Exercise Training Reduces Inflammatory Mediators in the Intestinal Tract of Healthy Older Adult Mice*

Nicholas Packer and Laurie Hoffman-Goetz School of Public Health and Health Systems, University of Waterloo

RÉSUMÉ

Le vieillissement s'allie à une augmentation d'inflammation intestinale et le risque élevé de maladies chroniques, y compris les maladies inflammatoires de l'intestin et le cancer du côlon; nombreuses études épidémiologiques indiquent que l'exercice régulier réduit les risques. Cette étude a examiné les effets à long terme de l'exercice volontaire sur les médiateurs inflammatoires dans les intestins des souris âgées et en bonne santé C57BL/6 (âgées de 15–16 mois). On a désigné les animaux soit à quatre mois de roue d'exercice à souris (RES ; n - 20), soit à une groupe de contrôle « sédentaire » (NRL ; n = 20). Les lymphocytes intestinaux ont été récoltés et analysés pour la présence de (1) pro-inflammatoire (TNF-a, IL-1 β) et de cytokines pléotropes (IL-6), et (2) de pro-(caspase-3/-7) et d'anti-(Bcl-2) protéines apoptotiques. L'efficacité d'exercise a été confirmée par l'activité des enzymes dans les muscles squelettiques ; l'évidence de stress a été confirmée par un plasma 8-iso-PGF_{2a} et la corticostérone. Les RES souris ont réalisés une incidence inférieure de TNF- α , de la caspase-7, et de 8-isoprostanes (p < .05) par rapport aux contrôles sédentaires, ce qui suggère que l'exercice à long terme peut « protéger » l'intestin en réduisant la manifestation de cytokines inflammatoires et du protéine apoptotique.

ABSTRACT

Aging is associated with increased intestinal inflammation and elevated risk of chronic diseases including inflammatory bowel diseases and colon cancer; many epidemiologic studies show that regular exercise reduces risk. This study examined the effects of long-term voluntary exercise on inflammatory mediators expressed in the intestine of older (15–16 months), healthy C57BL/6 mice. Animals were assigned to four months of freewheel running (WR; n = 20) or to a "sedentary" no wheel running (NWR; n = 20) control group. Intestinal lymphocytes were harvested and analysed for expression of (1) pro-inflammatory (TNF- α , IL-1 β) and pleiotropic (IL-6) cytokines, and (2) pro-(caspase-3/-7) and anti-(Bcl-2) apoptotic proteins. Training was confirmed by skeletal muscle enzyme activity; stress was assessed by plasma 8-iso-PGF_{2 α} and corticosterone. The WR mice had a lower expression of TNF- α , caspase-7, and 8-isoprostanes (p < .05) compared to sedentary controls, suggesting that long-term exercise may "protect" the bowel by reducing inflammatory cytokine and apoptotic protein expression.

* We thank Julia Guan for technical assistance. Research was supported by a grant from the Natural Sciences and Engineering Research Council of Canada. Nicholas Packer was the recipient of a Canadian Institutes of Health Research post-graduate scholarship.

Manuscript received: / manuscrit reçu : 02/06/11

Manuscript accepted: / manuscrit accepté : 23/12/11

Mots clés: exercice, lymphocytes intestinales, vieillissement, cytokine inflammatoire, apoptose, modèles animales

Keywords: exercise, intestinal lymphocytes, aging, inflammatory cytokines, apoptosis, animal models

Correspondence and requests for offprints should be sent to / La correspondance et les demandes de tirés-à-part doivent être adressées à:

Laurie Hoffman-Goetz, Ph.D., M.P.H. School of Public Health and Health Systems Faculty of Applied Health Sciences University of Waterloo 200 University Avenue West Waterloo, ON N2L 3G1 (lhgoetz@uwaterloo.ca)

Canadian Journal on Aging / La Revue canadienne du vieillissement 31 (2) : 161–171 (2012) doi:10.1017/S0714980812000104

Introduction

Colorectal cancer (CRC) is the second leading cause of cancer mortality among older Canadians (Canadian Cancer Society's Steering Committee on Cancer Statistics, 2011). Despite the complex etiology of CRC, chronic inflammation is a fundamental risk factor (Demarzo et al., 2008). This is evidenced by elevated CRC risk among persons with inflammatory bowel disease (IBD), an increase proportional to the extent and duration of colitis (Kulaylat & Dayton, 2010). Canadians have among the highest IBD incidence rates worldwide (Bernstein et al., 2006). IBD is particularly debilitating among seniors, as 10-15 per cent of incident cases are diagnosed in patients aged 60 or older (Softley et al., 1988), and the presence of age-related co-morbidities may interfere with effective diagnosis and treatment (Swaroop, 2007).

Researchers have hypothesized that CRC pathogenesis may be influenced by inflammatory immune mechanisms (Terzić, Grivennikov, Karin, & Karin, 2010). Two inflammatory cytokines, tumour necrosis factor-alpha (TNF- α) (Grimm et al., 2010) and interleukin-1 beta (IL-1β) (Szkaradkiewicz et al., 2009), are elevated in the bowel of CRC patients, and elevated TNF- α predicts both the extent of carcinogenesis and likelihood of tumour recurrence (Grimm et al., 2010). Furthermore, immunosenescence (age-related immune changes) may increase cancer risk as a result of increases in chronic low-grade inflammation (Singh & Newman, 2010). Even healthy, aged individuals have demonstrated increased expression of interleukin-6 (IL-6), TNF- α , and IL-1 β in peripheral mononuclear cells (Fagiolo et al., 1993) and increased IL-6 and TNF-r1 in plasma (Stowe, Peek, Cutchin, & Goodwin, 2010).

Colorectal cancer has a long asymptomatic period, and physical symptoms typically appear only in advanced stages of disease when the prognosis is largely predetermined and treatment is limited (Gonzalez-Hermoso, Perez-Palma, Marchena-Gomez, Lorenzo-Rocha, & Medina-Arana, 2004). Accordingly, screening programs and strategies for risk factor reduction are critical avenues to reduce disease burden in aged individuals (Edwards et al., 2010). Since 40–50 per cent of CRC patients die within five years of initial diagnosis (Compton et al., 1999), even a modest delay in disease onset could significantly improve individual and population health outcomes (Luo, Bradley, Dahman, & Gardiner, 2010).

Physical activity is a promising lifestyle intervention identified to reduce CRC incidence (Colditz, Cannuscio, & Frazier, 1997) with a relative risk reported of 0.76 (95% CI 0.72 – 0.81) (Wolin, Yan, Colditz, & Lee, 2009) and 0.79 (95% CI 0.72 – 0.87) (Samad, Taylor, Marshall, & Chapman, 2005) among regular exercisers compared to inactive individuals. The relationship between increased physical activity and lowered CRC risk follows a strong dose-response curve (Thune & Furberg, 2001), and it is possible that regular exercise may reduce the risk of CRC recurrence (Halle & Schoenberg, 2009).

The mechanisms whereby regular physical activity protects against inflammatory bowel diseases and CRC in older people are not well-known. One possibility is that physical activity prevents age-related increases in inflammation by altering the intestinal inflammatory cytokine and apoptotic milieu. In young animals, longterm voluntary physical activity reduces intestinal TNF-α expression (Hoffman-Goetz, Pervaiz & Guan, 2009). This finding has recently been confirmed in aged humans: long-term (8.6 ± 0.3 months) resistance training reduced plasma TNF-α levels (Córdova et al., 2011). Moreover, Gomez-Merino et al. (2007) found that exercise training reduced IL-1 β in rat adipose tissue and skeletal muscle (Lira et al., 2009). In response to long-term physical activity, IL-6 increases in the intestinal compartment (Hoffman-Goetz, Pervaiz, Packer, & Guan, 2010) and in skeletal muscle (Pedersen et al., 2003) of young animals but decreases in older individuals (Nicklas et al., 2008).

The intestinal compartment is populated by a number of immunologically active cells. Lamina propria (LP), and intraepithelial (iIEL) and intestinal (IL) lymphocytes, play an important role in homeostasis (Lefrançois & Lycke, 2001), thereby modulating the inflammatory process (Iliev, Mileti, Matteoli, Chieppa, & Rescigno, 2009) and regulating the secretion of key cytokines (Powrie, 2004).

It is difficult to evaluate the extent of lymphocyte involvement in intestinal homeostasis in healthy adults. Utilizing an animal model, however, circumvents the problem of invasive tissue sampling from healthy people. Moreover, tissue biopsies are typically limited to individuals with intestinal pathologies and are restricted anatomically to the large intestine (Pironti et al., 2010). Major advantages of using an animal model include: (1) the ability to obtain healthy intestinal tissue for analysis; (2) sufficient tissue collection for assessment of multiple biomarkers and mediators; and (3) detailed physiological measurement and validation of "training" status. An animal model also allows precise age determination, a key consideration when examining the effects of training in an "aged" cohort.

The purpose of this study was to describe the effects of long-term voluntary physical activity on the expression of inflammatory cytokines and apoptotic proteins in intestinal lymphocytes of healthy older adult animals. The cytokines chosen were the classic pro-inflammatory (TNF- α , IL-1 β) and pleiotropic (IL-6) cytokines implicated in intestinal inflammation, IBD, and CRC progression. In addition, the expression of two apoptotic

proteins (caspase-3 and -7) in intestinal lymphocytes was assessed because they are activated by TNF- α , IL-1 β , and IL-6 (Wyllie, 2010) and induce cell death. We hypothesized that long-term, low-stress physical activity in mice would decrease the intestinal expression of pro-inflammatory (TNF- α , IL-1 β) and pleiotropic (IL-6) cytokines and apoptotic proteins (caspase-3, caspase-7) relative to untrained (sedentary) age-matched controls.

Materials and Methods

Animals

Female C57BL/6 mice (n = 40) (Harlan, Indianapolis, IN, U.S.), 11–12 months old, were individually housed at 21 ± 1 °C on a 12hr/12hr reversed light/dark cycle. Mice had *ad libitum* access to standard rodent diet (Lab Rodent Chow, PMI Feeds, IN, U.S.) and tap water throughout the experiments. All experiments were conducted in accordance with the ethical guidelines and protocols of the Canadian Council on Animal Care and were approved by the University Animal Research Ethics Committee.

Exercise-Training Protocol

Mice were matched by weight and randomized to an exercise-training condition: access to in-cage running wheels (wheel running, WR; n = 20) or to a control cage (no wheel running, NWR; n = 20) for 4 months. A magnetic switch attached to each wheel (23 cm in diameter) and an automated computer monitoring system (Vital View Application software, Mini-Mitter, Sunriver, Oregon, U.S.) captured the number of completed revolutions. Activity during the dark cycle was recorded as the number of revolutions completed per 15-min interval, converted to distance run (km), and summed by day, week, and month. Total running volume was monitored as an indicator of training status, in combination with skeletal muscle cytochrome *c* oxidase activity and body weight. All WR mice were sacrificed 24 hours after the last training session as in-cage running wheels were locked in order to prevent any carry-over effects of voluntary exercise.

Skeletal Muscle Cytochrome c Oxidase (CO) Activity

Cytochrome *c* oxidase (CO) plays an important role in determining the mitochondrial respiratory capacity of skeletal muscle as it constitutes the last step of ATP generation. Following sacrifice by sodium pentobarbital (0.6 - 0.8 cc per mouse, i.p.) overdose, soleus (SOL), and plantaris (PLANT) muscle samples were isolated from WR and NWR mice, frozen in liquid nitrogen, and stored at -80 °C until assayed. Muscles were cut into 5–10-mg segments, mashed and homogenized in buffer

[glycerol (50%), sodium phosphate buffer (20 mM), 2-mercaptoethanol (5 mM), ethylenediaminetetraacetic acid (EDTA, 0.5 mM), BSA (10%)] to yield a 50:1 dilution, and sonicated (using a 3-m tip, 2 sec on, 5 sec off for a total of 20 sec at 60 Hz; Vibra Cell, Sonics and Materials, Danbury, Connecticut, U.S.). Protein content was determined by Lowry assay (Lowry, Rosebrough, Farr, & Randall, 1951). Muscle homogenates were diluted to 1:500 dilutions in a 10-mM potassium phosphate buffer. Next, 20 μ L of reduced cytochrome *c* and 10 μ L of diluted homogenate were combined with 970 μ L of warmed (37 °C) phosphate buffer. Cytochrome *c* absorbance was determined spectrophotometrically at 550 nm.

Biomarkers of Stress (Plasma 8-iso-PGF $_{2\alpha}$ and Corticosterone)

We measured plasma 8-iso-PGF_{2 α} and corticosterone as biomarkers indicative of oxidative and psychogenic stress respectively. This procedure and rationale have been described extensively elsewhere (Hoffman-Goetz et al., 2010). In brief, immediately following sacrifice, we used a 1-mL syringe containing heparin to collect blood via cardiac puncture. Plasma was separated by centrifugation (6 min at 400 rpm) and frozen at -80 °C. Plasma 8-iso-PGF_{2 α} was assessed by a commercially available kit using direct enzyme immunoassay (EIA) as per the manufacturer's specifications (Cayman Chemical, Ann Arbor, Michigan, U.S.). Samples (100-µL sample, 25 µL of 10N NaOH) were hydrolyzed for 2 hr at 45 °C, neutralized to pH 6-8 with 12N HCl, centrifuged at 14,000 g for 5 min, and incubated with 8-iso-PGF_{2 α} antibody for 24 hr at 4 °C. Per cent absorbance was read at 412 nm at room temperature using a PowerWave 340 microplate spectrophotometer (Biotek Instruments, Winooski, Vermont, U.S.). The intra-assay percent coefficient of variation (% CV) was 11.7 per cent.

We measured corticosterone using a commercially available EIA kit, according to the manufacturer's instructions (Cayman Chemical). The cold spike protocol was used for the purification of plasma samples and corticosterone concentrations measured using a PowerWave 340 microplate spectrophotometer (Biotek Instruments) at 412 nm. The intra-assay coefficient of variation (% CV) was 12.5 per cent.

Determination of Cytokines and Apoptotic Proteins Expression in Intestinal Lymphocytes

Intestinal Lymphocyte (IL) Isolation

Intestinal lymphocyte (IL) isolation was performed as described (Hoffman-Goetz & Quadrilatero, 2003). Following sacrifice by sodium phenobarbital overdose, we used cold phosphate buffered saline (PBS) to wash the excised intestinal compartment; Peyer's patches and visible fat were removed and single-cell IL suspensions prepared by isolation over a pre-washed nylon wool (0.3 g) column. The eluted cells were layered over a Lympholyte-M density gradient medium (Cedarlane Laboratories, Burlington, Ontario) and centrifuged to remove debris. The remaining cell pellet (containing IEL and LP lymphocytes) was suspended in 400 μ L of PBS. Turk's staining solution (99 μ L) was used to enumerate intestinal lymphocytes (1 μ L) by light microscopy. This technique yields high lymphocyte recovery (i.e., 90.9 ± 0.5% CD45⁺ by flow cytometry analysis).

Western Blot Analysis of Cytokines and Apoptotic Proteins Intestinal lymphocytes (IL) were fractionated in lysis buffer on ice for 45 min. Lysates (1×10^5) were centrifuged (15 min, 10,000 g) and supernatant extracted. A BCA assay was used to determine protein concentration (Lowry et al., 1951). Protein (40 µg) and selected molecular weight markers (Full Range Rainbow, Amersham Biosciences, Pittsburgh, Pennsylvania, U.S.) were electrophoresed on a 12 per cent SDS-PAGE gel before transfer to PVDF membrane. Membranes were stained with Ponceau S to confirm quality of transfer and equal loading. After electrophoresis, membranes were incubated for 1 hr with primary antibody (1:200 in 10% milk-TBST): TNF-α (clone: N-19; goat antihuman polyclonal IgG), IL-1-β (clone: Fx02l; mouse anti-rat monoclonal), IL-6 (clone: M-19; goat anti-rat polyclonal IgG), caspase-3 (clone: H-277; rabbit antihuman polyclonal IgG), caspase-7 (clone: 10-1-62; mouse anti-human monoclonal IgG₁), and Bcl-2 (clone: C-2; mouse anti-human monoclonal IgG₁) (Santa Cruz Biotechnology, Santa Cruz, California, U.S.). Membranes were incubated for 1 hr with secondary antibody: biotin-conjugated rabbit anti-goat IgG-B (TNF-α; IL-6), and horseradish peroxidise-conjugated goat anti-mouse IgG-HRP (IL-1 β , caspase-7, Bcl-2), or goat anti-rabbit IgG-HRP (caspase-3) IgG at a concentration of 1:2000 in 10% milk-TBST. ECL Plus detection reagent (Amersham Biosciences) and the ChemiGenius 2 Bioimaging System were used for protein determination. We used a biotinylated protein ladder to identify the molecular weight of selected proteins (Cell Signalling Technology, Millipore [Canada] Ltd., Etobicoke, Ontario). Recombinant standards (Cedarlane Laboratories) were run on each gel. We ran samples from the two exercise conditions on each immunoblot and normalised band densities to control bands on each immunoblot (reported as arbitrary densitometric units [A.U.]) for each group.

Experimental Design and Statistical Analysis

All variables were analysed by one-way ANOVA with exercise training (i.e., WR, NWR) as the independent factor, and cytokine and apoptotic proteins as the dependent factors (SPSS for Windows Version 19; SPSS Inc, Chicago, Ilinois, U.S.). Running volumes (distances) by month were analysed by repeated measures ANOVA. Levene's test was used to check for homogeneity of variance. Significant differences were accepted if p < 0.05. Values are group means ± 1 SEM for respective units (e.g., µmol/min/g; arbitrary densitometric units [A.U.]; g).

Results

Physiological Indicators of Training and Exercise

Table 1 shows the physiologic characteristics [distance run (km), body weight (g), cytochrome c oxidase (μ Mol/min/g protein) activity in soleus and plantaris muscles] of healthy older C57BL/6 female mice after

Table 1: Body weight (g), running distance (km), 8-iso-PGF _{2a} (pg/ml), corticosterone (ng/ml), and cytochrome c oxidase activity	
(μMol/min/g protein) in C57Bl/6 of healthy older adult mice after 4 months of exercise training ^α	

Group	Running distance (km)						
	Month 1	Month 2	Month 3		Month 4		
WR NWR	151.4 ± 10.8 -	166.3 ± 9.6 -	133.6 ± 7.0 - (ml) Cytochrome C Oxidase		126.8 ± 11.6 - Body weight (g)		
Group	8-iso-PGF₂α (pg∕ml)	Corticosterone (ng/ml)					
			Soleus	Plantaris	Initial	Final	
WR NWR	167.6 ± 9.2* 209.4 ± 10.1	53.1 ± 9.2 45.1 ± 7.8	13.5 ± .8* 9.5 ± .5	12.3 ± .5* 8.0 ± .3	30.4 ± .5 30.7 ± .7	31.1 ± .6* 34.0 ± .8	

^a Values reported as group means ± one standard error

* denotes significant (p < .05) group effects

WR = wheel running

NWR = no wheel running

4 months of low-intensity exercise training. WR and NWR mice did not differ in initial body weights (p > .05). However, after 4 months of exercise training, WR mice were significantly lighter than NWR mice (p < .05). Over the 4 months of wheel running activity, the mean distance run per animal was 578.1 km. The average monthly distance run per mouse remained steady for the first two months and thereafter declined by 20–30 km per month. WR mice also had significantly higher cytochrome *c* oxidase activity in SOL and PLANT compared to NWR mice (p < .05).

Biomarkers of Stress

Plasma 8-iso-PGF2a and Corticosterone

Table 1 also shows the plasma concentration of 8-iso-PGF_{2a} and corticosterone in trained and untrained aged mice. WR mice had significantly lower plasma 8-iso-PGF_{2a} levels compared with NWR mice [F(1, 29) = 9.36, p < .05]. WR mice did not differ from NWR controls in their plasma corticosterone levels [F(1, 31) = 0.423, p > .05]. Given that plasma 8-iso-PGF_{2a} is an indicator of oxidative stress, it can be inferred that exercise training was associated with lower resting levels of cellular oxidant stress (Roberts & Morrow, 2000). The lack of a difference in corticosterone levels between the two groups suggests that voluntary low-intensity wheel running did not significantly influence psychogenic stress in the animals (Girard & Garland, 2002).

Western Blot Analysis of Cytokines and Apoptotic Proteins

Pro-inflammatory and Pleiotropic Cytokines

Figure 1 (Panels A-C) shows the effect of long-term (4 months) voluntary wheel running (i.e., exercise training) on mouse intestinal lymphocyte expression of two pro-inflammatory cytokines, TNF- α and IL-1 β , and the pleiotropic cytokine IL-6 (units expressed in arbitrary densitometric units [A.U.]). Long-term physical activity was associated with significantly lower expression of TNF- α [F (1, 29) = 9.51, p < .05] in WR compared to NWR mice. Training was also associated with non-significant decreases in IL-1 β [F (1, 34) = 2.63, p = .12] and IL-6 [F (1, 37) = 2.21, p = .15] expression in the intestinal lymphocytes of older mice.

Figure 2 (Panels A-C) shows the effect of long-term (4 months) voluntary wheel-running (i.e., exercise training) on intestinal lymphocyte expression of the pro-apoptotic proteins caspase-3 and caspase-7, and the anti-apoptotic protein Bcl-2. Long-term physical activity was associated with a significant reduction in caspase-7 [F(1, 34) = 7.19, p < .05] and a non-significant decrease in caspase-3 [F(1, 33) = 1.62, p = .21] expression in intestinal lymphocytes of healthy older mice. Voluntary wheel running for 4 months also led to a

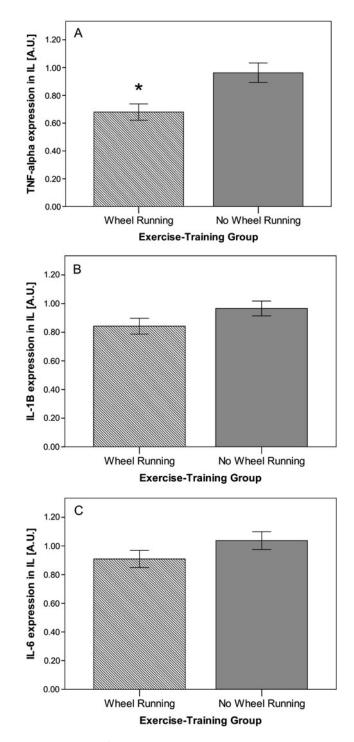


Figure 1: Pro-inflammatory and pleiotropic cytokine production in intestinal lymphocytes (IL) of aged mice.

non-significant increase of the expression in intestinal lymphocytes of the anti-apoptotic protein, Bcl-2, [F(1, 36) = 3.17, p = .08] in healthy older animals.

Discussion

The purpose of this study was to describe the effects of long-term, voluntary aerobic exercise training on

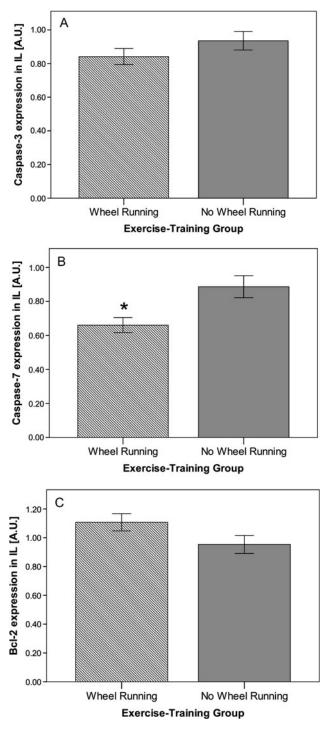


Figure 2: Pro- and anti-apoptotic protein expression in intestinal lymphocytes (IL) of aged mice.

inflammatory cytokine and apoptotic protein expression in the intestinal tract of healthy older adult mice. We hypothesized that intestinal lymphocytes of trained mice would display lower levels of apoptotic proteins (caspase-3, caspase-7) and pro-inflammatory (TNF- α , IL-1 β) and pleiotropic (IL-6) cytokines, and higher levels of anti-apoptotic proteins (Bcl-2) compared to Nicholas Packer and Laurie Hoffman-Goetz

"untrained" age-matched sedentary controls. We found that 4 months of wheel running was associated with decreased intestinal lymphocyte expression of the pro-inflammatory cytokine TNF-α and the apoptotic protein caspase-7 in this aged cohort. To our knowledge, no other studies have examined the effect of voluntary long-term physical activity on the intestinal lymphocyte expression of pro-inflammatory and pleiotropic cytokines and apoptotic proteins in healthy aged individuals (either animals or people). Collectively, the results suggest that regular physical activity may protect against senescent increases in intestinal inflammation though a mechanism of decreased expression of TNF- α and caspase-7 in lymphocytes resident in the bowel. Caspase-7 is preferentially activated by caspase-1 inflammasomes within an inflammatory environment; therefore, reduced caspase-7 expression supports the anti-inflammatory effects of regular physical activity. In addition, Lamkanfi and Kanneganti (2010) have shown that caspase-7 deficient mice are resistant to endotoxemia, suggesting that interference in caspase-7 activation may be of benefit in conditions where inflammation contributes directly to disease.

The cytokine TNF- α , a potent mediator of inflammation and carcinogenesis, induces the inflammatory effects of IL-6 (and increases production of IL-1 β) by up-regulating the transcription factor NF_k β (Wilson, 2008). Elevated TNF- α is found in the bowel of CRC patients (Grimm et al., 2010), and therapeutic application of anti-TNF-α antibodies prevents CRC development in animal models of colitis (Popivanova, Kitamura, & Wu, 2008). IL-1 β is a pro-inflammatory cytokine that promotes tumour formation by inducing cellular proliferation (Jung, Isaacs, Lee, Trepel, & Neckers, 2003), apoptosis (Wang, Zhang, Zhao, & Fei, 2009), and vasculogenesis (Jung et al., 2003). IL-6 has been shown to prevent apoptosis in myeloid LP cells (Grivennikov et al., 2009) and to promote tumour progression in animal models of colitis (Strassmann et al., 1993). Although IL-6 demonstrates both pro- and anti-inflammatory functions (Pedersen et al., 2003; Pedersen & Fischer, 2007), its effects are primarily pro-inflammatory with increasing age (Dobbs et al., 1999) or in the presence of inflammatory disease conditions (Ridker, Rifai, Stampfer, & Hennekens, 2000). Thus, decreased intestinal lymphocyte expression of these cytokines (TNF-α, IL-1β, IL-6) in response to long-term freewheel training contributes to our understanding of how regular physical activity in older people may decrease the risk of inflammatory diseases of the bowel and colorectal cancer.

Intestinal lymphocyte TNF- α expression was markedly lower in the trained, older animals. Regular physical activity may also decrease intestinal inflammation by reductions in adipose tissue, a major tissue source of non-immune-cell-derived endogenous TNF- α (Coppack, 2001), or via suppression of TNF- α transcription or translation. Intestinal TNF- α levels are thought to be predictive of risk for later CRC development (Popivanova et al., 2008). Thus, lower TNF- α may reduce the risk of developing intestinal pathology. This training-induced reduction in TNF- α is remarkable given the reported elevated expression of TNF- α and TNF-r1 in peripheral mononuclear cells (Fagiolo et al., 1993) and plasma (Stowe et al., 2010) of healthy older individuals.

Despite the older age of our animals, the results mirror those of previous studies demonstrating that training decreases baseline intestinal lymphocyte expression of TNF- α in young animals (Hoffman-Goetz et al., 2009). This was observed despite the fact that young and old mice also differ markedly in their physiologic response to training. In response to 4.5 weeks of resistance exercise (i.e., 14 sessions of electrically evoked training), young (3 months) rats showed a 15.6 per cent increase in wet muscle (tibialis anterior) weight, whereas old (30 months) rats showed no increase (Murlasits et al., 2006). Although both young and old rats showed increased (+968.8% and +409.1% respectively) HSP72 expression in tibialis muscle in response to prolonged resistance exercise, old rats demonstrated an augmented training response in spite of equal resistancetraining conditions. Moreover, since TNF-a has been shown to promote carcinogenesis (Grimm et al., 2010), our novel finding may explain, in part, the nature of the protective effect of regular exercise, even at advanced ages. Specifically, exercise training may reduce or buffer against intestinal inflammation in aged individuals despite age-related increases in inflammation and TNF-α expression (Fagiolo et al., 1993).

No differences in the intestinal lymphocyte expression of IL-1 β or IL-6 were observed although there were non-significant decreases among exercise trained older mice. This is an important observation given the lower TNF- α expression in response to training and the role of TNF- α in inducing IL-1 β and IL-6 production (Wilson, 2008). The direction of the trends for both IL-1 β and IL-6 support the possibility that TNF- α may be functioning in this manner. Further, the direction of these exercise training effects, albeit not significant, must be considered in light of the observation that healthy elderly adults have elevated expression of IL-6 and IL-1 β in immunologically active blood cells (Fagiolo et al., 1993) and a higher concentration of IL-6 in plasma (Stowe et al., 2010). Previous studies show that exercise training decreases baseline expression of IL-1β (Gomez-Merino, Drogou, Guezennec, & Chennaoui, 1997) in white adipose tissue of healthy young mice, while long-term resistance training reduces circulating IL-6 in healthy older people (Córdova et al., 2011).

Inflammation induces both the intrinsic and extrinsic pathways of apoptosis (cell death). The former (intrinsic) is mediated by mitochondrial activation of caspases and cytoplasmic translocation, subsequently leading to cell lysis (Wyllie, 2010). These effects are antagonized by cellular anti-apoptotic proteins, such as Bcl-2, which prevent the release, activation, and translocation of mitochondrial caspases (Kuwana & Newmeyer, 2003). Alternatively, the extrinsic pathway proceeds via cytokine (i.e., TNF- α , IL-1 β) induced activation of membrane death receptors (Mignini, Traini, Tomassoni, Vitali, & Streccioni, 2008). Collectively, these biological processes may help to explain how cytokine and apoptotic dysregulation can contribute to gastrointestinal pathology and uncontrolled inflammation. Nevertheless, whether apoptotic dysregulation in the bowel is "beneficial" or "harmful" to the individual depends on the specific physiological situation and cell types involved. For example, resistance to apoptosis is a hallmark of colorectal cancer cell survival (Liu, 2010) whereas excessive apoptosis of intestinal epithelium may contribute to the etiology of inflammatory diseases such as ulcerative colitis (Levine, 2000).

Our study shows that there was a significant reduction in the expression of the pro-apoptotic protein, caspase-7, in the intestinal lymphocytes of "trained" older mice. This training-induced reduction in caspase-7 may be protective in buffering aging-associated increases in the expression of this apoptotic mediator (Zhang, Chong, & Herman, 2002). Furthermore, lower caspase-7 expression may be protective, as elevated caspase-7 predicts a higher likelihood of oral squamous cell carcinoma recurrence (Coutinho-Camillo, Lourenço, Nishimoto, Kowalski, & Soares, 2010). However, the contribution of elevated caspase-7 as a risk factor for other cancers, such as colonic adenocarcinoma, remains to be determined. We did not find a significant reduction in the expression of caspase-3 in intestinal lymphocytes of older healthy animals; however, the direction of the difference between trained and untrained mice was supportive of the finding in young mice given long-term exercise training (e.g., Hoffman-Goetz & Spagnuolo, 2007). The reason for this lack of a significant training effect on caspase-3 expression is not clear from our data; age-dependent kinetics of the caspase cascade may be involved (Jiang, Walker, & Steinle, 2009).

Similar to observations others have made in young animals (Davidson, Burnett, & Hoffman-Goetz, 2006), we confirmed that 4 months of low intensity, voluntary wheel running was sufficient to induce physical, physiological, and biochemical changes indicative of "training" in aged mice. The skeletal muscles of "trained" mice showed increased cytochrome *c* oxidase enzyme activity. Cytochrome c oxidase plays a crucial role in cellular energy regulation and directly influences mitochondrial ATP production (Fontanesi, Soto, Horn, & Barrientos, 2006). The enzymatic activity of cytochrome c oxidase is reduced with increasing age (Bagh, Thakurta, Biswas, Behera, & Chakrabarti, 2011) but, in young animals, has been shown repeatedly to increase in response to long-term training (Hoffman-Goetz et al., 2010).

Training-induced increases in skeletal muscle cytochrome c oxidase can also provide an antioxidant function "buffering" against the damaging effects of aging (De Lisio et al., 2011). During the first two months of training, the total running distances of aged mice increased before falling sharply during the third and fourth months. In young mice, there was a sequential increase in running distances across the entire training duration (Hoffman-Goetz et al., 2010). The decrease in wheel running volume over the 4 months among the aged cohort confirms the findings of Turner, Kleeberger, and Lightfoot (2005) and may reflect an age-related loss of aerobic capacity (Waters et al., 2008) or an impaired endocrine response to environmental, psychological, or physiological stressors (Waters et al., 2010).

Plasma 8-iso-PGF_{2 α} is derived from membrane-derived arachidonic acid, a reaction catalyzed by reactive oxygen intermediates (Roberts & Morrow, 2000). Corticosterone is a steroid hormone produced by the adrenal gland in response to acute physiological or psychological stress. It has a crucial role in the "fight or flight" response by stimulating gluconeogenesis and, thereby, in the capability of a subject's accessing energy reserves (Girard & Garland, 2002). Both plasma 8-iso-PGF_{2 α} and corticosterone increase throughout the lifespan (Garrido, de Blas, Del Arco, Segovia, & Mora, 2010; Montine et al., 2011). We observed that WR and NWR mice did not differ in baseline stress as measured by the biomarker plasma corticosterone. Moreover, since corticosterone did not differ between WR and NWR, the observed lower levels of plasma 8-iso-PGF_{2 α} in WR mice suggests that regular exercise lowers baseline oxidative or cellular stress in older individuals. Given that 8-iso-PGF_{2 α} is a biomarker of inflammation, it possible that this phenomenon might be one mechanism whereby the beneficial effects of exercise are realized, particularly since elevated 8-iso-PGF_{2 α} predicts chronic disease progression - including some cancers (Dai & Zhu, 2009).

Collectively, our study findings of decreased TNF- α , caspase-7, and 8-iso-PGF2 α in response to long-term wheel running activity support the hypothesis that regular physical activity can modulate inflammation, even in aged individuals. Moreover, given that TNF- α

is a cytokine that "regulates" the function of other cytokines – and, indirectly – apoptosis, training may confer protection against disease initiation and progression by disrupting the harmful activity of this key cytokine. The effects of exercise on TNF- α appear to influence the direction of the caspase-7 and 8-iso-PGF_{2 α} effects particularly since (1) TNF- α binding to tumour necrosis factor-related apoptosis-inducing ligand (TRAIL) increases pro-caspase-7 cleavage and apoptosis (Zauli et al., 2003), and (2) reductions in TNF- α reduce baseline oxidative stress, which is reflected in lower levels of 8-iso-PGF2 α in plasma (Rizzo et al., 2008).

This study has five important limitations. First, we are limited by the extent to which these results can be generalized to a human population. However, animal models are appropriate to identify basic "biomarkers of aging" and improve the understanding of human disease mechanisms (Huber & Sierra, 2009). Accordingly, gerontological research using animal models is an important step in establishing scientific evidence to guide clinical geriatrics and future practice recommendations. Second, 15- to 16-month-old C57BL/6 mice were not extremely aged; however, given a life expectancy of 23 months for this strain (Anisimov, 2009) and an average age of 63 years in humans for initial diagnosis of sporadic CRC (Lynch & de la Chappelle, 1999), this age cohort should provide a useful basis for comparison. A third study limitation is that we measured protein expression and not intestinal lymphocyte mRNA for the cytokines and apoptotic proteins selected for analysis; it is possible that some proteins may have originated, not from lymphocytes, but from other cells in the intestinal tract (such as epithelial cells). Fourth, this study was done in healthy animals in order to investigate possible mechanisms contributing to the effects of "training" in a healthy population. Future research will be needed to determine if this physical-activity-associated "beneficial" effect applies in animal models of human IBD. Finally, our study population comprised only female mice. Female C57BL/6 mice have been shown to be better runners than males (Turner et al., 2005); additional studies with male C57BL/6 mice will be necessary to ensure that gender does not confound the protective effect of training.

In summary, freewheel running for 4 months in healthy aged female C57BL/6 mice was associated with decreased intestinal lymphocyte expression of the proinflammatory cytokine TNF- α , the apoptotic protein caspase-7, and the plasma marker of oxidative stress 8-iso-PGF_{2 α}. These "anti-inflammatory" effects of regular exercise raise the possibility that physical activity is protective against the development of intestinal pathology and colorectal cancer, even at later ages. This study used an animal model to explore potential biological mechanisms between physical activity and inflammatory mediators known to be involved in the etiology of chronic intestinal diseases. This animal research should be considered as hypothesis-generating, and future research, focused on biomarkers of aging and IBD, will be needed to determine if physical activity reduces inflammatory cytokines in the bowel of older, healthy humans.

References

- Anisimov, V.N. (2009). Carcinogenesis and aging 20 years after: Escaping horizon. *Mechanisms of Ageing and Development*, 130(1–2), 105–121.
- Bagh, M.B., Thakurta, I.G., Biswas, M., Behera, P., & Chakrabarti, S. (2011). Age-related oxidative decline of mitochondrial functions in rat brain is prevented by long term oral antioxidant supplementation. *Biogerontology*, 12(2), 119–131.
- Bernstein, C.N., Wajda, A., Svenson, L.W., MacKenzie, A., Koehoorn, M., Jackson, M., et al. (2006). The epidemiology of inflammatory bowel disease in Canada: A population-based study. *American Journal of Gastroenter*ology, 101(7), 1559–1168.
- Canadian Cancer Society's Steering Committee on Cancer Statistics. (2011). *Canadian cancer statistics 2011*. Toronto, Ontario, Canada: Canadian Cancer Society.
- Colditz, G.A., Cannuscio, C.C., & Frazier, A.L. (1997). Physical activity and reduced risk of colon cancer: Implications for prevention. *Cancer Causes & Control*, 8(4), 649–667.
- Compton, C.C., Fielding, L.P., Burgart, L.J., Conley, B., Cooper, H.S., Hamilton, S.R., et al. (1999). Prognostic factors in colorectal cancer. College of American Pathologists Consensus Statement. Archives of Pathology & Laboratory Medicine, 124(7), 979–994.
- Coppack, S.W. (2001). Pro-inflammatory cytokines and adipose tissue. *The Proceedings of the Nutrition Society*, 60, 349–356.
- Córdova, C., Lopes-E-Silva, F., Jr., Pires, A.S., Souza, V.C., Brito, C.J., Moraes, C.F., et al. (2011). Long-term resistance training is associated with reduced circulating levels of IL-6, IFN-gamma and TNF-alpha in elderly women. *Neuroimmunomodulation*, *18*(3), 165–170.
- Coutinho-Camillo, C.M., Lourenço, S.V., Nishimoto, I.N., Kowalski, L.P., & Soares, F.A. (2010). Caspase expression in oral squamous cell carcinoma. *Head & Neck*, Nov 12. [Epub ahead of print].
- Dai, Q., & Zhu, X. (2009). F2-isoprostanes and metabolite, and breast cancer risk. *North American Journal of Medicine* & Science, 2(3), 106–108.

- Davidson, S.R., Burnett, M., & Hoffman-Goetz, L. (2006). Training effects in mice after long-term voluntary exercise. *Medicine and Science in Sports and Exercise*, 38, 250–255.
- De Lisio, M., Kaczor, J.J., Phan, N., Tarnopolsky, M.A., Boreham, D.R., & Parise, G. (2011). Exercise training enhances the skeletal muscle response to radiationinduced oxidative stress. *Muscle & Nerve*, 43(1), 58–64.
- Demarzo, M.M., Martins, L.V., Fernandes, C.R., Herrero, F.A., Perez, S.E., Turatti, A., et al. (2008). Exercise reduces inflammation and cell proliferation in rat colon carcinogenesis. *Medicine and Science in Sports and Exercise*, 40(4), 618–621.
- Dobbs, R.J., Charlett, A., Purkiss, A.G., Dobbs, S.M., Weller, C., & Peterson, D.W. (1999). Association of circulating TNF-alpha and IL-6 with ageing and parkinsonism. *Acta Neurologica Scandinavica*, 100, 34–41.
- Edwards, B.K., Ward, E., Kohler, B.A., Eheman, C., Zauber, A.G., Anderson, R.N., et al. (2010). Annual report to the nation on the status of cancer, 1975-2006, featuring colorectal cancer trends and impact of interventions (risk factors, screening, and treatment) to reduce future rates. *Cancer*, 116(3), 544–573.
- Fagiolo, U., Cossarizza, A., Scala, E., Fanales-Belasio, E., Ortolani, C., Cozzi, E., et al. (1993). Increased cytokine production in mononuclear cells of healthy elderly people. *European Journal of Immunology*, 23(9), 2375–2378.
- Fontanesi, F., Soto, I.C., Horn, D., & Barrientos, A. (2006). Assembly of mitochondrial cytochrome c-oxidase, a complicated and highly regulated cellular process. *American Journal of Physiology. Cell Physiology*, 291(6), C1129–C1147.
- Garrido, P., de Blas, M., Del Arco, A., Segovia, G., & Mora, F. (2010). Aging increases basal but not stress-induced levels of corticosterone in the brain of the awake rat. *Neurobiology of Aging*, Apr 21. [Epub ahead of print].
- Girard, I., & Garland, T., Jr. (2002). Plasma corticosterone response to acute and chronic voluntary exercise in female house mice. *Journal of Applied Physiology*, 92(4), 1553–1161.
- Gomez-Merino, D., Drogou, C., Guezennec, C.Y., & Chennaoui, M. (2007). Effects of chronic exercise on cytokine production in white adipose tissue and skeletal muscle of rats. *Cytokine*, 40, 23–29.
- Gonzalez-Hermoso, F., Perez-Palma, J., Marchena-Gomez, J., Lorenzo-Rocha, N., & Medina-Arana, V. (2004). Can early diagnosis of symptomatic colorectal cancer improve the prognosis? *World Journal of Surgery*, 28(7), 716–720.
- Grimm, M., Lazariotou, M., Kircher, S., Höfelmayr, A., Germer, C.T., von Rahden, B.H., et al. (2010). Tumor necrosis factor-α is associated with positive lymph node status in patients with recurrence of colorectal cancer indications for anti-TNF-α agents in cancer treatment. *Analytical Cellular Pathology*, *33*(3), 151–163.

- Grivennikov, S., Karin, E., Terzic, J., Mucida, D., Yu, G.Y., Vallabhapurapu, S., et al. (2009). IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. *Cancer Cell*, *15*(2), 103–113.
- Halle, M., & Schoenberg, M.H. (2009). Physical activity in the prevention and treatment of colorectal carcinoma. *Deutsches Ärzteblatt International*, 106(44), 722–727.
- Hoffman-Goetz, L., Pervaiz, N., & Guan, J. (2009). Voluntary exercise training in mice increases the expression of antioxidant enzymes and decreases the expression of TNF-alpha in intestinal lymphocytes. *Brain, Behaviour, and Immunity*, 23, 498–506.
- Hoffman-Goetz, L., Pervaiz, N., Packer, N., & Guan, J. (2010). Freewheel training decreases pro- and increases antiinflammatory cytokine expression in mouse intestinal lymphocytes. *Brain, Behaviour, and Immunity*, 24(7), 1105–1115.
- Hoffman-Goetz, L., & Quadrilatero, J. (2003). Treadmill exercise in mice increases intestinal lymphocyte loss via apoptosis. *Acta Physiologica Scandinavica*, 179(3): 289–297.
- Hoffman-Goetz, L., & Spagnuolo, P.A. (2007). Freewheel exercise training modifies pro- and anti-apoptotic protein expression in mouse splenic lymphocytes. *International Journal of Sports Medicine*, 28(9), 787–791.
- Huber, W.R., & Sierra, F. (2009). The longevity dividend: Why invest in basic aging research? *Canadian Journal on Aging*, 28(4), 391–394.
- Iliev, I.D., Mileti, E., Matteoli, G., Chieppa, M., & Rescigno, M. (2009). Intestinal epithelial cells promote colitisprotective regulatory T-cell differentiation through dendritic cell conditioning. *Mucosal Immunology*, 2(4), 340–350.
- Jiang, Y., Walker, R.J., & Steinle, J.J. (2009). Age-associated increase in cleaved caspase 3 despite phosphorylation of IGF-1 receptor in the rat retina. *The Journals of Gerontology. Series A, Biological Sciences and Medical Sciences*, 64(11), 1154–1159.
- Jung, Y.J., Isaacs, J.S., Lee, S., Trepel, J., & Neckers, L. (2003). IL-1beta-mediated up-regulation of HIF-1alpha via an NFkappaB/COX-2 pathway identifies HIF-1 as a critical link between inflammation and oncogenesis. *The FASEB Journal*, *17*(14), 2115–2117.
- Kulaylat, M.N., & Dayton, M.T. (2010). Ulcerative colitis and cancer. *Journal of Surgical Oncology*, 101(8), 706–712.
- Kuwana, T., & Newmeyer, D.D. (2003). Bcl-2-family proteins and the role of mitochondria in apoptosis. *Current Opinion in Cell Biology*, *15*, 691–699.
- Lamkanfi, M., & Kanneganti, T.D. (2010). Caspase-7: A protease involved in apoptosis and inflammation. *International Journal of Biochemistry & Cell Biology*, 42, 21–24.
- Lefrançois, L., & Lycke, N. (2001). Isolation of mouse small intestinal intraepithelial lymphocytes, Peyer's patch,

and lamina propria cells. *Current Protocols in Immu*nology, Chapter 3: Unit 3.19.

- Levine, A.D. (2000). Apoptosis: Implications for inflammatory bowel disease. *Inflammatory Bowel Diseases*, 6(3), 191–205.
- Liu, K. (2010). Role of apoptosis resistance in immune evasion and metastasis of colon cancer. *World Journal of Gastrointestinal Oncology*, 2(11), 309–406.
- Lira, F.S., Koyama, C.H., Yamashita, A.S., Rosa, J.C., Zanchi, N.E., Batista, M.L., Jr., et al. (2009). Chronic exercise decreases cytokine production in healthy rat skeletal muscle. *Cell Biochemistry and Function*, 27(7), 458–461.
- Lowry, O.H., Rosebrough, N.J., Farr, A.L., & Randall, R.J. (1951). Protein measurement with the folin phenol reagent. *The Journal of Biological Chemistry*, 193, 265–275.
- Luo, Z., Bradley, C.J., Dahman, B.A., & Gardiner, J.C. (2010). Colon cancer treatment costs for Medicare and dually eligible beneficiaries. *Health Care Financing Review*, 31(1), 35–50.
- Lynch, H.T., & de la Chapelle, A. (1999). Genetic susceptibility to non-polyposis colorectal cancer. *Journal of Medical Genetics*, 36, 801–818.
- Mignini, F., Traini, E., Tomassoni, D., Vitali, M., & Streccioni, V. (2008). Leucocyte subset redistribution in a human model of physical stress. *Clinical and Experimental Hypertension*, 30(8), 720–731.
- Montine, T.J., Peskind, E.R., Quinn, J.F., Wilson, A.M., Montine, K.S., & Galasko, D. (2011). Increased cerebrospinal fluid F2-isoprostanes are associated with aging and latent Alzheimer's disease as identified by biomarkers. *Neuromolecular Medicine*, 13(1), 37–43.
- Murlasits, Z., Cutlip, R.G., Geronilla, K.B., Rao, K.M., Wonderlin, W.F., & Always, S.E. (2006). Resistance training increases heat shock protein levels in skeletal muscle of young and old rats. *Experimental Gerontology*, 41(4), 398–406.
- Nicklas, B.J., Hsu, F.C., Brinkley, T.J., Church, T., Goodpaster, B.H., Kritchevsky, S.B., et al. (2008). Exercise training and plasma C-reactive protein and interleukin-6 in elderly people. *Journal of the American Geriatrics Society*, 56(11), 2045–2052.
- Pedersen, B.K., & Fischer, C.P. (2007). Physiological roles of muscle-derived interleukin-6 in response to exercise. *Current Opinion in Clinical Nutrition and Metabolic Care*, 10(3), 265–271.
- Pedersen, B.K., Steensberg, A., Keller, P., Keller, C., Fischer, C., Hiscock, N., et al. (2003). Muscle-derived interleukin-6: lipolytic, anti-inflammatory and immune regulatory effects. *Pflugers Archiv*, 446(1), 9–16.
- Pironti, A., Tadeu, V., Pedroni, A., Porcu, A., Manca, A., Massarelli, G., et al. (2010). Role of routine small intestinal biopsy in adult patient with irritable bowel syndromelike symptoms. *Minerva Medica*, 101, 129–134.

- Popivanova, B.K., Kitamura, K., & Wu, Y. (2008). Blocking TNF-α in mice reduces colorectal carcinogenesis associated with chronic colitis. *The Journal of Clinical Investigation*, *118*, 560–570.
- Powrie, F. (2004). Immune regulation in the intestine: A balancing act between effector and regulatory T cell responses. *Annals of the New York Academy of Science*, 1029, 132–141.
- Ridker, P.M., Rifai, N., Stampfer, M.J., & Hennekens, C.H. (2000). Plasma concentration of interleukin-6 and the risk of future myocardial infarction among apparently healthy men. *Circulation*, 101, 1767–1772.
- Rizzo, M.R., Abbatecola, A.M., Barbieri, M., Vietri, M.T., Cioffi, M., Grella, R., et al. (2008). Evidence for antiinflammatory effects of combined administration of vitamin E and C in older persons with impaired fasting glucose: impact on insulin action. *Journal of the American College of Nutrition*, 27(4), 505–511.
- Roberts, L.J., & Morrow, J.D. (2000). Measurement of F(2)-isoprostanes as an index of oxidative stress in vivo. *Free Radical Biology & Medicine*, 28(4), 505–513.
- Samad, A.K., Taylor, R.S., Marshall, T., & Chapman, M.A. (2005). A meta-analysis of the association of physical activity with reduced risk of colorectal cancer. *Colorectal Disease*, 7(3), 204–213.
- Singh, T., & Newman, A.B. (2010). Inflammatory markers in population studies of aging. *Ageing Research Reviews*, Dec 8. [Epub ahead of print].
- Softley, A., Myren, J., Clamp, S.E., Bouchier, I.A., Watkinson, G., & de Dombal, F.T. (1988). Inflammatory bowel disease in the elderly patient. *Scandavian Journal of Gastroenterology*. *Supplement*, 144, 27–30.
- Stowe, R.P., Peek, M.K., Cutchin, M.P., & Goodwin, J.S. (2010). Plasma cytokine levels in a population-based study: Relation to age and ethnicity. *The Journals of Gerontology. Series A, Biological Sciences and Medical Sciences*, 65(4), 429–433.
- Strassmann, G., Jacob, C.O., Fong, M., & Bertolini, D.R. (1993). Mechanisms of paraneoplastic syndromes of colon-26: involvement of interleukin 6 in hypercalcemia. *Cytokine*, 5(5), 463–468.
- Swaroop, P.P. (2007). Inflammatory bowel diseases in the elderly. *Clinics in Geriatric Medicine*, 23(4), 809–821, vi.
- Szkaradkiewicz, A., Marciniak, R., Chudzicka-Strugała, I., Wasilewska, A., Drews, M., Majewski, P., et al. (2009). Proinflammatory cytokines and IL-10 in inflammatory

bowel disease and colorectal cancer patients. *Archivum Immunologiae et therapie experimentalis*, 57(4), 291–294.

- Terzić, J., Grivennikov, S., Karin, E., & Karin, M. (2010). Inflammation and colon cancer. *Gastroenterology*, 138(6), 2101–2114.
- Thune, I., & Furberg, A.S. (2001). Physical activity and cancer risk: Dose-response and cancer, all sites and site-specific. *Medicine and Science in Sports and Exercise*, 33(6 Suppl), S530–S550; discussion S609–S610.
- Turner, M.J., Kleeberger, S.R., & Lightfoot, J.T. (2005). Influence of genetic background on daily running-wheel activity differs with aging. *Physiological Genomics*, 22(1), 76–85.
- Wang, Q., Zhang, H., Zhao, B., & Fei, H. (2009). IL-1beta caused pancreatic beta-cells apoptosis is mediated in part by endoplasmic reticulum stress via the induction of endoplasmic reticulum Ca2+ release through the c-Jun N-terminal kinase pathway. *Molecular and Cellular Biochemistry*, 324(1–2), 183–190.
- Waters, R.P., Renner, K.J., Pringle, R.B., Summers, C.H., Britton, S.L., Koch, L.G., et al. (2008). Selection for aerobic capacity affects corticosterone, monoamines and wheel-running activity. *Physiology & Behavior*, 93(4–5), 1044–1054.
- Waters, R.P., Renner, K.J., Summers, C.H., Watt, M.J., Forster, G.L., Koch, L.G., et al. (2010). Selection for intrinsic endurance modifies endocrine stress responsiveness. *Brain Research*, 1357, 53–61.
- Wilson, J.A. (2008). Tumor necrosis factor alpha and colitisassociated colon cancer. *New England Journal of Medicine*, 358(25), 2733–2734.
- Wolin, K.Y., Yan, Y., Colditz, G.A., & Lee, I.M. (2009). Physical activity and colon cancer prevention: A meta-analysis. *British Journal of Cancer*, 100(4), 611–616.
- Wyllie, A.H. (2010). "Where, O death, is thy sting?" A brief review of apoptosis biology. *Molecular Neurobiology*, 42, 4–9.
- Zauli, G., Milani, D., Rimondi, E., Baldini, G., Nicolin, V., Grill, V., et al. (2003). TRAIL activates a caspase 9/7dependent pathway in caspase 8/10-defective SK-N-SH neuroblastoma cells with two functional end points: Induction of apoptosis and PGE2 release. *Neoplasia*, 5(5), 457–466.
- Zhang, Y., Chong, E., & Herman, B. (2002). Age-associated increases in the activity of multiple caspases in Fisher 344 rat organs. *Experimental Gerontology*, 37(6), 777–789.