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Author for correspondence: Amanda Ayala, Email: ayala2ap@gmail.com

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Superparasitism and fitness parameters in three native wasp parasitoids (Braconidae: Opiinae) of the Mexican fruit fly, *Anastrepha ludens* (Diptera: Tephritidae)

Amanda Ayala^{1,2} ⁽ⁱ⁾, Jorge Toledo¹ ⁽ⁱ⁾, Gabriela Pérez-Lachaud³ ⁽ⁱ⁾, Pablo Liedo¹ ⁽ⁱ⁾ and Pablo Montova² ⁽ⁱ⁾

¹El Colegio de la Frontera Sur, Agricultura, Sociedad y Ambiente, Carretera Antiguo Aeropuerto Km 2.5, Tapachula, Chiapas, 30700, México; ²Programa Moscafrut SENASICA-SADER, Camino a los Cacaotales S/N, Metapa de Domínguez, Chiapas, 30860, México and ³El Colegio de la Frontera Sur, Conservación de la Biodiversidad, Avenida Centenario Km 5.5, Chetumal, Quintana Roo, 77014, México

Abstract

Knowledge on reproductive strategies and host use in insect parasitoids is fundamental for biological control purposes. Superparasitism and multiparasitism, oviposition in a previously parasitized host by a female of the same or different species, respectively, may impact pest management decisions. Our objective was to determine the occurrence of superparasitism and multiparasitism in three species of native larval-pupal solitary endoparasitoids that attack Anastrepha Shiner species (Diptera: Tephritidae) in the Neotropical region, and the possible effect on offspring fitness parameters. Doryctobracon crawfordi (Viereck), Utetes anastrephae (Viereck), and Opius hirtus (Fischer) occur in sympatry in Mexico, and are currently under consideration for use as biocontrol agents. Experiments were conducted under laboratory conditions with females acting alone (self-superparasitism), females in groups of the same species (conspecific superparasitism), and females in mixed groups (multiparasitism). Our results showed that self-superparasitism is an uncommon strategy in the three native species and is rare under conditions of intraspecific competition. In the case of multiparasitism, a higher number of immature stages of U. anastrephae was observed, compared to those of D. crawfordi and O. hirtus. However, it is not clear yet if this was due to some adult female trait or to the competitive ability of the larvae. We conclude that most females of the native species studied appeared to avoid superparasitism, specifically when acting alone, suggesting a high discrimination ability, which is probably a result of a close relationship and evolutionary history with Anastrepha hosts.

Introduction

Reproductive success in parasitoid wasps depends largely on the decisions made by females during their foraging activities (Outreman and Pierre, 2005). The acceptance of a host for oviposition becomes the most important step once the host has been found, and this is mainly dependent upon the quality of the host (Visser *et al.*, 1992). In solitary parasitoids, where only one individual can complete its development in a given host, a determining factor for host acceptance is its condition of being previously parasitized or not (Ueno, 1994), since an already parasitized host will be, in general, of lower quality for the foraging female (Bakker *et al.*, 1985; Nelson and Roitberg, 1995). Sympatric wasp parasitoids using the same resources may compete for hosts both at the adult stage (when females forage in the same patch) and at the larval stage, when hosts are superparasitized or multiparasitized (within-host competition) (Harvey *et al.*, 2013; Cusumano *et al.*, 2016).

Superparasitism is a reproductive strategy that occurs when a female parasitoid parasitizes a host that has been previously parasitized, either by herself or by another female of the same species (van Alphen and Visser, 1990). However, many species of solitary parasitoids are able to distinguish between parasitized and non-parasitized hosts, which is known as discrimination ability (van Lenteren, 1981; Mackauer, 1990). This depends on the capacity of female parasitoids for identifying external or internal cues on the hosts (Vinson, 1976; Roitberg and Mangel, 1988). In solitary parasitoids, supernumerary individuals are eliminated through direct, physical combat or through physiological suppression (Bakker *et al.*, 1985; Hubbard *et al.*, 1987; Mackauer *et al.*, 1992).

Superparasitism occurs at different levels: (1) self-superparasitism, when a female solitary parasitoid oviposits more than once on or in the same host; (2) conspecific superparasitism, when two or more females of the same species parasitize one host (van Dijken and Waage,

1987); and (3) multiparasitism, when two or more females of different species parasitize the same host (Vinson and Ables, 1980; Pschorn-Walcher, 1987; Pijls *et al.*, 1995). Multiparasitism events may lead either to interspecific competition among parasitoids, or to a facilitation process in those situations in which a given species may benefit from the presence of the other species (Poelman *et al.*, 2014; Cusumano *et al.*, 2016).

Under both mass rearing and field conditions, superparasitism has been reported in *Diachasmimorpha longicaudata* (Ashmead) (Hymenoptera: Braconidae), which was introduced into Mexico to control fruit flies of the genus *Anastrepha* Schiner (Diptera: Tephritidae) (González *et al.*, 2007, 2010; Montoya *et al.*, 2011, 2013). Such reproductive strategy has been related to a femalebiased sex ratio without adversely affecting other fitness parameters of the offspring (González *et al.*, 2007; Montoya *et al.*, 2011). On the contrary, little is known of the effect of superparasitism and multiparasitism in the guild of native *Anastrepha* parasitoids.

In the Neotropics, the guild of native parasitoids associated with fruit flies of this genus comprises mainly members of the family Braconidae (Hymenoptera), for the most part, Opiinae solitary, larval-pupal endoparasitoids (Ovruski et al., 2000). Among the most important species are Doryctobracon crawfordi Viereck, Utetes anastrephae Viereck, and Opius hirtus Fischer (López et al., 1999; Ovruski et al., 2000), which are polyphagous and associated with several species of the genus Anastrepha. Doryctobracon crawfordi is distributed from central Mexico to Argentina (Ovruski et al., 2005) and is associated mainly with larvae of Anastrepha ludens Loew, A. fraterculus Wiedemann, A. striata Schiner, A. serpentina Wiedemann, and A. obligua Macquart (López et al., 1999). The distribution of U. anastrephae extends from northern Florida to northern Argentina (Sivinski et al., 1997; Ovruski et al., 2000), while O. hirtus is distributed mainly in Mexico, Costa Rica, and the Dominican Republic (Ovruski et al., 2000). Opius hirtus has been related to Anastrepha species not attacked by the other parasitoid species in the region (López et al., 1999; Sivinski et al., 2000), such as Anastrepha cordata Aldrich, which develops in Tabernaemontana alba Mill (Apocynaceae) and other small-sized fruits (Hernández-Ortíz et al., 1994). Opius hirtus has the narrowest host range of these three species and has a great capacity to forage for rare, lowdensity hosts (García-Medel et al., 2007).

The purpose of this study was to determine, under laboratory conditions, the presence and level of superparasitism in the three native parasitoid species of *Anastrepha* flies, and its possible effects on fitness parameters such as offspring sex ratio, survival, and fecundity. This basic knowledge could contribute to deciding on the use of these species in augmentative biocontrol programs.

Materials and methods

Biological material

The experiments were carried out at the Biological Control Laboratory of the Methods Development Unit of the Moscafrut Program (SENASICA-SADER), located in Metapa de Domínguez, Chiapas, Mexico. Environmental conditions were: temperature of 25 ± 1 °C, relative humidity of $75 \pm 5\%$, and photoperiod of 12:12 h (light: darkness). Eight-day-old *A. ludens* host larvae were provided by the Moscafrut facility where this species is mass reared at a rate of 75 million per week (Orozco-Dávila *et al.*, 2017). Doryctobracon crawfordi, U. anastrephae, and O. hirtus adults were provided by the Biological Control laboratory

where they are currently maintained at the colony level using the methods described by Aluja *et al.* (2009).

Determination of superparasitism

Self-superparasitism

Individual 5-day-old mated females were individually placed in $30 \times 30 \times 30$ cm Plexiglass cages and 20 A. ludens larvae introduced into each cage as hosts. Females were sexually mature (see Lawrence et al., 1978; Ovruski et al., 2003) and naive (i.e., had no previous experience of oviposition). The three parasitoid species in this study are known to be synovigenic, i.e. they emerge with few or no eggs, but egg-load significantly increases after 24 h (Cicero et al., 2011). The host larvae were mixed with artificial larval diet (see Orozco-Dávila et al., 2017) and exposed to each female parasitoid in the oviposition units, which consisted of a Petri dish $(8.5 \times 1.5 \text{ cm})$ covered with fine organza mesh secured in place by an elastic band to prevent escape of the larvae. After 2 h of exposure, the larvae were removed and placed in plastic containers (~ 100 ml) to continue their development to the pupal stage. After 24 h, the diet was removed by washing with water and replaced with moist coconut fiber as substrate for pupation. After 72 h, the exposed hosts were observed under a Discovery V8 Karl Zeiss stereomicroscope (Göttingen, Germany) in order to quantify the number of oviposition scars present on the fly puparium as an indicator of superparasitism (as in Montoya et al., 2000, 2003). A 10% subsample of the hosts was dissected to corroborate and correlate the presence of immature parasitoid stages with the number of oviposition scars observed on the puparium. The non-dissected hosts were placed in plastic containers (~100 ml) labeled according to the number of scars, and held till adult parasitoid or fly emergence to determine the relationship between offspring sex ratio and number of scars. Fifteen replicates were conducted per parasitoid species.

Conspecific superparasitism and offspring fitness

Groups of five mated females of the same species and age (5-day-old), with no oviposition experience, were placed in Plexiglass cages of the type described above, and provided with 20 larvae in an oviposition unit. The same methods described in the previous section were followed for these tests.

In order to determine the effect of superparasitism on the longevity and fecundity of the progeny of each species, we formed pairs with the adult parasitoids that emerged from hosts with different number of scars (1 scar, 2–3 scars, and >3 scars). Survival was evaluated under two conditions: (a) without food or water, and (b) with food (honey) and water (as in FAO/IAEA/USDA, 2019). For the first condition, only mortality of both sexes was recorded every day, while for the second condition, mortality and fecundity were recorded daily. For fecundity, twenty 8-dayold *A. ludens* larvae were exposed to each adult pair for 2 h a day, starting from the fifth day until the female parasitoid died. There were 15 replicates per species.

Multiparasitism

Groups of three females (one female of each species) were introduced into the Plexiglass cages described above, where 20 A. *ludens* larvae were exposed to them in a Petri dish oviposition unit for 2 h. The Petri dishes with larvae were then removed and the larvae placed in recycled diet in a plastic container (~100 ml) with a lid, in order to continue their development to the pupal stage. The diet was removed after 24 h of exposure and replaced with a moist substrate of coconut fiber. The puparia were maintained under laboratory conditions until adult emergence. The percentage of emergence (observed level of parasitism) and sex ratio of each species was recorded. To corroborate the presence of immature stages of the parasitoid species in competition, a subsample of 10% of the parasitized hosts was dissected three days after exposure. The larvae were identified by morphological differences among species according to Murillo *et al.* (2016). A total of 15 replicates were conducted.

Statistical analyses

The relationship between number of oviposition scars per puparium and number of immature stages found within each host was analyzed by a simple linear regression in the self-superparasitism experiment, and by a binary logistic regression in the case of the conspecific superparasitism experiment, according to the nature of the data. The possible influence of superparasitism (number of scars per host) on the sex of the emerging adult was analyzed with a logistic regression using a logit link function. Survival curves were compared using the log-rank test and a Cox proportional hazards model (Lee and Wang, 2003). Fecundity was estimated from the number of daughters produced daily by each female for each species of parasitoid, and the data analyzed using a one-way ANOVA with a Tukey test. Multiparasitism data were analyzed using a one-way ANOVA with a Tukey test. The logistic regression was conducted using Statgraphics Centurion ver. XV (2008). For the one-way ANOVA analysis, linear regression and log rank test we used JMP version 7.0.1 (SAS, 2007). Data were checked for the assumption of normality with Kolmogorov-Smirnov test.

Results

Self-superparasitism

A total of 87 (29%) of the hosts exposed to *D. crawfordi* (n = 300) exhibited oviposition scars, of which 33 (38%) had more than one scar. There was no relationship between the number of scars present per host and the number of immature stages found within the host (0–1 parasitoid larva/host) (logistic regression, $\chi^2 =$ 1.04, d.f. = 1, P = 0.31, fig. 1a). The percentage of observed emergence of D. crawfordi was 33%. For the hosts parasitized by U. anastrephae, 147 puparia (49%) had scars, of which 73 (50%) had more than one scar. No relationship was observed between number of scars and quantity of immature stages ($\chi^2 = 1.22$, d.f. = 1, P = 0.26, fig. 1b). The percentage of observed emergence in this species was 34%. A similar tendency was observed for O. hirtus, where the relationship between number of immature stages and number of scars on the host puparia was also not significant $(\chi^2 = 1.53, d.f. = 1, P = 0.21, fig. 1c)$, with 81 (27%) of the hosts exhibiting scars, of which 34 (42%) had more than one scar. Observed adult emergence was 23%. The sex of the resulting adults was not influenced by the number of scars per host in any of the three species: D. crawfordi (logistic regression, $\chi^2 =$ 0.16, d.f. = 1, P = 0.68), U. anastrephae ($\chi^2 = 0.05$, d.f. = 1, P =0.82), and O. hirtus ($\chi^2 = 0.01$, d.f. = 1, P = 0.93).

Conspecific superparasitism and offspring fitness

Of the larvae exposed to *D. crawfordi*, 73% (219/300) exhibited oviposition scars, of which 79% (173/219) had more than one

scar. The relationship between number of oviposition scars on the puparia and number of immature stages within them was at the limit of significance ($R^2 = 0.25$; F = 4.3, d.f. = 1,13; P = 0.05, fig. 1d). The number of immature stages was never greater than four and the percentage of adults that emerged was 27%. In U. anastrephae, 58% (174/300) of the hosts exhibited oviposition scars, of which 84% (146/174) exhibited more than one scar. In this species, a strong positive and significant relationship was observed between number of scars and number of immature stages ($R^2 = 0.72$; F = 56.13, d.f. = 1,21; P < 0.0001, fig. 1e). Adult emergence was 36%. In O. hirtus, 57% of hosts (170/300) exhibited oviposition scars, of which 80% exhibited more than one scar (135 hosts). However, most of the dissected hosts contained only one larva and the relationship between number of scars per host and number of immature stages was not significant ($R^2 = 0.059$, F = 1.84, d.f. = 1,29, P = 0.18, fig. 1f). The percentage of adult emergence in this species was 13%.

Under conspecific superparasitism conditions, the probability of an emerging parasitoid being a female was not related to the number of scars present on the host for any of the species evaluated: *D. crawfordi* (logistic regression, $\chi^2 = 1.97$, d.f. = 1, *P* = 0.16), *U. anastrephae* (logistic regression $\chi^2 = 0.96$, d.f. = 1, *P* = 0.32), and *O. hirtus* (logistic regression $\chi^2 = 0.035$, d.f. = 1, *P* = 0.85).

Survival

In the test without food and water, the number of oviposition events per host (1 scar, 2–3 scars, and >3 scars) had no effect on female and male survival for any of the three species [*D. crawfordi* (log-rank test, $\chi^2 = 9.20$, d.f. = 2, P = 0.32; $\chi^2 = 0.67$, d.f. = 2, P = 0.71), *U. anastrephae* (log-rank test, $\chi^2 = 17.95$, d.f. = 1, $P \le 0.05$; $\chi^2 = 1.98$, d.f. = 2, P = 0.36), and *O. hirtus* (log-rank test, $\chi^2 = 7.24$, d.f. = 2, P = 0.29; $\chi^2 = 2.93$, d.f. = 2, P = 0.23)] for females and males, respectively. However, when comparing among species, *O. hirtus* was the species with the longest survival (5.26 ± 0.24 days) (mean \pm SE; n = 43) (log-rank test $\chi^2 = 12.81$, d.f. = 2, P = 0.0001). *Doryctobracon crawfordi* and *U. anastrephae* lived for shorter times, 4.43 ± 0.25 days (\pm SE, n = 24) and $4.38 \pm$ 0.17 days (\pm SE, n = 28), respectively.

When food and water were provided, survival was not significantly different among females of each species reared from hosts with different number of scars: *D. crawfordi* (log-rank test $\chi^2 =$ 5.78, d.f. = 2, *P* = 0.06), *U. anastrephae* (log-rank test $\chi^2 =$ 4.64, d.f. = 2, *P* = 0.09), and *O. hirtus* (log-rank test $\chi^2 =$ 0.71, d.f. = 2, *P* = 0.7) (fig. 2a–c). The number of scars present on the puparium had no effect on the survival of the emerged females and males, *D. crawfordi* (log-rank test $\chi^2 =$ 5.01, d.f. = 1, *P* = 0.08), *U. anastrephae* (log-rank test $\chi^2 =$ 1.98, d.f. = 1, *P* = 0.36), and *O. hirtus* (logrank test $\chi^2 =$ 2.93, d.f. = 1, *P* = 0.23).

Fecundity

Females of *D. crawfordi* and *O. hirtus* that emerged from hosts with a low number of scars (1–3 scars) produced a significantly higher number of daughters (F = 1.66, d.f. = 2,70, P = 0.01 and F = 5.05, d.f. = 2,66, P = 0.001, respectively), compared with those emerged from hosts with more than three scars (Table 1). In contrast, the number of daughters in the progeny of *U. anastrephae* females was unaffected by the number of scars present on the host from which they emerged (F = 1.48, d.f. = 2,69, P = 0.23) (Table 1). The observed sex ratio in the three species of native parasitoids studied was 1:1.



Figure 1. Relationship between number of oviposition scars per host and number of immature stages in self-superparasitism and conspecific superparasitism conditions in *Doryctobracon crawfordi* (a, d), *Utetes anastrephae* (b, e), and *Opius hirtus* (c, f). Logistic regression (a, b, c and f) and linear regression (d, e).



Figure 2. Survival of fed females of Doryctobracon crawfordi (a), Utetes anastrephae (b), and Opius hirtus (c) that emerged from host larvae with different numbers of oviposition scars.

Multiparasitism

A greater number of *U. anastrephae* immature stages was observed in the dissected puparia (1 ± 0.06) (±SE) (one-way ANOVA, *F* = 22.7, d.f. = 2,87, *P* < 0.01), followed by *D. crawfordi* and *O. hirtus*. Only one host was found concurrently parasitized by the three species of parasitoids. The combination of immature stages most frequently observed per host was *D. crawfordi* and *U. anastrephae* (*n* = 4 hosts) (fig. 3). Of the non-dissected hosts, there were no significant differences in observed adult emergence between *U. anastrephae* and *D. crawfordi* (*F* = 2.15, d.f. = 1,33, *P* = 0.13). However, the number of observed *O. hirtus* adults that emerged (0.58 ± 0.22) (±SE) was significantly lower than that of

D. crawfordi and *U. anastrephae* $(3.08 \pm 0.86 \text{ and } 2.83 \pm 0.78, \text{ respectively})$ (±SE) (*F* = 4.01, d.f. = 2,42, *P* = 0.03).

Discussion

Our results show that, in the three native parasitoid species studied, superparasitism does not appear to be a commonly adopted strategy, since this phenomenon was practically non-existent when females foraged in solitary. Under intraspecific competition conditions, it occurred only infrequently. Furthermore, multiparasitism of the host was observed only in nine instances (n = 300). Avoidance of superparasitism in native parasitoid females

Table 1. Mean daily daughters per female (mean \pm SE). Females emerged from host puparia with a different number of scars under the condition of conspecific superparasitism.

		Parasitoid species		
Number of scars	D. crawfordi	U. anastrephae	O. hirtus	
1	4.7 ± 0.37a	3.02 ± 0.6a	2.3 ± 0.40a	
2–3	3.5 ± 0.60ab	2.02 ± 0.3a	1.6 ± 0.2ab	
>3	2.8 ± 0.43b	2.1 ± 0.5a	$1.1\pm0.17b$	

Different letters in a single column indicate a statistically significant difference. One-way ANOVA and Tukey test, P < 0.05.



Figure 3. Number of dissected hosts (*n* = 30) that contained immature stages of one, two, and three wasp species under multiparasitism conditions. Species: (a) *Opius hirtus*, (b) *Doryctobracon crawfordi*, (c) *Utetes anastrephae*, (d) *U. anastrephae/O. hirtus*, (e) *D. crawfordi/O. hirtus*, (f) *D. crawfordi/U. anastrephae*, and (g) *D. crawfordi/U. anastrephae/O. hirtus*.

foraging in solitary was suggested in a previous study by Ayala *et al.* (2018) and is confirmed here.

Interestingly, when the females of any of the three parasitoid species foraged alone, more than 40% of the hosts exposed exhibited more than one scar, even though unparasitized hosts were still available. This high percentage of hosts with more than one scar can be attributed to the probing activity of females during host discrimination and not to an act of self-superparasitism, since no correlation was observed between number of scars per puparium and number of immature stages found within them. This contrasts with what has been reported in the exotic species *Di. longicaudata* (Montoya *et al.*, 2000; González *et al.*, 2007) and *D. tryoni* (Cameron) (Ayala *et al.*, 2014), which tend to superparasitize hosts even when foraging alone.

The findings described above lend particular importance since we can infer that females of these native species have an intrinsic capacity for host discrimination that makes them very selective, so they do not appear to be influenced by external factors such as the number of non-parasitized hosts available. According to Godfray (1994), egg load in female parasitoids is a factor that normally correlates with discrimination capacity, where a limitation in the production of eggs makes females more selective. However, the females of the species in this study are synovigenic, and are categorized by Cicero et al. (2011) as species limited by time. Such species often produce a high number of eggs or have the ability to replace them quickly, thereby making their reproductive success proportional to the number of hosts they can attack during their life-time (Stephens and Krebs, 1986; Charnov and Stephens, 1988). This type of species can use the strategy of superparasitizing their hosts or reabsorb their eggs, which increases their life expectancy (Rivero-Lynch and Godfray, 1997).

Chemical signals, such as marking pheromones, or different chemical secretions that accompany the process of oviposition can actively influence the process of host selection and facilitate the recognition of previously parasitized hosts (Vet and Dicke, 1992). The female selection process observed in these three species could be related to the detection of marking pheromones or substances injected during the process of oviposition (e.g., venoms and associated viruses), which seemingly altered or even diminished the quality of the host by causing internal changes (Mackauer, 1990; Gauthier and Monge, 1999; Outreman et al., 2001). This contrasts with an interspecific facilitation effect of superparasitism suggested in some studies (Cusumano et al., 2016). The substances introduced into the hosts during oviposition play roles in conditioning the physiology of the host in order to facilitate the development of the parasitoids (Moreau and Asgari, 2015). Another effect of these substances is

the permanent or temporary paralysis of the hosts (Vinson and Iwantsch, 1980), as is the case of the three studied species, which produced mean transient paralysis from 13 to 24 min in stung hosts (Ayala *et al.*, 2018). This facilitates oviposition without the parasitoid wasps being subjected to the defensive behavior of the hosts (Vinson and Iwantsch, 1980), although it could also be a strategy to avoid competition by delaying the moment at which a host can be detected and attacked again (Ayala *et al.*, 2018). It is also possible that the experience of first oviposition contact with a non-parasitized larva, and subsequently with a previously parasitized larva, is another factor that favored the capacity for discrimination observed in the females, since it was demonstrated that *Di. longicaudata* females with oviposition experience tended to superparasitize hosts less frequently than naive ones (Montoya *et al.*, 2003).

Competition for hosts in the presence of conspecifics seems to promote superparasitism in some cases. Females of Di. longicaudata and D. tryoni increase their superparasitic activity when faced with competition (Montoya et al., 2000; Ayala et al., 2014), with no adverse effects on the longevity and fecundity of their progeny (González et al., 2010; Avala et al., 2014). In the case of the native species in this study, no direct physical interaction between females was observed during the experiments, and the presence of conspecific females foraging in the same patch had no effect on superparasitism in O. hirtus. However, in D. crawfordi and U. anastrephae, a direct relationship was observed between number of scars and number of immature stages within the puparia of A. ludens, which suggests an increase in the propensity to superparasitize hosts in these two species when conspecifics are present in the same patch. Two nonexclusive alternative strategies that may favor superparasitism are infanticide (including ovicide) and the fact that the host' immune response may be compromised by the presence of multiple eggs (van Alphen and Visser, 1990; Godfray, 1994). In addition to discriminating a previously parasitized host, females of some species have the capacity to identify the sex of the previously oviposited egg and assign the sex of her progeny accordingly, as is the case of the species Anisopteromalus calandrae Howard (Hymenoptera: Pteromalidae) (Lebreton et al., 2010).

According to King (1993), sex ratio in parasitoid wasps can vary depending on three conditions: (a) size and quality of host, (b) superparasitism, and (c) incidence of various females foraging in the same site. In some instances, superparasitism may favor the proportion of females. This is explained by the better ability of female larvae to compete, as in the case of *Eupelmus vuilleti* (Hymenoptera: Eupelmidae) (Darrouzet *et al.*, 2003). Van Baaren *et al.* (1999) reported similar results in *Anaphes victus* Huber (Hymenoptera: Mymaridae), where females were better competitors than males in superparasitism conditions.

For the exotic species *Di. longicaudata* and *D. tryoni*, superparasitism has been positively related to the production of females, with a resulting female-biased sex ratio at the population level (González *et al.*, 2007; Montoya *et al.*, 2011), whereas in our study with three native species, the observed superparasitism had no effect on the production of females. Our results also show that, in the cases of *O. hirtus* and *U. anastrephae*, the number of oviposition scars per host had no effect on the longevity and fecundity of the progeny, unlike in *D. crawfordi*, where lower longevity and lower fecundity were reported in females emerging from puparia with more than one scar (Ayala *et al.*, 2014).

Interspecific competition in parasitoids can arise as a result of exploitation of a common resource (May and Hassell, 1981; Schoener, 1983; Bográn et al., 2002; Wang et al., 2008), and may occur while both females forage for hosts and later while larvae develop within the host (Harvey et al., 2013, Cusumano et al., 2016). The greater the similarities existing among foraging habits, the higher the probability of competition caused by oviposition and feeding of closely related parasitoid species. According to the results of our multiparasitism experiment, the species with apparently higher competitive abilities under intrinsic conditions was U. anastrephae, since most of the dissected hosts contained a single immature stage corresponding to that species, although similar percentages of subsequent emergence of adults of U. anastrephae and D. crawfordi were observed. Traits indirectly related to competition of adult females (host discovery efficiency, attack rate, egg load) might have also affected the observed results. The more frequent presence of U. anastrephae larvae in the dissected hosts could be attributed to higher female competitive abilities. For instance, the instantaneous rate of discovery and handling time reported by Poncio et al. (2016) for this species were greater than those reported by Montoya et al. (2000) for Di. longicaudata. Ayala et al. (2018) also noted that U. anastrephae attacked more hosts than D. crawfordi and O. hirtus under similar experimental conditions. Moreover, the first larval instar of U. anastrephae exhibits larger mandibles than the other two species, a trait that could allow this species to outcompete better the others (Murillo et al., 2016). Extrinsic competition depends mainly on adult density and female wasp: host larvae ratio (Poncio et al., 2016). We did not observe consistent interactions among foraging females of the different species, possibly due to the low female density in our experiments (three females, one female per parasitoid species) and a higher number of available hosts in the experimental arena. According to Harvey et al. (2013), multiparasitism conditions are more advantageous for species with a rapid development, or species that attack their hosts during earlier stages of development.

Even when superparasitism in parasitoid wasps is common in nature and can be adaptive under specific conditions, the native species studied here tended to avoid superparasitizing hosts, probably because of the close relationship they maintain with hosts of the genus *Anastrepha*. The tendency to superparasitize has also been observed in species such as *Trichogramma* spp. under mass rearing conditions, where a high proportion of females leads to superparasitism, and the sexual proportion of the progeny favors the production of males (Smith, 1996). Parasitoid: host ratios and superparasitism have been related as key factors influencing sex ratio when insects are reared under laboratory conditions (Waage *et al.*, 1985; King, 1993; Montoya *et al.*, 2011). Thus, this kind of studies are relevant when natural enemies are evaluated as biocontrol agents. Our study also provides sound data on the potential and limitations of mass rearing of each species.

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References

- Aluja M, Sivinski J, Ovruski S, Guillen L, Lopez M, Cancino J, Torres Anaya A, Gallegos-Chan G and Ruiz L (2009) Colonization and domestication of seven species of native New World hymenopterous larval-prepupal and pupal fruit fly (Diptera: Tephritidae) parasitoids. *Biocontrol Science and Technology* 19, 49–79.
- Ayala A, Martinez AM, Figueroa I, Pineda S, Miranda M, Liedo P and Montoya P (2014) Superparasitism strategies by a native and an exotic parasitoid species attacking the Mexican fruit fly, Anastrepha ludens (Diptera: Tephritidae). Biocontrol Science and Technology 24, 925–935.
- Ayala A, Pérez-Lachaud G, Toledo J, Liedo P and Montoya P (2018) Host acceptance by three native braconid parasitoid species attacking larvae of the Mexican fruit fly, *Anastrepha ludens* (Diptera, Tephritidae). *Journal of Hymenoptera Research* 63, 33–49.
- Bakker K, van Alphen JJM, van Batenburg FHD, van der Hoeven N, Nell HW, van Strien-van Liempt WTFH and Turlings TCJ (1985) The function of host discrimination and superparasitization in parasitoids. Oecologia 67, 572–576.
- Bográn CE, Heinz KM and Ciomperlik MA (2002) Interspecific competition among insect parasitoids: field experiments with whiteflies as hosts in cotton. *Ecology* 83, 653–668.
- Charnov EL and Stephens DW (1988) On the evolution of host selection in solitary parasitoids. *The American Naturalist* **132**, 707–722.
- Cicero L, Sivinski J, Rull J and Aluja M (2011) Effect of larval host food substrate on egg load dynamics, egg size and adult female size in four species of braconid fruit fly (Diptera: Tephritidae) parasitoids. *Journal of Insect Physiology* 57, 1471–1479.
- Cusumano A, Peri E and Colazza S (2016) Interspecific competition/facilitation among insect parasitoids. Current Opinion in Insect Science 14, 12–16.
- Darrouzet E, Imbert E and Chevrier C (2003) Self-superparasitism consequences for offspring sex ratio in the solitary ectoparasitoid *Eupelmus vuilleti. Entomologia Experimentalis et Applicata* 109, 167–171.
- FAO/IAEA/USDA (2019) Product Quality Control for Sterile Mass-Reared and Released Tephritid Fruit Flies, Version 7.0. Vienna, Austria: International Atomic Energy Agency. 148 pp.
- García-Medel D, Sivinski J, Díaz-Fleischer F, Ramírez-Romero R and Aluja M (2007) Foraging behavior by six fruit fly parasitoids (Hymenoptera: Braconidae) released as single- or multiple-species cohorts in field cages: influence of fruit location and host density. *Biological Control* 43, 12–22.
- Gauthier N and Monge JP (1999) Could the egg itself be the source of the oviposition deterrent marker in the ectoparasitoid *Dinarmus basalis*? *Journal of Insect Physiology* 45, 393–400.
- **Godfray HCJ** (1994) *Parasitoids: Behavioral and Evolutionary Ecology.* Princeton: Princeton University Press.
- González PI, Montoya P, Pérez-Lachaud G, Cancino J and Liedo P (2007) Superparasitism in mass reared *Diachasmimorpha longicaudata* (Ashmead) (Hymenoptera: Braconidae), a parasitoid of fruit flies (Diptera: Tephritidae). *Biological Control* **40**, 320–326.

- González PI, Montoya P, Pérez-Lachaud G, Cancino J and Liedo P (2010) Host discrimination and superparasitism in wild and mass-reared Diachasmimorpha longicaudata (Hym.: Braconidae) females. Biocontrol Science and Technology 20, 137–148.
- Harvey JA, Poelman EH and Tanaka T (2013) Intrinsic inter- and intraspecific competition in parasitoid wasps. *Annual Review of Entomology* 58, 333–351.
- Hernández-Ortíz V, Perez-Alonso R and Wharton RA (1994) Native parasitoids associated with the genus *Anastrepha* (Diptera: Tephritidae) in Los Tuxtlas, Veracruz, México. *Entomophaga* **39**, 171–178.
- Hubbard SF, Marris G, Reynolds A and Rowe GW (1987) Adaptive patterns in the avoidance of superparasitism by solitary parasitic wasps. *Journal of Animal Ecology* 56, 387–401.
- King BH (1993) Sex ratio manipulation by parasitoid wasps. In Wrensch DL and Ebbert MA (eds), *Evolution and Diversity of Sex Ratio in Insects and Mites*. New York: Chapman & Hall, pp. 418–441.
- Lawrence PO, Greany PD, Nation JL and Baranowski RM (1978) Oviposition behavior of *Biosteres longicaudatus*, a parasite of the Caribbean fruit fly, *Anastrepha suspensa*. *Annals of the Entomological Society of America* **71**, 253–256.
- Lebreton S, Chevrier C and Darrouzet E (2010) Sex allocation strategies in response to conspecifics' offspring sex ratio in solitary parasitoids. *Behavioral Ecology* **21**, 107–112.
- Lee ET and Wang JW (2003) Statistical Methods for Survival Data Analysis. New York, NY: Wiley.
- López M, Aluja M and Sivinski J (1999) Hymenopterous larval-pupal and pupal parasitoids of *Anastrepha* flies (Diptera: Tephritidae) in Mexico. *Biological Control* 15, 119–129.
- Mackauer, M. (1990) Host discrimination and larval competition in solitary endoparasitoids. In Mackauer M, Ehler LE and Roland J (eds), *Critical Issues in Biological Control*, Andover: Intercept, pp. 41–62.
- Mackauer M, Bai B, Chow A and Danyk T (1992) Asymmetric larval competition between two species of solitary parasitoid wasps: the influence of superparasitism. *Ecological Entomology* 17, 233–236.
- May RM and Hassell MP (1981) The dynamics of multiparasitoid-host interactions. *The American Naturalist* 117, 234–261.
- Montoya P, Liedo P, Benrey B, Cancino J, Barrera JF, Sivinski J and Aluja M (2000) Biological control of Anastrepha spp. (Diptera: Tephritidae) in mango orchards through augmentative releases of Diachasmimorpha longicaudata (Ashmead) (Hymenoptera: Braconidae). Biological Control 18, 216–224.
- Montoya P, Benrey B, Barrera JF, Zenil M, Ruiz L and Liedo P (2003) Oviposition behavior and conspecific host discrimination in *Diachasmimorpha longicaudata* (Hymenoptera: Braconidae), a fruit fly parasitoid. *Biocontrol Science Technology* **13**, 683–690.
- Montoya P, Cancino J, Pérez-Lachaud G and Liedo P (2011) Host size, superparasitism and sex ratio in mass-reared *Diachasmimorpha longicaudata*, a fruit fly parasitoid. *BioControl* 56, 11–17.
- Montoya P, Ruiz L, Pérez-Lachaud G, Cancino J and Liedo P (2013) Field superparasitism by *Diachasmimorpha longicaudata* attacking *Anastrepha* spp. larvae on mango fruits. *Biological Control* **64**, 160–165.
- **Moreau SJM and Asgari S** (2015) Venom proteins from parasitoid wasps and their biological functions. *Toxins* 7, 2385–2412.
- Murillo FD, Liedo P, Nieto-López MG, Cabrera-Mireles H, Barrera JF and Montoya P (2016) First instar larvae morphology of Opiinae (Hymenoptera: Braconidae) parasitoids of *Anastrepha* (Diptera: Tephritidae) fruit flies. Implications for interspecific competition. *Arthropod Structure & Development* 45, 294–300.
- Nelson JM and Roitberg BD (1995) Flexible patch time allocation by the leafminer parasitoid, *Opius dimidiatus*. *Ecological Entomology* **20**, 245–252.
- Orozco-Dávila D, Quintero L, Hernández E, Solís E, Artiaga T, Hernández R and Montoya P (2017) Mass rearing and sterile insect releases for the control of *Anastrepha* spp. (Diptera: Tephritidae) pests in Mexico a review. *Entomologia Experimentalis et Applicata* **164**, 76–187.
- **Outreman Y and Pierre JS** (2005) Adaptive value of host discrimination in parasitoids: when host defenses are very costly. *Behavioural Process* **70**, 93–103.
- Outreman Y, Le Ralec A, Plantegenest M, Chaubet B and Pierre JS (2001) Superparasitism limitation in an aphid parasitoid: cornicle secretion

avoidance and host discrimination ability. *Journal of Insect Physiology* 47, 339–348.

- **Ovruski S, Aluja M, Sivinski J and Wharton RA** (2000) Hymenopteran parasitoids on fruit-infesting Tephritidae (Diptera) in Latin America and the southern United States: diversity, distribution, taxonomic status and their use in fruit fly biological control. *Integrated Pest Management Reviews* **5**, 81–107.
- **Ovruski S, Colin C, Soria A, Oroño LE and Schliserman P** (2003) Introducción y producción en laboratorio de *Diachasmimorpha tryoni* y *Diachasmimorpha longicaudata* (Hymenoptera: Braconidae) para el control biológico de *Ceratitis capitata* (Diptera: Tephritidae) en la Argentina. *Revista de la Sociedad Entomologica de Argentina* **62**, 49–59.
- **Ovruski S, Wharton RA, Schliserman P and Aluja M** (2005) Abundance of *Anastrepha fraterculus* (Diptera: Tephritidae) and its associated native parasitoids (Hymenoptera) in "feral" guavas growing in the endangered northernmost Yungas forests of Argentina with an update on the taxonomic status of Opiinae parasitoids previously reported in this country. *Environmental Entomology* **34**, 807–818.
- Pijls JWAM, Hofker KD, van Staalduinen MJ and van Alphen JJM (1995) Interspecific host discrimination and competition in *Apoanagyrus* (*Epidinocarsis*) lopezi and A. (E.) diversicornis, parasitoids of the cassava mealybug *Phenacoccus manihoti. Ecological Entomology* 20, 326–332.
- Poelman EH, Gols R, Gumovsky AV, Cortesero AM, Dicke M and Harvey JA (2014) Food plant and herbivore host species affect the outcome of intrinsic competition among parasitoid larvae. *Ecological Entomology* 39, 693–702.
- Poncio S, Montoya P, Cancino J and Nava DE (2016) Determining the functional response and mutual interference of Utetes anastrephae (Hymenoptera: Braconidae) on Anastrepha obliqua (Diptera: Tephritidae) larvae for mass rearing purposes. Annals of the Entomological Society of America 109, 518–525.
- Pschorn-Walcher H (1987) Interspecific competition between the principal larval parasitoids of the pine sawfly, *Neodiprion sertifer* (Geoff.) (Hym.: Diprionidae). *Oecologia* 73, 621–625.
- Rivero-Lynch AP and Godfray HCJ (1997) The dynamics of egg production, oviposition and resorption in a parasitoid wasp. *Functional Ecology* 11, 184–188.
- Roitberg BD and Mangel M (1988) On the evolutionary ecology of marking pheromones. *Evolutionary Ecology* **2**, 289–315.
- **SAS Institute** (2007) *JMP Statistical Discovery Software*, version 7.0.1. Cary, NC: SAS Institute Inc.
- Schoener TW (1983) Field experiments on interspecific competition. The American Naturalist 122, 240–285.
- Sivinski J, Aluja M and López M (1997) Spatial and temporal distributions of parasitoids of Mexican Anastrepha species (Diptera: Tephritidae) within the canopies of fruit trees. Annals of the Entomological Society of America 90, 604–618.
- Sivinski J, Piñero J and Aluja M (2000) The distribution of parasitoids (Hymenoptera) of *Anastrepha* fruit flies (Diptera: Tephritidae) along an altitudinal gradient in Veracruz, Mexico. *Biological Control* **18**, 258–269.
- Smith SM (1996) Biological control with *Trichogramma*: advances, successes, and potential of their use. *Annual Review of Entomology* 41, 375–406.
- Statgraphics (2008) Statgraphics Centurion XV User Manual. Herndon, VA: Statgraphics.
- Stephens DW and Krebs JR (1986) Foraging Theory. Princeton, NJ: Princeton University Press.
- **Ueno T** (1994) Self-recognition by the parasitic wasp *Itoplectis naranyae* (Hymenoptera: Ichneumonidae). *Oikos* **70**, 333–339.
- van Alphen JJ and Visser ME (1990) Superparasitism as an adaptive strategy for parasitoids. Annual Review of Entomology 35, 59–79.
- van Baaren J, Landry BL and Boivin G (1999) Sex allocation and larval competition in a superparasitizing solitary egg parasitoid: competing strategies for an optimal sex ratio. *Functional Ecology* **13**, 66–71.
- van Dijken MJ and Waage JK (1987) Self and conspecific superparasitism by the egg parasitoid *Trichogramma evanescens*. *Entomologia Experimentalis et Applicata* 43, 183–192.
- van Lenteren JC (1981) Host discrimination by parasitoids. In Nordlund AD, Jones RL and Lewis WL (eds), *Semiochemicals: Their Role in Pest Control*. New York: John Wiley and Sons, pp. 153–179.

- Vet LEM and Dicke M (1992) Ecology of infochemical use by natural enemies in a tritrophic context. Annual Review of Entomology **37**, 141–172.
- Vinson SB (1976) Host selection by insect parasitoids. Annual Review of Entomology 21, 109–133.
- Vinson SB and Ables JR (1980) Interspecific competition among endoparasitoids of tobacco budworm larvae (Lep.: Noctuidae). *Entomophaga* 25, 357–362.
- Vinson SB and Iwantsch GF (1980) Host regulation by insect parasitoids. *The Quarterly Review of Biology* 55, 143–165.
- Visser ME, Luyckx B, Nell HW and Boskamp GJF (1992) Adaptive superparasitism in solitary parasitoids: marking of parasitized hosts in relation to the pay-off from superparasitism. *Ecological Entomology* **17**, 76–82.
- Waage JK, Carl KP, Mills NJ and Greathead DJ (1985) Rearing entomophagous insects. In Singh P and Moore RF (eds), *Handbook of Insect Rearing*, vol. 1. Amsterdam: Elsevier, pp. 45–46.
- Wang XY, Yang ZQ, Wu H and Gould JR (2008) Effects of host size on the sex ratio, clutch size, and size of adult *Spathius agrili*, an ectoparasitoid of emerald ash borer. *Biological Control* **44**, 7–12.