

Shells of *Patella aspera* as ‘islands’ for epibionts

GUSTAVO M. MARTINS^{1,2}, JOÃO FARIA^{1,2}, MIGUEL FURTADO^{2,3} AND ANA I. NETO^{1,2}

¹CIIMAR/CIMAR, Interdisciplinary Centre for Marine and Environmental Research, University of Porto, Rua dos Bragas 289, 4050-123 Porto, Portugal, ²Centre for Research in Natural Resources, Department of Biology, University of Azores, 9501-801 Ponta Delgada, S. Miguel, Açores, Portugal, ³Faculty of Sciences, University of Lisbon, Campo Grande 1749-016 Lisboa, Portugal

*In this study we examined the epibiont assemblage on shells of the living limpet *Patella aspera*. Limpets were collected at two sites at each of the nine islands of the Azores, totalling 707 individuals examined. Shells were measured and all the epibiota identified to the lowest taxonomic resolution possible. 190 taxa were recorded, of which 97% were algae, including 17 new records for the Azores. Only five shells were devoid of fouling organisms. The assemblage was dominated by a few algal taxa, whereas the majority of species occurred on less than 10% of the shells. A significant and positive relationship was generally found between basibiont size (shell length) and epibiota richness. The strength (slope) of the relationship, however, varied between islands and sites. These results suggest that a range of processes operating at multiple spatial scales influenced epibiont assemblages. Many features identified in these assemblages resemble, in many ways, those examined in island biogeography, suggesting that basibionts may be considered as ‘islands’ and may provide a suitable model system to test ecological hypotheses about ecosystems that are not so amenable to experimentation.*

Keywords: epibiosis, limpets, species richness, spatial scales, Azores

Submitted 9 January 2014; accepted 5 March 2014; first published online 8 April 2014

INTRODUCTION

Epibiosis is defined as a non-symbiotic spatial association between a substratum organism (basibiont) and a sessile organism (epibiont), with the latter growing attached to the basibiont's outer surface and, if present, trophic exchange with the substratum organism is facultative (Wahl, 1989, 2010). Epibiosis is predominantly found in the aquatic environment, with water providing an effective food vector for many sessile organisms (Harder, 2008). In fact, there are many described examples of marine epibiosis (Morton, 1988; Wahl & Mark, 1999) and in some habitats, for example, soft-bottoms, biogenic or living surfaces may actually comprise the largest proportion of hard stable substrata available for colonization (e.g. Creed, 2000; Harder, 2008; Munguia & Miller, 2008). The list of marine basibionts is long but generally includes slow-moving or sessile, non-burrowing, long-lived and large organisms (Wahl, 1989; Wahl & Mark, 1999). Most epibionts are facultative, occurring on both living and non-living substrata (Wahl & Mark, 1999).

Ecologically, epibiosis can entail a range of benefits and disadvantages for both the epi- and basibiont, which are summarized in Wahl (1989). According to Wahl & Mark (1999), some groups of species, including macro- and microalgae, and sessile filter-feeders, such as bryozoans, cnidarians and porifera, are more frequently found as epibionts. In contrast, molluscs and crustaceans are more frequently found as basibionts (see figure 2 in Wahl & Mark, 1999).

Patellid limpets are considered key organisms capable of structuring rocky habitats and in areas where grazing is intense few species but the most grazing-resistant forms of algae (i.e. crustose algae) are able to grow (Hawkins & Hartnoll, 1983; Jenkins *et al.*, 2008; Martins *et al.*, 2010). In contrast, the shells of limpets can support a dense cover of algae (including those that are grazed off of the rock) and animals.

Patellid limpets are traditionally an important food species in the archipelagos of Macaronesia (north-east Atlantic), where they are collected and consumed, especially during summer months (Martins *et al.*, 2011). As a consequence of intense harvesting, populations of these species are considered overexploited, at least at some islands (Hawkins *et al.*, 2000; Navarro *et al.*, 2005; Martins *et al.*, 2008a). *Patella aspera* Röding (1798) occurs predominantly in the shallow subtidal of Macaronesia and often supports a dense cover of algal turf on the shells. In a previous account of the epibiota found on the shells of *P. aspera* collected from two sites of the Azores, Fralick *et al.* (1985) identified a total of 20 species of macroalgae. In this study we add to the previous work by examining the epibiota on a much larger sample covering the entire Azores archipelago. In addition, we explore the data for evidence of the processes responsible for structuring these epibiotic communities.

MATERIALS AND METHODS

Sampling

A total of 784 individuals of *Patella aspera* were sampled on two different populations in each of the nine islands of the

Corresponding author:
G.M. Martins
Email: gmartins@uac.pt

Table 1. Summary information about sampling sites and the numbers of individuals examined.

Island	Site		N
Corvo	Baixa da Lomba	39°41'37"N 31°05'04"W	29
	Baía Pão de Açúcar	39°40'50"N 31°07'06"W	29
Flores	Fajãzinha	39°26'11"N 31°15'46"W	28
	Ponta da Caveira	39°25'27"N 31°08'12"W	35
Faial	Porto do Salão	38°37'31"N 28°39'24"W	52
	Castelo Branco	38°31'01"N 28°42'31"W	47
Pico	São João	38°24'47"N 28°19'55"W	44
	Santa Luzia	38°33'30"N 28°23'28"W	40
São Jorge	Ponta Furada	38°41'15"N 28°05'21"W	37
	Velas	38°40'49"N 28°12'48"W	48
Terceira	São Mateus	38°39'07"N 27°16'40"W	45
	Cabo da Praia	38°42'30"N 27°02'48"W	52
Graciosa	Ponta da Barca	39°05'40"N 28°02'59"W	36
	Carapacho	39°00'42"N 27°57'20"W	57
São Miguel	Lagoa	37°44'39"N 25°35'41"W	47
	João Bom	37°54'03"N 25°47'49"W	48
Santa Maria	Castelete	36°57'10"N 25°01'03"W	45
	Baía da Cré	37°00'19"N 25°08'26"W	52
Total			707

Azores (Table 1). The soft part of each individual was removed for molecular studies (in preparation), and the shells were frozen or preserved in ethanol. Upon inspection, shell integrity was verified and broken shells were discarded from the analysis. Those that were in good condition were retained (N = 707, see Table 1 for details on sampling sites and numbers), hydrated in water, and all the epibiota present on the shells were identified to the lowest possible taxonomic level using a dissecting and optical microscope. Total cover of sessile epibiota on each shell was estimated using the semi-quantitative scale DAFOR, while mobile animals (i.e. limpets) were enumerated. In addition, shell length was measured using a Vernier calliper with a precision of 0.1 mm.

Data analysis

The relationship between basibiont shell length and epibiota richness was examined using linear regression. Spatial variation in shell size and epibiota richness was examined using a two-way fully nested design with the following factors: island (nine levels, random) and site (two levels, random and nested within island). The analyses were run on PERMANOVA (Anderson, 2001, 2005) based on Euclidean distances, with 999 permutations which produce estimates of variation analogous to the classical ANOVA (Anderson, 2001). For the analysis of the spatial variation in richness, shell length was included as a covariate to standardize for spatial variation in shell size.

Table 2. A two-way hierarchical PERMANOVA examining the spatial variability in mean basibiont size (shell length) among islands and sites.

Source	df	MS	F	P
Island	8	50.708	5.15	0.025
Site (Island)	9	9.934	21.93	0.001
Residual	689	0.045		

RESULTS

Variation in basibiont size

Basibiont shell length ranged between 16.4 and 74.3 mm, with a mean shell length of 40.5 mm (± 0.4 standard error (SE), N = 707). Of the 707 individuals selected, only 74 (~10%) were larger than the minimum legal catch size (50.0 mm), an indication of the high level of exploitation of this species. In addition, there was significant spatial variation in limpet shell length at the scale of the islands and sites (Table 2).

Variation in epibiota richness

A total of 190 taxa were identified of which 97% were algae with only five animal taxa recorded. Only five shells out of the 707 (0.7%) examined were completely devoid of epibionts. A total of 17 new records for the Azorean algal flora were found on the epibiota species identified (four Chlorophyta, two Heterokontophyta (Phaeophyceae) and 11 Rhodophyta, Table 3). Encrusting coralline algae (i.e. *Lithothamnion* spp.) was the most frequent taxa, occurring on 54% of all shells (Figure 1). Small turf-forming algal species were the dominant erect taxa, amongst which *Gelidium* sp., *Ceramium rubrum*, *Jania* sp. and *Polysiphonia denudata* were the most common, being recorded on at least 30% of the shells (Figure 1). The large majority of the taxa identified (>85% of the species) was recorded in less than 10% of the shells (Figure 1).

Table 3. New algal additions to the Azorean flora found as epibiota on shells of *Patella aspera*.

Taxa	No. of species
RHODOPHYTA	11
<i>Aglaothamnion cordatum</i> (Børgesen)	
Feldmann-Mazoyer	
<i>Antithamnionella floccosa</i> (O.F. Müller) Whittick	
<i>Antithamnionella spirographidis</i> (Schiffner)	
E.M.Wollaston	
<i>Ceramium tenerrimum</i> (G. Martens) Okamura	
<i>Diplothamnion jolyi</i> C.Hoek	
<i>Herposiphonia secunda</i> f. <i>secunda</i> (C. Agardh)	
Falkenberg	
<i>Lejolisia mediterranea</i> Bornet	
<i>Neosiphonia sphaerocarpa</i> (Børgesen) M.-S. Kim & I.K. Lee	
<i>Spermothamnion flabellatum</i> Bornet in Bornet & Thuret	
<i>Spermothamnion strictum</i> (C. Agardh) Ardissonne	
<i>Stichothamnion cymatophilum</i> Børgesen	
HETEROKONTOPHYTA (Phaeophyceae)	2
<i>Asteronema rhodochortonoides</i> (Børgesen) D.G. Müller & E.R. Parodi	
<i>Hincksia rallsiae</i> (Vickers) P.C. Silva in Silva, Meñez & Moe	
CHLOROPHYTA	4
<i>Cladophora dalmatica</i> Kützting	
<i>Cladophoropsis macromeres</i> W.R. Taylor	
<i>Ernodesmis verticillata</i> (Kützting) Børgesen	
<i>Rhizoclonium tortuosum</i> (Dillwyn) Kützting	
Total	17

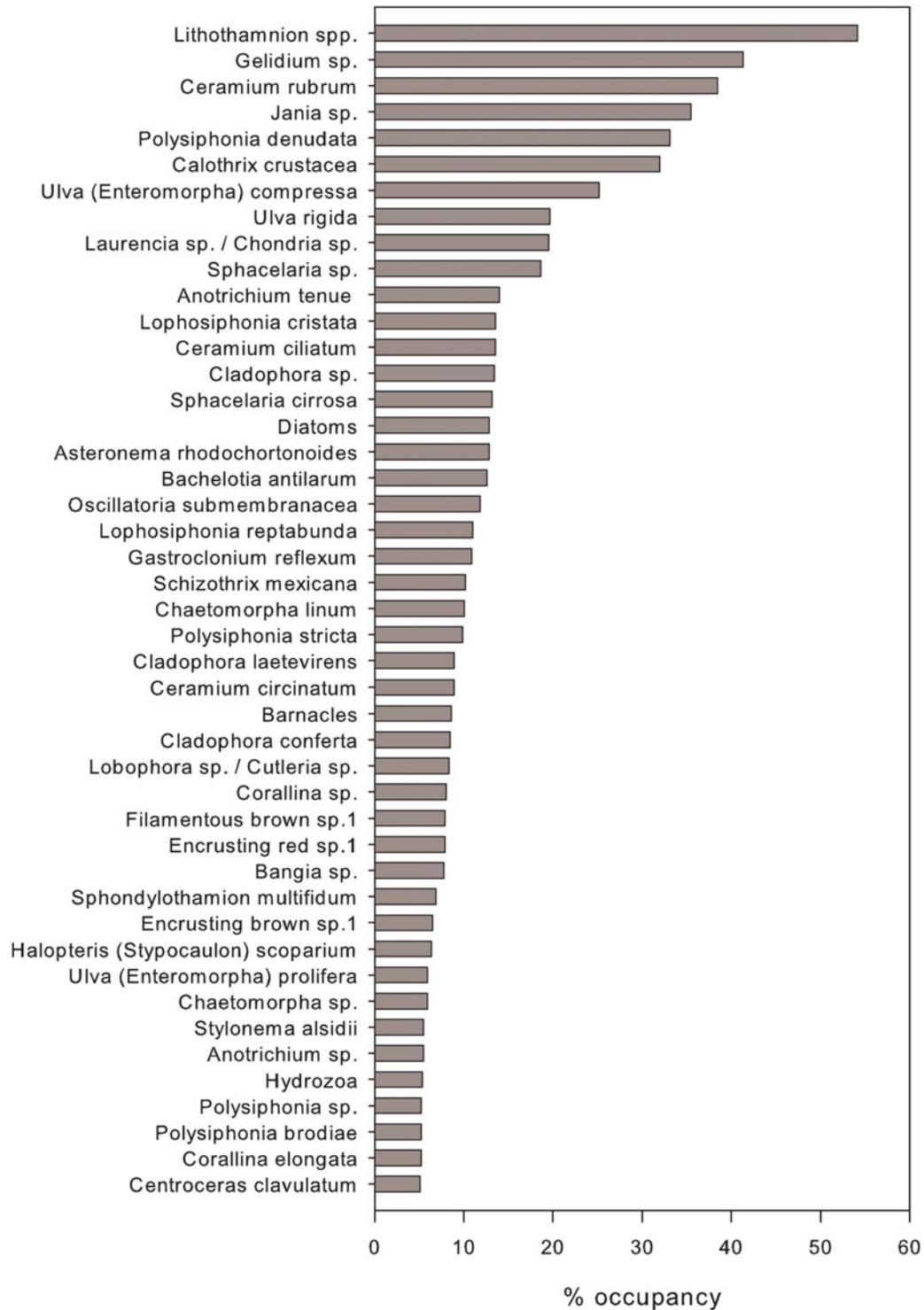


Fig. 1. Epibiont percentage occupancy on shells of *Patella aspera*. Taxa that were not present on at least 5% of the shells are not included for clarity.

The mean number of epibionts found on shells of *Patella aspera* was 9.00 ± 0.15 (mean \pm SE) and ranged between 0 and 26 taxa. A significant and positive linear regression was found between basibiont shell length and epibiont richness (Figure 2) indicating that larger (and older) limpets support a richer epibiota.

Analysis of the spatial variation in epibiota richness showed significant interactions between the covariate (shell length), island and site (Table 4). Analysis of Figure 3

shows that a positive and significant relationship between basibiont size and epibiont richness was spatially inconsistent, being present at some islands, but not at others. Similarly, the relationship was variable between sites within islands. These results suggest that the relationship between the basibiont and epibiota was influenced by processes other than the basibiont size alone, including processes operating at large- (island) and intermediate- (site) spatial scales.

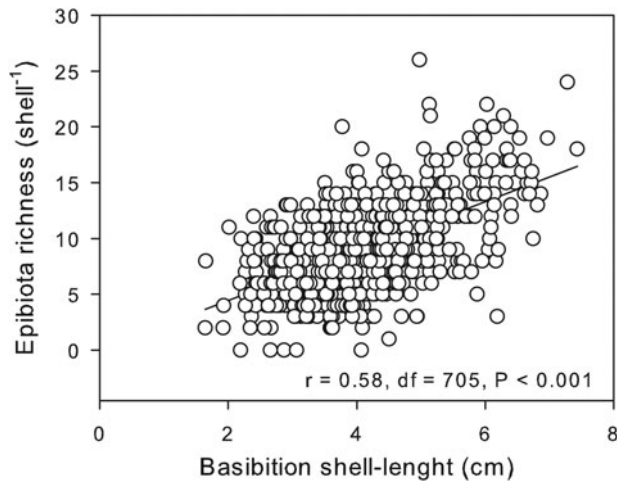


Fig. 2. Relationship between basibiont size (limpet shell length) and epibiota richness.

Table 4. A two-way PERMANOVA examining differences in epibiota richness on shells of *Patella aspera* among islands and sites. Individual shell length was used as the covariate.

Source	df	MS	F	P
Covariate	1	3761.70	37.95	0.001
Island	8	162.67	2.39	0.076
Site (island)	9	67.43	8.91	0.001
Covariate × island	8	20.51	2.71	0.006
Covariate × site (island)	9	17.31	2.29	0.014
Residual	671	7.56		

DISCUSSION

An overwhelmingly large number of epibiota was found on shells of *Patella aspera*, unlike the earlier record by Fralick *et al.* (1985), which is not surprising given the restricted sample examined by the latter. This large diversity found on the shells highlights the idea that *P. aspera*, despite being a key herbivore, also plays an important habitat for many of the species found and on which it feeds. Apart from the new records (see below), for which there is no information regarding their distribution in the Azores, all taxa found on shells have also been recorded on a natural rocky substratum (authors, personal observation). This suggests that: (1) shells of limpets do not provide a 'unique' habitat; and (2) no obligatory epibiont exists among the taxa here found. Nevertheless, the shells of *P. aspera* may provide a key microhabitat that increases the survivorship of colonizing algae, especially where they occur in areas devoid of conspicuous macroalgae (barrens) created by the sea urchin *Arbacia lixula*.

The new additions to the Azorean algal flora reported in this study correspond to species that have probably gone unnoticed until now due to their small size and/or obscure life cycle, although a northern extension of such taxa due to climate changes in the last decades (i.e. increase in sea temperature) cannot be excluded. In fact, all the new additions here reported are known from southern locations in the Macaronesian region (Guiry & Guiry, 2013), with their presence in the Azores representing a hypothetical recent northern extension in their distribution. Distribution shifts in marine

species associated with recent increases in sea temperature have been referred to before for the north-east Atlantic, with several examples showing that rising sea temperatures or climate related changes can result in southern species shifting northward (Hawkins *et al.*, 2003; Perry *et al.*, 2005).

A substantially lower number of completely unfouled shells was generally found (0.74%) compared to other mollusc basibionts, such as turbo snails (Wernberg *et al.*, 2010) and fan mussels (Addis *et al.*, 2009). Wahl & Sönnichsen (1992) found that epibiosis on the shells of *Littorina littorea* was spatially variable. In this case, intertidal populations from Helgoland rarely carried any epibiota, whereas subtidal populations from the Kiel Bight were frequently fouled. They found that the extent of shell fouling was primarily driven by littorinid density (probably associated with mucus secretion, bulldozing or mutual grazing), whereas differences in habitat conditions played a secondary role. The overall low density of *P. aspera* (as a consequence of exploitation), and the fact that herbivore epibionts were rarely found, may thus explain the insignificant proportion of unfouled shells. There are, however, other possibilities. Many studies have shown that epibiosis can influence predator–prey interactions, either by facilitating or hampering the success of predators (Wahl *et al.*, 1997; Laudien & Wahl, 1999; Enderlein *et al.*, 2003). For instance, Silva (2008) and Silva *et al.* (2008) found that limpets can generally be important prey for crabs. A strong and selective preference of crabs for unfouled shells (Wahl *et al.*, 1997) could thus also lead to a generally low number of unfouled shells. Moreover, selective harvesting by man (e.g. unfouled shells are more easily detected) could also add to the effects of predation. Without rigorous experimental work, however, it is impossible to determine the reason why so few shells remain free of epibionts.

As in most studies examined (e.g. Wernberg *et al.*, 2010), there was a positive relationship between basibiont size and epibiota richness. This is not surprising, given the ubiquitous species–area relationship (see Connor & McCoy, 1979; McGuinness, 1984; Lomolino, 2000 for reviews). However, in the case of living substrata, this positive relationship could also be the result of temporal accumulation of species, as larger shells are also generally associated with older individuals.

Results also indicated that the relationship between basiont size and epibiota richness was variable between islands and between locations within islands. Not only did slopes differ, but also there were islands and sites where the relationship was non-existent. Wernberg *et al.* (2010) also found spatially variable results between basibiont size and epiflora richness, and attributed these differences to spatial variation in the density of epizooic herbivores. In our case, epizooic herbivores were rare and thus unlikely to produce such results. However, these results do indicate that epibiota richness is not only driven by basibiont size, but also that other processes (e.g. regional and local species pool, local environmental and biotic conditions) have the ability to influence these assemblages. In this regard, epibiont assemblages are no different from the generality of marine communities (e.g. Fraschetti *et al.*, 2005; Martins *et al.*, 2008b).

In conclusion, we found a highly diverse assemblage living on the shells of *P. aspera*. Epibiota richness was positively influenced by basibiont size, but there was evidence that larger-scale processes operating at the scales of island and location also played a role in structuring these assemblages.

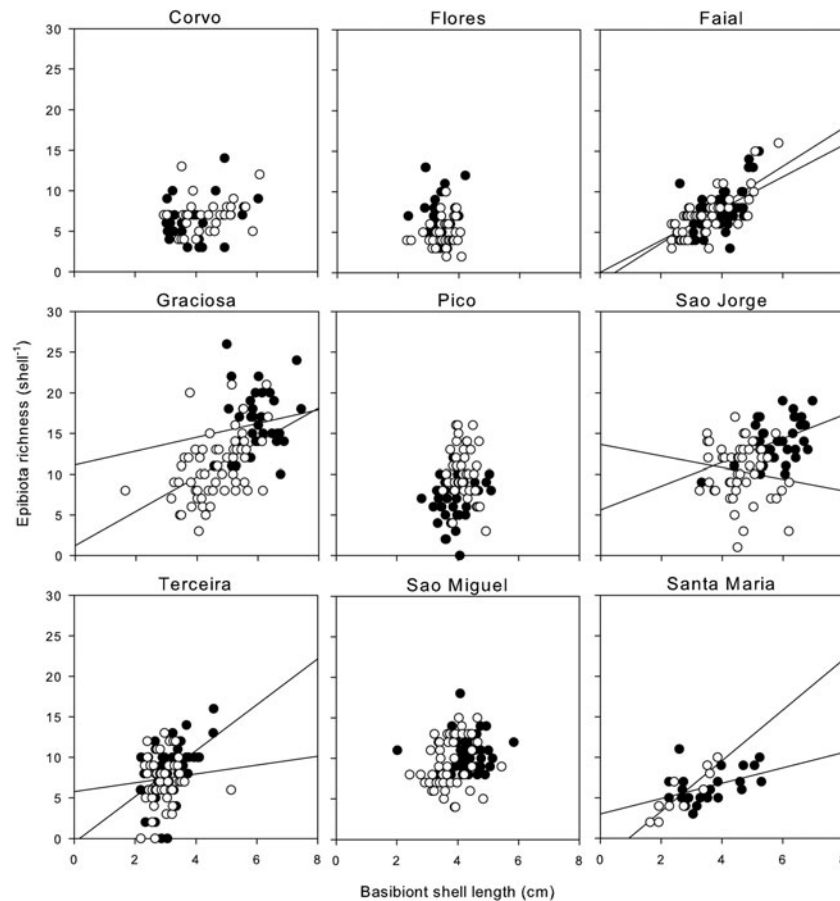


Fig. 3. Relationship between basibiont size (limpet shell length) and epibiota richness broken down between islands and sites. Filled and open circles represent each of the two sites sampled on each island, respectively. Significant correlation lines are indicated and extended to the axes for clarity.

The basibiont–epibiont relationship resembles, in many ways, the studies of island biogeography (e.g. MacArthur & Wilson, 1963; Whittaker *et al.*, 2008) suggesting that it might be a good model system in which to experimentally test hypotheses about ecosystems that are not so amenable to experimentation (e.g. islands and lakes).

FINANCIAL SUPPORT

G.M.M. was supported by a post-doctoral grant awarded by Fundação para a Ciência e Tecnologia (FCT), Portugal (SFRH/BDP/63040/2009). J.F. was supported by a PhD grant awarded by FRCT, Azores (M3.1.2/F/021/2011). This research was funded by the project Patélgene (PTDC/BIA-BIC/115837/2009, FCT) and by the European Regional Development Fund (ERDF) through the COMPETE—Operational Competitiveness Programme and national funds through FCT—Foundation for Science and Technology, under the project 'PEst-/MAR/LA0015/2011'.

REFERENCES

Addis P., Secchi M., Brundu G., Manunza A., Corrias S. and Cau A. (2009) Density, size structure, shell orientation and epibiontic colonization of the fan mussel *Pinna nobilis* (Mollusca: Bivalvia) in their

contrasting habitats in an estuarine area of Sardinia (W Mediterranean). *Scientia Marina* 73, 143–152.

Anderson M.J. (2001) A new method for non-parametric multivariate analysis of variance. *Austral Ecology* 26, 32–46.

Anderson M.J. (2005) *PERMANOVA: permutational multivariate analysis of variance*. Auckland: Department of Statistics, University of Auckland.

Connor E.F. and McCoy E.D. (1979) Statistics and biology of the species–area relationship. *American Naturalist* 113, 791–833.

Creed J.C. (2000) Epibiosis on cerith shells in a seagrass bed: correlation of shell occupant with epizoite distribution and abundance. *Marine Biology* 137, 775–782.

Enderlein P., Moorthi S., Rohrscheidt H. and Wahl M. (2003) Optimal foraging versus shared doom effects: interactive influence of mussel size and epibiosis on predator preference. *Journal of Experimental Marine Biology and Ecology* 292, 231–242.

Fralick R.A., Hehre E.J. and Mathieson A.C. (1985) Observations on the marine algal flora of the Azores I: notes on the epizoic algae occurring on the marine molluscs *Patella* spp. *Arquipélago* VI, 39–43.

Fraschetti S., Terlizzi A. and Benedetti-Cecchi L. (2005) Patterns of distribution of marine assemblages from rocky shores: evidence of relevant spatial scales. *Marine Ecology Progress Series* 296, 13–29.

Guiry M.D. and Guiry G.M. (2013) *AlgaeBase*. Galway: University of Ireland, World-wide electronic publication. Available at: <http://www.algaebase.org> (accessed 13 March 2014).

- Harder T.** (2008) Marine epibiosis: concepts, ecological consequences and host defence. In Flemming H.C., Murthy P.S., Venkatesan R. and Cooksey K. (eds) *Marine and industrial biofouling*. Berlin: Springer, pp. 219–231.
- Hawkins S.J., Côrte-Real H.B.S.M., Pannacciulli F.G., Weber L.C. and Bishop J.D.D.** (2000) Thoughts on the ecology and evolution of the intertidal biota of the Azores and other Atlantic islands. *Hydrobiologia* 440, 3–17.
- Hawkins S.J. and Hartnoll R.G.** (1983) Grazing of intertidal algae by marine invertebrates. *Oceanography and Marine Biology: an Annual Review* 21, 195–282.
- Hawkins S.J., Southward A.J. and Genner M.J.** (2003) Detection of environmental change in a marine ecosystem—evidence from the western English Channel. *Science of the Total Environment* 310, 245–256.
- Jenkins S.R., Moore P., Burrows M.T., Garbat D.J., Hawkins S.J., Ingólfsson A., Sebens K.P., Snelgrove P.V.R., Wetthey D.S. and Woodin S.A.** (2008) Comparative ecology of North Atlantic shores: do differences in players matter for process? *Ecology* 89, S3–S23.
- Laudien J. and Wahl M.** (1999) Indirect effects of epibiosis on host mortality: Seastar predation on differently fouled mussels. *Marine Ecology* 20, 35–47.
- Lomolino M.V.** (2000) Ecology's most general, yet protean pattern: the species–area relationship. *Journal of Biogeography* 27, 17–26.
- MacArthur R.H. and Wilson E.O.** (1963) An equilibrium theory of insular zoogeography. *Evolution* 17, 373–387.
- Martins G.M., Jenkins S.R., Hawkins S.J., Neto A.I. and Thompson R.C.** (2008a) Exploitation of rocky intertidal grazers: population status and potential impacts on community structure and functioning. *Aquatic Biology* 3, 1–10.
- Martins G.M., Thompson R.C., Hawkins S.J., Neto A.I. and Jenkins S.R.** (2008b) Rocky intertidal community structure in oceanic islands: scales of spatial variability. *Marine Ecology Progress Series* 356, 15–24.
- Martins G.M., Thompson R.C., Neto A.I., Hawkins S.J. and Jenkins S.R.** (2010) Exploitation of intertidal grazers as a driver of community divergence. *Journal of Applied Ecology* 47, 1282–1289.
- Martins G.M., Jenkins S.R., Hawkins S.J., Neto A.I. and Thompson R.C.** (2011) Illegal harvesting affects the success of fishing closure areas. *Journal of the Marine Biological Association of the United Kingdom* 91, 929–937.
- McGuinness K.A.** (1984) Equations and explanations in the study of species area curves. *Biological Reviews of the Cambridge Philosophical Society* 59, 423–440.
- Morton B.** (1988) *Partnerships in the sea: Hong Kong's marine symbioses*. Hong Kong: Hong Kong University Press.
- Munguia P. and Miller T.E.** (2008) Habitat destruction and metacommunity size in marine systems. *Journal of Animal Ecology* 77, 1175–1182.
- Navarro P.G., Ramirez R., Tuya F., Fernandez-Gil C., Sanchez-Jerez P. and Haroun R.J.** (2005) Hierarchical analysis of spatial distribution patterns of patellid limpets in the Canary Islands. *Journal of Molluscan Studies* 71, 67–73.
- Perry A.L., Low P.J., Ellis J.R. and Reynolds J.D.** (2005) Climate change and distribution shifts in marine fishes. *Science* 308, 1912–1915.
- Silva A.C.F.** (2008) *Predation by crabs on rocky shores in north-east Atlantic*. PhD thesis. University of Plymouth, UK.
- Silva A.C.F., Hawkins S.J., Boaventura D.M. and Thompson R.C.** (2008) Predation by small aquatic predators regulates populations of the intertidal limpet *Patella vulgata* (L.). *Journal of Experimental Marine Biology and Ecology* 367, 259–265.
- Wahl M.** (1989) Marine epibiosis. I. Fouling and antifouling: some basic aspects. *Marine Ecology Progress Series* 58, 175–189.
- Wahl M.** (2010) Epibiosis. In Dürr S. and Thomasson J.C. (eds) *Biofouling*. Oxford: Wiley–Blackwell, pp. 100–108.
- Wahl M., Hay M.E. and Enderlein P.** (1997) Effects of epibiosis on consumer–prey interactions. *Hydrobiologia* 355, 49–59.
- Wahl M. and Mark O.** (1999) The predominantly facultative nature of epibiosis: experimental and observational evidence. *Marine Ecology Progress Series* 187, 59–66.
- Wahl M. and Sönnichsen H.** (1992) Marine epibiosis. IV. The periwinkle *Littorina littorea* lacks typical antifouling defences—why are some populations so little fouled? *Marine Ecology Progress Series* 88, 225–235.
- Wernberg T., Tuya F. and Thomsen M.** (2010) Turban snails as habitat for foliose algae: contrasting geographical patterns in species richness. *Marine and Freshwater Research* 61, 1237–1242.
- and
- Whittaker R.J., Triantis K.A. and Ladle R.J.** (2008) A general dynamic theory of oceanic island biogeography. *Journal of Biogeography* 35, 977–994.

Correspondence should be addressed to:

G.M. Martins
 CIIMAR/CIMAR, Interdisciplinary Centre for Marine and Environmental Research
 University of Porto Rua dos Bragas 289, 4050-123 Porto Portugal
 email: gmartins@uac.pt