

Changes in renal hemodynamics of undernourished fetuses appear earlier than IUGR evidences

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Original Article

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Abstract

The present study used a sheep model of intrauterine growth restriction, combining maternal undernutrition and twinning, to determine possible markers of early damage to the fetal kidney. The occurrence of early deviations in fetal hemodynamics which may be indicative of changes in blood perfusion was assessed by Doppler ultrasonography. A total of 24 sheep divided in two groups were fed with the same standard grain-based diet but fulfilling either their daily maintenance requirements for pregnancy (control group; $n = 12$, six singleton and six twin pregnancies) or only the 50% of such quantity (food-restricted group; $n = 12$; four singleton and eight twin pregnancies). All the fetuses were assessed by both B-mode and Doppler ultrasonography at Day 115 of pregnancy. Fetal blood supply was affected by maternal undernutrition, although there were still no evidences of brain-sparing excepting in fetuses at greatest challenge (twins in underfed pregnancies). However, there were early changes in the blood supply to the kidneys of underfed fetuses and underfed twins evidenced decreases in kidney size.

Introduction

Intrauterine growth restriction (IUGR) is the failure of the fetus to reach its genetically established growth rate. IUGR is mainly due to inadequate supply of nutrients and oxygen,¹ either by maternal malnutrition/hypoxia and/or placental insufficiency.^{2,3} Occurrence of IUGR at low-altitude in developed countries (i.e. related to placental insufficiency) is estimated to be around 6%,^{4,5} but increases up to 15% in less favored areas and even to around 17% in case of maternal hypobaric hypoxia at high altitude.^{6–9} IUGR is a concerning health issue because of its implications in perinatal mortality and morbidity and its long-term consequences on health and disease risk of the individuals; mainly on neurological, metabolic, immune, cardiovascular and renal features.¹⁰

Prenatal programming inherent to IUGR has rapidly emerged as a plausible cause for postnatal disorders, in spite of little direct evidence in humans.¹¹ Most of the translational research performed in the area has been traditionally focused on the cardiometabolic consequences of IUGR. However, there is also increasing evidence that prenatal programming following IUGR may impair nephrogenesis, causing a decrease in the glomerular number but a compensatory glomerular enlargement. The consequences are reduced nephron endowment, hypertension and renal diseases in adulthood.^{10,12–14} Hence, there is a strong necessity of preclinical and clinical research on improved detection methods and biomarkers as an optimal antenatal surveillance may be highly beneficial for early detection of IUGR and alleviation of its postnatal effects.

Currently, ultrasonographic monitoring of fetal anatomy and growth is a routinely clinical procedure in which IUGR is suspected in case of abnormal fetal size; after that, evaluation of symmetry, structural and/or chromosomal anomalies, and Doppler hemodynamics are used to differentiate asymmetric IUGR fetuses secondary to maternal and/or placental disorders and oxygen from those symmetric IUGR fetuses secondary to chromosomal and genetic syndromes and intrauterine infections.^{15,16} However, choosing appropriate monitoring and intervention tools and intervals still remains as a main clinical challenge as adequate antenatal diagnosis, treatment and timely delivery may significantly diminish the risks of the disease.¹⁷

Preclinical studies in animal models are an important source of information for a systematic analysis of pregnancy disturbances and IUGR.¹⁸ Models have been traditionally based on laboratory rodents, especially rats and mice.^{19–21} However, rodents have marked differences with humans in developmental patterns, metabolic and endocrine routes and physiology of organs and systems.^{22,23} The use of large animal species may overcome these limitations

and offer numerous profitable characteristics for preclinical research.¹⁸ Specifically in sheep, the effects of exposure to under-nutrition on fetal growth patterns and the occurrence of IUGR were early described²⁴ and therefore the model has been traditionally used for studies on IUGR.^{25,26} Finally, the temperament and size of sheep facilitate fetal screening by non-invasive techniques like B-mode and Doppler ultrasonography.^{27,28}

The present study used a sheep model of IUGR, combining maternal undernutrition and twinning, to determine a possible marker of early damage to the fetal kidney. Our hypothesis was based on the 'brain-sparing' effect occurring during IUGR processes, which consists of a redistribution of the blood circulation to maximize the supply of oxygen and nutrients to the brain.²⁹ In consequence, the growth of the brain is increased to the expenses of the growth of the body and other organs, like the kidney. Hence, we assessed, by Doppler ultrasonography, the occurrence of early deviations in fetal hemodynamics which may be indicative of changes in blood perfusion.

Methods

Animals and experimental procedures

The experiment involved a total of 24 multiparous pregnant ewes (Sarda breed) from the experimental flock of AGRIS Sardegna (Italy). These females became pregnant after natural breeding following cycle synchronization with intravaginal pessaries impregnated with progestagens [20 mg of fluorogestone acetate (FGA), Chronogest[®] CR; MSD-AH, Madison, NJ, USA) for 12 days plus a single i.m. injection of 200 IU of eCG (Folligon[®]; MSD-AH), concurrent with pessary insertion. The day of mating was considered Day 0 for experimental purposes. At Day 24 after mating (around 15% of the total length of ovine pregnancy, estimated in a mean of 150 days), pregnancy diagnosis was performed by transrectal ultrasonography, with a real-time B-mode scanner (Aloka SSD 500; Aloka Co., Tokyo, Japan) fitted with a 7.5 MHz linear-array probe. The ewes were pair matched in two groups (control and food-restricted) according to age, body weight and prolificacy (singleton or twins). All the sheep were fed with the same standard grain-based diet but fulfilling either their daily maintenance requirements for pregnancy (control group; $n = 12$, six singleton pregnancies and six twin pregnancies) or only the 50% of such quantity (food-restricted group; $n = 12$; four singleton pregnancies and eight twin pregnancies). Inappropriate maternal nutrition during early and mid-pregnancy can significantly disrupt placental development, which reaches a maximum growth by approximately Day 75–80 of gestation.³⁰

Ultrasonographic biometry and Doppler evaluation of fetal hemodynamics

All the fetuses were assessed by ultrasonography at Day 115 of pregnancy (around 75% of the total length of ovine pregnancy), just before the overt growth arrest which becomes apparent between 120 and 130 days of pregnancy.²⁵ Ultrasonographic scans were performed with a Voluson-i ultrasound machine (GE, Tiefenbach, Austria) equipped with an automatic 2–5 MHz 4D convex probe. Scans were recorded using the 'cine-loop' option and measurements were obtained in all the fetuses with built-in electronic calipers. As fetus size was too large for viewing the entire body-length at this pregnancy stage, measurements included the thoracic diameter (TD), the biparietal diameter (BPD)

and the length and volume of the left kidney (KL and KV; Fig. 1). The acquisition of kidney volume was performed using the 3D ultrasound mode. Scans of satisfactory quality and without artifacts, after examining the multiplanar display obtained to ensure that the whole kidney had been captured, were used to calculate the volume of the organ by the Virtual Organ Computer Aided analysis (VOCAL).

The blood flow parameters from umbilical cord (UA), middle cerebral (MCA) and renal arteries (RA) were determined in all the fetuses (Fig. 2). Briefly, after identifying the vessels by using color Doppler (UAs were found at the free-floating UA proximal to the placental insertion; MCAs were located after Circle of Willis identification; RAs were assessed proximal to kidney insertion), the sample pulsed Doppler gate was placed over the vessels. Then, the waveforms of three consecutive cardiac cycles in each vessel were recorded, disregarding views with insonation angles between 0° and 50°. Measurements were obtained once the entire examination was recorded and included resistance index (RI), pulsatility index (PI) and systolic-to-diastolic peak velocity ratio (SD-ratio). Assessment of brain-sparing was performed by determining the cerebro-umbilical ratios (i.e. the ratios between MCA and UA values) for RI, PI and SD-ratio.

Statistical analysis

The effects from independent variables (i.e. maternal diet and prolificacy) and their interaction on dependent variables related to offspring phenotype (morphometric and hemodynamic parameters) were assessed using two-way analysis of variance (ANOVA). Maternal diet was categorized in control *v.* restricted diets and prolificacy was categorized in singleton *v.* twin pregnancies. Morphometric and hemodynamic parameters included TD, BPD, length and volume of the left kidney (KL and KV) and blood flow parameters (RI, PI and SD-ratio) from UA, MCA and RA. Possible relationships among morphometric and hemodynamic data of the offspring were determined by Pearson correlation procedures. All data were reported as means \pm S.E.M. and probabilities were considered significant at $P < 0.05$.

Results

There were no significant differences in the mean values of biparietal and thoracic diameters of the fetuses when comparing control *v.* singleton pregnancies and food-restricted *v.* twin pregnancies at Day 115 of gestation (Table 1). The same was found for the volume of the kidneys although the value for kidney length was numerically higher in restricted pregnancies. Within restricted pregnancies, kidney length was significantly higher in singleton than in twin pregnancies (33.1 ± 0.9 *v.* 28.2 ± 0.6 mm, respectively; $P < 0.01$).

Assessment of fetal hemodynamics (Table 2) at the UA showed that maternal food restriction was related to a higher SD-ratio (3.37 ± 0.09 *v.* 2.86 ± 0.20 in the control group; $P < 0.05$), without effects on RI and PI. There were no effects when evaluating these parameters at the MCA or when evaluating the cerebro-umbilical (MCA/UA) ratios.

On the other hand, there were no effects from twinning in any of the absolute hemodynamic parameters at UA and MCA of both control and restricted fetuses. Assessment of the cerebro-umbilical ratios showed no effects of twinning in the control pregnancies, but twinning in restricted pregnancies was associated to significantly higher cerebro-umbilical ratios for SD-ratio and RI ($P < 0.05$ for both).

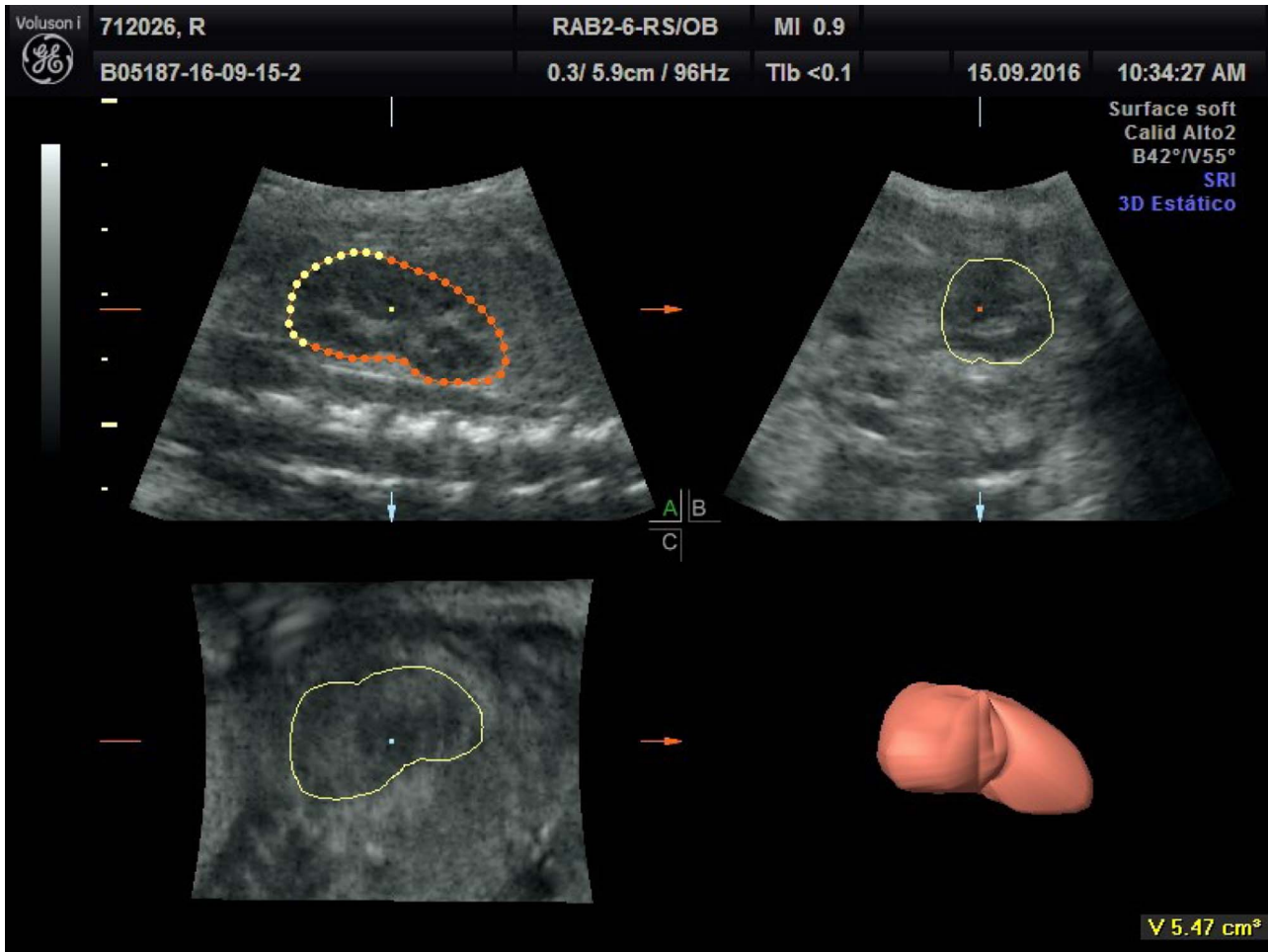


Fig. 1. Integration of multiplanar ultrasound scans and measurements (longitudinal, transversal and coronal planes) for three-dimensional assessment of renal volume in a control sheep fetus.

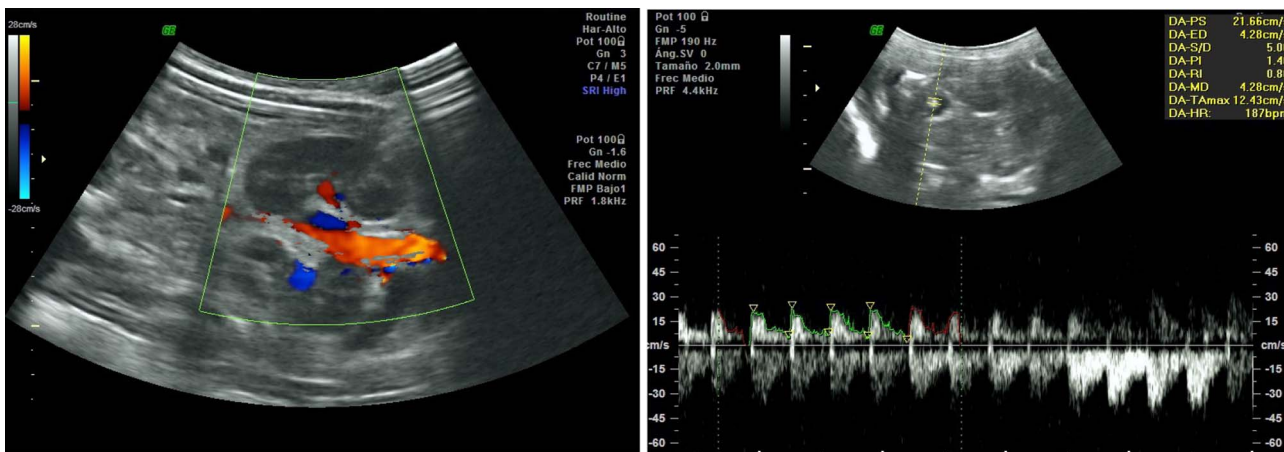


Fig. 2. Identification of the renal artery by using color Doppler (left image) and detection of the waveform in the portion proximal to kidney insertion (right image) in a control sheep fetus.

The assessment of the kidney showed that maternal food restriction, despite the lack of significant effects on size of the organ, induced a significant decrease in all the hemodynamic parameters at the RA ($P < 0.05$ for all) and, consequently, in the corresponding reno-umbilical ratios (SD-ratio: $P < 0.005$; RI: $P < 0.05$; PI: $P < 0.01$). Twinning in control pregnancies was

associated to lower reno-umbilical SD-ratio and IR ($P < 0.05$ for both) but, in contrast, there were no significant differences between singletons and twins in restricted pregnancies.

The assessment of possible relationships between fetal size and hemodynamic features obtained by the Pearson procedure showed a lack of effects at the level of both UA and MCA in both

Table 1. Mean values (\pm s.e.m.) for biparietal (DBP) and thoracic diameters (DTC), and volume and length of the kidney (RVOL and RLENGTH, respectively) in singleton and twin control and food-restricted fetuses at Day 115 of pregnancy

	Control			Food-restricted			<i>P</i>
	Singleton	Twins	<i>P</i>	Singleton	Twins	<i>P</i>	
DBP	4.89 \pm 0.07			4.84 \pm 0.08			0.450
	4.59 \pm 0.04	4.82 \pm 0.09	0.132	4.71 \pm 0.13	4.87 \pm 0.09	0.425	
DTC	6.69 \pm 0.19			6.33 \pm 0.15			0.151
	6.76 \pm 0.24	6.66 \pm 0.28	0.829	5.80 \pm 0.48	6.47 \pm 0.13	0.071	
RVOL	5.98 \pm 0.27			5.59 \pm 0.26			0.317
	6.16 \pm 0.68	5.90 \pm 0.26	0.678	5.89 \pm 0.42	5.55 \pm 0.30	0.682	
RLENGTH	2.73 \pm 0.16			2.91 \pm 0.07			0.141
	2.89 \pm 0.17	2.65 \pm 0.11	0.218	3.31 \pm 0.09	2.83 \pm 0.06	0.005	

Table 2. Mean values (\pm s.e.m.) for fetal hemodynamics assessment [systolic-to-diastolic peak velocity ratio (SD), resistance index (RI) and pulsatility index (PI)] performed at the umbilical cord artery (UA), middle cerebral artery (MCA) and renal artery (RA) of singleton and twin control and food-restricted fetuses at Day 115 of pregnancy

	Control			Food-restricted			<i>P</i>
	Singleton	Twins	<i>P</i>	Singleton	Twins	<i>P</i>	
SD_UA	2.87 \pm 0.09			3.38 \pm 0.20			0.043
	2.86 \pm 0.27	2.87 \pm 0.09	0.935	3.75 \pm 0.59	3.29 \pm 0.22	0.385	
IP_UA	1.05 \pm 0.04			1.18 \pm 0.06			0.115
	1.02 \pm 0.12	1.06 \pm 0.03	0.622	1.28 \pm 0.19	1.16 \pm 0.07	0.503	
IR_UA	0.65 \pm 0.01			0.68 \pm 0.02			0.154
	0.64 \pm 0.03	0.65 \pm 0.01	0.703	0.72 \pm 0.04	0.67 \pm 0.02	0.416	
SD_MCA	2.69 \pm 0.31			2.34 \pm 0.15			0.271
	2.39 \pm 0.03	2.84 \pm 0.47	0.555	2.13 \pm 0.08	2.44 \pm 0.20	0.342	
IP_MCA	1.04 \pm 0.11			0.92 \pm 0.07			0.334
	0.91 \pm 0.06	1.11 \pm 0.15	0.461	0.84 \pm 0.06	0.96 \pm 0.09	0.447	
IR_MCA	0.61 \pm 0.03			0.56 \pm 0.02			0.223
	0.58 \pm 0.01	0.63 \pm 0.05	0.618	0.53 \pm 0.15	0.58 \pm 0.03	0.431	
SD_RA	4.11 \pm 0.50			2.97 \pm 0.14			0.033
	5.56 \pm 1.49	3.51 \pm 0.28	0.061	2.59 \pm 0.23	3.08 \pm 0.17	0.174	
IP_RA	1.30 \pm 0.07			1.09 \pm 0.05			0.017
	1.41 \pm 0.11	1.26 \pm 0.08	0.305	0.96 \pm 0.11	1.13 \pm 0.06	0.212	
IR_RA	0.72 \pm 0.02			0.65 \pm 0.02			0.015
	0.78 \pm 0.05	0.69 \pm 0.02	0.092	0.60 \pm 0.04	0.66 \pm 0.02	0.205	

control and restricted fetuses. However, comparison of fetal size and hemodynamics at the RA showed significant differences between the two nutritional regimes. There were no significant effects of fetal size in the control group, but restricted fetuses with larger BPD had higher reno-umbilical ratios for IP and IR ($r=0.974$, $P<0.05$ and $r=0.993$, $P<0.01$, respectively).

Discussion

The results from the present study indicate that maternal undernutrition is related to a decrease in the materno-fetal blood flow but an increase in in the blood supply to the offspring kidneys. These changes are even earlier to the blood flow

redistribution occurring during the 'brain-sparing' effect and even previous to changes in size of the fetuses, and the proper kidney, which are characteristics of IUGR processes. Fetuses at the greatest challenge (twins in underfed pregnancies) showed, in addition to hemodynamic changes in the cerebro-umbilical ratios indicating early stages of brain-sparing, morphological changes evidenced by a decrease in kidney size, which supports the notion that fetal renal excretory function is affected in risk pregnancies.³¹

Currently, ultrasonographic monitoring of fetal growth and symmetry, followed by Doppler assessment of fetal hemodynamics, is routinely used to determine occurrence and type of IUGR.^{15,16} Doppler sonography is used to detect changes in the uteroplacental and fetal perfusion through assessment of blood vessels of clinical relevance like the uterine, umbilical and middle cerebral arteries and the ductus venosus. The most common assessment is based on umbilical artery (UA) Doppler data which, however, cannot constitute a useful diagnosis because abnormal UA indexes are only found when irreversible adverse perinatal outcomes are established.³² On the contrary, UA values may be within normal range in IUGR fetuses with early cerebral vasodilatation and therefore Doppler measurements of MCA and cerebro-placental ratios (CPR) are most valuable tools.³³ CPR is considered the best predictor as it reflects not only the circulatory insufficiency of UA but also the adaptive changes resulting in modifications of the MCA hemodynamics.³⁴ Hence, even with normal UA Doppler indexes, abnormal CPR values are indicative of fetal distress, acidemia, neurologic disorders and adverse perinatal outcomes.^{33,35,36}

In any case, abnormal Doppler indexes at either brain or umbilical vessels have a poor predictive value,³⁷ as they are only found when fetuses already have damages and the only option is programmed delivery after weighing the risks of prematurity against the risks of adverse intrauterine condition. Hence, there is a strong need for earlier markers of changes in fetal hemodynamic which will likely lead to targeted monitoring intervals and to the implementation of new protocols for early diagnosis and management of IUGR.

In this scenario, the data of the present study, obtained just prior the overt growth arrest occurring in case of IUGR,³¹ have a significant value for both increasing the availability of tools for an adequate clinical follow-up of pregnancy and the knowledge of the pathophysiology of renal damage. In our study, maternal malnutrition was related to a higher UA SD-ratio which, even within a normal range, may indicate an increased risk of compromise to the fetus.³⁸ There were still no evidences of IUGR or brain-sparing since, except in the most compromised fetuses (twins in restricted pregnancies), which showed higher cerebro-umbilical indexes reflecting modifications of blood flow at the brain.³⁴ Conversely, we found clear evidences of changes in the blood supply to the kidneys, supporting that organs with a rich arterial blood supply (i.e. eye, kidney, heart and brain) are the primary target to blood pressure changes.³⁹ We found, unexpectedly, a significant decrease in all the hemodynamic parameters at the RA and, in consequence, in the corresponding reno-umbilical ratios. Overall, these changes are indicating an increased blood flow to the kidneys.

The existence and extent of hemodynamics changes at the RA of IUGR fetuses was the focus of an intense debate from earlier studies, with authors claiming a diminished blood flow⁴⁰⁻⁴² and other authors claiming no changes or even a reduction in downstream resistance.^{43,44} The common current idea is that IUGR affects kidney development and, hence, renal blood flow is decreased due to the brain-sparing effect; however, the differences

among the cited studies may be caused by differences in the timing of pregnancy and the degree of IUGR at the Doppler evaluation.

In fact, the increased blood flow to the fetal kidney found in the present study (performed at the beginning of the third trimester and just prior the overt growth arrest occurring during IUGR) reinforces early studies with Doppler ultrasound, which addressed that the renal flow response to hypoxia depends on the degree of hypoxia and IUGR.⁴⁵ First data were obtained by surgery and the microsphere technique,⁴⁶ and indicated that the blood flow to kidneys remains constant or increases during the transition from high to moderately low levels of arterial oxygen content and then decreases abruptly after more severe hypoxia. In turns, these evidences support earlier hypotheses addressing that the renal blood flow in the fetuses appears to be maintained by autoregulation independently of the blood flow redistribution to the brain in IUGR fetuses.^{47,48} By joining these data and the data in current study, we can conclude that assessment of renal hemodynamics can be used as a diagnostic tool for identifying fetuses at the earlier stages of IUGR.

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Conflicts of Interest. None.

Ethical Standards. The experimental procedures with animals (sheep, *Ovis aries*) were approved by the Animal Care and Use Committee of the University of Sassari, Italy. All the experimental work was carried out at the facilities of the Department of Animal Production of AGRIS (Bonassai, Sardegna, Italy). These facilities meet the requirements of the European Union for Scientific Procedure Establishments. The experimental procedures followed ethical guidelines for care and use of animals for research (European Union Directive 2010/63/UE for animal experiments).

References

1. Brodsky D, Christou H. Current concepts in intrauterine growth restriction. *J Intens Care Med.* 2004; 19, 307-319.
2. Nardoza LM, Araujo Júnior E, Barbosa MM, et al. Fetal growth restriction: current knowledge to the general Obs/Gyn. *Arch Gynecol Obstet.* 2012; 286, 1-13.
3. Cetin I, Mando C, Calabrese S. Maternal predictors of intrauterine growth restriction. *Curr Opin Clin Nutr Metab Care.* 2013; 16, 310-319.
4. Baschat AA. Fetal responses to placental insufficiency: an update. *BJOG.* 2004; 111, 1031-1041.
5. Ghidini A. Idiopathic fetal growth restriction: a pathophysiologic approach. *Obstet Gynecol Surv.* 1996; 51, 376-382.
6. Moore LG, Niermeyer S, Zamudio S. Human adaptation to high altitude: regional and life-cycle perspectives. *Am J Phys Anthropol.* 1998; (Suppl. 27), 25-64.
7. Jensen GM, Moore LG. The effect of high altitude and other risk factors on birthweight: independent or interactive effects? *Am J Public Health.* 1997; 87, 1003-1007.
8. Mortola JP, Frappell PB, Aguero L, Armstrong K. Birth weight and altitude: a study in Peruvian communities. *J Pediatr.* 2000; 136, 324-329.

9. Giussani DA, Niu Y, Herrera EA, *et al*. Heart disease link to fetal hypoxia and oxidative stress. *Adv Exp Med Biol*. 2014; 814, 77–87.
10. Ismail H, Chang YL. Sequelae of fetal growth restriction. *J Med Ultrasound*. 2012; 20, 191–200.
11. Saffery R. Epigenetic change as the major mediator of fetal programming in humans: are we there yet? *Ann Nutr Metab*. 2014; 64, 203–207.
12. Puddu M, Fanos V, Podda F, Zaffanello M. The kidney from prenatal to adult life: perinatal programming and reduction of number of nephrons during development. *Am J Nephrol*. 2009; 30, 162–170.
13. Ritz E, Amann K, Koleganova N, Benz K. Prenatal programming-effects on blood pressure and renal function. *Nat Rev Nephrol*. 2011; 7, 137–144.
14. Dötsch J, Alejandre-Alcazar M, Janoschek R, *et al*. Perinatal programming of renal function. *Curr Opin Pediatr*. 2016; 28, 188–194.
15. Miller J, Turan S, Baschat AA. Fetal growth restriction. *Semin Perinatol*. 2008; 32, 274–280.
16. Rizzo G, Arduini D. Intrauterine growth restriction: diagnosis and management. A review. *Minerva Ginecol*. 2009; 61, 411–420.
17. Kouskouti C, Regner K, Knabl J, Kainer F. Cardiotocography and the evolution into computerised cardiotocography in the management of intrauterine growth restriction. *Arch Gynecol Obstet*. 2017; 295, 811–816.
18. Gonzalez-Bulnes A, Astiz S, Parraguez VH, Garcia-Contreras C, Vazquez-Gomez M. Empowering translational research in fetal growth restriction: sheep and swine animal models. *Curr Pharm Biotechnol*. 2016; 17, 848–855.
19. Armitage JA, Taylor PD, Poston L. Experimental models of developmental programming: consequences of exposure to an energy rich diet during development. *J Physiol*. 2005; 565, 3–8.
20. Schroeder M, Shbiro L, Zagoory-Sharon O, Moran TH, Weller A. Toward an animal model of childhood-onset obesity: follow-up of OLETF rats during pregnancy and lactation. *Am J Physiol Regul Integr Comp Physiol*. 2009; 296, R224–R232.
21. Rosenfeld CS. Animal models to study environmental epigenetics. *Biol Reprod*. 2010; 82, 473–488.
22. Neitzke U, Harder T, Schellong K, *et al*. Intrauterine growth restriction in a rodent model and developmental programming of the metabolic syndrome: a critical appraisal of the experimental evidence. *Placenta*. 2008; 29, 246–254.
23. Neitzke U, Harder T, Plagemann A. Intrauterine growth restriction and developmental programming of the metabolic syndrome: a critical appraisal. *Microcirculation*. 2011; 18, 304–311.
24. Charlton V, Johengen M. Effects of intrauterine nutritional supplementation on fetal growth retardation. *Biol Neonate*. 1985; 48, 125–142.
25. Symonds ME, Budge H, Stephenson T, McMillen IC. Fetal endocrinology and development-manipulation and adaptation to long-term nutritional and environmental challenges. *Reproduction*. 2001; 121, 853–862.
26. Wallace JM, Aitken RP, Milne JS, Hay WW. Nutritionally mediated placental growth restriction in the growing adolescent: consequences for the fetus. *Biol Reprod*. 2004; 71, 1055–1062.
27. Morel O, Pachy F, Chavatte-Palmer P, *et al*. Correlation between utero-placental three-dimensional power Doppler indices and the uterine real blood flow: evaluation in a pregnant sheep experimental model. *Ultrasound Obstet Gynecol*. 2010; 36, 635–640.
28. Vonnahme KA, Lemley CO. Programming the offspring through altered uteroplacental hemodynamics: how maternal environment impacts uterine and umbilical blood flow in cattle, sheep and pigs. *Reprod Fertil Develop*. 2012; 24, 97–104.
29. Scherjon SA, Smolders-DeHaas H, Kok JH, Zondervan HA. The “brain-sparing” effect: antenatal cerebral Doppler findings in relation to neurologic outcome in very preterm infants. *Am J Obstet Gynecol*. 1993; 169, 169–175.
30. Osgerby JC, Wathes DC, Howard D, Gadd TS. The effect of maternal undernutrition on the placental growth trajectory and the uterine insulin-like growth factor axis in the pregnant ewe. *J Endocrinol*. 2004; 182, 89–103.
31. Zywicki M, Blohowiak SE, Magness RR, Segar JL, Kling PJ. Increasing fetal ovine number per gestation alters fetal plasma clinical chemistry values. *Physiol Rep*. 2016; 4, e12905.
32. Unterscheider J, Daly S, Geary MP, *et al*. Optimizing the definition of intrauterine growth restriction: the multicenter prospective PORTO study. *Am J Obstet Gynecol*. 2013; 208, 290.e1–6.
33. Mureşan D, Rotar IC, Stamatian F. The usefulness of fetal Doppler evaluation in early versus late onset intrauterine growth restriction. Review of the literature. *Med Ultrason*. 2016; 18, 103–109.
34. Shahinaj R, Manoku N, Kroj E, Tasha I. The value of the middle cerebral to umbilical artery Doppler ratio in the prediction of neonatal outcome in patient with preeclampsia and gestational hypertension. *J Prenat Med*. 2010; 4, 17–21.
35. DeVore GR. The importance of the cerebroplacental ratio in the evaluation of fetal well-being in SGA and AGA fetuses. *Am J Obstet Gynecol*. 2015; 213, 5–15.
36. Nassr AA, Abdelmagied AM, Shazly SA. Fetal cerebro-placental ratio and adverse perinatal outcome: systematic review and meta-analysis of the association and diagnostic performance. *J Perinat Med*. 2016; 44, 249–256.
37. Albu AR, Anca AF, Horhoianu VV, Horhoianu IA. Predictive factors for intrauterine growth restriction. *J Med Life*. 2014; 7, 165–171.
38. Devoe LD, Gardner P, Dear C, Faircloth D. The significance of increasing umbilical artery systolic-diastolic ratios in third-trimester pregnancy. *Obstet Gynecol*. 1992; 80, 684–687.
39. Mensah GA, Croft JB, Giles WH. The heart, kidney, and brain as target organs in hypertension. *Cardiol Clin*. 2002; 20, 225–247.
40. Vyas S, Nicolaidis KH, Campbell S. Renal artery flow-velocity waveforms in normal and hypoxemic fetuses. *Am J Obstet Gynecol*. 1989; 161, 168–172.
41. Arduini D, Rizzo G. Fetal renal artery velocity waveforms and amniotic fluid volume in growth-retarded and post-term fetuses. *Obstet Gynecol*. 1991; 77, 370–373.
42. Yoshimura S, Masuzaki H, Gotoh H, Ishimaru T. Fetal redistribution of blood flow and amniotic fluid volume in growth-retarded fetuses. *Early Hum Dev*. 1997; 47, 297–304.
43. Surányi A, Streitman K, Pál A, *et al*. Fetal renal artery flow and renal echogenicity in the chronically hypoxic state. *Pediatr Nephrol*. 2000; 14, 393–399.
44. Stigter RH, Mulder EJJ, Bruinse HW, Visser GHA. Doppler studies on the fetal renal artery in the severely growth-restricted fetus. *Ultrasound Obstet Gynecol*. 2001; 18, 141–145.
45. Arbeille P. Fetal arterial Doppler-IUGR and hypoxia. *Eur J Obstet Gynecol Reprod Biol*. 1997; 75, 51–53.
46. Peeters LLH, Sheldon RE, Douglas-Jones M, Makowski EL, Meschia G. Blood flow to fetal organs as a function of arterial oxygen content. *Am J Obstet Gynecol*. 1979; 135, 637–646.
47. Shepherd T, Abboud M. The renal circulation. In *Handbook of Physiology* (eds. Knox FG, Spielman WS), Section 2, vol. 3, 1983; pp. 205–209. American Physiological Society: Washington, DC.
48. Tanabe R. Doppler ultrasonographic assessment of fetal renal artery blood flow velocity waveforms in intrauterine growth retarded fetuses. *Kurume Med J*. 1992; 39, 203–208.