Endocrine disruption, parasites and pollutants in wild freshwater fish

S. JOBLING^{1*} and C. R. TYLER²

¹ Department of Biological Sciences, Brunel University, Uxbridge, Middlesex, UK
 ² School of Biological Sciences, Hatherly Laboratory, Exeter University, Exeter, Devon EX4 4PS, UK

SUMMARY

Disruption of the endocrine system has been shown to occur in wild freshwater fish populations across the globe. Effects range from subtle changes in the physiology and sexual behaviour of fish to permanently altered sexual differentiation, impairment of gonad development and/or altered fertility. A wide variety of adverse environmental conditions may induce endocrine disruption, including sub-optimal temperatures, restricted food supply, low pH, environmental pollutants, and/or parasites. Furthermore, it is conceivable that any/all of these factors could act simultaneously to cause a range of disparate or inter-related effects. Some of the strongest evidence for a link between an adverse health effect, as a consequence of endocrine disruption, and a causative agent(s) is between the condition of intersex in wild roach (*Rutlius rutilus*) in UK rivers and exposure to effluents from sewage treatment works. The evidence to indicate that intersex in roach (and other cyprinid fish living in these rivers) is caused by chemicals that mimic and/or disrupt hormone function/balance in treated sewage effluent is substantial. There are a few parasites that affect the endocrine system directly in fish, including the tape worm *Ligula intestinalis* and a few parasites from the micropsora phylum. *L. intestinalis* acts at the level of the hypothalamus restricting GnRH secretion (resulting in poorly developed gonads) and is one of the very few examples where an endocrine disrupting event has been shown to result in a population-level effect (reducing it). It is well established that many parasites affect the immune system and thus the most common effect of parasites on the endocrine system in fish is likely to be an indirect one.

Key words: parasite, pollutant, endocrine, fish, disruption, intersex.

INTRODUCTION

Disruption of the endocrine system of fish has become an increasingly important area of research over the past decade. Effects that have been reported in wild fish populations around the globe include compromised growth, disruptions in reproduction and altered sexual development (Kime, 1998; Vos et al. 2000). The most thoroughly documented examples of endocrine disruption in wild fish are in roach, Rutilus rutilus, and gudgeon, Gobio gobio, living in rivers in the UK, where the presence of vitellogenin (VTG) in the blood of male fish (VTG is an oestrogen-dependent, and normally femalespecific blood protein - see below) and intersexuality (ovotestis) are widespread (Jobling et al. 1998; van Aerle et al. 2001). These effects are not restricted to freshwater fish, but have also been documented in estuarine fish such as the euryhaline flounder (Platichthys flesus; Allen et al. 1999) and the sand goby (Pomatoschistus minutus; Matthiessen et al. 2002). Studies throughout mainland Europe (Flammarion et al. 2000; Vigano et al. 2001; Gercken & Sordyl, 2002; Hecker et al. 2002), the USA (Folmar et al. 1996, 2001; Harshbarger, Coffey & Young, 2000)

* Corresponding author. Tel: 01895 274000. Fax: 01895 274348. E-mail: susan.jobling@brunel.ac.uk

and in Japan (Hashimoto et al. 2000) have similarly documented evidence for feminization of male fish, albeit that the extent and severity generally appears to be less than in the UK. In the USA and Canada, and in Australia, masculinization of wild fish has also been reported (Howell, Black & Bortone, 1980; Munkitterick et al. 1991, 1998; McMaster et al. 1991; Van Der Kraak, 1992; Batty & Lim, 1999; Bortone & Cody, 1999); in these cases female fish develop secondary sex features characteristic of males. Few studies have shown that the endocrine changes reported in wild fish populations have resulted in adverse reproductive and/or developmental consequences. Furthermore, the effects of alterations in the endocrine system in fish on immunity, longevity or population growth and stability are little understood. Recent studies have shown that wild intersex fish have altered sex steroid hormone profiles (roach and bream, Abramis brama; Jobling et al. 1998; Hecker et al. 2002), and altered spawning time and reduced sperm production (roach; Jobling et al. 2002a). Furthermore, studies in the roach have shown that these intersex fish are reproductively compromised, producing sperm with poorer motility and with a lower fertilisation success than normal male fish (Jobling *et al.* 2002b). This suggests that endocrine disruption in individual fish, potentially, has consequences to fish populations as a whole.

Parasitology (2003), **126**, S103–S108. © 2003 Cambridge University Press DOI: 10.1017/S0031182003003652 Printed in the United Kingdom

Non-reproductive alterations in physiological function have also been reported in several wild populations of fish as a consequence of disruptions in their endocrine systems. Epizootics of thyroid hyperplasia and hypertrophy (affecting 100% of the population) have been reported in various species of salmonids in heavily polluted regions of the Great lakes in the USA (see Leatherland & Sonstegard, 1982; Leatherland et al. 1989; Leatherland, 1993). Similarly, long-term exposure of flounder in mesocosms to sediments heavily polluted with cocktails of contaminants has been shown to cause reduced retinoid levels in both the liver and the plasma (Besselink et al. 1998). Very recently, thyroid abnormalities were reported in mummichogs from a polluted site in the USA (Carletta, Weis & Weis, 2002). When taken together, these studies suggest that thyroid function in fish appears to be sensitive to contaminant exposure.

There are significant gaps in our understanding of the causes of endocrine disruption in wild fish and this has hindered the process of hazard identification. Furthermore, it is now clearly established that cocktails of chemicals can have interactive effects in mediating these endocrine changes in fish. Endocrine disruption in fish, however, does not always have a chemical aetiology and can also arise as a result of exposure to a variety of other adverse environmental conditions and/or factors, including sub-optimal temperatures, restricted food supply, low pH, environmental pollutants and/or parasites. Furthermore, it is conceivable that any/all of these factors could act simultaneously to cause a range of disparate or inter-related effects. This short review is focused on the effects of pollutants and parasites (and the interactions between them) on the endocrine system of fish.

POLLUTANTS AS CAUSATIVE AGENTS OF ENDOCRINE DISRUPTION IN FISH

Some of the major causative agents of endocrine disruption seen in fish are thought to be endocrine disrupting chemicals (EDCS). These are chemicals (including both natural and man-made chemicals) that can interfere with the normal functioning of the endocrine systems in humans and wildlife (Vos et al. 2000). Endocrine-disrupting chemicals can act by selectively binding to hormone receptors to generate (agonists) or block (antagonists) hormone-mediated responses. Agonists for the oestrogen receptor, for example, include natural oestrogens such as coumesterol and genistein, oestradiol and oestrone, and pharmaceuticals such as ethinyloestradiol, and industrial chemicals such as DDT, bisphenol-A and nonylphenol. Antagonists of the androgen receptor include metabolites of the fungicide vinclozolin and the DDT metabolite p,p'DDE, and thus they block testosterone-induced cellular responses. EDCs can

also act by interfering with hormone synthesis, metabolism or excretion. Compounds that have been shown to alter oestrogen biosynthesis include cyanoketone, ketoconazole, the herbicide atrazine (Sanderson *et al.* 2001), and the fungicide fenarimol (Hirsch *et al.* 1987). Most of the known endocrine disruptors studied to date appear to affect the reproductive system, although there are other environmental agents that are known to alter the thyroid hormone system.

In UK freshwaters, it is thought that endocrine disrupters, present in treated sewage effluents, are responsible for the widespread occurrence of feminization in roach (Jobling et al. 1998). Male roach (and other species of fish) living in the proximity of sewage treatment works effluent discharges synthesise and secrete the female specific yolk protein precursor, vitellogenin, and in the more heavily impacted regions, develop ovo-testes. Vitellogenin is normally produced in the liver of female fish as a result of the binding of endogenous oestrogen to the oestrogen receptors found there. VTG is then carried by the blood to the ovary where it is sequestered by the growing oocytes and cleaved to form yolk. Male fish would not normally produce vitellogenin (as they have no ovaries), but will if they are exposed to an exogenous source of oestrogens. The presence of vitellogenin in the blood of male fish is, therefore, a reliable indicator that they have been exposed to an unnaturally high level of oestrogen (which may be derived from an endogenous and/or exogenous source). Chemical fractionation of sewage effluents, together with screening and identification of hormonally active constituents of the effluents using recombinant yeasts containing the human oestrogen receptor, has identified several natural oestrogens (oestrone and oestradiol), the synthetic contraceptive pill hormone ethinyloestradiol (from human and animal waste), as well as oestrogen-mimicking chemicals, such as 4-nonylphenol (derived from the manufacture and use of surfactants), in most of the effluents examined (Desbrow et al. 1998). Sex steroid oestrogens are extremely potent at low concentrations. Alkylphenols are comparably weak oestrogens, albeit they are more persistent, and bioaccumulate in animals. The fact that these chemicals are present in treated sewage effluent discharges that continually enter UK rivers means that fish are constantly exposed to them. Laboratory-based studies have shown that both steroid oestrogens and alkylphenols, such as 4NP, are present in effluents at concentrations sufficient to induce VTG synthesis in fish (Sheahan et al. 1994; Jobling et al. 1996; Routledge et al. 1998; Gray, Teather & Metcalfe, 1999) and, for steroid oestrogens, at concentrations close to those that will induce feminization of structural features in the testis (induction of an ovarian cavity; van Aerle et al. 2002). Furthermore, it is well established that exposure of fish eggs, embryos or

young fry to pharmacological doses of steroidal oestrogens can cause partial or complete sex reversal. Similarly, very high concentrations of 4NP will induce gonadal sex reversal in fish (Yokota et al. 2001). Whilst this does not prove that there is a direct causal link between intersexuality, seen in wild fish and the presence of these particular chemicals, the weight of evidence is very strong indeed. It is likely, therefore, that oestrogenic chemicals and their mimics are at least partially responsible for the occurrence of hermaphrodite roach in rivers that receive sewage effluents (which almost all do). Notwithstanding this, there are no studies in which a specific endocrine disruption event in wild fish has been causally linked with exposure to a specific chemical. Furthermore, effluents are highly complex mixtures of chemicals and studies investigating the interactive effects of binary mixtures of oestrogenic chemicals in fish have shown that combinations of steroid oestrogens, alkylphenolic chemicals and pesticides are additive in their effects. This highlights the fact that even chemicals that are weakly active as hormone mimics need to be taken into consideration when assessing exposure of fish populations to complex mixtures of chemicals such as those found in effluents. Complicating the issue of sexual disruption in wild fish even further is the fact that recent studies have found that effluents from STWs also have androgenic activity (Kirk et al. 2002).

PARASITES AS CAUSATIVE AGENTS OF ENDOCRINE DISRUPTION IN FISH

The relationships between parasites and their hosts are complex, and neurohormones, growth factors and hormones play prominent roles in these relationships. In vertebrates, such as fish, parasites may secrete hormones, neuropeptides, or cytokinelike molecules that influence the hosts physiological and immunological responses. Alternatively, the parasites may secrete factors that directly alter the host's hormone levels (Beckage, 1993). Having said all of this, relatively few parasites are known to affect the endocrine system of fish and cause reproductive and/or developmental dysfunction. The best known of these is the tapeworm Ligula intestinalis. Ligula acts by interfering with the pituitary-gonadal axis of its fish host so that parasitised fish are unable to reproduce (Arme, 1997; Williams et al. 1998). During its development, Ligula intestinalis grows rapidly to a large size in the fish's body cavity, characteristically distending the abdomen. There are also recent reports of parasite-induced gigantism in a single population of ligulosed roach, although whether these growth effects represent an adaptive response by the host or a manipulation by L. intestinalis of the host, remains to be elucidated (Loot et al. 2002).

Two other parasites, from the phylum Microsporidia are known to infest the gonadal tissue of fish

and can cause endocrine disruption. These parasites, Pleisistophora mirandellae, a parasite that infests the gonadal tissue, and Myxozoa (Myxobolus sp. a parasite that infests the connective tissue of the gonad and other tissues) are identified by their characteristic spores. P. mirandellae has been suggested to be the possible causative agent of intersex roach from Finnish brackish waters (Wiklund et al. 1996). These authors report the degeneration of the ovarian tissue and the appearance of testicular tissue in fish infected with the parasite. In our own extensive studies of roach in UK rivers, however, there is no evidence to suggest a link between intersexuality and P. mirandellae infection, as the two phenomena have never been found to occur simultaneously in sufficient numbers of fish to present a convincing argument. Infection with P. mirandellae is characterized by the appearance of cysts containing spores in both ovarian stroma and in the cytoplasm of the oocytes. The presence of parasitic stages in early development in primary oocytes and mature spores in secondary oocytes (those undergoing vitellogenesis - yolk uptake) provides evidence that the development of the parasite is synchronized with the development of the oocyte. In heavily infected individuals, the host or immune response is so intense that the degeneration of the ovarian tissue occurs causing a reduction in reproductive capacity. Although P. mirandellae is a highly specific parasite that normally targets oocytes (Pekkarinen, 1995), infections of testicular tissue have also been recorded. Although the infected testicular cysts did displace the testicular germ cells, the parasite does not infect the germ cells themselves.

In contrast with *P. mirandellae*, *Myobolus* sp. occurs scattered throughout the connective tissue of infected individuals. Indeed, *Myxobolus* sp. in general, display little specificity in the tissues they infect and can be found in the gill lamellae, kidney and other connective tissues within the peritoneal cavity. In addition, the host response to *Myxobolus* sp. is not as pronounced as seen in *P. mirandellae*. Notwithstanding this, there are reports of *Myxobolus* sp. inducing destruction of the ovaries leading to complete castration of parasitised individuals (Gbankoto *et al.* 2001).

When taken together, the various reports of parasitism and endocrine disruption suggest that parasitism can lead to disruption of reproduction and development in some species of fish, although the frequency with which these effects have been reported suggests that parasitism alone could not explain the widespread incidence of endocrine disruption in wild fish populations found in most parts of the world.

THE INTERACTION BETWEEN PARASITES AND POLLUTANTS

The interactions of the immune and endocrine systems means that endocrine disruption and

S. Jobling and C. R. Tyler

immunotoxicity (and therefore susceptibility to disease/infection) can occur simultaneously, and probably do in many environments where fish are challenged with pollutants, parasites and other environmental stressors (Yada & Nakanishi, 2002). Pollution of the aquatic environment with industrial or agricultural sewage is known as an important immunosuppressing factor resulting in higher susceptibility to infectious diseases (Fatima et al. 2001; Lacroix et al. 2001; Kollner et al. 2002). Recent reports suggest that sewage-caused organic enrichment of sediments may lead to an increased prevalence of infection by parasites in the vicinity of sewage treatment works outfalls (Marcogliese & Cone, 2001). These data, however, are not substantial and were not supported by studies on the immune function of these animals. In conclusion, there are clearly some chemicals and some parasites that alone can cause physiological disturbances as a consequence of inducing disruptions in the hormone balance of fish, and some cases these interactions lead to adverse health effects. To develop a clearer understanding of how parasitism and exposure to pollutants act together to impact the endocrine system in fish needs carefully targeted studies to do so, and to date these have not been forthcoming.

REFERENCES

- ALLEN, Y., MATTHIESSEN, P., HAWORTH, S., THAIN, J. E. & FEIST, S. (1999). Survey of estrogenic activity in United Kingdom estuarine and coastal waters and its effects on gonadal development of the flounder Platichthys flesus. *Environmental Toxicology and Chemistry* **18**, 1791–1800.
- ARME, C. (1997). Ligula intestinalis: Interactions with the pituitary-gonadal axis of its fish host. Journal of Helminthology 71, 83-84.
- BATTY, J. & LIM, R. (1999). Morphological and reproductive characteristics of male mosquitofish (*Gambusia affinis holbrooki*) inhabiting sewage-contaminated waters in New South Wales, Australia. Archives of Environmental Contamination and Toxicology **36**, 301–307.
- BECKAGE, N. E. (1993). Endocrine and neuro-endocrine host-parasite relationships. *Receptor* **3**, 233–245.
- BESSELINK, H. T., FLIPSEN, E. M. T. E., EGGENS, M. L., VETHAAK, A. D., KOEMAN, J. H. & BROUWER, A. (1998). Alterations in plasma and hepatic retinoid levels in flounder (*Platichthys flesus*) after chronic exposure to contaminated harbour sludge in a mesocosm study. *Aquatic Toxicology* 42, 271–285.
- BORTONE, S. A. & CODY, R. (1999). Morphological masculinization in poeciliid females from a paper mill effluent receiving tributary of the St. Johns River, Florida, USA. *Bulletin of Environmental Contamination* and Toxicology **63**, 150–156.
- CARLETTA, M. A., WEIS, P. & WEIS, J. S. (2002). Development of thyroid abnormalities in mummichogs, *Fundulus heteroclitus*, from a polluted site. *Marine Environmental Research* 54, 201–604.
- DESBROW, C., ROUTLEDGE, E. J., BRIGHTY, G. C., SUMPTER, J. P. & WALDOCK, M. (1998). Identification of estrogenic chemicals in STW effluent. 1. Chemical fractionation

and *in vitro* biological screening. *Environmental Science* and *Technology* **32**, 1549–1558.

- FATIMA, M., AHMAD, I., SIDDIQUI, R. & RAISUDDIN, S. (2001). Paper and pulp mill effluent-induced immunotoxicity in freshwater fish Channa punctatus (Bloch). Archives of Environmental Contamination and Toxicology 40, 271–276.
- FLAMMARION, P., BRION, F., BABUT, M., BARRIC, J., MIGEON, B., NOURY, P., THYBAUD, E., TYLER, C. R. & PALAZZI, X. (2000). Induction of fish vitellogenin and alterations in testicular structure: Preliminary results of estrogenic effects in chub (Leuciscus cephalus). *Ecotoxicology* 9, 127–135.
- FOLMAR, L. C., DENSLOW, N. D., KROLL, K., ORLANDO, E. F., ENBLOM, J., MARCINO, J., METCALF, C. & GUILETTE, L. J. (2001). Altered serum sex steroids and vitellogenin induction in walleye (*Stizostedion vitreum*) collected near a metropolitan sewage treatment plant. *Archives* of *Environmental Contamination and Toxicology* 40, 392–398.
- FOLMAR, L. C., DENSLOW, N. D., RAO, V., CHOW, M., CRAIN, D. A., ENBLOM, J., MARCINO, J. & GUILLETTE, L. J. (1996).
 Vitellogenin induction and reduced serum testosterone concentrations in feral male carp (*Cyprinus carpio*) captured near a major metropolitan sewage treatment plant. *Environmental Health Perspectives* 104, 1096–1101.
- GBANKOTO, A., PAMPOULIE, C., MARQUES, A. & SAKITI, G. N. (2001). Myxobolus dahomeyensis infection in ovaries of Tilapia species from Benin (West Africa). Journal of Fish Biology 58, 883–886.
- GERCKEN, J. & SORDYL, H. (2002). Intersex in feral marine and freshwater fish from northeastern Germany. *Marine Environmental Research* 54, 651–655.
- GRAY, M. A., TEATHER, K. L. & METCALFE, C. D. (1999). Reproductive success and behavior of Japanese medaka (Oryzias latipes) exposed to 4-tert-octylphenol. Environmental Toxicology and Chemistry 18, 2587–2594.
- HARSHBARGER, J. C., COFFEY, M. J. & YOUNG, M. Y. (2000). Intersexes in Mississippi River shovelnose sturgeon sampled below Saint Louis, Missouri, USA. *Marine Environmental Research* **50**, 247–250.
- HASHIMOTO, S., BESSHO, H., HARA, A., NAKAMURA, M., IGUCHI, T. & FUJITA, K. (2000). Elevated serum vitellogenin levels and gonadal abnormalities in wild male flounder (*Pleuronectes yokohamae*) from Tokyo Bay, Japan. *Marine Environmental Research* **49**, 37–53.
- HECKER, M., TYLER, C. R., HOFFMANN, M., MADDIX, S. & KARBE, L. (2002). Plasma biomarkers in fish provide evidence for endocrine modulation in the Elbe River, Germany. *Environmental Science and Technology* 36, 2311–2321.
- HIRSCH, K. S., WEAVER, D. E., BLACK, L. J., FALCONE, J. F. & MACLUSKY, N. J. (1987). Inhibition of central nervous system aromatase activity – a mechanism for fenarimol-induced infertility in the male rat. *Toxicology and Applied Pharmacology* **91**, 235–245.
- HOWELL, W. M., BLACK, D. A. & BORTONE, S. A. (1980). Abnormal expression of secondary sex characters in a population of mosquitofish, *Gambusia affinis holbrooki* – evidence for environmentally-induced masculinization. *Copeia* 980, 676–681.
- JOBLING, S., BERESFORD, N., NOLAN, M., RODGERS-GRAY, T., BRIGHTY, G. C., SUMPTER, J. P. & TYLER, C. R. (2002 *a*).

Disruption of fish endocrines by parasites and pollutants

Altered sexual maturation and gamete production in wild roach (*Rutilus rutilus*) living in rivers that receive treated sewage effluents. *Biology of Reproduction* **66**, 272–281.

JOBLING, S., COEY, S., WHITMORE, J. G., KIME, D. E., VAN
LOOK, K. J. W., MCALLISTER, B. G., BERESFORD, N.,
HENSHAW, A. C., BRIGHTY, G., TYLER, C. R. & SUMPTER, J. P.
(2002b). Wild intersex roach (*Rutilus rutilus*) have
reduced fertility. *Biology of Reproduction* 67, 515–524.

JOBLING, S., NOLAN, M., TYLER, C. R., BRIGHTY, G. & SUMPTER, J. P. (1998). Widespread sexual disruption in wild fish. *Environmental Science and Technology* 32, 2498–2506.

JOBLING, S., SHEAHAN, D., OSBORNE, J. A., MATTHIESSEN, P. & SUMPTER, J. P. (1996). Inhibition of testicular growth in rainbow trout (*Oncorhynchus mykiss*) exposed to estrogenic alkylphenolic chemicals. *Environmental Toxicology and Chemistry* **15**, 194–202.

KIME, D. E. (1998). *Endocrine Disruption in Fish*. Boston, Kluwer Academic Publishers, pp. 396.

- KIRK, L. A., TYLER, C. R., LYE, C. M. & SUMPTER, J. P. (2002). Changes in estrogenic and androgenic activities at different stages of treatment in wastewater treatment works. *Environmental Toxicology and Chemistry* **21**, 972–979.
- KOLLNER, B., WASSERRAB, B., KOTTERBA, G. & FISCHER, U. (2002). Evaluation of immune functions of rainbow trout (Oncorhynchus mykiss) – how can environmental influences be detected? Toxicology Letters 131, 83–95.

LACROIX, A., FOURNIER, M., LEBEUF, M., NAGLER, J. J. & CYR, D. G. (2001). Phagocytic response of macrophages from the pronephros of American plaice (*Hipoglossoides platessoides*) exposed to contaminated sediments from Baie des Anglais, Quebec. *Chemosphere* **45**, 599–607.

LEATHERLAND, J. F. (1993). Field observations on reproductive and developmental dysfunction in introduced and native salmonids from the Great Lakes. *Journal of Great Lakes Research* **19**, 737–751.

LEATHERLAND, J. F., LIN, L., DOWN, N. E. & DONALDSON, E. M. (1989). Thyroid hormone content of eggs and early developmental stages of 3 stocks of goitered coho salmon (*Oncorhynchus kisutch*) from the Great-Lakes of North-America, and a comparison with a stock from British Columbia. *Canadian Journal of Fisheries and Aquatic Science* **46**, 2146–2152.

LEATHERLAND, J. F. & SONSTEGARD, R. A. (1982). Bioaccumulation of organochlorines by yearling coho salmon (*Oncorhynchus kisutch walbaum*) fed diets containing great-lakes coho salmon, and the pathophysiological responses of the recipients. *Comparative Biochemistry and Physiology* **72C**, 91–100.

LOOT, G., POULIN, R., LEK, S. & GUEGAN, J. F. (2002). The differential effects of *Ligula intestinalis* (L.) plerocercoids on host growth in three natural populations of roach, *Rutilus rutilus* (L.). *Ecology of Freshwater Fish* **11**, 168–177.

- MARCOGLIESE, D. J. & CONE, D. K. (2001). Myxozoan communities parasitizing *Notropis hudsonius* (Cyprinidae) at selected localities on the St. Lawrence River, Quebec: Possible effects of urban effluents. *Journal of Parasitology* **87**, 951–956.
- MATTHIESSEN, P., ALLEN, Y., BAMBER, S., CRAFT, J., HURST, M., HUTCHINSON, T., FEIST, S., KATSIADAKI, I., KIRBY, M., ROBINSON, C., SCOTT, S., THAIN, J. & THOMAS, K. (2002).

The impact of oestrogenic and androgenic contamination on marine organisms in the United Kingdom – summary of the EDMAR programme. *Marine Environmental Research* **54**, 645–649.

MUNKITTRICK, K. R., McMASTER, M. E., McCARTHY, L., SERVOS, M. & VAN DER KRAAK, G. (1998). An overview of recent studies on the potential of pulp-mill effluents to alter reproductive parameters in fish. *Journal of Toxicology and Environmental Health B* **1**, 347–371.

MUNKITTRICK, K. R., PORTT, C. B., VAN DER KRAAK, G. J., SMITH, I. R. & ROKOSH, D. A. (1991). Impact of bleached kraft mill effluent on population characteristics, liver mfo activity, and serum steroid-levels of a Lake Superior white sucker (*Catostomus commersoni*) population. *Canadian Journal of Fisheries and Aquatic* Science **48**, 1371–1380.

- McMASTER, M. E., VAN DER KRAAK, G. J., PORTT, C. B., MUNKITTRICK, K. R., SIBLEY, P. K., SMITH, I. R. & DIXON, D. G. (1991). Changes in hepatic mixed-function oxygenase (mfo) activity, plasma steroid-levels and age at maturity of a white sucker (*Catostomus commersoni*) population exposed to bleached kraft pulp-mill effluent. *Aquatic Toxicology* **21**, 199–218.
- PEKKARINEN, M. (1995). Pleistophora mirandellae Vaney & Conte, 1901 (Protozoa: Microspora) infection in the ovary of the roach, Rutilus rutilus. Memoranda of the Society of Fauna Flora Fennica **71**, 19–32.
- ROUTLEDGE, E. J., SHEAHAN, D., DESBROW, C., BRIGHTY, G. C., WALDOCK, M. & SUMPTER, J. P. (1998). Identification of estrogenic chemicals in STW effluent. 2. *In vivo* responses in trout and roach. *Environmental Science and Technology* **32**, 1559–1565.
- SANDERSON, J. T., LETCHER, R. J., HENEWEER, M., GIESY, J. P. & VAN DEN BERG, M. (2001). Effects of chloro-s-triazine herbicides and metabolites on aromatase activity in various human cell lines and on vitellogenin production in male carp hepatocytes. *Environmental Health Perspectives* **109**, 1027–1031.

SHEAHAN, D. A., BUCKE, D., MATTHIESSEN, P., SUMPTER, J. P., KIRBY, M. F., NEALL, P. & WALDOCK, M. (1994). Chapter 9. In Sublethal and Chronic Effects of Pollutants on Freshwater Fish (ed. Muller, R. & Lloyd, R.), pp. 99–112. Cambridge, FAO, Fishing News Books, Blackwell Scientific.

VAN AERLE, R., NOLAN, M., JOBLING, S., CHRISTIENSEN, L. B., SUMPTER, J. P. & TYLER, C. R. (2001). Sexual disruption in a second species of wild cyprinid fish (the Gudgeon, *Gobio Gobio*) in United Kingdom freshwaters. *Environmental Toxicology and Chemistry* **20**, 2841–2847.

VAN AERLE, R., POUNDS, N., HUTCHINSON, T. H., MADDIX, S. & TYLER, C. R. (2002). Window of sensitivity for the estrogenic effects of ethinylestradiol in early life-stages of fathead minnow, *Pimephales promelas. Ecotoxicology* **11**, 423–434.

VAN DER KRAAK, G. J., MUNKITTRICK, K. R., MCMASTER, M. E., PORTT, C. B. & CHANG, J. P. (1992). Exposure to bleached kraft pulp-mill effluent disrupts the pituitary-gonadal axis of white sucker at multiple sites. *Toxicology and Applied Pharmacology* **115**, 224–233.

VIGANO, L., ARILLO, A., BOTTERO, S., MASSARI, A. & MANDICH, A. (2001). First observation of intersex cyprinids in the Po River (Italy). *Science of the Total Environment* **269**, 189–194.

S. Jobling and C. R. Tyler

- VOS, G., DYBING, E., GREIM, H. A., LADEFOGED, O., LAMBRE,
 C., TARAZONA, J. V., BRANDT, I. & VETHAAK, A. D. (2000).
 Health effects of endocrine-disrupting chemicals on wildlife, with special reference to the European situation. *Critical Reviews in Toxicology* 30, 71–133.
- WIKLUND, T., LOUNASHEIMO, L., LOM, J. & BYLUND, G. (1996). Gonadal impairment in roach *Rutilus rutilus* from Finnish coastal areas of the northern Baltic sea. *Diseases of Aquatic Organisms* **26**, 163–171.
- WILLIAMS, M. A., PENLINGTON, M. C., KING, J. A., HOOLE, D. & ARME, C. (1998). *Ligula intestinalis* (Cestoda) infections of roach (*Rutilus rutilus*) (Cyprinidae):

immunocytochemical investigations into the salmon- and chicken-II type gonadotrophin-releasing hormone (GnRH) systems in host brains. *Acta Parasitologica* **43**, 232–235.

- YADA, T. & NAKANISHI, T. (2002). Interaction between endocrine and immune systems in fish. *International Reviews in Cytology* **220**, 35–92.
- YOKOTA, H., SEKI, M., MAEDA, M., OSHIMA, Y., TADOKORO, H., HONJO, T. & KOBAYASHI, K. (2001). Life-cycle toxicity of 4-nonylphenol to medaka (*Oryzias latipes*). *Environmental Toxicology and Chemistry* **20**, 2552–2560.