


Genetic diversity and structure analysis of *Vigna unguiculata* L. (Walp.) landraces from southeastern Mexico using ISSR markers

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Abstract

Cowpea (*Vigna unguiculata* L. Walp.) is an important grain legume in tropical and subtropical regions. It requires low resource inputs and has a high nutritional value. Therefore, cowpea can play an important role in the development of agriculture. In southern Mexico, Mayan farmers have conserved and developed cowpea landraces for centuries. Nevertheless, information on their genetic diversity, conservation status and potential use is minimal. To generate information toward sustainable use, management and conservation of this species, we evaluated the genetic diversity and structure of 20 cowpea landraces from southeast Mexico using 10 inter-simple sequence repeat (ISSR) molecular markers. These ISSR markers generated 68 loci with a 67.7% polymorphism rate and average polymorphic information content of 0.36. The results of Bayesian assignment and the UPGMA analysis suggest the formation of two main groups defined by their genetic origin in southeast Mexico. High levels of genetic structure were found with a moderate level of genetic diversity distributed mainly between landraces. Low levels of intra-landrace variability were observed. Two landraces (P5 and P12) from Calakmul resulted in the high levels of genetic diversity. The selected markers were efficient at assessing genetic variability among Mexican cowpea landraces, providing valuable information that can be used in local conservation and participatory breeding programmes.

Keywords: Cowpea, genetic resources, landrace conservation, molecular markers, polymorphism

Introduction

Cowpea (*Vigna unguiculata* L. Walp.) is an important food legume worldwide (Tan *et al.*, 2012). It is cultivated in tropical regions, usually inter-cropped with cereals, but also in rotation as a sole crop (Gajera *et al.*, 2014). Cowpea plants are well adapted to high temperatures and drought and it is known for its high nutritional value. Therefore, it is useful for agricultural development globally (Carvalho *et al.*,

2017). This species, although of African origin, has been cultivated for centuries in Mexico (Hernández and Delgado, 1992). It is known as ‘xpelón’ in Maya-Yucatec and is cultivated in small areas for self-consumption within a traditional agroecosystem known as *milpa* (Castillo-Caamal, 2006; Morales-Morales *et al.*, 2019).

The phenotypic variability observed in *V. unguiculata* in the Yucatan Peninsula (Morales-Morales *et al.*, 2019) provides an opportunity to explore the genetic diversity in this region. A landrace is defined by Camacho *et al.* (2005) as a dynamic population of cultivated plants that has a historical origin, distinct identity and lacks formal

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crop improvement, as well as being genetically diverse, locally adapted and associated with traditional farming systems. Local farmers recognize cowpea landraces by their differences in lifespan, growth habit and pod and grain colours (Castillo-Caamal, 2006; Morales-Morales *et al.*, 2019). Nevertheless, farmers use the same names across similar landraces. This suggests either repeating genotypes or misclassification (Araújo *et al.*, 2019). Few efforts have been made to characterize the genetic diversity and conservation status of cowpea germplasm in Mexico (Lagunes-Espinoza *et al.*, 2007; Morales-Morales *et al.*, 2019). Conservation of cowpea landraces requires an understanding of the genetic variation of these local varieties. Careful characterization of landraces is a first step to guide efforts to conserve biodiversity and provide farmers with quality seeds of improved cultivars (Ghalmi *et al.*, 2010).

Traditionally, genetic diversity among cowpea genotypes is estimated by measuring phenotypic variation. Nevertheless, the expression of qualitative or quantitative traits is subject to environmental influences and limits knowledge of the germplasm structure (Wamalwa *et al.*, 2016). Alternatively, molecular markers such as randomly amplified polymorphic DNA (RAPD) (Gajera *et al.*, 2014), simple sequence repeat (SSR) (Ali *et al.*, 2015; Wamalwa *et al.*, 2016; Chen *et al.*, 2017) and inter-simple sequence repeat (ISSR) (Anatala *et al.*, 2014; Igwe *et al.*, 2017; Kumar *et al.*, 2017) have been applied to characterize genetic diversity, genetic relationships and germplasm management and conservation in *V. unguiculata*. Among the molecular techniques, ISSR markers are one of the simplest and most widely used techniques (Ghalmi *et al.*, 2010). Furthermore, ISSR markers do not require prior information on the genome of the species, making them useful for studies of genetic diversity, phylogeny, genomic mapping and evolutionary biology (Tan *et al.*, 2012). The lack of collections in many areas of Mexico has prevented a precise assessment of genetic diversity and structure of cowpea landraces. Therefore, our work aimed to assess the genetic diversity and structure of 20 cowpea landraces from south-east Mexico with ISSR molecular markers.

Material and methods

Sample collection and DNA extraction

In total, 20 cowpea landraces from Yucatan Peninsula in the southeast of Mexico were sampled. These landraces were collected from farmers in areas where the agroecosystem *milpa* has been maintained as an important economic activity (Martínez-Castillo *et al.*, 2004). The list of cowpea landraces used and their origins are described in Table 1.

To obtain fresh leaf samples to extract high-quality DNA, seeds were sown in the greenhouse at the

Campus de Ciencias Biológicas y Agropecuarias at the Universidad Autónoma de Yucatán. Five individual 15–20 d old plantlets per landrace were randomly chosen and approximately 100 mg of fresh leaves collected for DNA purification using a modified CTAB protocol described by Falcón and Valera (2007). We analysed only five plantlets per landrace because in self-pollinated crops such as *V. unguiculata*, the genetic variation is mainly found between populations and not within populations (Martínez-Castillo *et al.*, 2014; Menssen *et al.*, 2017). The concentration of extracted DNA in the individual samples was measured using a NanoDrop Lite Spectrophotometer (Thermo Scientific) and the quality of the genomic DNA samples was determined by electrophoresis in a 0.8% agarose gel. All samples were then diluted with DNase free water to 25 ng/μl and frozen until used for ISSR analysis.

PCR amplification and ISSR analysis

Twelve ISSR primers, previously used in cowpea genetic diversity studies (Ghalmi *et al.*, 2010; Gajera *et al.*, 2014; Igwe *et al.*, 2017; Araújo *et al.*, 2019), were tested for their ability to detect polymorphisms in Mexican cowpea landraces. After a preliminary test, 10 primers that yielded good amplification and high levels of polymorphism were selected for ISSR analysis (Table 2).

Polymerase chain reaction (PCR) was performed in a final volume of 15 μl containing 30 ng template DNA, 1X PCR buffer, 0.2 mM MgCl₂ (50 mM), 0.2 mM dNTPs (2.5 mM), 0.8 μM primer (10 μM) (T4Oligo) and 1 U of Taq DNA polymerase (5 U/μl) (Invitrogen Standard Taq DNA polymerase recombinant kit) suspended in ultrapure water. DNA amplification was performed in a Select Cycler II thermocycler (Select BioProducts, Edison N.J., EE.UU.) programmed for 4 min denaturation at 94°C, followed by 35 cycles of 1 min at 94°C, 1 min annealing with temperatures ranging from 46 to 53°C depending on the primer used, 1 min at 72°C and final extension for 10 min at 72°C. Amplification products were separated by electrophoresis in 1.5% agarose gel with 1X Sodium Boric acid buffer (Brody and Kern, 2004), visualized and photographed over a UV transilluminator after staining with SYBR Safe (Thermo Fisher Scientific). Amplicon lengths were estimated with a 100 bp DNA Ladder (New England BioLabs, Ipswich, MA, USA).

Analysis of genetic diversity and population structure

Clear and distinct bands amplified by ISSR primers were scored for presence (1) and for absence (0) of individual allele among populations to obtain the data matrix.

Table 1. Code, origin and local name of 20 cowpea landraces from southeast Mexico

Code	State	Municipality	Village	Local name
P1	Yucatán	Tahdziú	Tahdziú	xnuc xpelon
P2	Yucatán	Santa Elena	Santa Elena	xpelon de mata
P3	Campeche	Calkiní	Becal	xpelon
P4	Campeche	Chamotón	Felipe Carrillo Puerto	xpelon de guía
P5	Campeche	Calakmul	Cristóbal Colón	pelon
P6	Campeche	Chamotón	Felipe Carrillo Puerto	vaina morada
P7	Quintana Roo	Bacalar	Bacalar	xpelon
P8	Quintana Roo	Chetumal	Chetumal	xpelon grande
P9	Yucatán	Dzizantún	Dzizantún	espeón
P10	Yucatán	Oxuczacab	Yaxhom	yax xpelon
P11	Quintana Roo	José María Morelos	Dziuché	dominga
P12	Campeche	Calakmul	Concepción	pelon
P13	Quintana Roo	Felipe Carrillo Puerto	Tihosuco	xpelon grande
P14	Quintana Roo	Felipe Carrillo Puerto	Tepich	dominga
P15	Yucatán	Tixmehuac	Tixmehuac	xmejen xpelon
P16	Quintana Roo	José María Morelos	Sabán	xpelon grande
P17	Campeche	Escárcega	Ejido Pimental	pelon
P18	Yucatán	Tahdziú	Tahdziú	xmejen xpelon
P19	Campeche	Escárcega	Ejido Don Samuel	pelon
P20	Quintana Roo	Felipe Carrillo Puerto	Chunhuhub	xpelon

Population structure was assessed with STRUCTURE 2.3.4 software based on a no admixture model (Evanno *et al.*, 2005). Models were tested for K -values ranging from 1 to 6, with 10 independent runs each and 100,000 Markov chain Monte Carlo interactions. The optimal K value was chosen according to the ΔK statistic proposed

by Evanno *et al.* (2005) using STRUCTURE HARVESTER software (Earl Dent and vonHoldt, 2012). Once the number of genetic clusters was established, each individual was assigned to a cluster and the overall membership of each sampled individual in the cluster was estimated. The genetic diversity and population structure of the cowpea

Table 2. Number of alleles, allele frequency, gene diversity, percentage of polymorphism and polymorphism information content of 10 ISSR's primers in cowpea landraces from southeast Mexico

ISSR Primer	Annealing temperature (°C)	No of alleles	Mayor allele frequency	Gene diversity	% P	PIC	Range of fragment size (pb)
811 (GA) ₈ C	46	6	0.53	0.49	33	0.37	[350–1500]
812 (GA) ₈ A	50	7	0.65	0.43	86	0.33	[300–1000]
816 (CA) ₈ T	51	6	0.62	0.46	100	0.35	[270–1000]
825 (AC) ₇ T	51	7	0.55	0.49	71	0.37	[300–900]
826 (AC) ₈ C	53	8	0.66	0.39	100	0.31	[500–1300]
834 (AG) ₈ YT	49	8	0.56	0.47	25	0.36	[300–1500]
835 (AG) ₈ YC	47	6	0.65	0.41	67	0.32	[200–1000]
842 (GA) ₈ YG	49.5	7	0.52	0.50	43	0.37	[250–1200]
856 (AC) ₈ YA	51	7	0.60	0.45	57	0.35	[150–1200]
857 (AC) ₈ YC	50	6	0.57	0.47	83	0.36	[200–1000]
Mean	–	6.8	0.59	0.46	67.7	0.35	[282–1160]

% P, percentage of polymorphism; PIC, polymorphic information content.

landraces were further investigated by Wright's differentiation index (F_{ST}) using the Bayesian approach proposed by Zhivotovsky (1999) with AFLP-Surv 1.0 software (Vekemans, 2002). Because cowpea is treated as a highly self-pollinated plant (Menssen *et al.*, 2017), the diversity statistics were calculated with the assumption that the populations are mostly selfing ($FIS = 0.95$). An analysis of molecular variance (AMOVA) was also performed with GenAlEx 6.5 software (Peakall and Smouse, 2012).

To assess the genetic relationships among genotypes, a dendrogram was constructed using Nei's genetic distance modified by Lynch and Milligan (1994) and UPGMA. The dendrogram was assessed by bootstrap analysis with 1000 replicates using CONSENSUS software from the PHYLIP package (Felsenstein, 2005). The dendrogram was displayed and edited with MEGA version X software (Kumar *et al.*, 2018).

For each primer, the number of alleles, allele frequency, gene diversity, percentage of polymorphism (% P) and polymorphic information content (PIC), were calculated using PowerMarker version 3.25 software (Liu and Muse, 2005). Genetic diversity indices were calculated at two levels – to the entire sample from Yucatan Peninsula and at individual landrace level. Given the dominant nature of ISSR markers, genetic diversity was also calculated by the Shannon-Weaver index (D). In addition, % P, the number of effective alleles (N_e) and expected heterozygosity (H_e), were calculated with PopGen version 1.31 software (Yeh *et al.*, 1999).

Results

Molecular diversity of V. unguiculata with ISSR

In this study, 10 ISSR markers were used to assess the genetic diversity of 20 cowpea landraces in Mexico. These ISSR markers generated a total of 68 loci with 67.7% P. The primers 816 and 826 resulted with 100% P. The number of alleles per locus ranged from 6 to 8 with a mean of 6.8 alleles per locus that ranged in size from 282 to 1160 base pairs (Table 2). The major allele frequency ranged from 0.53 to 0.66 with a mean value of 0.59 while genetic diversity ranged from 0.39 in the ISSR 826 to 0.50 in the ISSR 842 with a mean value of 0.46. In addition, the PIC values ranged from 0.31 in the ISSR 826 to 0.37 in the primers ISSR 811, 825 and 842 with a mean of 0.35 (Table 2).

Genetic structure of 20 cowpea landraces from Yucatan Peninsula, Mexico

The analysis of population structure suggested two genetically distinct groups ($K = 2$) for the entire sample from the Yucatan Peninsula (online Supplementary Fig. S1). The

assignment of landraces in the two groups is shown in the bar plot of Fig. 1 with different colours. The red group composed of 12 landraces (P1, P2, P3, P4, P5, P6, P7, P8, P9, P10, P11 and P12). The green group composed of eight landraces (P13, P14, P15, P16, P17, P18, P19 and P20). Focusing on percentages of membership shared within each of the observed groups, the landrace P5 clearly shared more admixture out of the 20 cowpea landraces, with 61 and 39% membership with groups red and green, respectively (Fig. 1). The geographical distribution of the 20 cowpea landraces in Yucatan Peninsula is shown in the Fig. 3.

The genetic differentiation of cowpea landraces from southeast Mexico is shown in online Supplementary Table S1. The total diversity (H_t) was 0.24 and diversity within populations (H_s) was 0.14. In addition, the genetic differentiation was high ($F_{st} = 0.42$) and gene flow low ($N_m = 0.19$), indicating that 42% of total genetic diversity was explained by differences among landraces (online Supplementary Table S1). These results agree with the AMOVA which indicated that 66% of total molecular variance was distributed among landraces, while 34% of genetic variance was within landraces (online Supplementary Table S2).

Genetic relationships among 20 cowpea landraces from Yucatan Peninsula, Mexico

The analysis of genetic relationships generated with the UPGMA method showed the formation of two main different clusters (A and B) with 10 landraces each, congruent in part, with the STRUCTURE analysis (Fig. 2). Each branch was colour-coded according to the groups identified by STRUCTURE. Therefore, cluster A contained all landraces from the red group in the STRUCTURE analysis. The cluster B contained all landraces from the green group in the STRUCTURE analysis plus, the landraces P5 and P8 from the red group.

Genetic diversity of 20 cowpea landraces from Yucatan Peninsula, Mexico

The genetic diversity was evaluated at two levels, across the entire sample and at landrace level. Both genetic diversity indices evaluated (H_e and I) showed a moderate level of genetic diversity for the entire sample of cowpea landraces ($H_e = 0.19$ and $I = 0.30$ respectively). Also, the % P and the N_e were 67.7% and 1.32, respectively (Table 3).

At landrace level, values of H_e ranged from 0.01 to 0.10 and the I ranged from 0.01 to 0.15 (Table 3). Overall, the landraces P12 and P5 both from the state of Campeche, municipality of Calakmul resulted with the highest genetic diversity parameters % P, N_e , H_e and I . Contrarily, the landraces P17 from the state of Campeche, municipality of

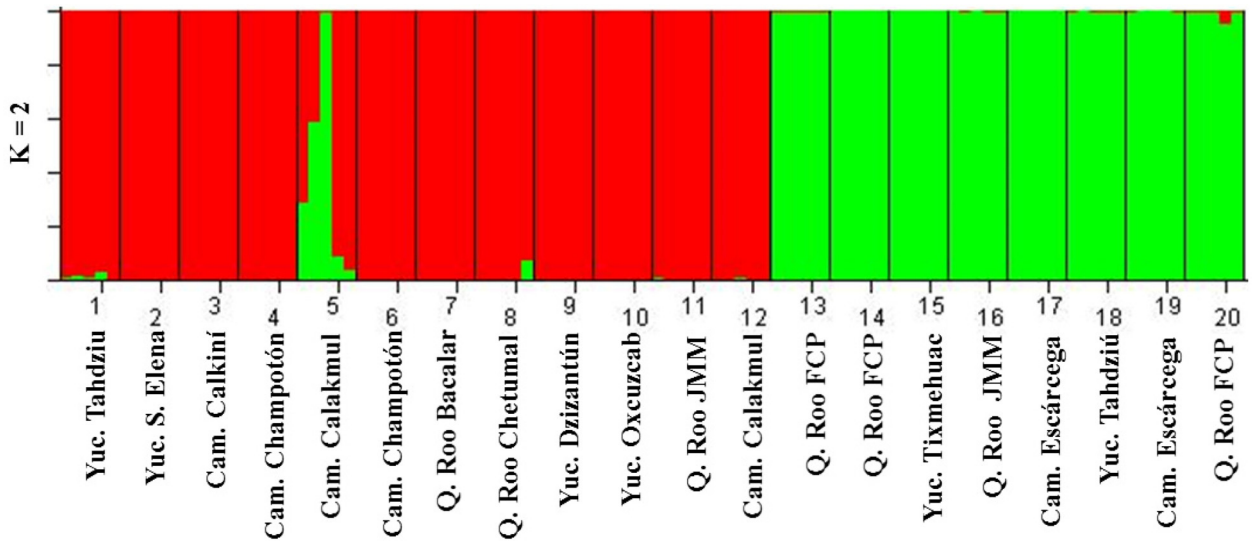


Fig. 1. Inferred ancestry of 20 cowpea landraces for $K = 2$ groups coloured red and green. Each individual is represented by a thin vertical line, divided into coloured segments that represent the individual estimated membership to each group.

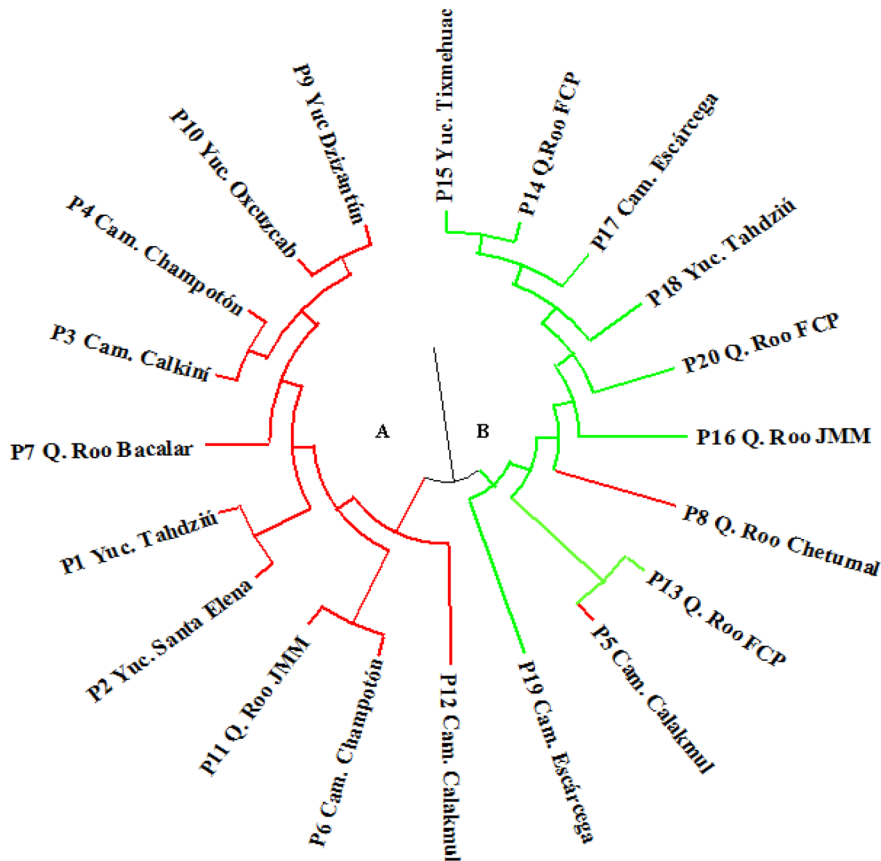


Fig. 2. UPGMA dendrogram of genetic relationship of 20 cowpea landraces from Yucatan Peninsula, México. Each branch was colour-coded according to membership into the $K = 2$ groups identified by STRUCTURE (same colours as in Fig. 1).

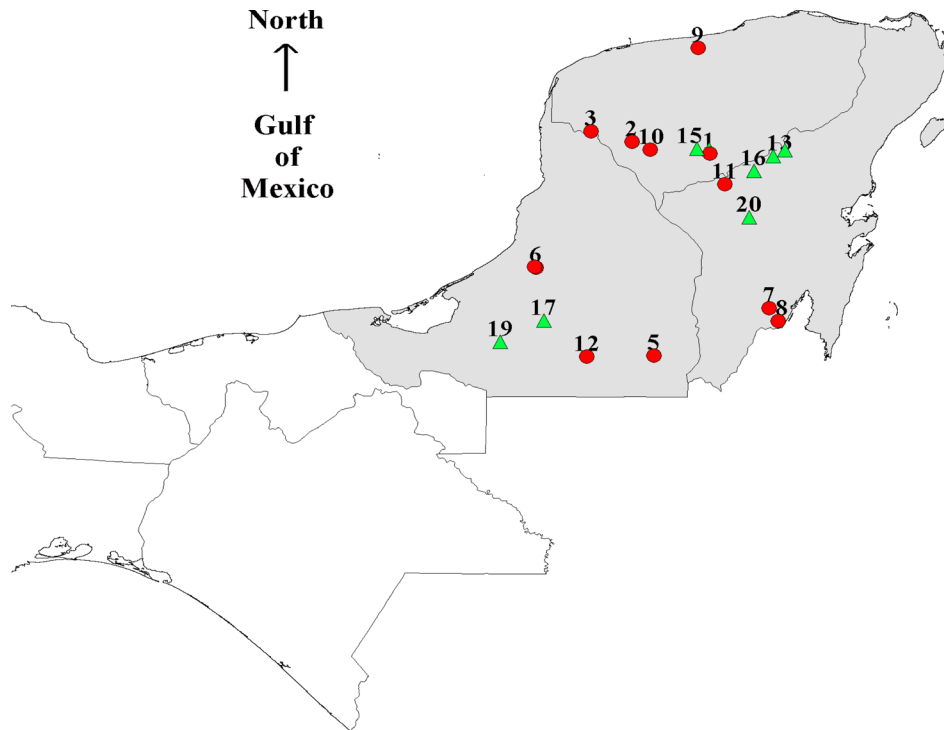


Fig. 3. Geographical distribution and population structure of 20 cowpea landraces from Yucatan Peninsula, México. Red circles and green triangles correspond to the red and green groups identified by STRUCTURE.

Escárcega resulted with the lowest diversity indices with 1.5% P, 1.01 Ne, 0.01 He and 0.01 of I.

Discussion

Analysis of molecular diversity of V. unguiculata with ISSR

Understanding genetic variation has important implications both for the conservation of existing genetic resources and breeding programs for new varieties. In the present study, the genetic diversity and structure analysis of 20 cowpea landraces from southeast Mexico were determined using ISSR molecular markers.

The ISSR markers used in this study successfully assessed the genetic diversity of cowpea landraces, showing high values of % P and PIC. The high levels of polymorphism detected by ISSR primers reflect their coverage of the genome, since microsatellites, besides being abundant, are well distributed (Araújo *et al.*, 2019). It has been shown that high polymorphism identifiable by molecular markers in cowpea hinged on the presence of repeated sequences of AC, CA, AG and GA, as adopted in this study (Igwe *et al.*, 2017). The mean PIC value observed here (0.35) was similar with previously reported values in cowpea (Chen *et al.*, 2017; Araújo *et al.*, 2019). PIC value, as measured by Botstein *et al.* (1980), showed that a mean PIC value ≥ 0.5

is highly informative, 0.25–0.50 reasonably informative and <0.25 is slightly informative, where loci with many alleles and a PIC value near 1 are most desirable (Ali *et al.*, 2015).

When assessing genetic diversity in Algerian cowpea landraces using ISSR markers, Ghalmi *et al.* (2010) found 104 alleles, a mean of 8.6 alleles per marker and 65% P. Recently, Araújo *et al.* (2019) studying 52 landrace populations from Brazil found 80 alleles, a mean of 5.7 alleles per marker with a polymorphism of 76%. These results were similar to what was observed here and higher than that was observed by Anatala *et al.* (2014) studying 10 cowpea genotypes with ISSR markers in India. In that study, the authors found a total of 103 alleles, a mean of 5.7 alleles per marker and 47% P (Anatala *et al.*, 2014). These differences in the % P can be explained by the origin and size of populations used and the ISSR primers selected for the analysis.

Genetic structure and diversity of cowpea landraces from Yucatan Peninsula, Mexico

In contrast to many other important world crops, relatively little is understood about the domestication history, worldwide dispersal and distribution of genetic variation of cowpea. Two major gene pools (West African versus East African gene pools) are reported in Africa where domestication has occurred (Huynh *et al.*, 2013). In addition to

Table 3. Genetic diversity parameters generated from 20 cowpea landraces from southeast Mexico with ISSR markers

Sample code	Sample size	% P	Ne (SD)	He (SD)	<i>I</i> (SD)
Yucatan Peninsula	100	67.7	1.32 (0.34)	0.19 (0.19)	0.30 (0.27)
P1	5	5.9	1.04 (0.17)	0.02 (0.10)	0.03 (0.14)
P2	5	10.3	1.08 (0.23)	0.04 (0.13)	0.06 (0.19)
P3	5	11.8	1.09 (0.26)	0.05 (0.14)	0.07 (0.20)
P4	5	19.1	1.13 (0.29)	0.08 (0.16)	0.11 (0.23)
P5	5	23.5	1.18 (0.34)	0.10 (0.18)	0.14 (0.26)
P6	5	14.7	1.11 (0.28)	0.06 (0.15)	0.09 (0.22)
P7	5	19.1	1.16 (0.35)	0.09 (0.18)	0.12 (0.26)
P8	5	14.7	1.11 (0.28)	0.06 (0.15)	0.09 (0.22)
P9	5	16.2	1.11 (0.27)	0.06 (0.15)	0.09 (0.22)
P10	5	16.2	1.13 (0.31)	0.07 (0.16)	0.10 (0.23)
P11	5	16.2	1.10 (0.25)	0.06 (0.14)	0.09 (0.21)
P12	5	25.0	1.18 (0.34)	0.10 (0.19)	0.15 (0.27)
P13	5	5.9	1.04 (0.17)	0.02 (0.10)	0.03 (0.14)
P14	5	10.3	1.08 (0.23)	0.04 (0.13)	0.06 (0.19)
P15	5	14.7	1.11 (0.28)	0.06 (0.15)	0.09 (0.22)
P16	5	4.4	1.03 (0.14)	0.02 (0.08)	0.02 (0.12)
P17	5	1.5	1.01 (0.11)	0.01 (0.06)	0.01 (0.09)
P18	5	14.7	1.11 (0.25)	0.06 (0.14)	0.08 (0.21)
P19	5	13.2	1.08 (0.24)	0.05 (0.14)	0.08 (0.20)
P20	5	11.8	1.09 (0.26)	0.05 (0.14)	0.07 (0.20)

% P, percentage of polymorphism; Ne, number of effective alleles; He, expected heterozygosity; *I*, Shannon's Information index; SD, standard deviation.

those gene pools, one additional gene pool was identified by Carvalho *et al.* (2017), corresponding to the regions of North Africa and South Europe.

The results of Bayesian assignment and the UPGMA cluster analysis are congruent in part, with the formation of two main groups. These genetically distinct groups observed in the Yucatan Peninsula could correspond to West and East African gene pools, introduced in this region during the sixteenth and seventeenth centuries. Most cowpeas in North America did not move directly from West Africa, in contrast to the popular view that cowpea was introduced directly from this region during the slave-trading period (Huynh *et al.*, 2013). In Mexico, *V. unguiculata* likely was brought independently from three different continents in different periods. It was probably first introduced in the Yucatan Peninsula by the Spaniards simultaneous with the Iberian cowpea in the Caribbean in the sixteenth century (Carvalho *et al.*, 2017). At the same time, cowpea was brought from eastern Africa to the New World, mostly on slave ships (Herniter, 2019).

In Yucatan Peninsula, cowpea is known by a Maya word, xpelón, and used in the traditional Maya cuisine, mostly eaten in pibes, tamales and broths (Cázares and Duch, 2004) especially in the traditional festivity of Hanan

pixan, the day of the deaths, celebrated in November, which could suggest a very early consumption. However, when and how cowpea arrived in the area is unclear. It remains an open question that requires further studies. Among the exceptions in the UPGMA analysis, the landraces P5 and P8 that belong to the red group in the STRUCTURE analysis were placed in the green cluster in the UPGMA dendrogram. In the first case, the landrace P5 showed a high degree of admixture and shared a great percentage of membership with the green group (Fig. 1). The complexity of the seed exchange network among rural farming communities facilitates access to seeds from other families and communities. In these cases, seed mixture likely occurs, especially when the seed coat exhibits the same colour, increasing the gene flow among landraces (Martínez-Castillo *et al.*, 2008). Alternatively, this accession could be an old variety developed by breeders in Mexico and adopted by farmers in the past. In the second case, despite the low levels of gene flow observed in P8 with the populations of the green group in the STRUCTURE analysis, it seems to be enough for the inclusion of P8 within the green cluster in the UPGMA or could be due to differences between both methods. Also, the landrace P8 is known by local farmers by 'xpelón grande' similar to the landraces

P13 and P16 both placed in the green cluster, in reference to a long vegetative cycle.

The red group formed in the STRUCTURE analysis correspond with the geographical regions of northeastern and southwestern of Campeche, eastern Yucatan and southwestern of Quintana Roo. Likewise, the green group was found across the geographical regions of southern Yucatan and central-eastern of Quintana Roo (Fig. 3). The geographical distribution of both genetic groups in the Yucatan Peninsula likely results from a complex network of seed exchange between communities in this region and not agroecological patterns, environment or political divisions. Also, the geographical regions of southern Yucatan and central-eastern of Quintana Roo included cowpea landraces from both genetic groups. Therefore, these regions are priority areas to develop *in-situ* conservation programs of cowpea landraces. Nevertheless, to develop a complete understanding of the spatial distribution of cowpea genetic structure in the Yucatan Peninsula, the analyses of a larger number of accessions are necessary.

Further, the level of genetic differentiation was relatively high ($F_{st}=0.42$), in agreement with the results of the AMOVA, which indicated that 66% of the total variance was distributed among landraces. These results indicate a high genetic divergence between the cowpea landraces of Yucatan Peninsula and can be explained by the low level of gene flow ($N_m = 0.19$) and levels of endogamy presented in the studied landraces. These results also indicate that local farmers efficiently conserved cowpea landraces with a high level of genetic identity and these landraces should, therefore, be preserved as different accessions in any germplasm collection (Gómez *et al.*, 2004).

Genetic diversity of 20 cowpea landraces from Yucatan Peninsula, Mexico

Cowpea was domesticated in West Africa and low levels of genetic diversity both within and between landraces of cultivated cowpea are related to a severe genetic bottleneck that occurred during the cowpea domestication (Chen *et al.*, 2017). The level of genetic diversity identified in this study ($H_t=0.24$, $H_s=0.14$) was lower than what was observed earlier by Gajera *et al.* (2014) and Igwe *et al.* (2017) in cowpea genotypes from India and Nigeria, respectively. Both authors reported values of total gene diversity (H_t) close to 0.4 and $H_s=0.3$. These results may be due to the presence of Iberian cowpea in southeastern Mexico. In the study of Carvalho *et al.* (2017) the authors reported a low genetic diversity in the Iberian cowpea germplasm with $H_e=0.1$.

Genetic parameters including Nei's genetic diversity (H_e), numbers of effective alleles (N_e) as well as Shannon's information index (I) are crucial in the study of genetic diversity in plant species (Igwe *et al.*, 2017).

Assessing the data from the entire sample of cowpea landraces from Yucatan Peninsula, the level of genetic diversity observed in our study ($N_e = 1.32$, $H_e = 0.19$ and $I = 0.30$), it is relatively moderate, compared to other genetic diversity studies with ISSR markers. In India, Gajera *et al.* (2014) assessed the genetic variability among 11 cowpea genotypes that were selected by a range of variability in seed phenotypic and biochemical characteristics, the authors reported values of N_e , H_e and I of 1.7, 0.42 and 0.61, respectively. Later, Igwe *et al.* (2017) investigated the genetic diversity of 18 cowpea accessions from different regions in Nigeria using ISSR and SCoT markers, data from ISSR markers revealed that mean values of N_e , H_e and I were 1.85, 0.45 and 0.64, respectively.

At the landrace level, low levels of intra-landrace variability were observed. Self-pollinated crops like cowpea often possess low intra-population variability (Fikiru *et al.*, 2010; Etminan *et al.*, 2016; Henareh *et al.*, 2016). In the study conducted by Ghalmi *et al.* (2010) with cowpea landraces, no differences were observed between individuals of the same landrace with molecular markers. The selection pressure exerted by Mayan farmers over the years likely led to a reduction in genetic diversity strengthening specific agro-ecological adaptations to the region (Gómez *et al.*, 2004; Ghalmi *et al.*, 2010; López *et al.*, 2019). Also, the small number of individuals studied may not be representative of the genetic diversity present in the accessions, masking the results (López *et al.*, 2019).

Interestingly, in the present study, two landraces (P5 and P12) that were collected in the Calakmul biosphere reserve in the estate of Campeche showed high levels of genetic diversity. These accessions comprise a valuable plant genetic resource that should be protected by conservation programs in the region and is a resource to be used in genetic improvement efforts.

In summary, our results contribute to the knowledge regarding the genetic diversity and structure of *V. unguiculata* in the Yucatan Peninsula, Mexico and should be considered when formulating criteria for sampling and conservation strategies of cowpea landraces in southern Mexico. Our results also identified cowpea landraces that are available for genetic improvement. Further studies including more landraces and with different molecular markers are suggested to obtain more detailed information and a precise assessment of genetic diversity and structure of cowpea landraces. We recommend to develop *in-situ* and *ex-situ* conservation programs of cowpea landraces in the region and to distinctly identify accessions with environmental tolerances, productivity and nutritional value for use in participatory breeding programs.

Supplementary material

To view supplementary material for this article, please visit <https://doi.org/10.1017/S147926212000026X>

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