diet in electroshockinduced seizures in mice. *Metab Brain Dis* 32, 351–358.
- Ziegler DR, Araujo E, Rotta LN, Perry ML & Goncalves CA (2002) A ketogenic diet increases protein phosphorylation in brain slices of rats. *J Nutr* **132**, 483–487.
- Ziegler DR, Oliveira DL, Pires C, *et al.* (2004) Ketogenic diet fed rats have low levels of S100B in cerebrospinal fluid. *Neurosci Res* 50, 375–379.
- 178. Blaise H, Ruskin D, Koranda J & Masino S (2015) Effects of a ketogenic diet on hippocampal plasticity in freely moving juvenile rats. *Physiol Rep* **3**, e12411.
- Masino S, Freedgood N, Reichert H, *et al.* (2019) Dietary intervention for canine epilepsy: two case reports. *Epilepsia Open* 4, 193–199.
- 180. Kephart WC, Mumford PW, Mao XS, *et al.* (2017) The 1-week and 8-month effects of a ketogenic diet or ketone salt supplementation on multi-organ markers of oxidative stress and mitochondrial function in rats. *Nutrients* 9, 1019.

- Mohamed H, El-Swefy S, Rashed L & Abd El-Latif S (2010) Biochemical effect of a ketogenic diet on the brains of obese adult rats. *J Clin Neurosci* 17, 899–904.
- 182. Hargrave S, Davidson T, Lee T & Kinzig K (2015) Brain and behavioral perturbations in rats following Western diet access. *Appetite* **93**, 35–43.
- Kim DY, Hao J, Liu R, *et al.* (2012) Inflammation-mediated memory dysfunction and effects of a ketogenic diet in a murine model of multiple sclerosis. *PLoS One* 7, e35476.
- Stumpf S, Berghoff S, Trevisiol A, *et al.* (2019) Ketogenic diet ameliorates axonal defects and promotes myelination in Pelizaeus-Merzbacher disease. *Acta Neuropathol* **138**, 147–161.
- Myers T, Langston J (2011) Diet composition exacerbates or attenuates soman toxicity in rats: implied metabolic control of nerve agent toxicity. *Neurotoxicology* **32**, 342–349.
- Bernardo-Colon A, Vest V, Clark A, *et al.* (2018) Antioxidants prevent inflammation and preserve the optic projection and visual function in experimental neurotrauma. *Cell Death Dis* 9, 1097.
- Harun-Or-Rashid M, Pappenhagen N, Palmer PG, *et al.* (2018) Structural and functional rescue of chronic metabolically stressed optic nerves through respiration. *J Neurosci* 38, 5122–5139.
- 188. Harun-Or-Rashid M & Inman DM (2018) Reduced AMPK activation and increased HCAR activation drive anti-inflammatory response and neuroprotection in glaucoma. *J Neuroinflammation* **15**, 313.
- Zarnowski T, Choragiewicz T, Schuettauf F, *et al.* (2015) Ketogenic diet attenuates NMDA-induced damage to rat's retinal ganglion cells in an age-dependent manner. *Ophthalmic Res* 53, 162–167.
- 190. Cheng B, Yang X, An L, *et al.* (2009) Ketogenic diet protects dopaminergic neurons against 6-OHDA neurotoxicity via upregulating glutathione in a rat model of Parkinson's disease. *Brain Res* **1286**, 25–31.
- Yang X & Cheng B (2010) Neuroprotective and anti-inflammatory activities of ketogenic diet on MPTP-induced neurotoxicity. *J Mol Neurosci* 42, 145–153.
- 192. Cooper M, McCoin C, Pei D, *et al.* (2018) Reduced mitochondrial reactive oxygen species production in peripheral nerves of mice fed a ketogenic diet. *Exp Physiol* **103**, 1206–1212.
- 193. Cooper M, Menta B, Perez-Sanchez C, *et al.* (2018) A ketogenic diet reduces metabolic syndrome-induced allodynia and promotes peripheral nerve growth in mice. *Exp Neurol* **306**, 149–157.
- 194. Liskiewicz A, Wlaszczuk A, Gendosz D, *et al.* (2016) Sciatic nerve regeneration in rats subjected to ketogenic diet. *Nutr Neurosci* **1**, 116–124.
- 195. Kong G, Huang Z, Ji W, *et al.* (2017) The ketone metabolite beta-hydroxybutyrate attenuates oxidative stress in spinal cord injury by suppression of Class I histone deacetylases. *J Neurotrauma* 34, 2645–2655.
- 196. Lu Y, Yang Y-Y, Zhou M-W, *et al.* (2018) Ketogenic diet attenuates oxidative stress and inflammation after spinal cord injury by activating Nrf2 and suppressing the NF-κB signaling pathways. *Neurosci Lett* **683**, 13–18.
- 197. Streijger F, Plunet WT, Lee JH, *et al.* (2013) Ketogenic diet improves forelimb motor function after spinal cord injury in rodents. *PLoS One* **8**, e78765.
- 198. Wang X, Wu X, Liu Q, *et al.* (2017) Ketogenic metabolism inhibits histone deacetylase (HDAC) and reduces oxidative stress after spinal cord injury in rats. *Neuroscience* **366**, 36–43.
- 199. Guo M, Wang X, Zhao Y, *et al.* (2018) Ketogenic diet improves brain ischemic tolerance and inhibits NLRP3 inflammasome

activation by preventing Drp1-mediated mitochondrial fission and endoplasmic reticulum stress. *Front Mol Neurosci* **11**, 86.

- Puchowicz M, Zechel J, Valerio J, et al. (2008) Neuroprotection in diet-induced ketotic rat brain after focal ischemia. J Cereb Blood Flow Metab 28, 1907–1916.
- 201. Rahman M, Muhammad S, Khan MA, *et al.* (2014) The β-hydroxybutyrate receptor HCA2 activates a neuroprotective subset of macrophages. *Nat Commun* 5, 1–11.
- Deng-Bryant Y, Prins M, Hovda D & Harris N (2011) Ketogenic diet prevents alterations in brain metabolism in young but not adult rats after traumatic brain injury. *J Neurotrauma* 28, 1813–1825.
- Greco T, Glenn T, Hovda D & Prins M (2016) Ketogenic diet decreases oxidative stress and improves mitochondrial respiratory complex activity. *J Cereb Blood Flow Metab* 36, 1603–1613.
- 204. Hu Z, Wang H, Jin W & Yin H (2009) Ketogenic diet reduces cytochrome c release and cellular apoptosis following traumatic brain injury in juvenile rats. *Ann Clin Lab Sci* **39**, 76–83.
- Hu Z, Wang H, Qiao L, *et al.* (2009) The protective effect of the ketogenic diet on traumatic brain injury-induced cell death in juvenile rats. *Brain Inj* 23, 459–465.
- Prins M, Fujima L & Hovda D (2005) Age-dependent reduction of cortical contusion volume by ketones after traumatic brain injury. *J Neurosci Res* 82, 413–420.
- Prins ML & Hovda DA (2009) The effects of age and ketogenic diet on local cerebral metabolic rates of glucose after controlled cortical impact injury in rats. *J Neurotrauma* 26, 1083–1093.
- 208. Salberg S, Weerwardhena H, Collins R, Reimer R & Mychasiuk R (2019) The behavioural and pathophysiological effects of the ketogenic diet on mild traumatic brain injury in adolescent rats. *Behav Brain Res* **376**, 112225.
- Schwartzkroin P, Wenzel H, Lyeth B, et al. (2010) Does ketogenic diet alter seizure sensitivity and cell loss following fluid percussion injury? *Epilepsy Res* 92, 74–84.
- Zhang F, Wu H, Jin Y & Zhang X (2018) Proton magnetic resonance spectroscopy (H1-MRS) study of the ketogenic diet on repetitive mild traumatic brain injury in adolescent rats and its effect on neurodegeneration. *World Neurosurg* **120**, e1193–e1202.
- Yang H, Shan W, Zhu F, Wu J & Wang Q (2019) Ketone bodies in neurological diseases: focus on neuroprotection and underlying mechanisms. *Front Neurol* 10, 585.
- 212. Morris G, Puri BK, Maes M, *et al.* (2020) The role of microglia in neuroprogressive disorders: mechanisms and possible neurotherapeutic effects of induced ketosis. *Prog Neuropsychopharmacol Biol Psychiatry* **99**, 109858.
- Spite M, Clària J & Serhan C (2014) Resolvins, specialized proresolving lipid mediators, and their potential roles in metabolic diseases. *Cell Metab* **19**, 21–36.
- 214. Totsch S, Waite M & Sorge R (2015) Dietary influence on pain via the immune system. In: Theodore JP & Gregory D, eds. *Progress in Molecular Biology and Translational Science*. Vol **131**. Cambridge: Academic Press, 435–469.

- 215. Farrell S, de Zoete R, Cabot P & Sterling M (2020) Systemic inflammatory markers in neck pain: a systematic review with meta-analysis. *Eur J Pain* **24**, 1666–1696.
- Schistad EI, Stubhaug A, Furberg A-S, Engdahl BL & Nielsen CS (2017) C-reactive protein and cold-pressor tolerance in the general population: the Tromsø Study. *Pain* **158**, 1280–1288.
- Dupuis N (2016) Anti- inflammatory effects of a ketogenic diet

 implications for new indications. In: Masino SA, ed.
 Ketogenic Diet and Metabolic Therapies: Expanded Roles in Health and Disease. Oxford, USA: Oxford University Press, Incorporated, 147–155.
- 218. Meeus M, Nijs J, Hermans L, Goubert D & Calders P (2013) The role of mitochondrial dysfunctions due to oxidative and nitrosative stress in the chronic pain or chronic fatigue syndromes and fibromyalgia patients: peripheral and central mechanisms as therapeutic targets? *Expert Opin Ther Targets* **17**, 1081–1089.
- Löffler M, Gamroth C, Becker S & Flor H (2020) Chronic pain as a neglected core symptom in mitochondrial diseases. *Neurology* 94, 357–359.
- 220. Van Den Ameele J, Fuge J, Pitceathly RD, *et al.* (2020) Chronic pain is common in mitochondrial disease. *Neuromuscul Disord* **30**, 413–419.
- 221. Sui B-d, Xu T-q, Liu J-w, *et al.* (2013) Understanding the role of mitochondria in the pathogenesis of chronic pain. *Postgrad Med J* **89**, 709–714.
- 222. Parker R, Lewis G, Rice D & McNair P (2016) Is motor cortical excitability altered in people with chronic pain? A systematic review and meta-analysis. *Brain Stimul* **9**, 488–500.
- 223. Neblett R, Cohen H, Choi Y, *et al.* (2013) The Central Sensitization Inventory (CSI): establishing clinically significant values for identifying central sensitivity syndromes in an outpatient chronic pain sample. *J Pain* **14**, 438–445.
- 224. Rezaei S, Abdurahman AA, Saghazadeh A, Badv RS & Mahmoudi M (2019) Short-term and long-term efficacy of classical ketogenic diet and modified Atkins diet in children and adolescents with epilepsy: a systematic review and meta-analysis. *Nutr Neurosci* **22**, 317–334.
- 225. Peek AL, Rebbeck T, Puts NAJ, *et al.* (2020) Brain GABA and glutamate levels across pain conditions: a systematic literature review and meta-analysis of 1H-MRS studies using the MRS-Q quality assessment tool. *Neuroimage* **210**, 116532.
- 226. Schugar RC, Huang X, Moll AR, Brunt EM & Crawford PA (2013) Role of choline deficiency in the fatty liver phenotype of mice fed a low protein, very low carbohydrate ketogenic diet. *PLoS One* **8**, e74806.
- 227. Anez-Bustillos L, Dao D, Finkelstein A, et al. (2019) Metabolic and inflammatory effects of an omega-3 fatty acid-based eucaloric ketogenic diet in mice with endotoxemia. J Parenter Enteral Nutr 43, 986–997.
- 228. Burma N, Leduc-Pessah H, Fan C & Trang T (2017) Animal models of chronic pain: advances and challenges for clinical translation. *J Neurosci Res* **95**, 1242–1256.
- Klinck MP, Mogil JS, Moreau M, *et al.* (2017) Translational pain assessment: could natural animal models be the missing link? *Pain* 158, 1633–1646.