

Role of Hand Hygiene Ambassador and Implementation of Directly Observed Hand Hygiene Among Residents in Residential Care Homes for the Elderly in Hong Kong

Vincent C. C. Cheng, MD;^{1,2} Hong Chen, MD;³ Shuk-Ching Wong, MNurs;² Jonathan H. K. Chen, PhD;¹ Wing-Chun Ng, MNurs;⁴ Simon Y. C. So, MMedSc;¹ Tuen-Ching Chan, MD;⁴ Sally C. Y. Wong, FRCPath;¹ Pak-Leung Ho, MD;¹ Lona Mody, MD;⁵ Felix H. W. Chan, MD;⁴ Andrew T. Y. Wong, MD;³ Kwok-Yung Yuen, MD¹

OBJECTIVE. Multidrug-resistant organisms (MDROs) are increasingly reported in residential care homes for the elderly (RCHEs). We assessed whether implementation of directly observed hand hygiene (DOHH) by hand hygiene ambassadors can reduce environmental contamination with MDROs.

METHODS. From July to August 2017, a cluster-randomized controlled study was conducted at 10 RCHEs (5 intervention versus 5 non-intervention controls), where DOHH was performed at two-hourly intervals during daytime, before meals and medication rounds by a one trained nurse in each intervention RCHE. Environmental contamination by MRDOs, such as methicillin-resistant *Staphylococcus aureus* (MRSA), carbapenem-resistant *Acinetobacter species* (CRA), and extended-spectrum β -lactamase (ESBL)-producing Enterobacteriaceae, was evaluated using specimens collected from communal areas at baseline, then twice weekly. The volume of alcohol-based hand rub (ABHR) consumed per resident per week was measured.

RESULTS. The overall environmental contamination of communal areas was culture-positive for MRSA in 33 of 100 specimens (33%), CRA in 26 of 100 specimens (26%), and ESBL-producing Enterobacteriaceae in 3 of 100 specimens (3%) in intervention and nonintervention RCHEs at baseline. Serial monitoring of environmental specimens revealed a significant reduction in MRSA (79 of 600 [13.2%] vs 197 of 600 [32.8%]; $P < .001$) and CRA (56 of 600 [9.3%] vs 94 of 600 [15.7%]; $P = .001$) contamination in the intervention arm compared with the nonintervention arm during the study period. The volume of ABHR consumed per resident per week was 3 times higher in the intervention arm compared with the baseline (59.3 ± 12.9 mL vs 19.7 ± 12.6 mL; $P < .001$) and was significantly higher than the nonintervention arm (59.3 ± 12.9 mL vs 23.3 ± 17.2 mL; $P = .006$).

CONCLUSIONS. The direct observation of hand hygiene of residents could reduce environmental contamination by MDROs in RCHEs.

Infect Control Hosp Epidemiol 2018;39:571–577

Transmission of multidrug-resistant organisms (MDROs) in long-term care facilities is an emerging challenge for infection control professionals. It has been made more complex by the significant amount of movement of residents between acute-care hospitals and long-term care facilities, where transmission and acquisition of hospital-acquired infections may occur frequently, including MDROs.¹ In our transmission dynamic analysis, the likelihood of methicillin-resistant *Staphylococcus aureus* (MRSA) transmission in long-term care facilities was 3 times higher than in hospital settings.² With the advances in laboratory diagnostics, resident-to-resident transmissions

have been documented more accurately by whole-genome sequencing, as illustrated in a cluster of MRSA isolates in a long-term care facility.³ Infection control measures against nosocomial transmission of MDROs currently focus on the most basic interventions, hand hygiene practice, and environmental cleanliness. Specifically, hand hygiene interventions have been shown to reduce infection transmission risks,^{4,5} but hand hygiene compliance has remained low in long-term care facilities. In Hong Kong, a multifaceted hand hygiene intervention with the provision of alcohol-based hand rub (ABHR), hand hygiene posters, training materials, and feedback was

Affiliations: 1. Department of Microbiology, Queen Mary Hospital, Hong Kong Special Administrative Region, China; 2. Infection Control Team, Queen Mary Hospital, Hong Kong West Cluster, Hong Kong Special Administrative Region, China; 3. Infection Control Branch, Centre for Health Protection, Department of Health, Hong Kong Special Administrative Region, China; 4. Community Geriatric Assessment Team, Fung Yiu King Hospital, Hong Kong West Cluster, Hong Kong Special Administrative Region, China; 5. Geriatric Research Education and Clinical Center, Ann Arbor Veteran Affairs Healthcare System, Ann Arbor, Michigan.

Received December 3, 2017; accepted January 10, 2018; electronically published February 27, 2018

© 2018 by The Society for Healthcare Epidemiology of America. All rights reserved. 0899-823X/2018/3905-0009. DOI: 10.1017/ice.2018.21

implemented in long-term care facilities.⁶ Although the hand hygiene compliance improved from 27% to 60% during the study period, the prevalence of MDROs continued to increase, especially MRSA, which has risen from 21.6% to 32.2% in our long-term care facilities.^{2,7} The effect of the WHO Five Moments hand hygiene campaign was not sustained, which may be attributed to the relatively low education level of staff in our long-term care facilities, where almost 50% of the staff have an education below the secondary level.⁸

Directly observed hand hygiene (DOHH) with delivery of ABHR to conscious patients before meals and medication rounds by a designated healthcare worker, namely a hand hygiene ambassador,⁹ has been evaluated in the hospital setting with proven success in the control of vancomycin-resistant enterococci (VRE) and multidrug-resistant *Acinetobacter baumannii*.^{10,11} Therefore, we performed a cluster-randomized controlled trial to evaluate the practicality and effect of DOHH for residents in long-term care facilities, also known as residential care homes for the elderly (RCHEs) in our locality. Colonization with MDROs on the hands of residents were significantly associated with environmental contamination,¹² and environmental contamination by MDROs have been shown to precede patients' acquisition of MDROs.^{13–15} We hypothesized that the degree of environmental contamination by MDROs would be significantly lower in the intervention arm than in the nonintervention arm during our serial monitoring throughout the study period. These findings may facilitate strategic planning of infection control programs in long-term care facilities.

METHODS

Setting

This cluster-randomized controlled study was conducted between July 1, 2017, and August 31, 2017, to assess the implementation of DOHH among 10 of the 65 RCHEs in a healthcare region in the Hong Kong Western District, which serves a population of 0.53 million. The study was conducted within a relatively short period in summer because the rate of hospitalization among residents is generally lower when the ambient temperature is higher in Hong Kong. The role of RCHEs was described previously.^{2,7} Briefly, an RCHE is a long-term care facility that provides daily nursing care to residents, including the use of feeding tubes, urinary catheters, and other medical devices in Hong Kong. Community geriatric assessment teams (CGATs) comprising of a team of geriatricians, nurses, and allied health professionals provide regular on-site visits to the RCHEs to perform comprehensive medical assessments. A hospital infection control team and the Infection Control Branch of the Center for Health Protection provide recommendations on infection control measures. This study was approved by the Institutional Review Board of the University of Hong Kong/Hospital Authority Hong Kong West Hospital Cluster. Written consent was obtained from recruited residents.

Participants

Based on the prevalence survey of MDROs, including MRSA, VRE and carbapenem-resistant *Acinetobacter* spp (CRA), among RCHEs in July and August 2015,⁷ RCHEs of comparable size with similar prevalence of MDROs were selected to participate in this study (Supplementary Figure 1). We stratified the RCHEs into 2 groups according to their prevalence of MDROs (MRSA colonization <35% vs ≥35%), and their size as reflected by number of resident registrations. An RCHE was classified as small if it had 25–60 residents, moderate if it had 61–120 residents, and large if it had 121–180 residents. In each group, the RCHEs were randomized and allocated into the intervention arm or the nonintervention arm in a 1:1 ratio. The investigators, staff, and residents of the RCHEs were not blinded to the allocations. To establish the updated MDRO prevalence among residents and environment, research nurses who received prior training from the infection control team obtained nasal swabs, axillary swabs, rectal swabs, and environmental samples using sponge swabs according to a standard protocol as previously described.^{7,16,17} Written consent forms were obtained from the participating residents. Subjects were excluded if they were not able to provide consent. Demographic information, dependency status, Barthel index, presence of indwelling devices and comorbidities were retrieved for each resident. History of hospitalization and antimicrobial usage in the prior 6 months were also analyzed. Risk factors for MDROs colonization among residents in RCHEs were assessed.

Intervention

In the intervention arm, DOHH was performed by a hand hygiene ambassador (a designated research nurse) immediately before meals and medication rounds in conscious residents and at 2-hour intervals between 9:00 AM and 6:00 PM during weekdays in each RCHE. The hand hygiene ambassador delivered 3 mL ABHR to the hands of residents per occurrence of DOHH either at the communal areas or at the bedside. A pocket-sized 60-mL ABHR container was used by the research nurse, and standard-sized 500-mL ABHR containers were placed in the cubicle, corridor, and communal areas of RCHEs for the residents, staff, and visitors. The World Health Organization (WHO) formulation (I and II) of ABHR was used. In the nonintervention arm, DOHH was not performed, but the WHO formulation of ABHR was provided at each RCHE. In both the intervention and nonintervention arms, the volume of ABHR consumption was measured by the research nurses in all recruited RCHEs on a weekly basis during the entire study period as an indirect surrogate marker of hand hygiene compliance as previously described.¹⁸ The baseline volume of ABHR consumption by each RCHE was estimated from the purchase invoices of ABHR.

Environmental samples were also collected from 10 selected sites in the communal areas (cuff of the portable blood

pressure machine, meal table top, activity table top, chair's armrest, hand rail of corridor, and remote control of television), and staff area (staff station table top, computer keyboard and mouse, trolley top and handle, and telephone handle), respectively, twice weekly immediately before environmental cleaning in each RCHE. Environmental samples were collected using Polywipe sponge swabs 5 cm × 10 cm in size (Medical Wire and Equipment, UK) as previously described.¹⁷ These swabs were sterile, premoistened, thin flexible sponges tailor-made for sampling environmental surfaces. Each sampled sponge swab was placed in an individual sterile plastic bag, which was sealed and clearly labeled before further processing in the laboratory. The protocol of twice-daily environmental cleaning with sodium hypochlorite 500 ppm in each RCHE remained unchanged throughout the study period. The serial environmental culture results of MDROs in the communal areas were analyzed.

Laboratory Investigation

The screening specimens from the residents were processed, and suspected isolates of MRSA, CRA, and extended-spectrum β-lactamase (ESBL)-producing Enterobacteriaceae from nasal, axillary, groin, and rectal swabs were identified using standard techniques described previously.^{7,19} We did not screen for VRE and CPE because of its low prevalence in our RCHE.^{7,20} For each sponge swab, 2 mL sterile normal saline (0.85% sodium chloride) was added to the sterile plastic bag. The sponge was squeezed repeatedly for proper mixing, then 100 μL suspension was transferred from the bag for processing as previously described.^{7,19}

Statistical Analysis

The Fisher exact test and Student *t* test were used where appropriate. SPSS version 20 software (SPSS Institute, Cary, NC) was used to perform the statistical analyses. All reported *P* values were 2-sided. A *P* value of <.05 was considered statistically significant.

RESULTS

Baseline Characteristics

In this study, we randomized 5 RCHEs to the intervention arm and 5 RCHEs to the nonintervention control arm. The post-randomization baseline characteristics of the intervention and nonintervention arms were not statistically different (Supplementary Table 1). Of the 774 residents registered in 10 RCHEs in the healthcare region of the Hong Kong Western District, 515 (66.5%) consented for MDROs screening. The cohort consisted of 291 women (56.5%) and 224 men (43.5%), with an overall mean age of 84.7 ± 9.4 years. At baseline, the prevalence of MRSA was 37.9%; the prevalence of CRA was 9.1%, and the prevalence of ESBL-producing Enterobacteriaceae was 55.9%. Overall, 369 of 515 residents (71.7%) were colonized with at least 1 MDRO. The bedside environmental samples from

the bedside rails of 515 residents were found to be contaminated by MRSA (14.4% of samples), CRA (7.6%), and ESBL-producing Enterobacteriaceae (0.6%). Samples from the table tops of these residents were also contaminated by MRSA (17.9% of samples), CRA (9.5%), and ESBL-producing Enterobacteriaceae (1.4%). For residents known to be colonized with MRSA, in their bedside environment, MRSA cultured positive for 26.7% of bedside rail samples and 33.3% of table top samples. For residents known to be colonized with CRA, the prevalence of CRA was 36.2% for bedside rails and 38.3% for table tops. For residents known to be colonized with ESBL-producing Enterobacteriaceae, the prevalence of ESBL-producing Enterobacteriaceae was 1.0% for bedside rails and 2.1% for table tops. At baseline, positive culture results were noted from the communal environment, where 33 of 100 swabs yielded MRSA (33%), 26 of 100 swabs yielded CRA (26%), and 3 of 100 swabs yielded ESBL-producing Enterobacteriaceae (3%). The epidemiological characteristics of these 515 residents, as well as the risk factors for MRSA, CRA, and ESBL-producing Enterobacteriaceae colonization are illustrated in Table 1 and Supplementary Table.

Reduction of Environmental Contamination

Serial environmental specimens in the communal areas revealed a significant reduction of environmental MRSA contamination in the intervention arm when compared with the nonintervention arm during the study period: 79 of 600 (13.2%) versus 197 of 600 (32.8%) (*P* < .001) (Table 2). In addition, a significant reduction of environmental CRA contamination was observed in the intervention arm compared with the nonintervention arm during the study period: 56 of 600 (9.3%) versus 94 of 600 (15.7%) (*P* = .001).

Consumption of Alcohol-Based Hand Rub

With the implementation of DOHH in the intervention arm during the study period, the volume of ABHR consumed per resident per week was 3 times higher in the intervention arm compared with the baseline (59.3 ± 12.9 mL vs 19.7 ± 12.6 mL; *P* < .001) and was significantly higher than the nonintervention arm (59.3 ± 12.9 mL vs 23.3 ± 17.2 mL; (*P* = .006) (Table 3). In the nonintervention arm, no significant difference was noted in ABHR consumption compared with the baseline (23.3 ± 17.2 mL vs 18.8 ± 15.5; *P* = .207).

DISCUSSION

With the implementation of DOHH by hand hygiene ambassadors in the RCHEs, we have demonstrated a significant reduction of environmental contamination of MRSA and CRA in the communal environment of the RCHEs. In fact, RCHEs are important reservoirs for MDROs. As illustrated in this study, environmental contamination rates of MRSA, CRA, and ESBL-producing Enterobacteriaceae were 33%, 26%,

TABLE 1. Epidemiological Characteristics of Residents Colonized With Multidrug-Resistant Organisms in the Residential Care Home for the Elderly in Both Intervention Arm (5 Homes) and Nonintervention Arm (5 Homes)

Patient Variables	MRSA			CRA			ESBL-E		
	Positive (n = 195)	Negative (n = 320)	P Value	Positive (n = 47)	Negative (n = 468)	P Value	Positive (n = 288)	Negative (n = 227)	P Value
Age, mean $\bar{y} \pm SD$	84.6 \pm 9.2	84.9 \pm 9.5	.756	84.6 \pm 8.6	84.8 \pm 9.5	.855	83.8 \pm 9.6	85.9 \pm 8.9	.072
Male sex, No. (%)	98 (50.3)	126 (39.4)	.016	32 (68.1)	192 (41.0)	<.001	134 (46.5)	90 (39.6)	.118
Presence of									
Nasogastric tube, No. (%)	12 (6.2)	6 (1.9)	.010	6 (12.8)	12 (2.6)	<.001	14 (4.9)	4 (1.8)	.057
Foley catheter, No. (%)	17 (8.7)	13 (4.1)	.029	2 (4.3)	28 (6.0)	.630	21 (7.3)	9 (4.0)	.110
Wound, No. (%)	11 (5.6)	12 (3.8)	.314	5 (10.6)	18 (3.8)	.032	14 (4.9)	9 (4.0)	.625
Bedsore, No. (%)	9 (4.6)	6 (1.9)	.073	3 (6.4)	12 (2.6)	.149	9 (3.1)	6 (2.6)	.747
Barthel index (mean total score \pm SD)	33.4 \pm 34.0	55.9 \pm 36.6	<.001	22.0 \pm 25.8	48.6 \pm 37.1	<.001	45.2 \pm 37.5	47.4 \pm 36.4	.507
Underlying diseases									
Chronic cardiac conditions, No. (%)	50 (25.6)	83 (25.9)	.941	11 (23.4)	122 (26.1)	.691	76 (26.4)	57 (25.1)	.742
Chronic cerebral conditions, No. (%)	33 (16.9)	47 (14.7)	.497	12 (25.5)	68 (14.5)	.057	42 (14.6)	38 (16.7)	.502
Chronic pulmonary conditions, No. (%)	43 (22.1)	60 (18.8)	.364	10 (21.3)	93 (19.9)	.818	53 (18.4)	50 (22.0)	.307
Diabetes mellitus, No. (%)	22 (11.3)	29 (9.1)	.413	6 (12.8)	45 (9.6)	.491	32 (11.1)	19 (8.4)	.301
Malignancies, No. (%)	13 (6.7)	21 (6.6)	.963	2 (4.3)	32 (6.8)	.758	18 (6.3)	16 (7.0)	.717
Use of antibiotics in the past 6 mo									
β -lactam/ β -lactamase inhibitors, No. (%)	21 (10.8)	15 (4.7)	.009	7 (14.9)	29 (6.2)	.036	24 (8.3)	12 (5.3)	.178
Cephalosporins, No. (%)	4 (2.1)	5 (1.6)	.736	2 (4.3)	7 (1.5)	.194	5 (1.7)	4 (1.8)	.982
Carbapenems, No. (%)	12 (6.2)	6 (1.9)	.010	7 (14.9)	11 (2.4)	<.001	13 (4.5)	5 (2.2)	.156
Fluoroquinolones, No. (%)	7 (3.6)	8 (2.5)	.476	5 (10.6)	10 (2.1)	.001	9 (3.1)	6 (2.6)	.747
History of hospitalization in the past 6 mo, No. (%)	99 (50.8)	122 (38.1)	.005	30 (63.8)	191 (40.8)	.002	127 (44.1)	94 (41.4)	.541

NOTE. CRA, carbapenem-resistant *Acinetobacter* spp; ESBL-E; extended-spectrum β -lactamase-producing Enterobacteriaceae; MDROs, multidrug-resistant organisms; MRSA, methicillin-resistant *Staphylococcus aureus*; SD, standard deviation.

and 3%, respectively, in the communal areas, while the rates of resident colonization by MRSA, CRA, and ESBL-producing Enterobacteriaceae, reached 38%, 9%, and 56%, respectively.

The burden of MDROs in Hong Kong is comparatively higher than in many European countries²¹ or in the United States.²² The high prevalence of MDROs in our RCHEs may be multifactorial. First, overcrowding is a significant problem in Hong Kong, and we have previously shown that the prevalence of MRSA in RCHEs was inversely associated with the average living area (square feet per person) per RCHE resident.² One-third of sites sampled from the communal area of the RCHE were shown to be culture positive for MRSA, which was twice the corresponding figure in nursing homes in the United States.²³ Because communal areas have a high contamination rate, secondary contamination of the surrounding patients may occur regardless of the patient's MRSA status. Data on environmental contamination by CRA in long-term care facilities or nursing homes remained limited in the literature. However, we noticed that CRA contamination was up to 26% in the RCHE environment. The degree of contamination was

even more astounding in the context of a lower resident colonization rate of 9%, and CRA has been an emerging threat in RCHEs in recent years.⁷ Second, the frequent movements of residents between RCHEs and acute-care hospitals may further increase the risk of MDRO acquisition and a previous study showed that a significant fraction of RCHE residents acquired MDRO colonization or infection during their stay in acute-care hospitals.²⁴ Nearly 50% of our residents have a history of hospitalization in the 6 months prior to the study. The use of broad-spectrum antibiotics, especially β -lactam- β -lactamase inhibitors and carbapenems, was also invariably a risk factor for MRSA and CRA colonization in this study. In addition, residents colonized with MRSA and CRA were more dependent in daily activity as evidenced by a significantly lower Barthel index, which was similar to another recent finding.²⁵

To control the emergence of MDROs in RCHEs, effective infection control programs are required in both RCHEs and acute-care hospitals. Proactive infection control measures have been promulgated in our acute-care hospitals to combat emerging VRE and CPE with success.^{26,27}

TABLE 2. Serial Monitoring of Environmental Contamination in Communal Areas by Multidrug-Resistant Organisms in 10 Residential Care Homes for the Elderly Under the Category of Intervention Arm (5 Homes) and Nonintervention Arm (5 Homes)

	Environmental Sample Collected per Time Point in Intervention Arm, n/N (%) ^a	Environmental Sample Collected per Time Point in Nonintervention Arm, n/N (%) ^a	P Value
Contamination of MRSA			
Baseline (time point 0)	16/50 (32) ^b	17/50 (34)	.248
Week 1 (time point 1)	5/50 (10)	10/50 (20)	.161
Week 1 (time point 2)	5/50 (10)	15/50 (30)	.012
Week 2 (time point 3)	8/50 (16)	13/50 (26)	.220
Week 2 (time point 4)	6/50 (12)	14/50 (28)	.046
Week 3 (time point 5)	7/50 (14)	15/50 (30)	.053
Week 3 (time point 6)	4/50 (8)	18/50 (36)	.001
Week 4 (time point 7)	5/50 (10)	17/50 (34)	.004
Week 4 (time point 8)	5/50 (10)	14/50 (28)	.022
Week 5 (time point 9)	5/50 (10)	18/50 (36)	.002
Week 5 (time point 10)	3/50 (6)	13/50 (26)	.006
Week 6 (time point 11)	5/50 (10)	18/50 (36)	.002
Week 6 (time point 12)	5/50 (10)	15/50 (30)	.012
Weeks 1–6 (time points 1–12)	79/600 (13.2) ^b	197/600 (32.8)	<.001
Contamination of CRA			
Baseline (time point 0)	16/50 (32) ^c	10/50 (20)	.171
Week 1 (time point 1)	3/50 (6)	6/50 (12)	.295
Week 1 (time point 2)	4/50 (8)	3/50 (6)	.695
Week 2 (time point 3)	8/50 (16)	12/50 (24)	.317
Week 2 (time point 4)	4/50 (8)	3/50 (6)	.695
Week 3 (time point 5)	5/50 (10)	9/50 (18)	.249
Week 3 (time point 6)	5/50 (10)	7/50 (14)	.538
Week 4 (time point 7)	4/50 (8)	9/50 (18)	.137
Week 4 (time point 8)	6/50 (12)	8/50 (16)	.564
Week 5 (time point 9)	4/50 (8)	3/50 (6)	.695
Week 5 (time point 10)	2/50 (4)	12/50 (24)	.004
Week 6 (time point 11)	5/50 (10)	13/50 (26)	.037
Week 6 (time point 12)	6/50 (12)	9/50 (18)	.401
Weeks 1–6 (time points 1–12)	56/600 (9.3) ^c	94/600 (15.7)	.001
Contamination of ESBL-E			
Baseline (time point 0)	2/50 (4.0) ^d	1/50 (2.0)	.999
Weeks 1–6 (time points 1–12)	8/600 (1.3) ^d	7/600 (1.2)	.795

NOTE. CRA, carbapenem-resistant *Acinetobacter* spp; ESBL-E; extended-spectrum β -lactamase-producing Enterobacteriaceae; MRSA, methicillin-resistant *Staphylococcus aureus*.

^a10 selected sites in the communal area (ie, cuff of the portable blood pressure machine, meal table top, activity table top, chair's armrest, hand rail of corridor, and remote control of television) and staff area (ie, staff station table top, computer keyboard and mouse, trolley top and handle, and telephone handle) twice weekly immediately before environmental cleanliness in each RCHE.

A total of 50 environmental samples were collected in 5 RCHEs in the intervention arm and another 50 environmental samples were collected in 5 RCHEs in the nonintervention arm at each time point.

^bA significant reduction in environmental MRSA contamination in the intervention arm compared with the baseline (79/600, 13.2% vs 16/50, 32.0%; $P = .005$).

^cA significant reduction in environmental CRA contamination in the intervention arm compared with the baseline (56/600, 9.3% vs 16/50, 32.0%; $P < .001$).

^dNo significant difference of environmental ESBL-producing Enterobacteriaceae contamination between baseline and study period in the intervention arm ($P = .176$).

This may explain the very low prevalence of VRE and CPE in our RCHEs.⁷ We believe that the promotion of proactive infection control measures have greatly reduced the risk of

hospital outbreaks due to epidemiologically important viruses and MDROs and that these measurements can be applied to our RCHEs.²⁸

TABLE 3. Consumption of Volume of Alcohol-Based Hand Rub (ABHR) per Conscious Resident per Week in 10 Residential Care Homes for the Elderly Under the Category of Intervention Arm (5 Homes) and Nonintervention Arm (5 Homes)

ABHR Consumption	ABHR per Resident per Week in the Intervention Arm, mean mL ± SD	ABHR per Resident per Week in the Nonintervention Arm, mean mL ± SD	P Value
Baseline	19.7 ± 12.6 ^a	18.8 ± 15.5 ^b	.921
Week 1	29.9 ± 11.0	10.1 ± 6.5	.008
Week 2	55.2 ± 15.2	18.6 ± 16.1	.006
Week 3	63.6 ± 12.6	25.6 ± 19.2	.006
Week 4	73.3 ± 21.2	27.1 ± 35.1	.036
Week 5	64.2 ± 22.1	24.0 ± 20.1	.018
Week 6	69.2 ± 35.1	34.5 ± 33.3	.148
Overall, weeks 1–6	59.3 ± 12.9 ^a	23.3 ± 17.2 ^b	.006

NOTE. SD, standard deviation.

^aThe volume of ABHR consumption per resident per week was statistically higher compared with the baseline ($P < .001$).

^bThe volume of ABHR consumption per resident per week was not statistically difference compared with the baseline ($P = .207$).

Of these proactive infection control measures, the use of DOHH in all conscious hospitalized patients before meal and medication rounds is a promising strategy.²⁹ The compliance of hand hygiene among hospitalized patients or residents in RCHes can be assured if the ABHR is delivered directly by the healthcare workers at these most critical times. Therefore, we implemented DOHH in the RCHes and evaluated its effect through this randomized control trial. A hand hygiene ambassador was responsible for delivering 3 mL ABHR to all conscious residents on a regular basis and before meal and medication rounds. This measure was well received by the residents. It was shown to reduce the risk of oral ingestion of MDROs via contaminated hands, and vice versa because there is lower risk of environmental contamination when fewer residents are carrying MDROs.¹² Throughout the study period, the proportion of environmental samples culture positive with MRSA and CRA was significantly lower in the RCHes with DOHH (intervention arm) than in those without (non-intervention arm). The volume of ABHR delivered per resident per week was also significantly higher in the intervention arm.

This study has several limitations. First, the study period was only 8 weeks, thus the long-term impact of this intervention could not be assessed. Second, although the reduction of environmental contamination by MRSA and CRA in the intervention arm showed an immediate and promising effect of DOHH, we could not exclude the possibility of spillage of ABHR onto the sampled surfaces resulting in reduced contamination by inadvertently disinfecting surfaces.³⁰ Third, the change of environmental contamination by ESBL-producing Enterobacteriaceae was not obvious in this study. Despite a high prevalence of residents colonized with

ESBL-producing Enterobacteriaceae, a disproportionately low culture-positive rate of ESBL-producing Enterobacteriaceae in the environment was noted. This could be due to the shorter duration of viability of Enterobacteriaceae on hands and inanimate surface, compared with *Staphylococcus aureus* and *Acinetobacter* spp.^{31,32} Finally, the infection control program required a hand hygiene ambassador assigned to facilitate DOHH. This may affect the practicality of routine implementation of such strategies in RCHes. Further study is warranted to investigate the cost-effectiveness of the implementation of DOHH in the control of MDROs in RCHes.

ACKNOWLEDGMENTS

Financial support: This study was partially supported by the Health and Medical Research Fund (HMRF), Food and Health Bureau, Hong Kong SAR Government (grant no. HKM-15-M12), and the Infection Control Branch of Centre for Health Protection, Department of Health, Hong Kong SAR Government.

Potential conflicts of interest: All authors report no conflicts of interest relevant to this article.

Address correspondence to Kwok-Yung Yuen, Department of Microbiology, Queen Mary Hospital, The University of Hong Kong, Hong Kong Special Administrative Region, China (kyyuen@hku.hk).

SUPPLEMENTARY MATERIAL

To view supplementary material for this article, please visit <https://doi.org/10.1017/ice.2018.21>.

REFERENCES

1. Smith PW, Bennett G, Bradley S, et al. SHEA/APIC guideline: infection prevention and control in the long-term care facility, July 2008. *Infect Control Hosp Epidemiol* 2008; 29:785–814.
2. Cheng VC, Tai JW, Wong ZS, et al. Transmission of methicillin-resistant *Staphylococcus aureus* in the long term care facilities in Hong Kong. *BMC Infect Dis* 2013;13:205.
3. Stine OC, Burrows S, David S, Johnson JK, Roghmann MC. Transmission clusters of methicillin-resistant *Staphylococcus aureus* in long-term care facilities based on whole-genome sequencing. *Infect Control Hosp Epidemiol* 2016;37:685–691.
4. Hocine MN, Temime L. Impact of hand hygiene on the infectious risk in nursing home residents: a systematic review. *Am J Infect Control* 2015;43:e47–e52.
5. Schweon SJ, Edmonds SL, Kirk J, Rowland DY, Acosta C. Effectiveness of a comprehensive hand hygiene program for reduction of infection rates in a long-term care facility. *Am J Infect Control* 2013;41:39–44.
6. Ho ML, Seto WH, Wong LC, Wong TY. Effectiveness of multifaceted hand hygiene interventions in long-term care facilities in Hong Kong: a cluster-randomized controlled trial. *Infect Control Hosp Epidemiol* 2012;33:761–767.
7. Cheng VC, Chen JH, Ng WC, et al. Emergence of carbapenem-resistant *Acinetobacter baumannii* in nursing homes with high background rates of MRSA colonization. *Infect Control Hosp Epidemiol* 2016;37:983–986.

8. Chan TC, Luk JK, Chu LW, Chan FH. Low education level of nursing home staff in Chinese nursing homes. *J Am Med Dir Assoc* 2013;14:849–850.
9. Cheng VC, Tai JW, Li WS, et al. Implementation of directly observed patient hand hygiene for hospitalized patients by hand hygiene ambassadors in Hong Kong. *Am J Infect Control* 2016;44:621–624.
10. Cheng VC, Tai JW, Chau PH, et al. Successful control of emerging vancomycin-resistant enterococci by territory-wide implementation of directly observed hand hygiene in patients in Hong Kong. *Am J Infect Control* 2016;44:1168–1171.
11. Cheng VC, Chen JH, Poon RW, et al. Control of hospital endemicity of multiple-drug-resistant *Acinetobacter baumannii* ST457 with directly observed hand hygiene. *Eur J Clin Microbiol Infect Dis* 2015;34:713–718.
12. Patel PK, Mantey J, Mody L. Patient hand colonization with MDROs is associated with environmental contamination in post-acute care. *Infect Control Hosp Epidemiol* 2017;38:1110–1113.
13. Hardy KJ, Oppenheim BA, Gossain S, Gao F, Hawkey PM. A study of the relationship between environmental contamination with methicillin-resistant *Staphylococcus aureus* (MRSA) and patients' acquisition of MRSA. *Infect Control Hosp Epidemiol* 2006;27:127–132.
14. Drees M, Snyderman DR, Schmid CH, et al. Prior environmental contamination increases the risk of acquisition of vancomycin-resistant enterococci. *Clin Infect Dis* 2008;46:678–685.
15. Cheng VCC, Wong SC, Chen JHK, et al. Control of multidrug-resistant *Acinetobacter baumannii* in Hong Kong: role of environmental surveillance in communal areas after a hospital outbreak. *Am J Infect Control* 2018;46:60–66.
16. Cheng VC, Li IW, Wu AK, et al. Effect of antibiotics on the bacterial load of methicillin-resistant *Staphylococcus aureus* colonisation in anterior nares. *J Hosp Infect* 2008;70:27–34.
17. Cheng VCC, Chen JHK, Wong SCY, et al. Hospital outbreak of pulmonary and cutaneous zygomycosis due to contaminated linen items from substandard laundry. *Clin Infect Dis* 2016;62:714–721.
18. Cheng VCC, Wong SC, Wong IWY, et al. The challenge of patient empowerment in hand hygiene promotion in health care facilities in Hong Kong. *Am J Infect Control* 2017;45:562–565.
19. Cheng VC, Chan JF, Lau EH, et al. Studying the transmission dynamics of methicillin-resistant *Staphylococcus aureus* in Hong Kong using spa typing. *J Hosp Infect* 2011;79:206–210.
20. Cheng VC, Chen JH, So SY, et al. A novel risk factor associated with colonization by carbapenemase-producing Enterobacteriaceae: use of proton pump inhibitors in addition to antimicrobial treatment. *Infect Control Hosp Epidemiol* 2016;37:1418–1425.
21. Aschbacher R, Pagani E, Confalonieri M, et al. Review on colonization of residents and staff in Italian long-term care facilities by multidrug-resistant bacteria compared with other European countries. *Antimicrob Resist Infect Control* 2016;5:33.
22. Cassone M, Mody L. Colonization with multi-drug resistant organisms in nursing homes: scope, importance, and management. *Curr Geriatr Rep* 2015;4:87–95.
23. Murphy CR, Eells SJ, Quan V, et al. Methicillin-resistant *Staphylococcus aureus* burden in nursing homes associated with environmental contamination of common areas. *J Am Geriatr Soc* 2012;60:1012–1018.
24. Kahvecioglu D, Ramiah K, McMaughan D, et al. Multidrug-resistant organism infections in US nursing homes: a national study of prevalence, onset, and transmission across care settings, October 1, 2010–December 31, 2011. *Infect Control Hosp Epidemiol* 2014;35(Suppl 3):S48–S55.
25. Rollnik JD, Bertram M, Bucka C, et al. Outcome of neurological early rehabilitation patients carrying multi-drug resistant bacteria: results from a German multi-center study. *BMC Neurol* 2017;17:53.
26. Cheng VC, Tai JW, Chen JH, et al. Proactive infection control measures to prevent nosocomial transmission of vancomycin-resistant enterococci in Hong Kong. *J Formos Med Assoc* 2014;113:734–741.
27. Cheng VC, Chan JF, Wong SC, et al. Proactive infection control measures to prevent nosocomial transmission of carbapenem-resistant Enterobacteriaceae in a non-endemic area. *Chin Med J (Engl)* 2013;126:4504–4509.
28. Cheng VC, Tai JW, Wong LM, et al. Effect of proactive infection control measures on benchmarked rate of hospital outbreaks: an analysis of public hospitals in Hong Kong over 5 years. *Am J Infect Control* 2015;43:965–970.
29. Cheng VC, Wong SC, Ho PL, Yuen KY. Strategic measures for the control of surging antimicrobial resistance in Hong Kong and mainland of China. *Emerg Microbe Infect* 2015;4:e8.
30. Adams CE, Smith J, Watson V, Robertson C, Dancer SJ. Examining the association between surface bioburden and frequently touched sites in intensive care. *J Hosp Infect* 2017;95:76–80.
31. Kampf G, Kramer A. Epidemiologic background of hand hygiene and evaluation of the most important agents for scrubs and rubs. *Clin Microbiol Rev* 2004;17:863–893.
32. Kramer A, Schwebke I, Kampf G. How long do nosocomial pathogens persist on inanimate surfaces? A systematic review. *BMC Infect Dis* 2006;6:130.