

## Research Article

**Cite this article:** Yasin M and Andreasen C (2019) The effect of oxygen concentration on the germination of some weed species under control conditions. *Weed Sci.* **67**: 580–588. doi: [10.1017/wsc.2019.37](https://doi.org/10.1017/wsc.2019.37)

Received: 17 September 2018

Revised: 24 January 2019

Accepted: 1 July 2019

**Associate Editor:**

Neha Rana, Bayer U.S. – Crop Science

**Keywords:**

Inhibition of germination; oxygen deficiency; seed germination; weed ecology

**Author for correspondence:**

Christian Andreasen, Department of Plant and Environmental Sciences, University of Copenhagen, Højbakkegaard Allé 13, DK 2630 Taastrup, Denmark. (Email: [can@plen.ku.dk](mailto:can@plen.ku.dk))

# The effect of oxygen concentration on the germination of some weed species under control conditions

Muhammad Yasin<sup>1</sup> and Christian Andreasen<sup>2</sup> 

<sup>1</sup>PhD Scholar, Department of Plant and Environmental Sciences, Faculty of Science, University of Copenhagen, Taastrup, Denmark; Lecturer, Department of Agronomy, College of Agriculture, University of Sargodha, Sargodha, Pakistan and <sup>2</sup>Associate Professor, Department of Plant and Environmental Sciences, Faculty of Science, University of Copenhagen, Taastrup, Denmark

**Abstract**

Continuous use of heavy machinery in fields and frequent farm traffic sometimes result in soil compaction. Soil compaction reduces the oxygen (O<sub>2</sub>) concentration in soil capillaries and hence lowers the O<sub>2</sub> availability for germinating seeds. We investigated how reduced O<sub>2</sub> levels changed germination behavior of weeds to elucidate their potential to adapt to O<sub>2</sub>-deficient soils (compacted, compressed, and waterlogged soils and soils with hard surfaces). Two similar laboratory experiments were conducted with five O<sub>2</sub> treatments (20.9%, 15%, 10%, 5%, and 2.5%). The germination percentage of the invasive weed hairy fiddleneck [*Amsinckia menziesii* (Lehm.) A. Nelson & J.F. Macbr. var. *menziesii*] and the common weeds common lambsquarters (*Chenopodium album* L.) and Persian speedwell (*Veronica persica* Poir) was not significantly reduced at 15% O<sub>2</sub>. The germination of scarlet pimpernel (*Anagallis arvensis* L. ssp. *arvensis*), silky windgrass [*Apera spica-venti* (L.) Beauv.], catchweed bedstraw (*Galium aparine* L.), and knawel (*Scleranthus annuus* L.) was significantly reduced at 15% O<sub>2</sub>. The highest germination was obtained at 20.9% O<sub>2</sub> for blackgrass (*Alopecurus myosuroides* Huds.), *A. spica-venti*, *G. aparine*, annual bluegrass (*Poa annua* L.), wild mustard (*Sinapis arvensis* L.), scentless chamomile [*Tripleurospermum inodorum* (L.) Sch. Bip.], field violet (*Viola arvensis* Murray) and the less common weeds *A. arvensis* and *S. annuus*. Distribution of flora in the landscape may change on O<sub>2</sub>-deficient soils by reducing germination of some species such as *A. arvensis* and *S. annuus* and favoring others like *A. menziesii* and *C. album*. The ability to germinate at 2.5% and 5% O<sub>2</sub> may contribute to explain why *A. myosuroides* and *A. menziesii* have become successful as weeds on O<sub>2</sub>-deficient soils, as they maintained a germination percentage between 34% and 58% at 2.5% O<sub>2</sub>.

**Introduction**

Agricultural development in many countries of the world has over the last 30 yr resulted in larger farms using bigger and heavier field implements. Continuous use of heavy machinery in fields and frequent field traffic often results in problems with soil compaction in the upper layers of the soil profile (Tullberg et al. 2007). Frequent traffic, even with lightweight tractors, sometimes compacts the topsoil and generates physical soil conditions that prevent seedling emergence (Botta et al. 2006). Soil compaction reduces oxygen (O<sub>2</sub>) concentration, influences crop growth, stunts plant root development, and affects the movement of water and chemicals in multiple ways (Lipiec et al. 2003). Soil compaction reduces the oxygen concentration in soil capillaries and hence reduces O<sub>2</sub> availability for germinating seeds.

Atmospheric air typically comprises 79% N<sub>2</sub>, 21% O<sub>2</sub>, and 0.03% CO<sub>2</sub>, while the average arable topsoil atmosphere contains 79% N<sub>2</sub>, 20.3% O<sub>2</sub>, and 0.15 to 0.65% CO<sub>2</sub>. The O<sub>2</sub> in topsoil (usually the top 5.1 cm [2 in.] to 20 cm [8 in.]) has been observed to be as low as 1%, and soil CO<sub>2</sub> has been noted to be as high as 10% under extreme conditions. In soil, O<sub>2</sub> percentage decreases with depth, and the rate of decrease is more rapid in clayey or silty soils than in sandy soils (Brady 1974). Silver et al. (1999) found that O<sub>2</sub> concentrations in soil decreased gradually. O<sub>2</sub> concentration was reduced significantly in 10- to 35-cm soil depths, and in some cases even below 3%. The O<sub>2</sub> level in the soil declined with increasing depth, and at 30-cm depth, the O<sub>2</sub> concentration was less than 0.01 kg m<sup>-3</sup> soil, which is inadequate for plant growth (Topp et al. 2000).

Under farming conditions, soil compaction is not only caused by frequent use of heavy machinery but also by cattle trampling pastures. Martinez and Zinck (2004) observed that in compact soils, O<sub>2</sub> penetration resistance ranged from 0.45 MPa under forest conditions to 4.25 MPa at 3- to 12-cm depth in 9-yr-old pastures.

In addition to trampling by livestock, rolling also causes soil compaction and limits O<sub>2</sub> concentration under field conditions. After sowing, rolling is often conducted to ensure good contact between seeds and soil particles and thereby improve the water supply and avoid desiccation of germinating seeds. The purpose of rolling can also be to press the stones and soil aggregates lying on the soil surface down into the soil to avoid damage to the combine harvester during harvest. Rolling may cause soil O<sub>2</sub> depletion due to increasing intensity of soil compaction and compression. Soil aggregates can exhibit steep O<sub>2</sub> gradients over tiny distances from the aggregate surface. Sexstone et al. (1985) found that the smallest aggregates with a radius of 4 mm had an anaerobic center. Larger aggregates with a radius >10 mm often had measurable anaerobic centers with irregular O<sub>2</sub> profiles, apparently due to O<sub>2</sub> intrusion often caused by old root channels. These irregular O<sub>2</sub> profiles created by soil aggregates in the topsoil may affect germination of many species like silky windgrass [*Apera spica-venti* (L.) Beauv.], with small seeds germinating from the upper 5 mm of the topsoil.

In compact soils, O<sub>2</sub> deficiency reduces respiration capability of seeds (Benvenuti and Macchia 1995). Deprived soil structure, compressed soil, excess soil water, and greater microbial activity may inhibit gaseous movement within the soil and cause depletion in soil O<sub>2</sub> (Drew 1992; Hodgson and MacLeod 1989; Ishii and Kadoya 1991). Seed germination depends on available O<sub>2</sub> concentration, water potential, and temperature, as well as the dormancy and physiological status of the seeds (Bradford et al. 2007). Most seeds need O<sub>2</sub> to germinate, but some seeds may germinate in the absence of O<sub>2</sub> (Rumpho and Kennedy 1981). Soil compaction may favor the germination of some aggressive weed species that germinate earlier and establish better than a crop. Seeds of velvetleaf (*Abutilon theophrasti* Medik.), tall morningglory [*Ipomoea purpurea* (L.) Roth], and wild mustard (*Sinapis arvensis* L.) buried at compressed soil under low O<sub>2</sub> concentrations switched from aerobic to anaerobic metabolism, resulting in reduced germination (Holm 1972). High O<sub>2</sub> levels were required for the germination of the weed species marsh dayflower [*Murdannia keisak* (Hassk.) Hand.-Maz.], barnyardgrass [*Echinochloa crus-galli* (L.) Beauv.], late watergrass [*Echinochloa oryzicola* (Vasinger.) Vasinger], and pale smartweed [*Persicaria lapathifolia* (L.) Delarbre], while monochoria [*Monochoria vaginalis* (Burm.f.) C. Presl ex Kunth] and rock bulrush [*Schoenoplectiella juncooides* (Roxb.) Lye] germinated better at reduced O<sub>2</sub> concentration. At low O<sub>2</sub> concentration, leaf greening and root and plumule elongation of all weeds were inhibited, whereas *S. juncooides* was inhibited by high O<sub>2</sub> concentration (Kataoka and Kim 1978). Seeds of the weed bur beggarticks (*Bidens tripartita* L.) germinated faster at 5% and 10% O<sub>2</sub>, and germination was delayed at 20.9% O<sub>2</sub> (Benvenuti and Macchia 1997). Seed germination of *E. crus-galli* was inhibited by the combination of exposure to 20.9% O<sub>2</sub> and light, which prevented surface soil germination (Boyd and VanAcker 2004).

National surveys (Andreasen and Streibig 2011; Andreasen and Stryhn 2008, 2012) indicated that the occurrence of some weed species was related to the O<sub>2</sub> levels in the soil. We observed that some of the most common weeds grew well even at very compressed soils, while some rarer weeds did not.

The study objective of our research was to investigate how decreasing O<sub>2</sub> concentrations changed germination behavior of some common and less common weed species. We tested the hypothesis that the germination of some successful weed species may be favored by low oxygen concentration, while other weed species, which have become rare, may suffer.

## Materials and Methods

### Seed Source

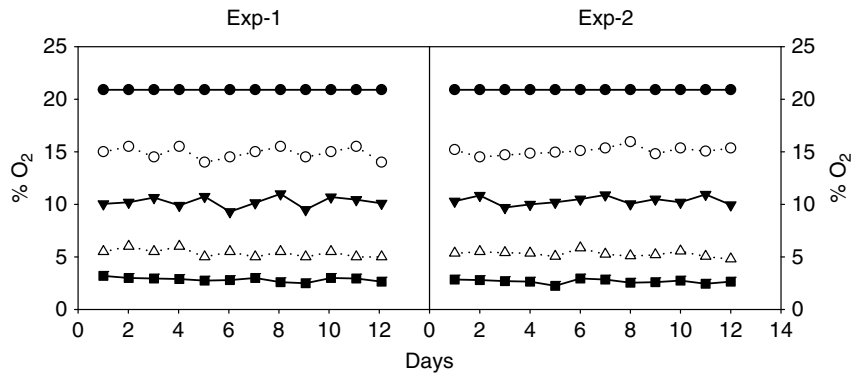
Nine broadleaf weeds were selected: one invasive weed (hairy fiddleneck [*Amsinckia menziesii* (Lehm.) A. Nelson & J.F. Macbr. var. *menziesii*]), two scarce weeds (scarlet pimpernel [*Anagallis arvensis* L. ssp. *arvensis*], knawel [*Scleranthus annuus* L.]), and six common weeds (common lambsquarters [*Chenopodium album* L.], catchweed bedstraw [*Galium aparine* L.], wild mustard (*Sinapis arvensis* L.), scentless chamomile [*Tripleurospermum inodorum* (L.) Sch. Bip.], Persian speedwell [*Veronica persica* Poir], field violet [*Viola arvensis* Murray]). We also selected three common grass weeds (blackgrass [*Alopecurus myosuroides* Huds.], *A. spica-venti*, annual bluegrass [*Poa annua* L.]).

A species was defined to be common if it occurred with a frequency above 5% in one of the most common crops in the latest national weed survey in Denmark (Andreasen and Stryhn 2008, 2012). A species was defined to be scarce if it occurred with a frequency below 1%. An invasive plant is a nonnative (alien) that is able to establish on many sites, grows quickly, spreads to the point of disrupting plant communities or ecosystems, and causes or is likely to cause economic or environmental harm or harm to human health. *Amsinckia menziesii* is such a plant in Scandinavia. *Poa annua*, *C. album*, *G. aparine*, *S. arvensis*, *T. inodorum*, *V. persica*, and *V. arvensis* are examples of weeds that have been common for many decades (Andreasen et al. 1996; Andreasen and Stryhn 2008, 2012) while *A. spica-venti* has become common during the last 25 yr (Andreasen and Stryhn 2008). *Alopecurus myosuroides* cannot be characterized as a common species according to the survey from 2004, but farm advisers now consider it a common weed in winter cereals. *Anagallis arvensis* and *S. annuus* are examples of species that have become scarce in common crops (Andreasen et al. 1996; Andreasen and Stryhn 2008, 2012) They have relatively small seed production, and it would probably take a long time before they could become common again, even if their growing conditions were improved.

Weed plants for the experimental study were collected and harvested at maturity from different crop fields at the experimental farms at Hojbjerggaard, Taastrup, Denmark (55.63°N, 12.28°E). Plants were dried in the shade for 3 wk at room temperature (14 ± 4 C). Seeds from plants were threshed using a threshing machine (Wintersteiger LD 350, 4705W, Amelia Earhart Drive, Salt Lake City, UT, USA) and cleaned using a seed cleaner (Westrup LA-LS, Slagelse, Denmark). Cleaned seeds were stored at 5 C for later use. The seed moisture was estimated to be 8% to 12% using a grain moisture meter (Wile 55, model W1910/FLFM, no.1, 3rd Shangdi, Haidian District, Beijing, China).

### Germination Experiment

Two similar experiments following a completely randomized design were performed at two separate times. Experiment 2 was performed after completion of Experiment 1. According to International Seed Testing Association (ISTA) procedures for seed testing, fewer than 400 seeds can be tested. In such cases, at least 100 seeds must be tested in replicates of 25 or 50 (ISTA 2011). We used 100 seeds in two replications per treatment level for each species in each germination bioassay. Fifty seeds of individual species were placed on moist filter paper in germination boxes (ISTA 2011) randomly placed in airtight glass containers (50-cm long, 30-cm wide, and 20-cm high) at 15 C under a



**Figure 1.** Fluctuation in oxygen concentrations for 12 weed species and eight crops at 20.9% O<sub>2</sub> (filled circles), 15% O<sub>2</sub> (open circles), 10% O<sub>2</sub> (filled triangles), 5% O<sub>2</sub> (open triangles), and 2.5% O<sub>2</sub> (filled squares) in airtight glass containers during germination Experiments 1 and 2.

16-h light/8-h dark photoperiod in a growth chamber (Termaks AS, Nino Lab, Køge, Denmark) containing tube lights on both sides, providing light intensity of 25,000 lx. Many of the weeds in this study were winter annuals (*A. arvensis*, *G. aparine*, *S. arvensis*, *V. persica*, *V. arvensis*, and the three grass weeds, *A. myosuroides*, *A. spica-venti*, and *P. anuua*). However, *C. album* is a summer annual in Denmark. In general, for a standard ISTA test, an alternating temperature regime of 20 C for 16 h and 30 C for 8 h is recommended for some of the species (ISTA 2011). We chose 15 C for all plant species, which is closer to the field conditions during summer and also appropriate for winter species that usually germinate during autumn in Denmark. Keeping the same temperature for all species made it possible to compare the response of the species to reduced O<sub>2</sub> concentrations. Each germination box was irrigated once with 60 ml of water, which was sufficient for every 14-d germination trial. Seeds were assumed to have germinated when either radicle tissue or the cotyledons protruded 2 mm beyond the seed coat. Germination was counted daily following a seedling evaluation protocol (AOSA 1990) until a constant count was achieved in each glass case. A total of 12,000 seeds were tested (5 O<sub>2</sub> treatment levels by 50 seeds by 2 replicates by 2 experiments by 12 plant species). Seeds and seedlings remained in the germination boxes in airtight glass containers for 14 d for observation, and O<sub>2</sub> was added as necessary to compensate for O<sub>2</sub> consumption and adjust the O<sub>2</sub>:N<sub>2</sub> balance.

### Oxygen Concentration

Five oxygen concentrations (20.9%, 15%, 10%, 5%, and 2.5%) were obtained by mixing N<sub>2</sub> gas with O<sub>2</sub> in airtight glass containers. The O<sub>2</sub> concentrations were kept constant during the experiments (Figure 1). The N<sub>2</sub> gas from a liquid N<sub>2</sub> cylinder was injected into the containers manually. One Gasman Personal Gas Monitor (Crowcon Detection Instruments, Rotterdam, Netherlands) was placed inside each glass container to monitor and keep the O<sub>2</sub> concentrations constant inside the containers during the experiments. N<sub>2</sub>:O<sub>2</sub> mixtures were monitored and adjusted daily, and the ratio was maintained inside each glass container. Ambient air contains about 78.09% N<sub>2</sub>, 20.95% O<sub>2</sub>, and 0.039% CO<sub>2</sub>. O<sub>2</sub> is required for germination of most species (Copeland and McDonald 2012). CO<sub>2</sub> concentrations higher than 0.039% retard germination, while N<sub>2</sub> has no influence (Copeland and McDonald 2012). In our experiments, we tried to isolate the effect of O<sub>2</sub> by continuous monitoring and adjustment to maintain a constant O<sub>2</sub> and N<sub>2</sub> ratio in order to find

differences between the sensibilities of the species to this specific germination factor. The glass containers used for these experiments had large volumes, and the CO<sub>2</sub> released from germinating seeds inside the glass containers was very small and was assumed to have no influence on the result.

### Data Analysis and Statistics

Experiments were analyzed separately. Statistical analyses were performed using the open-source program R v. 3.3.1 (<http://www.R-project.org>). The seed germination for each experiment was modeled using a cumulative distribution function of the standard three-parameter log-logistic model, allowing a different curve for each treatment using the add-on package drc (Ritz and Streibig 2005), as:

$$F(t) = \frac{d}{1 + \exp\{b[\log(t) - \log(t_{50})]\}} \quad (\text{Model 1})$$

where  $F(t)$  is the fraction of seeds germinated at time  $t$  (days). The upper-limit parameter,  $d$ , denotes the proportion of seeds that germinated during the experiment out of the total number of seeds. The parameter  $b$  is proportional to the slope of  $F$  at time  $t$  equal to the parameter  $t_{50}$ , where 50% of the total seeds germinated during the experimental period. The parameter  $t_{50}$  indicates the germination speed of the population measured in days. The estimation and model-checking procedures were based on treating the data as event times, that is, recording the time it takes for germination (the event of interest) to occur as described by Ritz et al. (2013). An overall test for any differences between  $d$  was made by comparing a model with all parameters allowed to vary between curves with a model in which only  $d$  was restricted to be the same for all curves using an ANOVA test. In a post hoc procedure, all pairwise comparisons for  $d$  were made, and significance levels were assessed by an LSD test.

### Results and Discussion

Generally, low O<sub>2</sub> concentrations changed the germination success (fraction of germinated seeds:  $d$ ) and germination speed (time to reach 50% germination:  $t_{50}$ ) (Tables 1 and 2). However, the germination percentage for some weed species did not decline at reduced O<sub>2</sub> concentration compared with ambient air (20.9% O<sub>2</sub>). In both experiments, the germination percentage of *C. album* was not significantly reduced when O<sub>2</sub> concentration fell to 15%. It declined when O<sub>2</sub> dropped from 15% to 2.5% (Table 1). The highest

**Table 1.** Estimated regression parameters of the germination curves (Model 1) with SE in parentheses for six weed species.<sup>a</sup>

Weed species	O <sub>2</sub>	t <sub>50</sub>		-b	
		Exp. 1	Exp. 2	Exp. 1	Exp. 2
	%				
<i>Alopecurus myosuroides</i>	20.9	4.48 (0.11)	5.28 (0.15)	7.83 (0.80)	6.58 (0.64)
	15	4.87 (0.12)	4.97 (0.22)	8.17 (0.85)	4.48 (0.46)
	10	4.96 (0.16)	4.92 (0.23)	6.55 (0.73)	4.58 (0.52)
	5	4.94 (0.17)	4.34 (0.30)	6.71 (0.83)	3.88 (0.57)
	2.5	6.73 (0.36)	4.84 (0.24)	5.75 (0.91)	6.14 (0.93)
<i>Amsinckia micranta</i>	20.9	4.54 (0.16)	5.78 (0.51)	5.36 (0.53)	3.06 (0.42)
	15	4.38 (0.15)	5.23 (0.29)	5.20 (0.51)	3.66 (0.41)
	10	4.91 (0.18)	6.74 (0.72)	5.11 (0.53)	3.01 (0.45)
	5	4.99 (0.25)	7.91 (2.06)	4.25 (0.49)	2.28 (0.48)
	2.5	7.20 (0.37)	5.90 (0.77)	5.28 (0.77)	2.83 (0.52)
<i>Angallis arvensis</i>	20.9	5.18 (0.20)	8.29 (0.55)	6.98 (1.01)	0.09 (2.27)
	15	5.60 (0.25)	7.72 (0.69)	9.44 (1.92)	6.58 (2.04)
	10	6.03 (0.44)	9.10 (1.32)	7.10 (1.88)	7.06 (3.50)
	5	53.02 (82.7)	19.05 (10.01)	1.43 (1.29)	4.67 (4.54)
	2.5	16.11 (3.27)	16.11 (3.27)	7.75 (5.13)	7.75 (5.13)
<i>Apera spica-venti</i>	20.9	5.57 (0.12)	9.05 (0.37)	10.83 (1.37)	9.80 (2.20)
	15	5.65 (0.13)	8.70 (0.39)	11.09 (1.53)	9.30 (2.15)
	10	5.92 (0.17)	9.61 (0.51)	9.76 (1.41)	10.28 (3.10)
	5	7.17 (0.24)	11.05 (0.93)	12.04 (2.55)	15.94 (9.00)
	2.5	9.41 (0.28)	15.54 (4.61)	19.82 (6.28)	8.37 (8.27)
<i>Chenopodium album</i>	20.9	6.13 (0.53)	9.03 (1.62)	3.63 (0.72)	3.34 (0.75)
	15	5.12 (0.33)	8.35 (1.87)	4.20 (0.62)	2.45 (0.50)
	10	4.94 (0.38)	6.97 (1.30)	4.24 (0.79)	2.61 (0.58)
	5	5.01 (0.57)	5.41 (0.63)	3.44 (0.73)	3.14 (0.65)
	2.5	5.86 (0.59)	9.19 (2.46)	4.45 (1.06)	3.75 (1.48)
<i>Galium aparine</i>	20.9	9.13 (0.30)	9.93 (0.58)	14.60 (3.80)	10.48 (3.28)
	15	9.20 (0.30)	10.40 (1.19)	17.78 (5.53)	8.80 (3.61)
	10	9.48 (1.19)	10.84 (1.02)	1.14 (2.72)	15.04 (9.37)
	5	1.04 (1.18)	1.04 (1.18)	1.25 (2.95)	1.25 (2.95)
	2.5	14.70 (6.37)	14.7 (6.37)	11.03 (7.65)	11.03 (7.65)

<sup>a</sup>t<sub>50</sub> is the time to 50% germination of those seeds that germinated in the research period. b expresses the slope at t<sub>50</sub>.

germination percentage was obtained at 15% O<sub>2</sub> in both experiments (Experiment 1:  $d = 0.46$ ; Experiment 2:  $d = 0.66$ ). *Veronica persica* followed the same trend (Table 2).

*Galium aparine*, *S. arvensis*, *T. inodorum*, and *V. arvensis* were all negatively affected when the O<sub>2</sub> concentration declined (Figures 2–4; Tables 1 and 2). *Tripleurospermum inodorum* exhibited maximum germination at 20.9% O<sub>2</sub> (Experiment 1:  $d = 0.87$ ;  $t_{50} = 3.17$ ; Experiment 2:  $d = 0.89$ ;  $t_{50} = 3.84$ ) (Figure 4; Table 2). The germination percentage decreased gradually with declining O<sub>2</sub> (Table 2; Figure 3) but sustained germination even at the lowest O<sub>2</sub> level (2.5%) (Experiment 1:  $d = 0.70$ ; Experiment 2:  $d = 0.71$ ) (Figure 3), which may give a competitive advantage over companion weed species sensitive to O<sub>2</sub>-deficient soils.

The germination of *A. menziesii* was significantly increased when the O<sub>2</sub> level dropped from 20.9% to 15% in Experiment 2, but declined when the O<sub>2</sub> concentration fell to 5% and 2.5% in both experiments. An O<sub>2</sub> level of 15% gave the fastest germination and highest germination percentage (Experiment 1:  $d = 0.90$ ;  $t_{50} = 4.38$ ; Experiment 2:  $d = 0.93$ ;  $t_{50} = 5.23$ ), followed by 20.9% O<sub>2</sub> (Figure 1; Table 1). Germination was significantly slower at 20.9% O<sub>2</sub> in Experiment 2, probably because of seed aging. The 2.5% O<sub>2</sub> resulted in slower germination (Experiment 1:  $t_{50} = 7.20$ ; Experiment 2:  $t_{50} = 5.90$ ) and reduced the germination percentage (Experiment 1:  $d = 0.58$ ; Experiment 2:  $d = 0.46$ ) (Figure 2; Table 1).

*Amsinckia menziesii* is an invasive species in Europe and is native to the states of Oregon and Washington, USA. It has become a widespread and aggressive weed in many parts of southern Scandinavia (Andreasen and Streibig 2011). Its success is probably caused by several factors such as fast growth and big seed

production over a long period of the growing season. Our experiments also showed that it germinates well at 15% O<sub>2</sub>. This may give *A. menziesii* a competitive advantage on O<sub>2</sub>-deficient soils compared with weed species that have maximum germination at 20.9% O<sub>2</sub> and reduced germination at 15% O<sub>2</sub> or below. Low O<sub>2</sub> concentration combined with a high CO<sub>2</sub> level in the soil may act as a signal for early completion of the germination phases in *A. menziesii* seeds and may promote synchronized seedling establishment. A similar effect has been reported for *E. crus-galli*, in which an increased level of CO<sub>2</sub> following rainfall acted as a germination signal (Yoshioka et al. 1998).

*Anagallis arvensis* and *S. annuus* were among the most sensitive species to any drop in O<sub>2</sub> concentration. The germination percentage of these species was relatively low, probably due to seed dormancy, but even if various methods were used to release the dormancy (see Yasin and Andreasen 2015), we would expect that the germination trend would remain the same at declining O<sub>2</sub> levels. For all individual species, the trend was the same in the two experiments, although the germination ability varied between the two experiments. The same trend has also been shown for horticultural species exposed to declining O<sub>2</sub> levels and subsequently having altered germination ability (Yasin and Andreasen 2016). *Scleranthus annuus* and *A. arvensis* have become less common (Andreasen and Stryhn 2008, 2012), probably due to several factors, such as their susceptibility to herbicides and sensitivity to low O<sub>2</sub> content in soil combined with relatively small biomass and seed production.

*Alopecurus myosuroides* and *P. annua* were less sensitive to decreasing O<sub>2</sub> concentrations (Figures 2 and 3). Germination

**Table 2.** Estimated regression parameters of the germination curves (Model 1) with SE in the parentheses for six weed species.<sup>a</sup>

Weed species	O <sub>2</sub>	t <sub>50</sub>		-b	
		Exp. 1	Exp. 2	Exp. 1	Exp. 2
<i>Poa annua</i>	20.9	5.47 (0.07)	9.39 (0.38)	16.06 (1.59)	7.00 (0.93)
	15	5.94 (0.11)	10.15 (0.24)	9.98 (0.96)	11.49 (1.68)
	10	6.07 (0.13)	15.11 (9.76)	8.77 (0.84)	4.90 (1.76)
	5	6.78 (0.20)	10.30 (0.69)	7.02 (0.75)	9.12 (2.58)
	2.5	8.01 (0.44)	11.27 (2.90)	6.77 (1.37)	6.04 (2.72)
<i>Scleranthus annuus</i>	20.9	8.63 (0.31)	9.24 (0.53)	11.99 (2.83)	11.00 (4.05)
	15	9.14 (0.61)	7.97 (0.61)	10.66 (4.07)	12.01 (5.88)
	10	19.05 (10.0)	9.05 (0.59)	4.67 (4.54)	16.72 (9.32)
	5	15.54 (4.61)	11.55 (4.54)	8.37 (8.27)	8.63 (8.00)
	2.5	16.11 (3.27)	11.42 (4.00)	7.75 (5.13)	11.36 (12.9)
<i>Sinapis arvensis</i>	20.9	3.04 (0.14)	3.45 (0.13)	5.83 (0.85)	7.40 (1.08)
	15	3.12 (0.12)	3.65 (0.15)	8.18 (1.39)	7.51 (1.19)
	10	3.90 (0.24)	4.36 (0.34)	6.51 (1.37)	5.49 (1.29)
	5	4.97 (0.59)	5.92 (0.42)	6.93 (2.70)	56.18 (302.)
	2.5	7.13 (1.72)	23.50 (16.53)	4.47 (2.37)	1.92 (1.27)
<i>Triplerospermum inodorum</i>	20.9	3.17 (0.09)	3.84 (0.14)	6.85 (0.70)	5.17 (0.50)
	15	3.35 (0.12)	4.22 (0.20)	5.15 (0.51)	4.22 (0.44)
	10	3.69 (0.14)	3.67 (0.11)	5.08 (0.51)	6.67 (0.70)
	5	4.29 (0.27)	4.50 (0.25)	3.77 (0.45)	3.84 (0.45)
	2.5	8.18 (1.43)	7.35 (0.88)	2.68 (0.49)	3.00 (0.49)
<i>Veronica persica</i>	20.9	4.88 (0.08)	5.54 (0.11)	10.65 (1.04)	8.66 (0.83)
	15	4.78 (0.07)	5.40 (0.08)	12.10 (1.17)	12.08 (1.16)
	10	5.55 (0.12)	5.85 (0.13)	8.86 (0.84)	8.07 (0.77)
	5	8.20 (0.17)	9.46 (0.48)	10.01 (1.10)	6.38 (0.91)
	2.5	9.16 (0.18)	10.68 (0.32)	21.11 (4.77)	15.92 (4.23)
<i>Viola arvensis</i>	20.9	4.38 (0.12)	4.97 (0.14)	8.41 (1.05)	9.88 (1.40)
	15	4.53 (0.10)	4.92 (0.18)	14.36 (2.12)	8.83 (1.44)
	10	4.68 (0.11)	4.34 (0.13)	12.66 (1.92)	11.55 (2.04)
	5	6.68 (0.53)	5.32 (0.30)	4.86 (0.96)	6.49 (1.21)
	2.5	8.04 (0.15)	9.50 (0.84)	67.1 (146.8)	9.53 (4.66)

<sup>a</sup>t<sub>50</sub> is the time to 50% germination of those seeds that germinated in the research period. *b* expresses the slope at t<sub>50</sub>.

was fastest and germination percentage was largest for *A. myosuroides* at 20.9% O<sub>2</sub> (Experiment 1: *d* = 0.82, t<sub>50</sub> = 4.48; Experiment 2: *d* = 0.89, t<sub>50</sub> = 5.28). There was a significant reduction in the germination speed and percentage when O<sub>2</sub> was reduced from 20.9% to 2.5% (Table 1; Figure 3). At the lowest O<sub>2</sub> concentration, 40% and 34% of the *A. myosuroides* seeds were able to germinate in Experiments 1 and 2, respectively.

*Alopecurus myosuroides* and *A. spica-venti* have posed increasing weed problems in Europe the last three decades, but their success seems more to be a result of the immense shift from summer to winter annual crops and the development of herbicide-resistant biotypes (Andreasen and Stryhn 2012) together with their ability to adapt to compacted soils. However, compact soils also ensure better water uptake and reduce soil evaporation, which can be an advantage for species sensitive to drought like *P. annua*, which has become the most common weed in many fields. Our results support the finding that common species like *P. annua* are favored by soil compaction (Warwick 1979) due to low O<sub>2</sub>. In our study, the three monocotyledons (*A. myosuroides*, *A. spica-venti*, and *P. annua*) were able to germinate even at 2.5% O<sub>2</sub> concentration. Heichel and Day (1972) also found that some monocotyledons were able to germinate even below 2 kPa O<sub>2</sub>. However, as our studies and that of Siegel and Rosen (1962) have shown, the capacity to germinate at low O<sub>2</sub> is not restricted to monocotyledons.

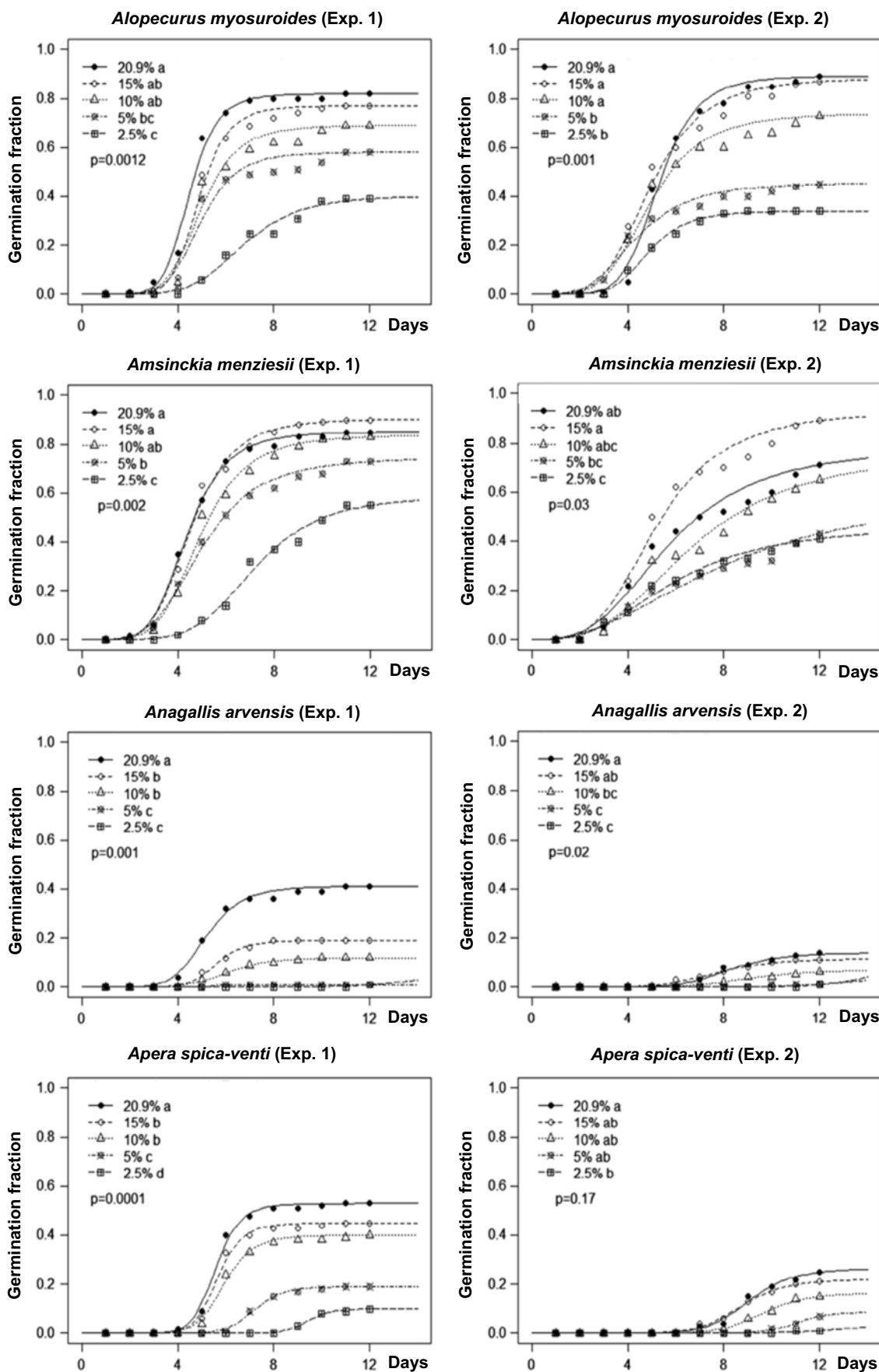
Our results cannot be compared directly with weed behaviors under field conditions, where CO<sub>2</sub> produced by root respiration and germinating plants, soil organisms, and microbial activities sometimes affects seed germination. Many factors, such as root respiration and respiration of soil-dwelling animals and

microorganisms, can affect germination conditions in the soil. CO<sub>2</sub> release may lead to the production of increased ethanol in cells, which is toxic to seed metabolism and inhibits seed germination (Thomson and Greenway 1991). We cannot expect the same response under field conditions, but testing under field conditions is normally unsatisfactory, as the results cannot be repeated with reliability (ISTA 2011). Consequently, it makes good sense to make germination tests under control conditions, even though we should keep in mind that many other factors affect germination locally in the field, and O<sub>2</sub> deficiency is often only temporary.

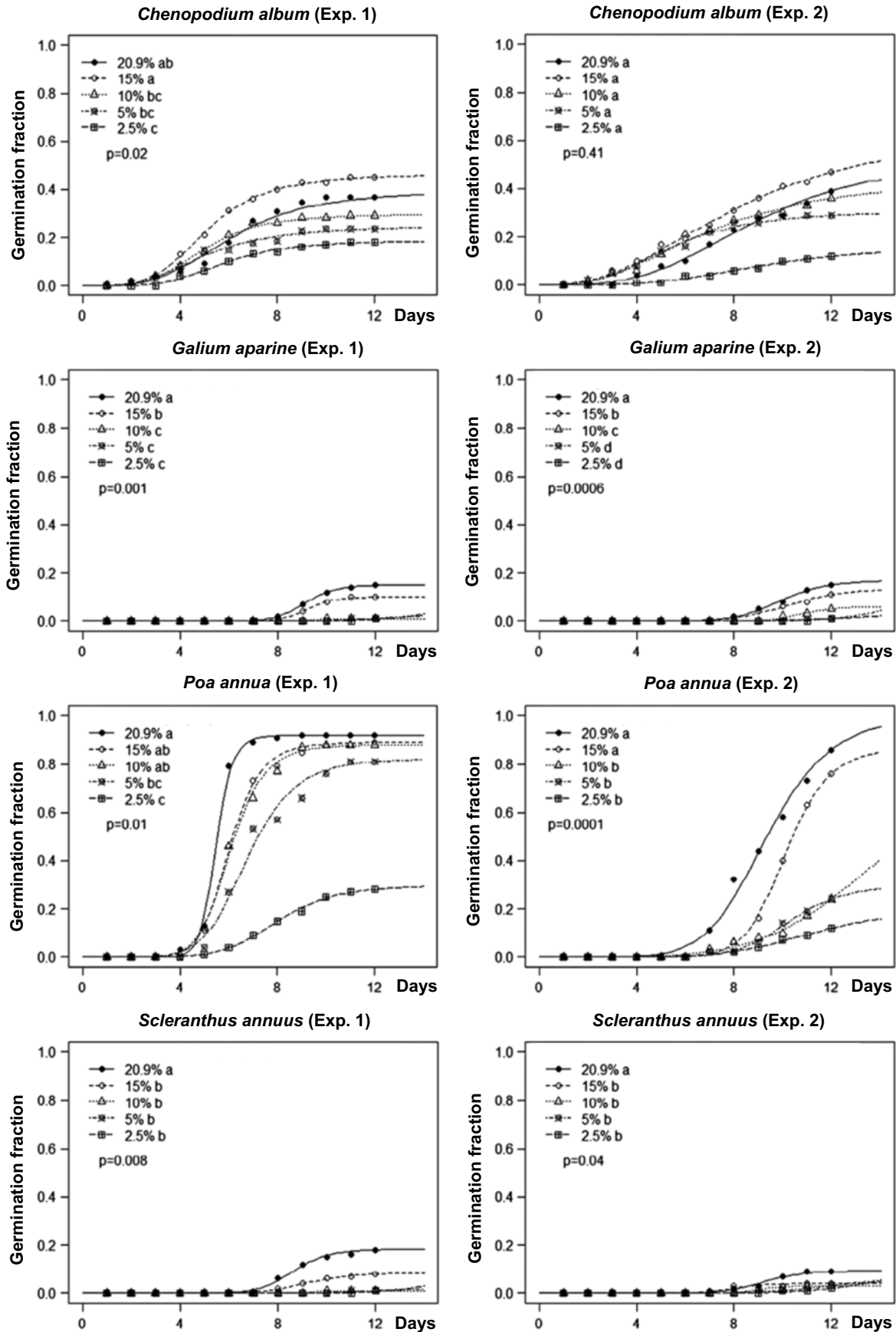
### Weed Flora Changes Over Time

Some species decline in abundance, whereas others increase due to the effect of a single factor (e.g., herbicide use, low O<sub>2</sub>) or multiple factors like improved agricultural management, changes in the farming system, and use of more broad-spectrum herbicides (Marshall et al. 2003). Boyd and VanAcker (2004) reported that germination of *S. arvensis* increased with increasing O<sub>2</sub> concentration. Fluctuating temperature, light, water, oxygen, ethylene, and nitrate are known to promote seed germination in many species (Yoshioka et al. 1998). The germination of *A. menziesii*, *C. album*, and *V. persica* was significantly favored by 15% O<sub>2</sub>. Recent modifications in agricultural practices, that is, enhanced levels of disturbance (soil compaction), favor the most generalist species, leading to biotic homogenization in arable landscapes (Fried et al. 2010).

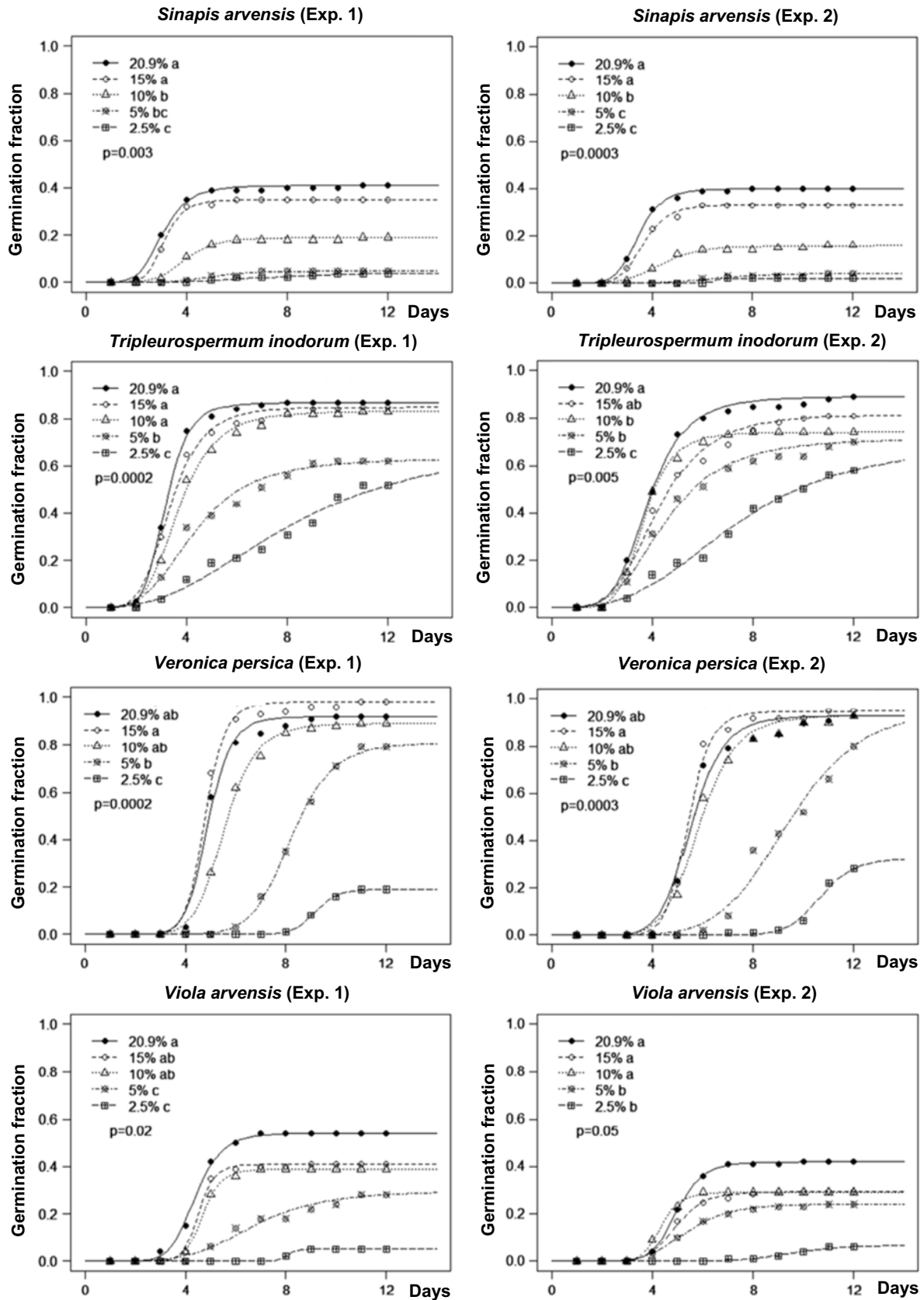
Our experiments have been able to show the effect of reduced O<sub>2</sub> concentrations on the germination of weed species under



**Figure 2.** Three-parameter log-logistic dose-response curves of germination over time for the weed species *Alopecurus myosuroides*, *Amsinckia menziesii*, *Anagallis arvensis*, and *Apera spica-venti* at five (20.9%, 15%, 10%, 5%, and 2.5%) O<sub>2</sub> concentrations from two independent experiments. Experiment 2 was performed after completion of Experiment 1. One-way ANOVA and LSD post hoc tests were performed. P-values and small letters beside each % O<sub>2</sub> treatment on the graphs show statistical significance.



**Figure 3.** Three-parameter log-logistic dose–response curves of germination over time for the weed species *Chenopodium album*, *Galium aparine*, *Poa annua*, and *Scleranthus annuus* at five (20.9%, 15%, 10%, 5%, and 2.5%) O<sub>2</sub> concentrations from two independent experiments. Experiment 2 was performed after completion of Experiment 1. One-way ANOVA and LSD post hoc tests were performed. P-values and small letters beside each % O<sub>2</sub> treatment on the graphs show statistical significance.



**Figure 4.** Three-parameter log-logistic dose–response curves of germination over time for the weed species *Sinapis arvensis*, *Tripleurospermum inodorum*, *Veronica persica*, and *Viola arvensis* at five (20.9%, 15%, 10%, 5%, and 2.5%) O<sub>2</sub> concentrations from two independent experiments. Experiment 2 was performed after completion of Experiment 1. One-way ANOVA and LSD post hoc tests were performed. P-values and small letters beside each % O<sub>2</sub> treatment on the graphs show statistical significance.



control conditions that make comparison possible. Our findings support the hypotheses that reduced O<sub>2</sub> concentrations due to soil compaction may be one reason why the invasive species *A. menziesii* and the aggressive species *C. album* and *T. inodorum* have become very common. It may also contribute to explain why the weed species *A. arvensis* and *S. annuus*, which are sensitive to reduces O<sub>2</sub> levels, have become rare. The ability of the two winter annual grass species *A. myosuroides* and *A. spica-venti* to germinate at 2.5% O<sub>2</sub> may also contribute to our understanding of why they have become extremely successful as weeds on O<sub>2</sub>-deficient soils during the last 20 yr in an area where autumn-sown crops have increased significantly in Europe.

**Acknowledgments.** We thank the University of Copenhagen, Denmark, for providing research support and facilities for the experiments, and the University of Sargodha, Pakistan, for awarding the Faculty Development Program (FDP) scholarship for doctoral study to MY. No conflicts of interest have been declared.

## References

- Andreasen C, Streibig JC (2011) Evaluation of changes in weed flora in arable fields of Nordic countries—based on Danish long-term surveys. *Weed Res* 51:214–226
- Andreasen C, Stryhn H (2008) Increasing weed flora in Danish arable fields and its importance for biodiversity. *Weed Res* 48:1–9
- Andreasen C, Stryhn H (2012) Increasing weed flora in Danish beet, pea and winter barley fields. *Crop Prot* 36:11–17
- Andreasen C, Stryhn H, Streibig JC (1996) Decline of the flora in Danish arable fields. *J Appl Ecol* 33:619–626
- [AOSA] Association of Official Seed Analysts (1990) Rules for testing seeds. *J Seed Technol* 12:1–112
- Benvenuti S, Macchia M (1995) Effect of hypoxia on buried weed seed germination. *Weed Res* 35:343–351
- Benvenuti S, Macchia M (1997) Germination ecophysiology of bur beggarticks (*Bidens tripartita*) as affected by light and oxygen. *Weed Sci* 45:696–700.
- Botta G, Jorajuria D, Rosatto H, Ferrero C (2006) Light tractor traffic frequency on soil compaction in the rolling Pampa region of Argentina. *Soil Tillage Res* 86:9–14
- Boyd N, VanAcker R (2004) Seed germination of common weed species as affected by oxygen concentration, light, and osmotic potential. *Weed Sci* 52:589–596
- Bradford KJ, Come D, Corbineau F (2007) Quantifying the oxygen sensitivity of seed germination using a population-based threshold model. *Seed Sci Res* 17:33–43
- Brady N (1974) The nature and properties of soils. 8th ed. New York: Macmillan. 639 p
- Copeland LO, McDonald M (2012) Principles of Seed Science and Technology. 4th ed. London: Kluwer Academic. 467 p
- Drew MC (1992) Soil aeration and plant-root metabolism. *Soil Sci* 154:259–268
- Fried G, Petit S, Reboud X (2010) A specialist-generalist classification of the arable flora and its response to changes in agricultural practices. *BMC Ecol* 10:1–11
- Heichel G, Day P (1972) Dark germination and seedling growth in monocots and dicots of different photosynthetic efficiencies in 2% and 20.9% O<sub>2</sub>. *Plant Physiol* 49:280–283
- Hodgson A, MacLeod D (1989) Oxygen flux, air-filled porosity, and bulk density as indices of vertisol structure. *Soil Sci Soc Am J* 53:540–543
- Holm RE (1972) Volatile metabolites controlling germination in buried weed seeds. *Plant Physiol* 50:293–297
- Ishii T, Kadoya K (1991) Continuous measurement of oxygen concentration in citrus soil by means of a waterproof zirconia oxygen sensor. *Plant Soil* 131:53–58
- [ISTA] International Seed Testing Association (2011) International Rules for Seed Testing, Germination Tests. Basserdorf, Switzerland: ISTA. 97 p
- Kataoka T, Kim S (1978) Oxygen requirement for seed germination of several weeds. *Weed Res Japan* 23:9–12
- Lipiec J, Arvidsson J, Murer E (2003) Review of modelling crop growth, movement of water and chemicals in relation to topsoil and subsoil compaction. *Soil Tillage Res* 73:15–29
- Marshall E, Brown V, Boatman N, Lutman P, Squire G, Ward L (2003) The role of weeds in supporting biological diversity within crop fields. *Weed Res* 43:77–89
- Martinez L, Zinck J (2004) Temporal variation of soil compaction and deterioration of soil quality in pasture areas of Colombian Amazonia. *Soil Tillage Res* 75:3–18
- Ritz C, Phipper CB, Streibig JC (2013) Analysis of germination data from agricultural experiments. *Eur J Agron* 45:1–6
- Ritz C, Streibig JC (2005) Bioassay analysis using R. *J Stat Softw* 12:1–22
- Rumpho ME, Kennedy RA (1981) Anaerobic metabolism in germinating seeds of *Echinochloa crus-galli* (barnyard grass) metabolite and enzyme studies. *Plant Physiol* 68:165–168
- Sextstone AJ, Revsbech NP, Parkin TB, Tiedje JM (1985) Direct measurement of oxygen profiles and denitrification rates in soil aggregates 1. *Soil Sci Soc Am J* 49:645–651
- Siegel S, Rosen L (1962) Effects of reduced oxygen tension on germination and seedling growth. *Physiol Plantarum* 15:437–444
- Silver WL, Lugo A, Keller M (1999) Soil oxygen availability and biogeochemistry along rainfall and topographic gradients in upland wet tropical forest soils. *Biogeochem* 44:301–328
- Thomson CJ, Greenway H (1991) Metabolic evidence for stelar anoxia in maize roots exposed to low O<sub>2</sub> concentrations. *Plant Physiol* 96:1294–1301
- Topp G, Dow B, Edwards M, Gregorich E, Curnoe W, Cook F (2000) Oxygen measurements in the root zone facilitated by TDR. *Can J Soil Sci* 80:33–41
- Tullberg J, Yule D, McGarry D (2007) Controlled traffic farming from research to adoption in Australia. *Soil Tillage Res* 97:272–281
- Warwick S (1979) The biology of Canadian weeds. 37. *Poa annua* L. *Can J Plant Sci* 59:1053–1066
- Yasin M, Andreasen C (2015) Breaking seed dormancy of *Alliaria petiolata* with phytohormones. *Plant Growth Regul* 77:307–315
- Yasin M, Andreasen C (2016) Effect of reduced oxygen concentration on the germination behavior of vegetable seeds. *Hort Environ Biotech* 57: 453–446
- Yoshioka T, Satoh S, Yamasue Y (1998) Effect of increased concentration of soil CO<sub>2</sub> on intermittent flushes of seed germination in *Echinochloa crus-galli* var. *crus-galli*. *Plant Cell Environ* 21:1301–1306