

# Olanzapine plus fluoxetine treatment alters mitochondrial respiratory chain activity in the rat brain

Agostinho FR, Réus GZ, Stringari RB, Ribeiro KF, Ferreira GK, Jeremias IC, Scaini G, Rezin GT, Streck EL, Quevedo J. Olanzapine plus fluoxetine treatment alters mitochondrial respiratory chain activity in the rat brain.

**Fabiano R. Agostinho<sup>1</sup>,  
Gislaine Z. Réus<sup>1</sup>, Roberto B. Stringari<sup>1</sup>,  
Karine F. Ribeiro<sup>1</sup>, Gabriela K. Ferreira<sup>2</sup>,  
Isabela C. Jeremias<sup>2</sup>, Giselli Scaini<sup>2</sup>,  
Gislaine T. Rezin<sup>2</sup>, Emílio L. Streck<sup>2</sup>,  
João Quevedo<sup>1</sup>**

<sup>1</sup>Laboratório de Neurociências and Instituto Nacional de Ciência e Tecnologia Translacional em Medicina (INCT-TM), Programa de Pós-Graduação em Ciências da Saúde, Unidade Acadêmica de Ciências da Saúde, Universidade do Extremo Sul Catarinense, Criciúma, Santa Catarina, Brazil; and <sup>2</sup>Laboratório de Fisiopatologia Experimental and Instituto Nacional de Ciência e Tecnologia Translacional em Medicina (INCT-TM), Programa de Pós-Graduação em Ciências da Saúde, Unidade Acadêmica de Ciências da Saúde, Universidade do Extremo Sul Catarinense, Criciúma, Santa Catarina, Brazil

Keywords: bipolar depression; bipolar disorder; fluoxetine; mitochondrial respiratory chain; olanzapine

Professor João Quevedo, MD, PhD, Laboratório de Neurociências and Instituto Nacional de Ciência e Tecnologia Translacional em Medicina (INCT-TM), Programa de Pós-Graduação em Ciências da Saúde, Unidade Acadêmica de Ciências da Saúde, Universidade do Extremo Sul Catarinense, Criciúma 88806-000, Santa Catarina, Brazil.  
Tel: +55 48 3443 4817;  
Fax: +55 48 3431 2736;  
E-mail: quevedo@unesc.net

**Background:** Evidence is emerging for the role of dysfunctional mitochondria in pathophysiology and treatment of mood disorders. In this study, we evaluated the effects of acute and chronic administration of fluoxetine (FLX), olanzapine (OLZ) and the combination of FLX/OLZ on mitochondrial respiratory chain activity in the rat brain.

**Methods:** For acute treatment, Wistar rats received one single injection of OLZ (3 or 6 mg/kg) and/or FLX (12 or 25 mg/kg) and for chronic treatment, rats received daily injections of OLZ (3 or 6 mg/kg) and/or FLX (12 or 25 mg/kg) for 28 days and we evaluated the activity of mitochondrial respiratory chain complexes I, II, II–III and IV in prefrontal cortex, hippocampus and striatum.

**Results:** Our results showed that both acute and chronic treatments with FLX and OLZ alone or in combination altered respiratory chain complexes activity in the rat brain, but in combination we observed larger alterations.

**Conclusions:** Finally, these findings further support the hypothesis that metabolism energy could be involved in the treatment with antipsychotics and antidepressants in combination to mood disorders.

## Significant outcomes

- Effects of FLX and OLZ alone or in combination on mitochondrial respiratory chain.
- Acute and chronic treatment of FLX and OLZ alone or in combination altered respiratory chain complexes activity.
- In combination, FLX and OLZ caused larger alterations.

## Limitations

- The results did not follow a pattern. Sometimes FLX and/or OLZ increased and sometimes decreased the activity of the complexes.

## Introduction

Mood disorders are among the most prevalent forms of mental illness. Severe forms of depression affect 2–5% of the US population, and up to 20% suffer from milder forms of the illness. Another roughly 1–2% are afflicted by bipolar disorder (BD) or its less severe variants (1,2) and are associated with higher rates of suicide and work loss (3–5).

Tissues with high energy demands, such as the brain, contain a large number of mitochondria and are therefore more susceptible to reduction of the aerobic metabolism (6). Mitochondrial disease results from a malfunction in biochemical cascade and the damage to the mitochondrial electron transport chain has been suggested to be an important factor in the pathogenesis of a range of neuropsychiatric disorders, such as BD, depression and schizophrenia (7,8). Several studies have shown that the abnormalities in energy metabolism lead to cellular degeneration (9). This effect may occur because when the mitochondrial dysfunction is severe it can lead to cell death by apoptosis or necrosis (10,11). In fact, mitochondria are involved in essential processes, such as apoptosis and calcium homeostasis (12–14), which are involved in cell death.

Mitochondria are intracellular organelles that play a crucial role in ATP production (9). Most cell energy is obtained through oxidative phosphorylation, a process requiring the action of various respiratory enzyme complexes located in a special structure of the inner mitochondrial membrane, the mitochondrial respiratory chain (15). In most organisms, the mitochondrial respiratory chain is composed of four complexes, where the electron transport couples with translocation of protons from the mitochondrial matrix to the intermembrane space. The generated proton gradient is used by ATP synthase to catalyse the formation of ATP by the phosphorylation of ADP (7,16).

A fixed combination of antipsychotic and antidepressant drugs was widely used in medicine and, at one time, was common in psychiatry. A generation ago, combinations of antidepressants with either antipsychotics [e.g. amitriptyline and perphenazine (Etrafon™; Schering–Bayer HealthCare Pharmaceuticals, Berlin, Germany and Triavil™; Merck & Company Inc., Whitehouse Station, NJ, USA)] or benzodiazepines [e.g. amitriptyline and chlordiazepoxide (Limbitrol™; Valeant Pharmaceuticals International, USA/Valeant Farmacêutica do Brasil, SP, Brazil)] were widely used by both psychiatrists and other medical practitioners (17). Recently, a fixed combination of the antipsychotic drug olanzapine (OLZ) and the antidepressant fluoxetine (FLX) (Symbyax™; Eli Lilly and Company, Indianapolis, IN, USA) has

been introduced for the treatment of BD (3,18). In a controlled study by Shelton et al. (19), subjects with treatment-resistant depression received OLZ alone, FLX alone or a combination of both; the combination was associated with significantly greater and faster improvement than was either drug alone. Although there is a clear clinical benefit from this combination, the precise neural mechanisms responsible for its efficacy are not clearly understood. Therefore, it is important to investigate the mechanisms of action of this combination in order to not only better understand the aetiology of the clinical syndromes, but also to eventually facilitate the development of improved drugs to treat them (4,20).

Considering the effects of OLZ, FLX and these combinations on brain energy metabolism are still unknown, we evaluated the effects of these drugs on mitochondrial respiratory chain in the rat prefrontal cortex, hippocampus and striatum. It is important to note that we chose the prefrontal cortex, hippocampus and striatum in this study because these brain areas are implicated in mood disorders (21,22).

## Material and methods

### Animals

Male adult Wistar rats (60 days old) were obtained from UNESC (Universidade do Extremo Sul Catarinense, Criciúma, SC, Brazil) breeding colony. They were housed five per cage with food and water available *ad libitum* and were maintained on a 12-h light/dark cycle (lights on at 7:00 h). All experimental procedures involving animals were performed in accordance with the NIH Guide for the Care and Use of Laboratory Animals and the Brazilian Society for Neuroscience and Behavior (SBNc) recommendations for animal care and with approval by local Ethics Committee under protocol number 510/2006.

### Drugs and treatments

OLZ (Zyprexa™) and FLX (Prozac™) were provided from Eli Lilly do Brasil Ltda, São Paulo, Brazil. Animals received daily intraperitoneal injections of OLZ (3 or 6 mg/kg), FLX (12 or 25 mg/kg) or combination of both drugs for 28 days in two protocols of the chronic model (A and B) and in the acute model for 1 day. Protocols and doses of drugs were performed in accordance with previous studies (23,24). All the drugs were dissolved in saline (0.9% NaCl) solution (vehicle). Control animals received saline (0.9% NaCl; 1.0 ml/kg). In the acute protocol, after the single injection, the animals were killed 2 h later by decapitation, and the prefrontal cortex, hippocampus and striatum were immediately removed. Similarly, in the chronic protocols, the animals were killed 2

(A) and 24 (B) h after the last injection, and the same areas were removed. The analysis was performed at different times of decapitation (2 and 24 h) after the last injection to be sure that the effects of the studied parameters were because of a chronic effect (23,24). After that, the activity of mitochondrial respiratory chain was measured ( $n = 5$  each).

#### Tissue and homogenate preparation

Hippocampus, striatum and prefrontal cortex were homogenised (1:10, w/v) in SETH (sucrose, EDTA, tris and heparin) buffer at pH 7.4 (250 mM sucrose, 2 mM ethylenediaminetetraacetic acid, 10 mM Trizma base and 50 IU/ml heparin). The homogenates were centrifuged at 800g for 10 min and the supernatants kept at  $-70^{\circ}\text{C}$  until used for mitochondrial respiratory chain activity determination. The maximal period between homogenate preparation and enzyme analysis was always <5 days. Protein content was determined by the method described by Lowry et al. (25) using bovine serum albumin as standard.

#### Respiratory chain enzyme activities

NADH dehydrogenase (complex I) was evaluated by the method described by Cassina and Radi (26) with the rate of NADH-dependent ferricyanide reduction at 420 nm. The activities of succinate-2,6-dichloroindophenol (DCIP) oxidoreductase (complex II) and succinate:cytochrome *c* oxidoreductase (complex II–III) were determined by the method described by Fischer et al. (27). Complex II activity was measured by following the decrease in absorbance due to the reduction in 2,6-DCIP at 600 nm. Complex II–III activity was measured by cytochrome *c* reduction from succinate at 550 nm. The activity of cytochrome *c* oxidase (complex IV) was assayed according to the method described by Rustin et al. (28) and measured by following the decrease in absorbance due to the oxidation of previously reduced cytochrome *c* at 550 nm. The activities of the mitochondrial respiratory chain complexes were calculated as nmol/min mg protein.

#### Statistical analysis

All data are presented as mean  $\pm$  SEM (standard error of the mean). Differences among experimental groups in the assessment of mitochondrial respiratory chain activity were determined by one-way ANOVA, followed by Tukey *post hoc* test when ANOVA was significant;  $p < 0.05$  was considered to be statistically significant.

## Results

As depicted in Fig. 1a, complex I activity increased in the prefrontal cortex of rats treated acutely with OLZ 6 mg/kg and OLZ 3 mg/kg plus FLX 12 mg/kg (Fig. 1a;  $F = 35.63$ ;  $p < 0.05$ ); in the hippocampus complex I activity increased with FLX 25 mg/kg (Fig. 1a;  $F = 180.27$ ;  $p < 0.05$ ); in the striatum complex I activity increased with OLZ 6 mg/kg (Fig. 1a;  $F = 3.69$ ;  $p < 0.05$ ). The complex II activity increased in prefrontal cortex (Fig. 1b;  $F = 2.28$ ;  $p < 0.05$ ) and hippocampus (Fig. 1b;  $F = 36.43$ ;  $p < 0.05$ ) after acute treatment with OLZ 6 mg/kg alone. The complex II–III activity increased in the prefrontal cortex (Fig. 1c;  $F = 17.03$ ;  $p < 0.05$ ) with OLZ 3 mg/kg plus FLX 25 mg/kg and OLZ 6 mg/kg plus FLX 25 mg/kg, in the hippocampus (Fig. 1c;  $F = 6.92$ ;  $p < 0.05$ ) with OLZ 3 mg/kg plus FLX 25 mg/kg and OLZ 6 mg/kg plus FLX 25 mg/kg and in the striatum (Fig. 1c;  $F = 15.71$ ;  $p < 0.05$ ) with OLZ 3 mg/kg plus FLX 12 or 25 mg/kg and OLZ 6 mg/kg plus FLX 25 mg/kg. After acute treatment, the complex IV activity increased in the prefrontal cortex (Fig. 1d;  $F = 5.7$ ;  $p < 0.05$ ) with OLZ 3 mg/kg plus FLX 25 mg/kg and OLZ 6 mg/kg plus FLX 12 or 25 mg/kg and in the striatum (Fig. 1d;  $F = 8.68$ ;  $p < 0.05$ ) with OLZ 3 mg/kg plus FLX 12 or 25 mg/kg. In the chronic treatment, when the animals were killed 2 h after the last injection (Fig. 2), there was an increase in complex I activity in the striatum after OLZ 6 mg/kg and FLX 25 mg/kg in combination (Fig. 2a;  $F = 4.02$ ;  $p < 0.05$ ). The complex II activity decreased in the striatum after chronic treatment with OLZ 6 mg/kg plus FLX 12 mg/kg (Fig. 2b;  $F = 1.87$ ;  $p < 0.05$ ) and it was not altered in the prefrontal cortex (Fig. 2b;  $F = 0.92$ ;  $p > 0.05$ ) and hippocampus (Fig. 2b;  $F = 1.93$ ;  $p > 0.05$ ). The complex II–III activity increased in the striatum with OLZ 3 mg/kg alone compared to the control group (Fig. 2c;  $F = 7.81$ ;  $p < 0.05$ ) and it was not altered in the prefrontal cortex (Fig. 2c;  $F = 1.87$ ;  $p > 0.05$ ) and hippocampus (Fig. 2c;  $F = 1.91$ ;  $p > 0.05$ ). The complex IV activity did not alter in the prefrontal cortex (Fig. 2d;  $F = 0.36$ ;  $p > 0.05$ ) and striatum (Fig. 2d;  $F = 3.52$ ;  $p < 0.05$ ) compared to the control group; in contrast, in the hippocampus, the complex IV activity increased after treatment with OLZ 6 mg/kg and FLX 25 mg/kg in combination compared to the control group (Fig. 2d;  $F = 2.4$ ;  $p < 0.05$ ).

In the chronic treatment, when the animals were killed 24 h after the last injection (Fig. 3), we showed that complex I activity decreased in the prefrontal cortex with FLX 12 mg/kg alone compared to the control group (Fig. 3a;  $F = 3.9$ ;  $p < 0.05$ );

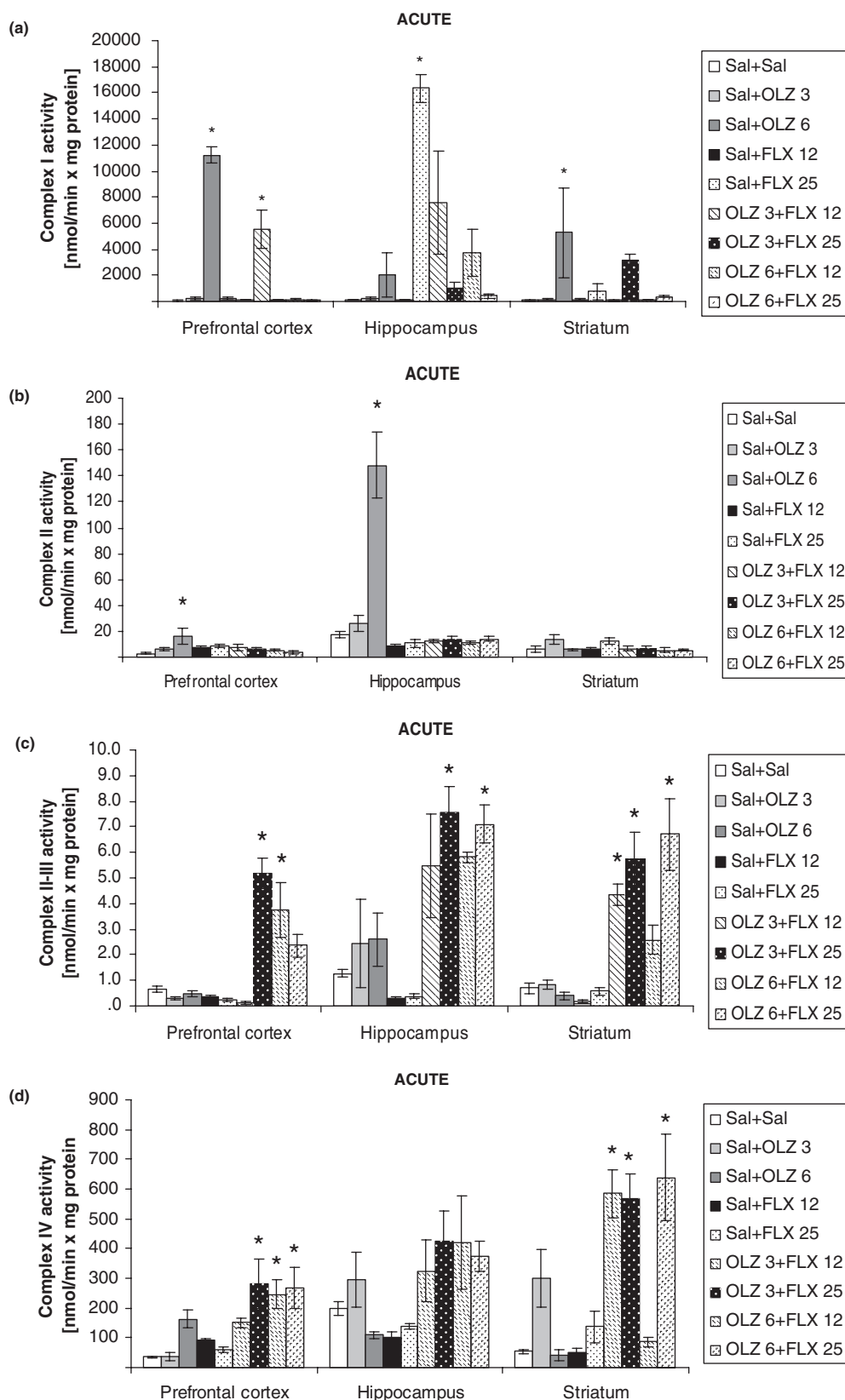


Fig. 1. Effects of the acute administration of olanzapine and fluoxetine on the complex I (a), II (b), II-III (c) and IV (d) activities in the rat prefrontal cortex, hippocampus and striatum. Bars represent means  $\pm$  SEM (standard error of the mean). \* $p < 0.05$  versus saline according to ANOVA followed by Tukey *post hoc* test.

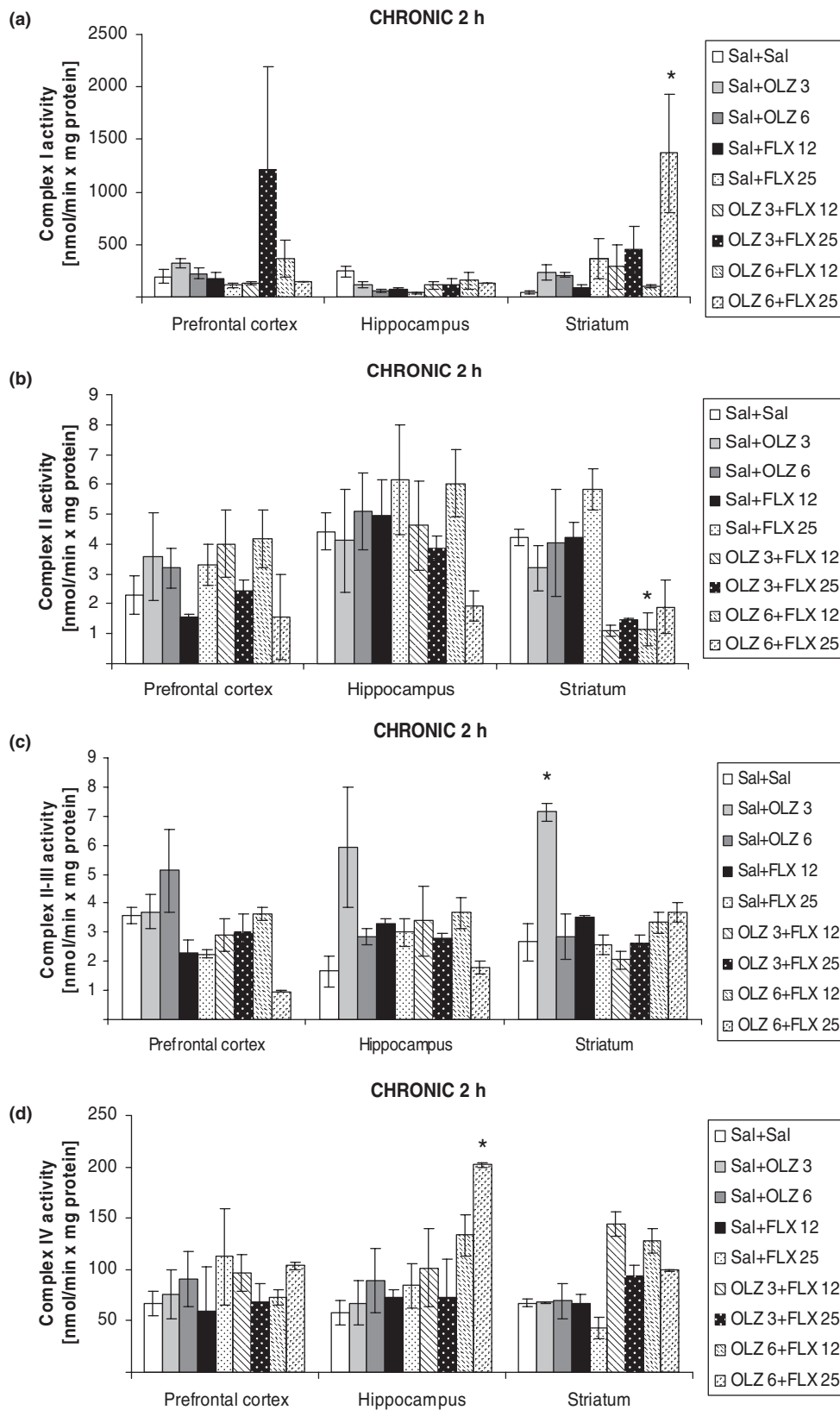


Fig. 2. Effects of the chronic administration of olanzapine and fluoxetine on the complex I (a), II (b), II–III (c) and IV (d) activities in the rat prefrontal cortex, hippocampus and striatum. The animals were killed 2 h after the last administration of the drugs. Bars represent means  $\pm$  SEM (standard error of the mean). \* $p < 0.05$  versus saline according to ANOVA followed by Tukey *post hoc* test.



however, the complex I activity did not alter in hippocampus (Fig. 3a;  $F = 2.21$ ;  $p > 0.05$ ) and striatum (Fig. 3a;  $F = 1.15$ ;  $p > 0.05$ ). The complex II activity (Fig. 2b) did not alter in the prefrontal cortex ( $F = 3.45$ ), hippocampus ( $F = 4.83$ ) and striatum ( $F = 1.39$ ) compared to the control group. Treatment with FLX 25 mg/kg alone decreased complex II–III activity in the striatum compared to the control group (Fig. 3c;  $F = 4.99$ ;  $p < 0.05$ ). In the hippocampus (Fig. 3c;  $F = 3.42$ ;  $p > 0.05$ ) and prefrontal cortex (Fig. 3c;  $F = 2.18$ ;  $p > 0.05$ ), we did not observe alteration in the complex II–III activity. Figure 3d shows that complex IV activity did not alter in the prefrontal cortex ( $F = 2.13$ ). In the striatum, the complex IV activity increased after treatment with OLZ 6 mg/kg and FLX 12 mg/kg in combination compared to the control group ( $F = 8.18$ ;  $p < 0.05$ ). In contrast, in the hippocampus the complex IV activity decreased after treatment with OLZ 3 and 6 mg/kg alone, as with FLX 12 and 25 mg/kg alone. In addition, OLZ 3 mg/kg plus FLX 12 or 25 mg/kg also decreased the complex IV activity compared to the control group ( $F = 6.87$ ;  $p < 0.05$ ).

## Discussion

In this study, we evaluated the effects of the antipsychotic OLZ and the antidepressant FLX (alone or in combination) on mitochondrial respiratory chain activity in the rat brain. We showed that both acute and chronic treatments with FLX and OLZ alone or in combination altered respiratory chain complex activity in the rat brain, but in combination we observed larger alterations. We showed that these alterations were related to treatment regime, complex, brain area and drug concentration.

Recent studies from our group showed that acute administration of FLX inhibited creatine kinase in the rat brain. This study also showed that chronic treatment, when the animals were killed 2 h after the last injection, showed a decrease in the creatine kinase activity after FLX administration, alone or in combination with OLZ. In contrast, when the animals were killed 24 h after the last injection we did not observe alterations in the enzyme (23). In addition, acute, but not chronic treatment with FLX and OLZ alone or in combination increased citrate synthase activity in the rat brain (24). Creatine kinase works as a buffering system of cellular ATP levels and citrate synthase has been used as a quantitative enzyme marker for the presence of intact mitochondria (29). Both enzymes play an important role in brain energy metabolism. In fact, several studies have been appointed to mitochondrial abnormalities in a

number of disorders, including depression, BD and schizophrenia (7,30,31).

Studies have identified that some brain regions from BD patients presented a decreased energy metabolism and abnormalities in mitochondrial DNA (32,33). Moreover, reductions of mitochondrial respiratory chain were found in patients with depression, schizophrenia and BD (30,34). Additionally, animal models evaluating the molecular pharmacology of mood stabilising drugs have implicated mitochondrial energy metabolism as a target for these drugs (35,36).

Dror et al. (37) showed alteration in complex I activity and in levels of mRNA and protein of the 24- and 51-kDa iron–sulphur flavoprotein subunits of the complex from platelets of schizophrenia patients, suggesting that these alterations may result in abnormal neural transmission, synaptic plasticity and connectivity, leading to abnormal behavioural symptoms in schizophrenia. Moreover, another study has shown abnormalities in energy metabolism in the basal ganglia of chronic schizophrenics (38). In addition, Iwamoto et al. (32) showed mitochondrial dysfunction in postmortem brains of schizophrenic patients; however, this dysfunction was due to the patients' medication, especially antipsychotics. Additionally, a study showed that OLZ, clozapine and haloperidol inhibited succinate dehydrogenase (an important enzyme of the Krebs cycle and part of the mitochondrial respiratory chain as an electron-transferring protein); however, aripiprazole antipsychotic increased the enzyme in the rat brain (39). Several studies have shown that antipsychotic drugs inhibited the respiratory electron transport chain (40–43). In this study, we showed that OLZ alone or in combination with FLX inhibited the complex IV activity in the hippocampus when the animals were killed 24 h after the last injection, and OLZ in combination with FLX inhibited the complex II activity in the striatum when the animals were killed 2 h after the last injection; however, in most cases, OLZ alone or in combination acted to increase the complex respiratory chain in the rat brain.

The effects of OLZ and FLX found in this study could be also related to oxidative stress. In fact, mitochondria can produce an excess of reactive oxygen species (ROS), which will cause oxidative damage to cellular constituents such as membrane lipids and proteins (44). In addition, mtDNA mutations in elevated production of ROS in turn proved to increase the number of mtDNA mutations (45). Several studies have generally suggested a compromised oxidative stress in psychiatric disorders such as BD, depression and schizophrenia (46–48). Additionally, chronic exposure to antipsychotics, haloperidol and

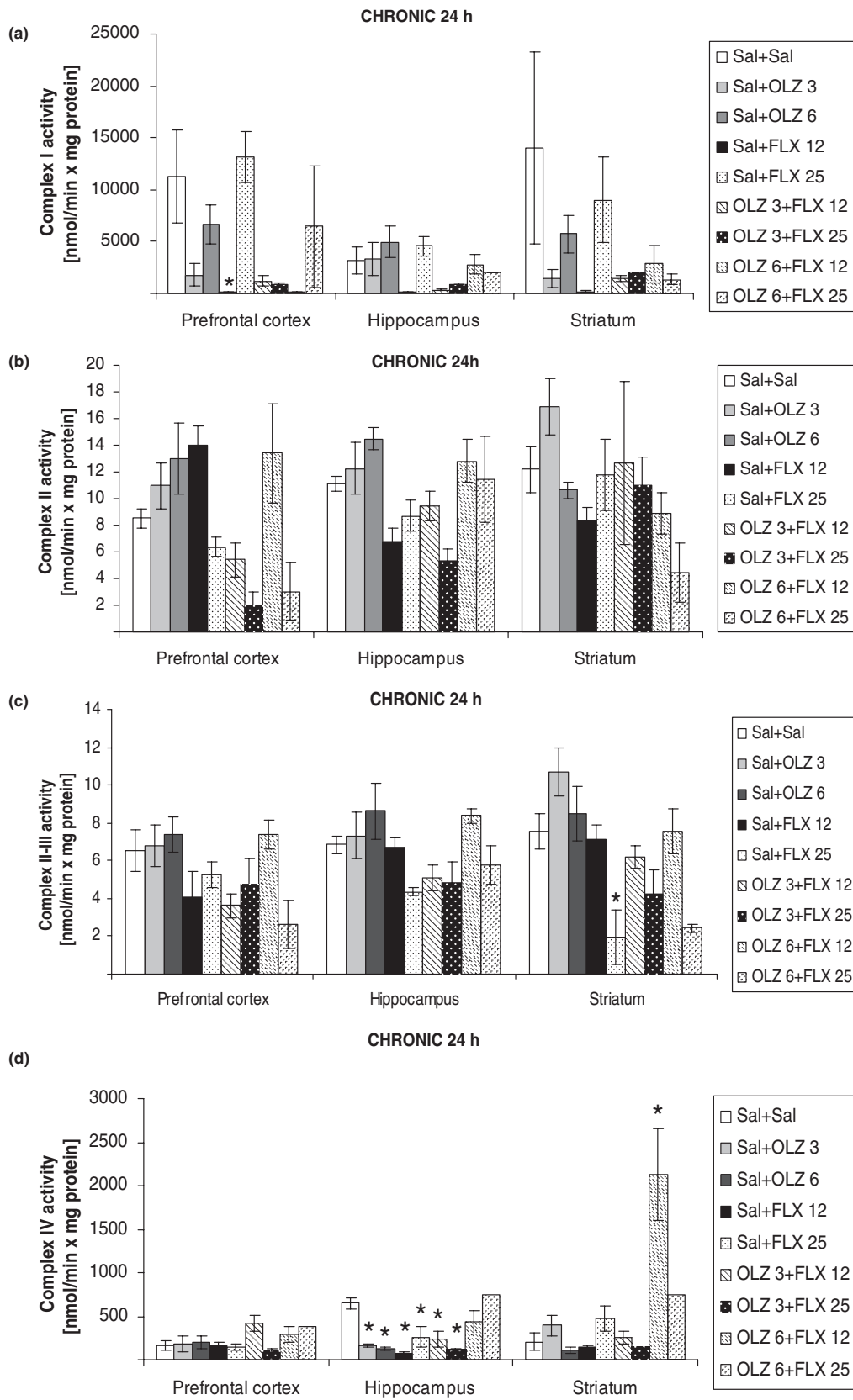


Fig. 3. Effects of the chronic administration of olanzapine and fluoxetine on the complex I (a), II (b), II-III (c) and IV (d) activities in the rat prefrontal cortex, hippocampus and striatum. The animals were killed 24 h after the last administration of the drugs. Bars represent means  $\pm$  SEM (standard error of the mean). \* $p < 0.05$  versus saline according to ANOVA followed by Tukey *post hoc* test.

clozapine, but not OLZ, caused changes in the activities of antioxidant enzymes and oxidative damage in the rat brain (49,50). Researchers have reported that some side effects of antipsychotics are associated with oxidative stress (51,52) and metabolism impairment (53). Recently, a study from our group showed that OLZ and FLX treatment inhibited creatine kinase activity (23), suggesting that inhibition of enzyme may be associated with the occurrence of some side effects of OLZ and FLX. However, OLZ exerted antioxidant effects through modulating ROS levels, superoxide dismutase activity and Bax expression to provide protective effects against *N*-methyl-4-phenylpyridinium-induced oxidative stress in PC12 cells (54). FLX has also shown an antioxidant effect (55–57).

Reductions in mRNA and proteins of complex I subunits NADH dehydrogenase ubiquinone flavoprotein (*NDUFV1*), NADH-ubiquinone oxidoreductase flavoprotein gene (*NDUFV2*) and NADH dehydrogenase (ubiquinone) Fe-S protein 1 (*NDUFS1*) have been shown in the cerebellum postmortem from patients with depression (30). Many animal models of mania and depression have revealed alterations in metabolism energy. Studies from our group showed reduced creatine kinase and citrate synthase activity in brain of rats submitted to the animal model of mania (35,58). Moreover, in another study from our group, it was shown that antidepressants imipramine (59) and paroxetine (60) increased creatine kinase activity in the rat brain, suggesting that the modulation of energy metabolism by antidepressants could be an important mechanism of action of these drugs. Nevertheless, our group also showed that mitochondrial respiratory chain complexes I, II–III and IV were inhibited after chronic mild stress in the cerebral cortex and cerebellum (61). Madrigal et al. (16) also reported that complexes I–III and II–III of mitochondrial respiratory chain were inhibited in rat brains after chronic stress (immobilisation for 6 h during 21 days). Hroudova and Fisar (62) showed that several antidepressant drugs inhibited complexes I and IV of the mitochondrial respiratory chain, suggesting that in pathophysiology of mood disorders therapeutic effects of antidepressant could have changes in energetic metabolism of cells determined by mitochondria.

In clinical practice, atypical antipsychotic drugs in combination with antidepressant drugs have been used as a strategy to treat (63) treatment-resistant depression (1,64,65) and psychotic depression (66). In an elegant controlled study, Matthews et al. (67) showed that subjects with treatment-resistant depression received OLZ and FLX alone or a combination of both; the combination was associated with significantly greater and faster improvement than was

either drug alone. In this study, we also showed greater effects of OLZ and FLX in combination under metabolism energy parameters, antidepressant FLX and antipsychotic OLZ alone or in combination increased or decreased mitochondrial respiratory chain, dependent on treatment regime, enzymatic complex, brain area and drug concentration. The reason for this different alteration in this study is unclear, but could be related to desensitisation of the effects of repeated OLZ and FLX administration or to the adaptation mechanism of mitochondria. The differences of OLZ and FLX found in these findings could be related to brain distribution of the drugs or differences in the toxicity of its metabolites.

In conclusion, taking together the present findings and evidence from the literature, we hypothesise that FLX and OLZ in combination could be involved in mitochondrial function, which is altered in several mood disorders. However, it remains to be seen if effects of the combination of drugs on the mitochondrial respiratory chain are related to the therapeutic or side effects of pharmacotherapy.

### Acknowledgements

This study was supported in part by grants from 'Conselho Nacional de Desenvolvimento Científico e Tecnológico' (CNPq-Brazil – JQ, ELS), the Instituto Cérebro e Mente (JQ) and UNESC (JQ and ELS). JQ and ELS are recipients of CNPq (Brazil) Productivity fellowships. GZR is holder of an FAPESC studentship.

### References

1. NELSON G, HALL GB, FORCHUK C. Current and preferred housing of psychiatric consumers/survivors. *Can J Commun Health* 2003;**22**:5–19.
2. KONRADI C, EATON M, MACDONALD ML, WALSH J, BENES FM, HECKERS S. Molecular evidence for mitochondrial dysfunction in bipolar disorder. *Arch Gen Psychiatry* 2004;**61**:300–308.
3. BELMAKER RH. Bipolar disorder. *N Engl J Med* 2004;**351**:476–486.
4. USTUN TB, AYUSO-MATEOS JL, CHATTERJI S, MATHERS C, MURRAY CJ. Global burden of depressive disorders in the year 2000. *Br J Psychiatry* 2004;**84**:386–392.
5. KUPFER DJ. The increasing medical burden in bipolar disorder. *JAMA* 2005;**293**:2528–2530.
6. BOEKEMA EJ, BRAUN HP. Supramolecular structure of the mitochondrial oxidative phosphorylation system. *J Biol Chem* 2007;**282**:1–4.
7. FATTAL O, BUDUR K, VAUGHAN AJ, FRANCO K. Review of the literature on major mental disorders in adult patients with mitochondrial diseases. *Psychosomatics* 2006;**47**:1–7.
8. PRABAKARAN S, SWATTON JE, RYAN MM et al. Mitochondrial dysfunction in schizophrenia: evidence for compromised brain metabolism and oxidative stress. *Mol Psychiatry* 2004;**9**:684–687.
9. CALABRESE V, SCAPAGNINI G, GIUFFRIDA-STELLA AM, BATES TE, CLARK JB. Mitochondrial involvement in brain



- function and dysfunction: relevance to aging, neurodegenerative disorders and longevity. *Neurochem Res* 2001;**26**: 739–764.
10. ARMSTRONG JS. The role of the mitochondrial permeability transition in cell death. *Mitochondrion* 2006;**6**:225–234.
  11. SCHAPIRA AH. Mitochondrial disease. *Lancet* 2006;**368**: 70–82.
  12. GUR RE, RESNICK SM, ALAVI A, CARROF S, KUSHNER M, REIVCH M. Regional brain function in schizophrenia II: repeated evaluation with positron emission tomography. *Arch Gen Psychiatry* 1987;**44**:126–129.
  13. CASTANIER C, ARNOULT D. Mitochondrial dynamics apoptosis. *Med Sci* 2010;**26**:830–835.
  14. HUNG CH, HO YS, CHANG RC. Modulation of mitochondrial calcium as a pharmacological target for Alzheimer's disease. *Ageing Res Rev* 2010;**9**:447–456.
  15. HORN D, BARRIENTOS A. Mitochondrial copper metabolism and delivery to cytochrome *c* oxidase. *IUBMB Life* 2008;**60**:421–429.
  16. MADRIGAL JLM, OLIVENZA R, MORO MA et al. Glutathione depletion, lipid peroxidation and mitochondrial dysfunction are induced by chronic stress in rat brain. *Neuropsychopharmacology* 2001;**24**:420–429.
  17. SHELTON RC. The return of fixed combinations in psychiatry: fluoxetine and olanzapine combination. *Ther Clin Risk Manag* 2006;**2**:187–192.
  18. TOHEN M, VIETA E, CALABRESE J. Efficacy of olanzapine and olanzapine-fluoxetine combination in the treatment of bipolar I depression. *Arch Gen Psychiatry* 2004;**60**: 1079–1088.
  19. SHELTON RC, TOLLEFSON GD, TOHEN M et al. A novel augmentation strategy for treating resistant major depression. *Am J Psychiatry* 2001;**158**:131–134.
  20. REZIN GT, CARDOSO MR, GONÇALVES CL et al. Inhibition of mitochondrial respiratory chain in brain of rats subjected to an experimental model of depression. *Neurochem Int* 2008;**53**:395–400.
  21. PITTENGER C, DUMAN RS. Stress, depression, and neuroplasticity: a convergence of mechanisms. *Neuropsychopharmacology* 2008;**33**:88–109.
  22. STRAKOWSKI SM, DELBELLO MP, ADLER CM. The functional neuroanatomy of bipolar disorder: a review of neuroimaging findings. *Mol Psychiatry* 2005;**10**:105–116.
  23. AGOSTINHO FR, SCAINI G, FERREIRA GK et al. Effects of olanzapine, fluoxetine and olanzapine/fluoxetine on creatine kinase activity in rat brain. *Brain Res Bull* 2009;**80**: 337–340.
  24. AGOSTINHO FR, RÉUS GZ, STRINGARI RB et al. Treatment with olanzapine, fluoxetine and olanzapine/fluoxetine alters citrate synthase activity in rat brain. *Neurosci Lett* 2011;**487**: 278–281.
  25. LOWRY OH, ROSEBOUGH NG, FARR AL, RANDALL RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem* 1951;**193**:265–275.
  26. CASSINA A, RADI R. Differential inhibitory action of nitric oxide and peroxynitrite on mitochondrial electron transport. *Arch Biochem Biophys* 1996;**328**:309–316.
  27. FISCHER JC, RUITENBEEK W, BERDEN JA et al. Differential investigation of the capacity of succinate oxidation in human skeletal muscle. *Clin Chim Acta* 1995;**153**:23–26.
  28. RUSTIN P, CHRETIEN D, BOURGERON T et al. Biochemical and molecular investigations in respiratory chain deficiencies. *Clin Chim Acta* 1994;**228**:35–51.
  29. MARCO R, PESTANA A, SEBASTIAN J, SOLS A. Oxaloacetate metabolic crossroads in liver. Enzyme compartmentation and regulation of gluconeogenesis. *Mol Cell Biochem* 1974;**3**:53–70.
  30. BEN-SHACHAR D, KARRY R. Neuroanatomical pattern of mitochondrial complex I pathology varies between schizophrenia, bipolar disorder and major depression. *PLoS One* 2008;**3**:3676.
  31. QUIROZ JA, GRAY AN, KATO T, MANJI HK. Mitochondrially mediated plasticity in the pathophysiology and treatment of bipolar disorder. *Neuropsychopharmacology* 2008;**33**:2551–2565.
  32. IWAMOTO K, BUNDO M, KATO T. Altered expression of mitochondria-related genes in postmortem brains of patients with bipolar disorder or schizophrenia, as revealed by large-scale DNA microarray analysis. *Hum Mol Genet* 2005;**14**: 241–253.
  33. KATO T. Mitochondrial dysfunction in bipolar disorder: from 31P-magnetic resonance spectroscopic findings to their molecular mechanisms. *Int Rev Neurobiol* 2005;**63**:21–40.
  34. ANDREAZZA AC, SHAO L, WANG JF, YOUNG LT. Mitochondrial complex I activity and oxidative damage to mitochondrial proteins in the prefrontal cortex of patients with bipolar disorder. *Arch Gen Psychiatry* 2010;**67**: 360–368.
  35. CORRÊA C, AMBONI G, ASSIS LC et al. Effects of lithium and valproate on hippocampus citrate synthase activity in an animal model of mania. *Prog Neuropsychopharmacol Biol Psychiatry* 2007;**31**:887–891.
  36. WANG FJ, SHAO L, SUN X, YOUNG LT. Glutathione S-transferase is a novel target for mood stabilizing drugs in primary cultured neuron. *J Neurochem* 2004;**88**:1477–1484.
  37. DROR E, KLEIN R, KARRY A et al. State-dependent alterations in mitochondrial complex I activity in platelets: a potential peripheral marker for schizophrenia. *Mol Psychiatry* 2002;**7**:995–1001.
  38. PRINCE JA, BLENNOW K, GOTTFRIES CG, KARLSSON I, ORELAND L. Mitochondrial function is differentially altered in the basal ganglia of chronic schizophrenics. *Neuropsychopharmacology* 1999;**21**:372–379.
  39. STRECK EL, REZIN GT, BARBOSA LM, ASSIS LC, GRANDI E, QUEVEDO J. Effect of antipsychotics on succinate dehydrogenase and cytochrome oxidase activities in rat brain. *Naunyn Schmiedeberg's Arch Pharmacol* 2007;**376**: 127–133.
  40. BURKHARDT C, KELLY JP, LIM YH, FILLEY CM, PARKER WD Jr. Neuroleptic medications inhibit complex I of the electron transport chain. *Ann Neurol* 1993;**33**:512–517.
  41. MODICA-NAPOLITANO JS, LAGACE CJ, BRENNAN WA, APRILLE JR. Differential effects of typical and atypical neuroleptics on mitochondrial function in vitro. *Arch Pharm Res* 2003;**26**:951–559.
  42. MAURER I, MOLLER HJ. Inhibition of complex I by neuroleptics in normal human brain cortex parallels the extrapyramidal toxicity of neuroleptics. *Mol Cell Biochem* 1997;**174**:255–259.
  43. JI B, LA Y, GAO L et al. A comparative proteomics analysis of rat mitochondria from the cerebral cortex and hippocampus in response to antipsychotic medications. *J Proteome Res* 2009;**8**:3633–3641.
  44. WU Y, WU S, LEE W, WEI Y. Mitochondrial respiratory dysfunction-elicited oxidative stress and posttranslational protein modification in mitochondrial diseases. *Ann N Y Acad Sci* 2010;**1201**:147–156.

45. PŁOSZAJ T, ROBASZKIEWICZ A, WITAS H. Oxidative damage of mitochondrial DNA: the result or consequence of enhanced generation of reactive oxygen species. *Postepy Biochem* 2010;**56**:139–146.
46. PADURARIU M, CIOBICA A, DOBRIN I, STEFANESCU C. Evaluation of antioxidant enzymes activities and lipid peroxidation in schizophrenic patients treated with typical and atypical antipsychotics. *Neurosci Lett* 2010;**479**: 317–320.
47. RÉUS GZ, STRINGARI RB, SOUZA B et al. Harmine and imipramine promote antioxidant activities in prefrontal cortex and hippocampus. *Oxid Med Cell Longev* 2010;**3**: 325–331.
48. VALVASSORI SS, PETRONILHO FC, RÉUS GZ et al. Effect of N-acetylcysteine and/or deferoxamine on oxidative stress and hyperactivity in an animal model of mania. *Prog Neuropsychopharmacol Biol Psychiatry* 2008;**32**:1064–1068.
49. POLYDORO M, SCHRÖDER N, LIMA MN et al. Haloperidol- and clozapine-induced oxidative stress in the rat brain. *Pharmacol Biochem Behav* 2004;**78**:751–756.
50. REINKE A, MARTINS MR, LIMA MS, MOREIRA JC, DALPIZZOL F, QUEVEDO J. Haloperidol and clozapine, but not olanzapine, induces oxidative stress in rat brain. *Neurosci Lett* 2004;**372**:157–160.
51. LOHR JB, CADET JL, LOHR MA et al. Vitamin E in the treatment of tardive dyskinesia: the possible involvement of free radical mechanisms. *Schizophr Bull* 1988;**14**:291–296.
52. PEET M, LAUGHARNE J, RANGARAJAN N. Tardive dyskinesia, lipid peroxidation, and sustained amelioration with vitamin E treatment. *Int Clin Psychopharmacol* 1993;**8**: 151–153.
53. ANDREASSEN OA, FERRANTE RJ, BEAL MF, JORGENSEN HA. Oral dyskinesias and striatal lesions in rats after long-term co-treatment with haloperidol and 3-nitropropionic acid. *Neuroscience* 1998;**87**:639–648.
54. PARK SW, LEE CH, LEE JG et al. Protective effects of atypical antipsychotic drugs against MPP<sup>+</sup>-induced oxidative stress in PC12 cells. *Neurosci Res* 2011;**69**:283–290.
55. GAŁECKI P, SZEMRAJ J, BIEŃKIEWICZ M, FLORKOWSKI A, GAŁECKA E. Lipid peroxidation and antioxidant protection in patients during acute depressive episodes and in remission after fluoxetine treatment. *Pharmacol Rep* 2009;**61**: 436–447.
56. KIRKOVA M, TZVETANOVA E, VIRCHEVA S, ZAMFIROVA R, GRYGIER B, KUBERA M. Antioxidant activity of fluoxetine: studies in mice melanoma model. *Cell Biochem Funct* 2010;**28**:497–502.
57. CHUNG YC, KIM SR, PARK JY et al. Fluoxetine prevents MPTP-induced loss of dopaminergic neurons by inhibiting microglial activation. *Neuropharmacology* 2011;**60**: 963–974.
58. STRECK EL, AMBONI G, SCAINI G et al. Brain creatine kinase activity in an animal model of mania. *Life Sci* 2008;**82**:424–429.
59. ASSIS IC, REZIN GT, COMIM CM et al. Effect of acute administration of ketamine and imipramine on creatine kinase activity in the brain of rats. *Rev Bras Psiquiatr* 2009;**31**:247–252.
60. SANTOS PM, SCAINI G, REZIN GT et al. Brain creatine kinase activity is increased by chronic administration of paroxetine. *Brain Res Bull* 2009;**80**:327–330.
61. REZIN GT, GONÇALVES CL, DAUFENBACH JF et al. Acute administration of ketamine reverses the inhibition of mitochondrial respiratory chain induced by chronic mild stress. *Brain Res Bull* 2009;**79**:418–421.
62. HROUDOVA J, FISAR Z. Activities of respiratory chain complexes and citrate synthase influenced by pharmacologically different antidepressants and mood stabilizers. *Neuro Endocrinol Lett* 2010;**31**:336–342.
63. HIRSCHFELD RM, MONTGOMERY SA, AGUGLIA E. Partial response and nonresponse to antidepressant therapy: current approaches and treatment options. *J Clin Psychiatry* 2002;**63**:826–837.
64. FAVA M. New approaches to the treatment of refractory depression. *J Clin Psychiatry* 2000;**61**:26–32.
65. MORISHITA S. Clonazepam as a therapeutic adjunct to improve the management of depression: a brief review. *Hum Psychopharmacol* 2009;**24**:191–198.
66. SCHATZBERG AF. New approaches to managing psychotic depression. *J Clin Psychiatry* 2003;**64**:19–23.
67. MATTHEWS JD, BOTTONARI KA, POLANIA LM. An open study of olanzapine and fluoxetine for psychotic major depressive disorder: interim analyses. *J Clin Psychiatry* 2002;**63**:1164–1170.