Neonatal DNA methylation and early-onset conduct problems: A genome-wide, prospective study

CHARLOTTE A. M. CECIL,^{*a*} ESTHER WALTON,^{*b*} SARA R. JAFFEE,^{*c*} TOM O'CONNOR,^{*d*} BARBARA MAUGHAN,^{*a*} CAROLINE L. RELTON,^{*b*} REBECCA G. SMITH,^{*e*} WENDY MCARDLE,^{*b*} TOM R. GAUNT,^{*b*} ISABELLE OUELLET-MORIN,^{*f*} AND EDWARD D. BARKER^{*a*}

^aKing's College London; ^bUniversity of Bristol; ^cUniversity of Pennsylvania; ^dUniversity of Rochester Medical Center; ^eExeter University; and ^fUniversity of Montreal

Abstract

Early-onset conduct problems (CP) are a key predictor of adult criminality and poor mental health. While previous studies suggest that both genetic and environmental risks play an important role in the development of early-onset CP, little is known about potential biological processes underlying these associations. In this study, we examined prospective associations between DNA methylation (cord blood at birth) and trajectories of CP (4–13 years), using data drawn from the Avon Longitudinal Study of Parents and Children. Methylomic variation at seven loci across the genome (false discovery rate < 0.05) differentiated children who go on to develop early-onset (n = 174) versus low (n = 86) CP, including sites in the vicinity of the monoglyceride lipase (*MGLL*) gene (involved in endocannabinoid signaling and pain perception). Subthreshold associations in the vicinity of three candidate genes for CP (monoamine oxidase A [*MAOA*], brain-derived neurotrophic factor [*BDNF*], and FK506 binding protein 5 [*FKBP5*]) were also identified. Within the early-onset CP group, methylation levels of the identified sites did not distinguish children who will go on to persist versus desist in CP behavior over time. Overall, we found that several of the identified sites correlated with prenatal exposures, and none were linked to known genetic methylation quantitative trait loci. Findings contribute to a better understanding of epigenetic patterns associated with early-onset CP.

Conduct problems (CP; e.g., fighting or stealing) are a major public health concern and a leading cause of youth treatment referral (Bywater, 2012). CP youth engage in antisocial and disruptive behaviors that result in significant distress to victims, impairment of life opportunities, and economic burden on judicial, healthcare, and social welfare services (Colman et al., 2009). In particular, an early-onset of CP (at or before the age of 10) designates a high-risk group of children, who show more severe and recidivistic patterns of antisocial behavior, higher levels of comorbid psychopathology (Barker, Oliver, & Maughan, 2010), increased risk for poor adult outcomes (Fergusson, Horwood, & Ridder, 2005; Odgers et al., 2007), and who account for the majority of criminal offences within a given community (Farrington, 2001). Consequently, early-onset CP are recognized as an important target for prevention and intervention efforts.

Like most complex phenotypes, early-onset CP are thought to result from the developmentally dynamic interplay of genetic and environmental influences. Studies have shown that the heritability of early-onset CP is moderate to high $(\sim 50\% - 80\%)$; see Barker, Cecil, Walton & Meehan, 2017, for a review) and that this genetic vulnerability interacts with environmental risk exposure (e.g., Jaffee et al., 2005). Exposure to adversity during pregnancy (e.g., maternal smoking, stress, and psychopathology) as well as the postnatal years (e.g., maltreatment and poverty) is frequent among this group of children, and thought to play an important role in the development and persistence of CP behavior (Barker & Maughan, 2009; Moffitt, 2006; Odgers et al., 2007, 2008). Yet, little is known about the biological mechanisms through which these influences confer increased risk for early-onset CP.

In recent years, DNA methylation (DNAm), an epigenetic mechanism that regulates gene expression (Jaenisch & Bird, 2003), has emerged as a potential mechanism underlying gene–environment interplay and disease susceptibility across

We are extremely grateful to all the families who took part in this study, the midwives for their help in recruiting them, and the whole Avon Longitudinal Study of Parents and Children (ALSPAC) team, which includes interviewers, computer and laboratory technicians, clerical workers, research scientists, volunteers, managers, receptionists, and nurses. We thank all involved with the ALSPAC DNA methylation, particularly the laboratory scientists and bioinformaticians who contributed considerable time and expertise to the data in this paper. The UK Medical Research Council and the Wellcome Trust (Grant 102215/2/13/2) and the University of Bristol provide core support for ALSPAC. This publication is the work of the authors, who will serve as guarantors for the contents of this paper. This research was specifically supported by National Institute of Child and Human Development Grant R01HD068437 (to E.D.B.). The first author (C.A.M.C.) is supported by Economic and Social Research Council Grant ES/N001273/1.

Address correspondence and preprint requests to: Charlotte A. M. Cecil or Edward D. Barker, Department of Psychology, King's College London, Institute of Psychiatry, Psychology and Neuroscience, De Crespigny Park, London, UK SE5 8AF; E-mail: charlotte.cecil@kcl.ac.uk or ted.barker@ kcl.ac.uk.

the life span. Specifically, studies have shown that (a) DNAm patterns are under significant genetic control, as evidenced by the discovery of a large number of methylation quantitative trait loci (mQTL; Gaunt et al., 2016; Jones, Fejes, & Kobor, 2013); and (b) DNAm is also sensitive to environmental influences (McGowan & Roth, 2015), including numerous nutritional, chemical, physical, and psychosocial exposures occurring in utero (e.g., Monk, Spicer, & Champagne, 2012; Richmond et al., 2015; Tobi et al., 2014) and postnatally (Lutz & Turecki, 2014; Szyf & Bick, 2013; Tyrka et al., 2015). In turn, DNAm patterns have been shown to play a central role in neurobiological and developmental processes (e.g., neurogenesis, synaptic plasticity, learning, and memory; Murgatroyd & Spengler, 2011), and aberrations in DNAm have been implicated in a range of negative outcomes, including childhood psychopathology and stress-related psychiatric disorders (Klengel, Pape, Binder, & Mehta, 2014; van Mil et al., 2014; Weder et al., 2014). Consequently, it has been suggested that DNAm may represent a molecular pathway through which environmental exposures become translated into phenotypic variation, conferring increased susceptibility to mental and physical health problems (Kofink, Boks, Timmers, & Kas, 2013; Lewis & Olive, 2014).

For example, experimental studies in animals have shown that exposure to early adversity (e.g., prenatal stress and low postnatal care) can cause alterations in offspring DNAm (e.g., in hypothalamus–pituitary–adrenal axis genes), driving differences in the offspring's behavioral response to future stressors (Turecki & Meaney, 2016; Weaver et al., 2004). Because DNAm marks can be passed on during cell division (and in some cases across generations), they can lead to long-term programming of gene expression, with downstream effects on stress sensitivity, behavior, and disease risk (Rodgers & Bale, 2015; Zannas & West, 2014). In the context of CP, epigenetic alterations may thus represent a mechanism through which genetic and environmental factors intersect, influencing developmental trajectories of childhood aggression and antisocial behavior (Tremblay & Szyf, 2010).

Thus far, only a handful of studies in humans have examined associations between DNAm and CP-related phenotypes. Of these, the majority have been carried out by a research group comparing patterns of DNAm (extracted from peripheral blood samples collected at ages 26–28) between adult males with a childhood history of chronic physical aggression (ages 6-15) and controls who maintained low levels of physical aggression during the same time period, based on data from a longitudinal cohort (Booij et al., 2010; Guillemin et al., 2014; Provençal et al., 2013, 2014; Wang et al., 2012). Using this case-control design, these studies reported that childhood aggression associated with (a) higher levels of solute carrier family C6, member 4 (SLC6A4) gene promoter methylation and lower in vivo levels of brain serotonin synthesis (Booij et al., 2010; Wang et al., 2012); (b) DNAm levels in a set of genes involved in cytokine function and inflammation (Provençal et al., 2013); and (c) across a large number of gene promoter regions in an epigenome-wide

scan (Provençal et al., 2014), a finding that was later extended to a small sample of adult females as well (Guillemin et al., 2014). Another set of studies based on a sample of antisocial boys diagnosed with oppositional defiant disorder or conduct disorder (4–16 years) have also documented an association between severity of callous–unemotional traits (e.g., low capacity for empathy, lack of guilt, and shallow affect) and DNAm levels (obtained from peripheral blood) across several candidate genes, including lower DNAm of the 5-hydroxytryptamine (serotonin) receptor 1B gene (*HTR1B*; Moul, Dobson-Stone, Brennan, Hawes, & Dadds, 2015), and higher DNAm of the oxytocin receptor gene (*OXTR*; Dadds et al., 2014), which in turn related to lower circulating oxytocin levels.

Despite these promising findings, research to date has been limited in a number of important ways. First, no study to our knowledge has specifically investigated DNAm patterns associated with early-onset CP, which designates a subgroup of children at high risk for negative adult outcomes. Second, the available research has been primarily cross-sectional or retrospective with regard to DNAm (i.e., associating behavioral phenotypes in childhood to DNAm later in development). As such, it is currently unclear whether observed alterations in DNAm patterns represent a risk factor for and/or a consequence of CP. In other words, it has not been possible to establish whether DNAm is a prospective risk for CP or, alternatively, the biological correlate of a chronic antisocial lifestyle. Third, there is a lack of studies integrating measures of the environment, which has precluded the possibility of assessing the epigenome in relation to *both* risk exposure and psychiatric phenotypes.

To address the above gaps, research from our group has begun to assess prospective associations between environmental risks, DNAm at repeated time points (birth and childhood), and CP-related outcomes, using data from the Avon Longitudinal Study of Parents and Children (ALSPAC; Fraser et al., 2013). For example, in children with early-onset CP and low internalizing problems, we found that DNAm levels in the promoter region OXTR at birth (but not at later points in childhood) associated with higher risk exposure in utero, reduced experience of victimization during childhood (i.e., indicative of an "evocative epigenetic-environment correlation") and more severe callous-unemotional traits at age 13 (Cecil et al., 2014). In addition, in early-onset CP, we found that a sugar- and fat-rich diet during pregnancy prospectively associated with higher DNAm of the intrauterine growth factor gene 2 (IGF2) at birth (but not during childhood), which, in turn, associated with more severe co-occurring attention-deficit/hyperactivity disorder symptoms (Rijslasdaam, Cecil, et al., 2016). The temporal specificity around birth observed across these and other studies (e.g., Cecil et al., 2016; Walton et al., 2016) may reflect a number of factors, including large-scale variability in DNAm patterns across developmental eras (Gaunt et al., 2016), differences between the tissues examined at each time point (i.e., cord blood at birth vs. whole blood in childhood), as well as the influence of developmentally specific genetic effects on symptom trajectories (e.g., Pingault, Rijsdijk, Zheng, Plomin, & Viding, 2015). In addition, findings may lend support for the developmental origins of health and disease hypothesis (Barker, 2007; Nigg, 2016), whereby methylation changes at birth may represent a more reliable proxy for intrauterine risk exposures (compared to DNAm in childhood), and associated perturbations in fetal development. However, the extent to which DNAm patterns across the genome at birth may prospectively associate with an early-onset of CP has yet to be examined.

In addition to advancing knowledge of potential biological differences between children with early-onset versus low CP, the study of DNAm may also help us to better understand why early-onset children themselves show considerable heterogeneity in their developmental trajectories of CP. Studies have found that, despite being considered a high-risk group as a whole, less than 50% of children with an early-onset of CP will actually continue to exhibit these problems into adolescence (i.e., early-onset persisting trajectory; EOP), while the rest will remit to near-typical levels of CP (i.e., childhood-limited trajectory [CL]; Barker & Maughan, 2009; Odgers et al., 2008). This heterogeneity poses a challenge for accurate diagnostic screening, clinical classification, and treatment formulation, as well as raising important questions about what factors may underlie these increasingly divergent trajectories of CP within early-onset children (i.e., in line with the concept of multifinality; Cicchetti & Rogosch, 1996). So far, evidence regarding early-life predictors of EOP versus CL trajectories has been somewhat mixed (see Moffitt et al., 2008, for a review), but generally suggests that the difference may be more quantitative than qualitative, with preand postnatal risk exposures found to be particularly high for EOP children, compared to their CL peers (i.e., EOP > CL > low CP; Barker & Maughan, 2009). However, it is currently unknown whether such a "graded" difference on the same risk factors may also exist at a biological level. The analysis of DNAm patterns between early-onsets who follow different developmental trajectories of CP (i.e., EOP vs. CL) may therefore help to refine the phenotype and contribute to a better understanding of heterogeneity within this highrisk group of children.

The Present Study

In light of the above research gaps, we made use of data from the ALSPAC cohort spanning pregnancy to late childhood to address three key aims. First, we examined whether DNAm patterns at birth (measured at >450,000 sites across the genome) prospectively differentiate children who will go on to develop early-onset (n = 174) versus low CP (n = 86). Second, to explore heterogeneity in CP trajectories within the early-onset group, we tested whether these DNAm patterns further distinguish between children who will persist (EOP; n = 91) versus desist (CL; n = 83) in CP over time. Third, we investigated potential genetic and environmental influences associated with the identified DNAm sites, including known genetic mQTLs and measures of prenatal adversity that have been previously linked to early-onset CP (e.g., maternal diet, smoking, and exposure to stressful events).

Methods

Sample

The Epigenetic Pathways to Conduct Problems Study consists of a subsample of youth (n = 321; 50% female) drawn from the ALSPAC and partially overlapping with a larger study of DNA methylation, the Accessible Resource for Integrated Epigenomics Studies (http://www.ariesepigenomics. org.uk; Relton et al., 2015). Specifically, children were included if they (a) have available measures of DNAm at two or more time points and (b) follow previously established trajectories of CP between 4 and 13 years of age (see Measures section below; Barker & Maughan, 2009). Children in this study were white Caucasian (6 removed from analyses due to missing ethnicity data).

ALSPAC is an ongoing epidemiological study of 14,541 pregnant women resident in Avon, United Kingdom, with expected dates of delivery April 1, 1991, to December 31, 1992. Of these initial pregnancies, there was a total of 14,676 fetuses, resulting in 14,062 live births and 13,988 children who were alive at 1 year of age (Fraser et al., 2013). When compared to 1991 National Census Data, the ALSPAC sample was found to be broadly similar to the United Kingdom population as a whole (Boyd et al., 2013). The study website contains details of all the data that are available (http://www.bris. ac.uk/alspac/researchers/data-access/data-dictionary). Ethical approval for the study was obtained from the ALSPAC Law and Ethics Committee and the Local Research Ethics Committees.

Measures

CP. CP trajectories were previously identified and validated in the whole ALSPAC sample using growth mixture models based on repeated measures of the Strengths and Difficulties Questionnaire conduct problems subscale (ages 4, 7, 8, 10, 12, and 13; Goodman, 2001), with boys and girls modeled in the same trajectories (Barker & Maughan, 2009). The original analysis identified four CP trajectories: (a) an early-onset persistent (EOP; 9%) group of children, who show high levels of CP early on and remain high across time; (b) a childhood-limited (CL; 15%) group who also show an early-onset of CP, but desist around adolescence; (c) an adolescent-onset group who only show CP behaviors later on (AO; 12%); and (d) a low CP group, who maintain low levels of CP throughout (low CP; 64%). The percentage of youth identified in the CP trajectories in ALSPAC is consistent with reported prevalence estimates in the general population (4%-20%); Egger & Angold, 2006; Kim-Cohen et al., 2003). In contrast to the overall ALSPAC sample, our epigenetic subsample featured a near equal number of youth in each trajectory, and was therefore enriched for CP (EOP: n = 91, 28%; CL: *n* = 83, 26%; AO: *n* = 61, 19%; low CP: *n* = 86, 27%). In this study, early-onset CP youth were defined as youth who follow either an EOP or CL trajectory. The AO group was excluded from the main analyses for two reasons. First, although AO and low children appear phenotypically similar in CP levels during the developmental period under investigation, they have been shown to differ in levels of environmental risk exposure and temperamental features (Barker & Maughan, 2009), so that we decided against combining AOs within the low (i.e., typical) group. Second, we did not include AOs as a separate comparison group in the epigenome-wide analyses due to concerns over statistical power and multiple testing burden resulting from the increase in number of comparisons.

DNA extraction and methylomic profiling. DNA was obtained from cord blood samples at birth. Each sample of 500 ng genomic DNA was bisulfite converted using an EZ-DNA methylation kit (Zymo Research, Orange, CA) and DNAm quantified using the Illumina HumanMethylation450 BeadChip (450K array; Illumina) with arrays scanned using an Illumina iScan (software version 3.3.28). The Illumina 450K array comprises >485,000 probes, each quantifying DNAm at a specific cytosine nucleotide-phosphate-guanine nucleotide (CpG) site, covering 99% of Reference Sequence genes, with an average of 17 CpG sites per gene region (distributed across promoter, 5' untranslated region (UTR), first exon, gene body, and 3'UTR regions). The BeadChip covers 96% of CpG islands, with additional coverage in island shores and their flanking regions. Initial quality control of data generated was conducted using GenomeStudio (Version 2011.1) to determine the status of staining, extension, hybridization, target removal, bisulfite conversion, specificity, nonpolymorphic, and negative controls. Furthermore, multiple checks for sample mismatch were carried out to ascertain that all cord blood samples in the study reflected infant DNA as opposed to maternal DNA. Specifically, samples were checked by calculating concordance with (a) genomewide association data from the same participants, (b) single nucleotide polymorphism (SNP) probes on the Illumina 450k array across mother and child, and (c) sex. Data were quantile normalized using the dasen function as part of the wateRmelon package (wateRmelon_1.0.3; Pidsley et al., 2013) within the R statistical analysis environment and batch corrected using the ComBat package (Johnson, Li, & Rabinovic, 2007). Probes previously reported to hybridize to multiple genomic regions or containing a SNP at the single base extension site were removed from subsequent analyses (Chen et al., 2013; Price et al., 2013), in addition to the 65 SNPs used for sample identification on the array (total probes removed 72,067). For each probe, DNAm levels were indexed by beta values (i.e., the ratio of methylated signal divided by the sum of the methylated and unmethylated signal [M/M + U]).

Prenatal environment. Multiple environmental influences were included that have been previously linked to early-onset CP, including maternal prenatal diet, smoking, alcohol use, and exposure to stressful events (Barker & Maughan, 2009). Information on dietary patterns was drawn from maternal ratings of the Food Frequency Questionnaire at 32 weeks gestation, where factor scores of "healthy" (i.e., high on fish, nonmeat protein, and vegetables) and "unhealthy" (i.e., high on processed and junk food) diet were extracted using confirmatory factor analysis, both of which showed acceptable model fit (for full details, see Barker, Kirkham, Ng, & Jensen, 2013). Maternal smoking and alcohol use during the first trimester of pregnancy were measured via maternal ratings, using a yes/no binary variable for smoking, and a 4-point scale for alcohol use (never to daily). With regard to stress exposure, we included cumulative risk scores of prenatal (18 to 32 weeks) adversity, based on 56 dichotomous items from the Life Events inventory (adapted in ALSPAC based on the work of Brown, Sklair, Harris, & Birley, 1973, and Barnett, Hanna, & Parker, 1983) and the Family Adversity Index (Wolke, Steer, & Bowen, 2004). Briefly, items were organized into four conceptually distinct but related risk domains, and summed to create the following cumulative scores: (a) life events (22 items; e.g., death in family or accident), (b) contextual risks (16 items; e.g., poor housing or financial problems), (c) maternal risks (10 items; e.g., psychopathology or criminal behavior), and (d) interpersonal risks (8 items; e.g., partner abuse or family conflict). The full list of items contained in these cumulative scores is provided in online-only supplementary Table S.1, and further information is available elsewhere (Cecil et al., 2014; Rijlaarsdam, Pappa, et al., 2016).

Statistical analyses

All analyses were performed within R (Version 3.0.1; R Core Team, 2014) on DNAm data regressed for sex and cell-type proportions (CD8 T lymphocytes, CD4 T lymphocytes, natural killer cells, B lymphocytes, and monocytes), estimated using the reference-based approach detailed in Houseman et al. (2012).

Step 1: Are there differences in neonatal patterns of DNAm between children who go on to develop early-onset versus low CP?

In the first step, a methylome-wide association analysis was performed using the CpGAssoc package (Barfield, Kilaru, Smith, & Conneely, 2012) to identify DNAm sites at birth that differentiate children who go on to develop early-onset (i.e., EOP and CL) versus low CP. False discovery rate (FDR) correction was implemented across all probes (i.e., genome-wide significance threshold set at q < 0.05). Effect sizes were calculated and interpreted following Cohen's guidelines, where an effect of 0.20 represents a small effect, an effect of 0.50 is a medium effect, and an effect of 0.80 is a large effect (Cohen, 1988).

Genome-wide significant loci were then uploaded to the UCSC genome browser (GRCh37/hg19 assembly; Kent et al., 2002) to explore their potential functional relevance, by comparing their genomic location to that of key regulatory elements recorded in the Encyclopedia of DNA Elements (ENCODE) database (http://genome.ucsc.edu/ENCODE/). These included (a) transcription factor binding sites, which are DNA regions where one or more specific proteins responsible for regulating transcription bind to (data generated on 161 transcription factors in 91 cell types via ChIP-seq); (b) DNase I hypersensitivity clusters, which tend to coincide with regulatory regions of the gene and indicate an open chromatin state that facilitates transcription (based on data from 125 cell types); and (c) *histone marks*, which are chemical modifications that influence how tightly packaged the DNA is around histone protein, thereby regulating how accessible DNA is for transcription (data available for seven cell types; however, given that DNAm patterns can be tissue specific, we only selected here the three cell types that are most relevant to cord blood; i.e., blood [GM12878, K562] and human umbilical vein endothelial [HUVEC] cells). To identify enriched biological pathways, we also analyzed probes that were significant at p < .001 using an optimized gene ontology method that corrects for multiple potential confounds, including background probe distribution and gene size (see onlineonly supplementary text).

Step 2: Within the early-onset group, do these DNAm sites also differentiate children who will persist versus desist in CP over time?

As a second step, we examined whether the identified DNAm sites may help to explain heterogeneity in developmental trajectories within the early-onset CP group. Specifically, we used one-way analysis of covariance models to test whether DNAm levels across the sites identified in Step 1 also differentiate early-onsets who will go on to follow a persisting (i.e., EOP) versus desisting (i.e., CL) trajectory of CP behavior over time, controlling for sex and cell type proportions.

Step 3: Are the identified DNAm sites associated with genetic and environmental factors?

As a third and final step, we explored potential genetic and environmental factors that may influence the DNAm sites associated with early-onset CP (i.e., identified in Step 1). As our sample was underpowered to directly examine genetic polymorphisms (i.e., SNPs) affecting DNAm, we used the mQTLdb resource (http://www.mqtldb.org/) to search for known mQTLs associated with our DNAm sites of interest. The mQTLdb database contains the results of a large-scale study based on the Accessible Resource for Integrated Epigenomics Studies sample in ALSPAC (from which our subsample is derived), characterizing genome-wide significant *cis* effects (i.e., SNP within ± 1000 base pairs of the DNAm site) and *trans* effects (i.e., ± 1 million base pairs) on DNAm levels across Illumina 450k probes at five different life stages, including cord blood DNAm at birth (Gaunt et al., 2016). Here, we searched for mQTLs based on results from the conditional genome-wide complex trait analysis, which was used to identify mQTLs with the most representative, independent effect on each DNAm site in order to account for linkage disequilibrium (Gaunt et al., 2016). With regard to environmental influences, we examined associations between DNAm levels in the identified sites and prenatal exposures (i.e., maternal diet, smoking, alcohol use, and stressful events) using Pearson bivariate correlations. Again, the significance threshold was set at an FDRcorrected level of q < 0.05 to adjust for multiple comparisons.

Results

Neonatal DNA methylation and early-onset CP

The Q-Q plot and Manhattan plot for the epigenome-wide association analysis at birth are displayed in Figure 1a and b. Table 1 shows the seven differentially methylated sites (q < 0.05) between early-onset versus low CP groups, as identified via our genome-wide analysis. Group differences were medium to large in effect size based on Cohen's guidelines (d = 0.66-0.84), and ranged from 4% to 8% in terms of mean DNAm difference. Of note, all seven sites remained differentially methylated after additionally adjusting for gestational age. DNAm levels across these sites were all significantly intercorrelated (both positive and negative correlations observed; see online-only supplementary Table S.2). Three of the sites had genic annotations (Figure 1c): (a) cg02503763, located in the promoter regulatory region of MGLL, a gene implicated in endocannabinoid signaling and nociperception (Iwasaki, Ishiguro, Higuchi, Onaivi, & Arinami, 2007); (b) cg21579239, annotated to the 5'UTR region of tau tubulin kinase 2 (TTBK2), a gene involved in tau protein phosphorylation, of which mutations have been linked to neurodegenerative disorders (Liao, Yang, Weng, Kuo, & Chang, 2015); and (c) cg16006965, located in the promoter regulatory region of germinal center expressed transcript 2 (GCET2), a gene implicated in immune function (Pan et al., 2007). The other four differentially methylated sites identified at birth were distal to annotated transcripts (see online-only supplementary Figure S.1 for boxplots): cg12628061 located between AK127270 (-407 kb) and phosphatidic acid phosphatase type 2B (PPAP2B; -507 kb), cg02131674 located closest to brain abundant membrane attached signal protein 2 (BASP1; -123 kb), cg15384400 located between ArfGAP with RhoGAP domain, ankyrin repeat and pH domain 2 (ARAP2; -12 kb) and death domain containing 1 (DTHD1; +26 kb), and cg07128473 located between tandom repeat S1 (TRPS1; -210 kb) and eukaryotic translation initiation factor 3 subunit (EIF3H; +876 kb). In all cases, except for probe cg15384400, methylation of these sites was *lower* in the early-onset versus low CP groups. These seven sites were then viewed in Genome Browser for functional characterization, based on ENCODE data on regu-



Figure 1. (Color online) Methylomic variation at birth prospectively associated with early-onset versus low conduct problems in childhood. Epigenome-wide associations between DNA methylation at birth and early-onset versus low conduct problems, including (a) Q–Q plot, (b) Manhattan plot, and (c) scatterplots of the top three differentially methylated sites with genic annotations. (d) Significantly enriched biological processes at birth are shown, based on gene ontology analysis of genes annotated to probes that associate with early-onset versus low conduct problems (p < .001). Circles represent gene ontology terms that survive false discovery rate correction and contain at least two genes. The *X* axis represents $-\log(10) p$ values. The opacity of the circles indicates level of significance (darker = more significant). The size of the circles indicates the percentage of genes in our results for a given pathway compared to the total number of genes in the same pathway (i.e., larger size = larger %).

Table 1. DNA methylation sites at birth that differentiate early-onset versus low CP children (genome-wide significance; q < 0.05)

Probe	Gene	Chr	Genomic Location	Position	р	q (FDR)	Direction	Early-Onset CP Mean	Low CP Mean	% Diff	Cohen d
cg12628061		1	_	56453730	2.47E-09	1.00E-03	Ļ	0.77	0.81	4	0.84
cg02131674		5		17341065	1.64E-08	3.00E-03	į	0.77	0.81	4	0.78
cg15384400		4	_	36257279	5.09E-08	7.00E-03	↑	0.26	0.21	5	0.79
cg07128473		8		116891579	1.00E-07	0.01	Ļ	0.83	0.86	3	0.76
cg02503763	MGLL	3	TSS1500	127542386	1.23E-07	0.01	į	0.38	0.46	8	0.73
cg21579239	TTBK2	15	5'UTR	43211292	1.40E-07	0.01	į	0.74	0.78	4	0.73
cg16006965	GCET2	3	TSS1500	111852325	7.85E-07	0.04	Ļ	0.53	0.6	7	0.66

Note: CP, conduct problems; FDR, false discovery rate. Arrows represent the direction of methylation differences, where \uparrow indicates higher methylation levels in early-onset versus low CP children, while \downarrow indicates lower methylation levels in early-onset versus low CP children. For the Cohen *d* values, 0.20 is a small effect size, 0.50 is a medium effect size, and 0.80 is a large effect size.

latory elements. Four of the DNAm loci coincided with transcription factor binding sites, including MGLLcg02503763 and GCET2_{cg16006965}, and implicated a number of shared transcription factors, including MAX (4/4 loci), POLR2A, RUNX3, and EP300 (3/4 loci). The majority of DNAm sites also overlapped with histone marks (6/7 loci) and DNAse I hypersensitive clusters (4/7 loci), suggesting that they are located in genomic regions that are likely to play regulatory roles in transcription. Gene ontology analysis ($n_{\text{probes}} =$ 1,101; $n_{\text{genes}} = 758$) further indicated that DNAm sites (at birth) that differentiate between CP groups are annotated to genes involved in a range of biological processes, including regulation of fat cell differentiation and immune system process, response to epinephrine and wounding, as well as forebrain generation of neurons and CNS neuron differentiation (2.95E-15 Figure 1d). GO results for thetop 20 enriched biological pathways (including those displayed in Figure 1d) are provided in online-only supplementary Table S.3.

In light of the above findings, we conducted two follow-up analyses to further investigate the relationship between DNAm at birth and early-onset CP.

Methylomic variation in MGLL, TTBK2, and GCET2. Our genome-wide analyses indicated that three of the seven sites identified as differentiating early-onset versus low CP children were annotated to genes, including MGLL, TTBK2, and GCET2 (one site each). As an additional step, we examined whether any other probes in the vicinity of these genes associated with early-onset CP at a nominal level. While none of the other probes in *TTBK2* ($n_{\text{total}} = 10$ probes) or GCET2 ($n_{\text{total}} = 19$ probes) associated with early-onset versus low CP (p > .05), four additional probes in MGLL were differentially methylated between groups, including another probe in the TSS1500 region (cg02116612; p = 1.95E-05; significant at gene level Bonferroni correction: 0.05/26 total annotated probes = 0.002) and three probes in the gene body region (cg14476212, p = .028; cg10319942, p = .034; and cg15694422, p = .048). The location of these probes and their relationship to ENCODE regulatory elements is displayed in Figure 2 (for more detailed information, see online-only supplementary text). Of note, two of these additional probes correlated significantly with the genome-wide significant site $MGLL_{cg02503763}$, including a stronger association with the most proximal probe in the TSS1500 region (cg02116612; r = .46, p = 6.04E-15) and a weaker association with one of the probes in the gene body (cg10319942; r = .15, p = .01).

Methylomic variation in candidate genes for CP. As shown in Table 1, none of the genome-wide significant loci identified resided in "traditional" candidate genes for CP. In order to maximize comparability with existing molecular studies, we carried out a follow-up analysis to investigate DNAm variation in the vicinity of genes that have been previously implicated in CP-relevant phenotypes. The selection of candidate genes was informed by a recent systematic review (Waltes, Chiocchetti, & Freitag, 2015) and meta-analysis (Pappa et al., 2015) of genetic influences on aggression. Specifically, we selected a total of 15 genes ($n_{\text{total}} = 387$ probes), across dopaminergic (monoamine oxidase A [MAOA, 14 probes], catechol-O-methyltransferase [COMT, 22 probes], SLC6A3 [52 probes], dopamine receptor D2 [DRD2; 22 probes], and DRD4 [21 probes]); serotonergic (SLC6A4 [14 probes], HTR1A [14 probes], HTR2A [25 probes], tryptophan hydroxylase 1 [TPH1; 4 probes], an TPH2 [18 probes]); and neuroendocrine and neurodevelopmental pathways (nuclear receptor subfamily 3, group C, member 1 [NR3C1; 35 probes], FK506 binding protein 5 [FKBP5, 32 probes], arginine vasopressin [AVP; 14 probes], OXTR [17 probes], and brain-derived neurotrophic factor [BDNF, 73 probes]). As done by Weder et al. (2014), we used a gene-level Bonferroni correction for these hypothesis-driven analyses, correcting for the total number of probes within each gene.

In total, 12% of probes were differentially methylated between groups at a nominal significance threshold (p < .05), with at least one probe associating with early-onset CP for each gene (see online-only supplementary Table S.4). How-



Monoglyceride Lipase (MGLL) gene

Figure 2. (Color online) Functional characterization of monoglyceride lipase (*MGLL*) DNA methylation sites associated with early-onset conduct problems. Expanded views from the UCSC genome browser of the *MGLL* gene showing the position of the DNA methylation sites associated with early-onset conduct problems relative to ENCODE regulatory elements. Track numbers are displayed on the left-hand site and represent the following: 1, genomic position of *MGLL* in chromosome 3; 2, genomic coordinates and scale; 3, location of all Illumina 450k probes that map onto the *MGLL* gene (n = 26); and 4, location of differentially methylated probes associated with early-onset versus low conduct problems (n = 5). These are highlighted in blue to facilitate comparison with the regulatory elements displayed in lower tracks (Tracks 6–8). In red online is the probe that survived genome-wide correction (*MGLL*_{cg02503763}; all other probes significant at p < .05); 5, schematic representation of the *MGLL* gene; 6, location of transcription factors (based on ChIP-seq data from 91 cell types), where darker shades indicate a stronger signal occupancy; 7, DNaseI hypersensitivity clusters (based on ChIP-seq data from 125 cell types), where darker shades also indicate a stronger signal; and 8, levels of enrichment of three histone marks (H3K27Ac, H3K4Me1, and H3K4Me3) across three cell types, including blood (GM12878 [in red online], K562 [in purple online]) and human umbilical vein endothelial (HUVEC [in blue online]) cells.

ever, only four of these loci survived gene-level Bonferroni correction (see Table 2), including sites annotated to *BDNF* (cg01225698, cg18354203), *FKBP5* (cg07061368), and the *MAOA* promoter region (i.e., TSS200; cg05443523). Effect sizes for these loci were small (d = 0.44–0.49), and mean group DNAm differences ranged from 1% to 3%. Based on ENCODE data, all four probes coincided with transcription factor binding sites and histone marks, with two of the probes coinciding with DNase I hypersensitivity clusters. As with the loci identified via

our epigenome-wide analysis, these four probes remained significantly associated with CP trajectory after controlling for gestational age.

Comparison of top DNAm probes between early-onsets who persist versus desist in CP

Within the early-onset CP group, none of the DNAm sites identified in the previous step (i.e., seven FDR-corrected ge-

Probe	Chr	Genomic Location	Position	d	q (Genome-Wide)	Bonferroni (Gene Level)	Direction	Early-Onset CP Mean	Low CP Mean	% Diff	Cohen d
3DNF	:										
cg01225698	11	Body; TSS1500	27742355	3.44E-04	0.30	Significant	←	0.16	0.15	1	0.47
cg18354203	11	Body; 5'UTR	27696004	4.97E-04	0.33	Significant	\rightarrow	0.72	0.75	ŝ	0.49
FKBP5											
cg07061368	9	5'UTR	35631736	9.96E-04	0.37	Significant	\rightarrow	0.81	0.83	7	0.44
NAOA											
cg05443523	X	TSS200	43515213	1.17E-03	0.38	Significant	←	0.36	0.33	с	0.44

nome-wide significant loci and four Bonferroni-corrected loci from the candidate gene follow-up analyses) were found to differentiate children who will go on to follow a persisting versus a desisting trajectory of CP over time (p > .05, see online-only supplementary Table S.5 for full results). This lack of differences suggest that, while methylomic variation at birth prospectively associates with an early-onset of CP (i.e., distinguishing between high vs. low CP early in childhood), this variation is not likely to contribute toward heterogeneity in developmental trajectories of CP observed within early-onset children.

Potential relevance of genetic and in utero environmental influences on the identified DNAm sites

The 11 DNAm sites associated with early-onset CP were carried forward to explore associations with potential genetic and environmental influences. Based on mQTLdb search, we found that none of the identified loci were associated with cis or trans mQTLs, suggesting that DNAm levels across these sites are not likely to be heavily influenced by known genetic polymorphisms. Bivariate correlations between the DNAm loci and prenatal exposures are shown in Table 3, controlling for sex and cell type. Of the 11 probes examined, 7 were nominally correlated with one or more exposures (p < p.05), but none of these associations remained significant after multiple correction (q > 0.05). The majority of associations involved maternal smoking and alcohol use during the first trimester of pregnancy. Around half of associations identified remained significant after controlling for all other prenatal exposures (i.e., unique association; see Table 3).

Discussion

This is the first study to investigate the epigenetic landscape of early-onset CP, using a prospective design. The study offers a number of strengths, including the examination of genome-wide data collected *prior* to the emergence of CP symptoms and the inclusion of the largest epigenetic CP sample to date, allowing for adequately powered group comparisons (Tsai & Bell, 2015), as well as the integration of environmental, DNAm, and phenotypic data. We highlight here three key findings: (a) methylomic variation across multiple loci at birth differentiated children who go on to develop early-onset versus low CP; (b) DNAm levels across these loci were comparable for early-onsets who persist versus desist in CP over time; and (c) several of the identified loci showed suggestive associations with prenatal exposures, while none were linked to known genetic mQTLs.

Genome-wide findings point to the potential importance of methylomic variation at birth

The use of a genome-wide, hypothesis-free approach enabled us to identify novel loci for CP, pointing to potential avenues for future investigation. Specifically, we found that seven loci

				Prenatal Environ	nent			
	Healthy Diet	Unhealthy Diet	Maternal Smoking	Maternal Alcohol Use	Life Events	Contextual Risks	Parental Risks	Interpersonal Risks
Genome-wide analyses								
cg12628061	0.09	-0.06	$-0.16 \ (p = .01)^a$	$0.17 \ (p = .01)$	0.09	0.02	-0.09	0.07
cg02131674	0.09	-0.07	-0.10	0.10	0.10	0.05	-0.09	0.04
cg15384400	-0.07	0.07	0.09	0.00	-0.05	-0.01	0.11	0.05
cg07128473	$0.14 \ (p = .03)^a$	-0.08	-0.10	0.12	0.05	0.05	-0.07	0.06
cg02503763 (MGLL)	0.03	-0.01	$-0.14 \ (p = .02)$	$0.13 \ (p = .03)^a$	-0.05	-0.04	-0.08	-0.02
cg21579239 (TTBK2)	0.08	-0.07	-0.16 (p = .01)	0.09	0.09	0.04	-0.12	0.02
cg16006965 (GCET2)	0.05	0.01	-0.10	$0.17 \ (p = .01)$	0.07	0.09	-0.07	0.04
Candidate gene follow-up a	ralyses							
cg01225698 (BDNF)	-0.10	0.08	0.05	-0.03	-0.02	0.04	0.03	0.02
cg18354203 (BDNF)	0.06	-0.12	-0.09	0.00	-0.01	-0.03	-0.06	0.07
cg07061368 (FKBP5)	0.00	-0.03	-0.08	$0.20 \ (p = .001)^a$	0.11	0.08	-0.03	0.10
cg05443523 (MAOA)	-0.09	0.10	0.03	-0.09	$-0.13 \ (p = .001)^a$	-0.02	-0.01	-0.04

we are not aware of studies directly investigating the link between *TTBK2* and CP-relevant phenotypes, it is noteworthy that associations between levels of tau phosphorylation in the brain and aggressive behavior have been previously reported in patients with Alzheimer disease (e.g., Guadagna, Esiri, Williams, & Francis, 2012). As such, it will be of interest in the future to examine whether *TTBK2* methylation may

The third gene identified, *GCET2*, plays a role in immune processes, including cytokine function (Pan et al., 2007). While an increasing number of studies have documented a link between inflammation and psychiatric phenotypes, including childhood aggression (e.g., Provençal et al., 2013), little is known about the function of this gene in relation to CP. It is interesting, however, that a genome-wide study recently identified *GCET2* expression levels as a potential biomarker for antidepressant treatment response (Hennings et al., 2015), pointing to a potential role of this gene in mood regulation.

contribute to early-onset CP via microtubule alterations in the

brain.

who go on to develop early-onset versus low CP. Functional characterization using ENCODE data indicated that the majority of the identified sites coincided with regulatory elements that play a key role in gene expression, including transcription factor binding sites, DNase I hypersensitivity clusters (i.e., regions of open chromatin), and histone marks. Six of the loci were significantly hypomethylated in early-onset versus low CP children, and three were annotated to genes. Of these, MGLL encodes a serine hydrolase that converts monoacylglycerides to free fatty acids and glycerol and acts as a major endocannabinoid metabolic enzyme (Iwasaki et al., 2007). It is interesting that previous studies have shown that MGLL activity associates with substance abuse (Hopfer et al., 2007), which is highly comorbid with CP (McAdams, Salekin, Marti, Lester, & Barker, 2014). Furthermore, CB1 cannabinoid receptor function (modulated indirectly by MGLL via 2-arachidonoylglycerol [2-AG] activation) has been shown to mediate aggression in animal knockout studies (Rodriguez-Arias et al., 2013). Together, these findings support a link between MGLL function and CP-relevant phenotypes, although more work will be needed to delineate the functional effects of MGLL methylation on broader endocannabinoid signaling and downstream consequences on behavior. We do note that in follow-up analyses, a number of additional DNAm sites associated with early-onset CP across the promoter region and gene body of MGLL, as well as coinciding with key regulatory elements, which further supports a link between epigenetic regulation of this gene and CP. Another DNAm site that showed CP-related DNAm at

birth was annotated to TTBK2. This gene is highly expressed

in subcortical brain structures (e.g., cerebellum and hippocampus) and is involved in the phosphorylation of tau and tu-

bulin, two key microtubule stabilization proteins that are robustly associated with multiple nervous system pathologies, including Alzheimer disease (Liao et al., 2015). Although

at birth prospectively differentiated CP groups, showing a moderate to large effect size difference between children

C. A. M. Cecil et al.

DNA methylation of candidate genes: Maximizing comparability with previous studies

Consistent with what has been previously reported in the psychiatric epigenetic literature (e.g., Cecil, Walton, & Viding, 2015), we found that none of the DNAm sites identified via our hypothesis-free, epigenome-wide analysis were annotated to genes that are typically examined in hypothesis-driven, candidate gene studies. In order to maximize comparability with existing molecular studies, we carried out an additional analysis focusing on DNAm in the vicinity of 15 genes that have been extensively investigated in relation to CP and related phenotypes (e.g., aggression; Pappa et al., 2015; Waltes et al., 2015). It is important to note that these studies have primarily focused on genetic (as opposed to epigenetic) contributions to CP, so that a direct test of replication was not possible. Rather, this follow-up analysis was designed to explore the potential role of these genes in early-onset CP from an epigenetic perspective. Although none of the DNAm sites annotated to candidate genes passed genome-wide correction, subthreshold associations were identified across three genes, including BDNF (two probes), MAOA (one probe), and FKBP5 (one probe). Of note, effect sizes were small but not trivial (>0.44), suggesting that these DNAm differences may hold relevance for understanding biological vulnerability for early-onset CP.

BDNF encodes for a neutrophic protein that performs a wide range of functions, including regulation of neurodevelopment, synaptic plasticity, and mood. While a link between aggression and BDNF activity has been documented by multiple lines of evidence, including findings from genetic, knock-out, endocrine, and pharmacological studies (Waltes et al., 2015), this is the first report, to our knowledge, of an association with BDNF DNAm levels. The role of MAOA activity on aggression has also received widespread attention from both the animal and human literature, due to its key role in the degradation of amine neurotransmitters in the brain, including dopamine, norepinephrine, and serotonin (Nelson & Trainor, 2007). Consistent with our findings, a recent study reported that hypermethylation of the MAOA promoter region (with an average DNAm difference between groups of 3%, the same as observed in the present study) was associated with antisocial personality disorder in a forensic population, as well as decreased gene expression and dysregulation of serotonin blood serum levels (Checknita et al., 2015). MAOA promoter DNAm in blood has also been shown to robustly predict MAOA enzymatic activity in the brain (Shumay, Logan, Volkow, & Fowler, 2012), suggesting that peripheral MAOA DNAm status may be a promising biomarker for functional levels in live central tissue, which is most relevant to CP. Finally, the FKBP5 gene, which codes a cochaperone of the glucocorticoid receptor that regulates its sensitivity, has been extensively investigated for its role in mediating stress, hormonal, and immune responses to adverse experiences, particularly in the context of gene-environment interactions (Zannas & Binder, 2014). For example, FKBP5

polymorphisms have been found to interact with childhood trauma (e.g., maltreatment) to predict a range of psychiatric outcomes, including childhood aggression (Bryushkova et al., 2016). While emerging evidence suggests that *FKBP5* methylation may mediate these observed Gene × Environment interactions (Zannas, Wiechmann, Gassen, & Binder, 2015), more work is needed to characterize its relevance to CP behavior.

Heterogeneity in developmental trajectories within the early-onset CP group

Whereas the identified DNAm sites at birth differentiated children who develop early-onset versus low CP, these same sites did not further discriminate between early-onsets who go on to follow a persisting (i.e., EOP) versus desisting (i.e., CL) trajectory of CP into adolescence. This lack of associations may reflect a number of possibilities. For example, it is possible that, at birth, EOP and CL children may be phenotypically and biologically similar. In other words, while they may both be at increased risk for early-onset CP relative to typical children, it may not yet be possible to separate those who will persist versus desist in CP behavior over time. Given that EOP versus CL children start to diverge in levels of CP around late childhood and predict different outcomes in adulthood (Farrington, Gallagher, Morley, Ledger, & West, 1988; Moffitt, Caspi, Harrington, & Milne, 2002), it will be of interest in the future to extend epigenetic analyses later in development, so as to investigate whether these behavioral differences may coincide with underlying epigenetic differences. In addition, the examination of DNAm at later time points would enable one to test potential associations with postnatal environmental influences, which are likely to play an important role in the divergence between EOP and CL trajectories. It is also possible that statistically, our analyses were underpowered to examine within-group differences in DNAm patterns among the early-onset group. Larger longitudinal studies will be needed in the future to enable a more fined-tuned comparison of DNAm patterns between EOP and CL children across multiple developmental periods.

Genetic and prenatal influences on DNAm patterns associated with early-onset CP

As a final step, we investigated whether DNAm loci at birth that differentiated between early-onset versus low CP children (both at the genome-wide and candidate-gene level) were also associated with potential genetic and environmental influences relevant to CP (Barker & Maughan, 2009; Salvatore & Dick, 2016). Based on findings from a large-scale study of genetic effects on DNAm (Gaunt et al., 2016), we found that none of the identified loci associated with known *cis* or *trans* mQTLs. While this suggests that the identified loci were unlikely to be heavily influenced by genetic structure, it is important to note that the heritability of DNAm patterns is greater than what can currently be ex-

plained using known mQTLs (Gaunt et al., 2016). As such, it is still possible that the identified loci may have been influenced by polygenic effects, involving many mQTL that each explain too little variance in isolation to be detected with currently available samples.

With regard to environmental influences, we identified suggestive associations between the identified DNAm sites and prenatal exposures. Of note, we found that proximal exposures (e.g., maternal smoking and alcohol use) correlated with DNAm more strongly than more distal exposures (e.g., life events). Albeit preliminary, these findings are consistent with previous studies showing that DNAm is associated with environmental influences (e.g., Lewis & Olive, 2014), with smoking-related effects, in particular, being among the most replicated across epigenetic studies (Gao, Jia, Zhang, Breitling, & Brenner, 2015). For example, we found that prenatal smoking correlated with hypomethylation of the MGLL probe, which in turn prospectively associated with early-onset CP. This is of interest given that activity of this gene has been shown to influence nicotine withdrawal in animals and humans (Muldoon et al., 2015), as well as interacting with environmental exposures to predict cannabis dependence symptoms and amygdala function (Carey et al., 2015). However, it is important to note that the associations identified in the present study were based on correlational analyses and did not survive multiple corrections. Consequently, they should be interpreted with caution and considered more as well-grounded hypotheses for further examination in larger longitudinal studies.

Limitations

Findings should be interpreted in light of a number of limitations. First, because results from this study were based on DNAm collected from peripheral samples, the extent to which they may be tissue specific is unclear. More research will be needed to assess the relevance of the identified DNAm sites to brain tissue, as well as characterizing their potential role in CP-related brain-based phenotypes. Furthermore, because we did not have access to RNA, functional characterization

References

- Barfield, R. T., Kilaru, V., Smith, A. K., & Conneely, K. N. (2012). CpGassoc: An R function for analysis of DNA methylation microarray data. *Bioinformatics*, 28, 1280–1281. doi:10.1093/bioinformatics/ bts124
- Barker, D. J. (2007). The origins of the developmental origins theory. *Journal of Internal Medicine*, 261, 412–417.
- Barker, E. D., Cecil, C. A. M., Walton, E., & Meehan, A. (2017). Genetic contributions to disruptive and impulse-control disorders. In J. Lochman & W. Matthys (Eds.), *The Wiley handbook of disruptive and impulse-control disorders*. Hoboken, NJ: Wiley.
- Barker, E. D., Kirkham, N., Ng, J., & Jensen, S. K. (2013). Prenatal maternal depression symptoms and nutrition, and child cognitive function. *British Journal of Psychiatry*, 203, 417–421. doi:10.1192/bjp.bp.113. 129486
- Barker, E. D., & Maughan, B. (2009). Differentiating early-onset persistent versus childhood-limited conduct problem youth. *American Journal of Psychiatry*, 166, 900–908. doi:10.1176/appi.ajp.2009.08121770

of the identified loci was performed using recorded ENCODE data. However, integration of transcriptomic data will mark an important step toward establishing the functional significance and downstream effects of the observed DNAm changes. Second, the current study was based on a community sample of children with relatively low rates of CP. In the future, it will be important to replicate our findings in high-risk populations who show more severe externalising problems (e.g., young offenders and psychiatric inpatients). Furthermore, the use of a larger number of cases versus controls will make it possible to examine potential moderators in the relationship between DNAm and early-onset CP (e.g., sex). Third, although we used a prospective design where DNAm was collected prior to the emergence of CP symptoms, the results from the study are correlational and cannot be taken to reflect causal pathways. As such, findings should be considered as hypothesis generating and in need of replication. In the future, the application of advanced inference methods (e.g., two-step Mendelian randomization; Pingault, Cecil, Murray, Munafò, & Viding, 2016; Relton & Davey Smith, 2012), will offer unique opportunities to establish causal relationship between prenatal environmental risk, DNAm, and CP.

Conclusion

In this study, we find that DNA methylation at multiple loci at birth prospectively associated with early-onset versus low conduct problems. These findings highlight the neonatal period as a potentially important window of biological vulnerability for conduct problems, as well as pinpointing novel potential risk markers for future investigation. Suggestive evidence of associations between the identified loci and prenatal exposures also lend preliminary support for a link between the early environment, DNA methylation, and the development of early-onset conduct problems.

Supplementary Material

To view the supplementary material for this article, please visit https://doi.org/10.1017/S095457941700092X.

- Barker, E. D., Oliver, B. R., & Maughan, B. (2010). Co-occurring problems of early onset persistent, childhood limited, and adolescent onset conduct problem youth. *Journal of Child Psychology and Psychiatry*, 51, 1217– 1226. doi:10.1111/j.1469-7610.2010.02240.x
- Barnett, B. E., Hanna, B., & Parker, G. (1983). Life event scales for obstetric groups. *Journal of Psychosomatic Research*, 27, 313–320.
- Booij, L., Tremblay, R. E., Leyton, M., Séguin, J. R., Vitaro, F., Gravel, P., ... Diksic, M. (2010). Brain serotonin synthesis in adult males characterized by physical aggression during childhood: A 21-year longitudinal study. *PLOS ONE*, 5, e11255.
- Boyd, A., Golding, J., Macleod, J., Lawlor, D. A., Fraser, A., Henderson, J., ... Davey Smith, G. (2013). Cohort profile: the "Children of the 90s"—The index offspring of the Avon Longitudinal Study of Parents and Children. *International Journal of Epidemiology*, 42, 111–127. doi:10.1093/ije/dys064
- Brown, G. W., Sklair, F., Harris, T. O., & Birley, J. L. T. (1973). Life-events and psychiatric disorders: Part 1. Some methodological issues. *Psychological Medicine*, *3*, 74–87.

- Bryushkova, L., Zai, C., Chen, S., Pappa, I., Mileva, V., Tiemeier, H., . . . Beitchman, J. H. (2016). FKBP5 interacts with maltreatment in a sample of children with extreme, pervasive, and persistent aggression. *Psychiatry Research*. Advance online publication.
- Bywater, T. J. (2012). Perspectives on the Incredible Years programme: Psychological management of conduct disorder. *British Journal of Psychiatry*, 201, 85–87. doi:10.1192/bjp.bp.111.107920
- Carey, C. E., Agrawal, A., Zhang, B., Conley, E. D., Degenhardt, L., Heath, A. C., . . . Bogdan, R. (2015). Monoacylglycerol lipase (MGLL) polymorphism rs604300 interacts with childhood adversity to predict cannabis dependence symptoms and amygdala habituation: Evidence from an endocannabinoid system-level analysis. *Journal of Abnormal Psychol*ogy, 124, 860–877. doi:10.1037/abn0000079
- Cecil, C. A., Lysenko, L. J., Jaffee, S. R., Pingault, J. B., Smith, R. G., Relton, C. L., . . . Barker, E. D. (2014). Environmental risk, oxytocin receptor gene (OXTR) methylation and youth callous-unemotional traits: A 13-year longitudinal study. *Molecular Psychiatry*, 19, 1071–1077. doi:10.1038/mp.2014.95
- Cecil, C. A. M., Walton, E., Smith, R. G., Viding, E., McCrory, E. J., Relton, C. L., ... Barker, E. D. (2016). DNA methylation and substance-use risk: A prospective, genome-wide study spanning gestation to adolescence. *Translational Psychiatry*, 6, e976.
- Cecil, C. A. M., Walton, E., & Viding, E. (2015). DNA methylation, substance use and addiction: A systematic review of recent animal and human research from a developmental perspective. *Current Addiction Reports*, 2, 331–346. doi:10.1007/s40429-015-0072-9
- Checknita, D., Maussion, G., Labonte, B., Comai, S., Tremblay, R. E., Vitaro, F., . . . Turecki, G. (2015). Monoamine oxidase A gene promoter methylation and transcriptional downregulation in an offender population with antisocial personality disorder. *British Journal of Psychiatry*, 206, 216–222. doi:10.1192/bjp.bp.114.144964
- Chen, Y. A., Lemire, M., Choufani, S., Butcher, D. T., Grafodatskaya, D., Zanke, B. W., . . . Weksberg, R. (2013). Discovery of cross-reactive probes and polymorphic CpGs in the Illumina Infinium HumanMethylation450 microarray. *Epigenetics*, 8, 203–209. doi:10.4161/epi.23470
- Cicchetti, D., & Rogosch, F. A. (1996). Equifinality and multifinality in developmental psychopathology. *Development and Psychopathology*, 8, 597–600.
- Cohen, J. (1988). *Statistical power analysis for the behavioral sciences* (2nd ed.). Hillsdale, NJ: Erlbaum.
- Colman, I., Murray, J., Abbott, R. A., Maughan, B., Kuh, D., Croudace, T. J., & Jones, P. B. (2009). Outcomes of conduct problems in adolescence: 40 year follow-up of national cohort. *British Medical Journal*, 338, a2981
- Dadds, M. R., Moul, C., Cauchi, A., Dobson-Stone, C., Hawes, D. J., Brennan, J., & Ebstein, R. E. (2014). Methylation of the oxytocin receptor gene and oxytocin blood levels in the development of psychopathy. *Development and Psychopathology*, 26, 33–40. doi:10.1017/s0954579413000497
- Egger, H. L., & Angold, A. (2006). Common emotional and behavioral disorders in preschool children: Presentation, nosology, and epidemiology. *Journal of Child Psychology and Psychiatry*, 47, 313–337.
- Farrington, D. P. (2001). Predicting adult official and self-reported violence. In G.-F. Pinard & L. Pagani (Eds.), *Clinical assessment of dangerousness: Empirical contributions*. Cambridge: Cambridge University Press.
- Farrington, D. P., Gallagher, B., Morley, L., Ledger, R. J. S., & West, D. J. (1988). Are there any successful men from criminogenic backgrounds? *Psychiatry*, 51, 116–130.
- Fergusson, D. M., Horwood, L. J., & Ridder, E. M. (2005). Show me the child at seven: The consequences of conduct problems in childhood for psychosocial functioning in adulthood. *Journal of Child Psychology* and Psychiatry, 46, 837–849.
- Fraser, A., Macdonald-Wallis, C., Tilling, K., Boyd, A., Golding, J., Davey Smith, G., . . . Lawlor, D. A. (2013). Cohort profile: The Avon Longitudinal Study of Parents and Children: ALSPAC mothers cohort. *International Journal of Epidemiology*, 42, 97–110. doi:10.1093/ije/dys066
- Gao, X., Jia, M., Zhang, Y., Breitling, L. P., & Brenner, H. (2015). DNA methylation changes of whole blood cells in response to active smoking exposure in adults: A systematic review of DNA methylation studies. *Clinical Epigenetics*, 7, 113. doi:10.1186/s13148-015-0148-3
- Gaunt, T. R., Shihab, H. A., Hemani, G., Min, J. L., Woodward, G., Lyttleton, O., . . . Ho, K. (2016). Systematic identification of genetic influences on methylation across the human life course. *Genome Biology*, 17, 1.
- Goodman, R. (2001). Psychometric properties of the Strengths and Difficulties Questionnaire. Journal of the American Academy of Child & Adoles-

cent Psychiatry, 40, 1337–1345. doi:10.1097/00004583-200111000-00015

- Guadagna, S., Esiri, M. M., Williams, R. J., & Francis, P. T. (2012). Tau phosphorylation in human brain: Relationship to behavioral disturbance in dementia. *Neurobiology of Aging*, 33, 2798–2806. doi:10.1016/j. neurobiolaging.2012.01.015
- Guillemin, C., Provencal, N., Suderman, M., Cote, S. M., Vitaro, F., Hallett, M., . . . Szyf, M. (2014). DNA methylation signature of childhood chronic physical aggression in T cells of both men and women. *PLOS ONE*, 9, e86822. doi:10.1371/journal.pone.0086822
- Hennings, J. M., Uhr, M., Klengel, T., Weber, P., Putz, B., Touma, C., . . . Lucae, S. (2015). RNA expression profiling in depressed patients suggests retinoid-related orphan receptor alpha as a biomarker for antidepressant response. *Translational Psychiatry*, 5, e538. doi:10.1038/tp.2015.9
- Hopfer, C. J., Lessem, J. M., Hartman, C. A., Stallings, M. C., Cherny, S. S., Corley, R. P., . . . Crowley, T. J. (2007). A genome-wide scan for loci influencing adolescent cannabis dependence symptoms: Evidence for linkage on chromosomes 3 and 9. *Drug and Alcohol Dependence*, 89, 34–41. doi:10.1016/j.drugalcdep.2006.11.015
- Houseman, E. A., Accomando, W. P., Koestler, D. C., Christensen, B. C., Marsit, C. J., Nelson, H. H., . . . Kelsey, K. T. (2012). DNA methylation arrays as surrogate measures of cell mixture distribution. *BMC Bioinformatics*, 13, 86. doi:10.1186/1471-2105-13-86
- Iwasaki, S., Ishiguro, H., Higuchi, S., Onaivi, E. S., & Arinami, T. (2007). Association study between alcoholism and endocannabinoid metabolic enzyme genes encoding fatty acid amide hydrolase and monoglyceride lipase in a Japanese population. *Psychiatric Genetics*, 17, 215–220. doi:10.1097/YPG.0b013e32809913d8
- Jaenisch, R., & Bird, A. (2003). Epigenetic regulation of gene expression: How the genome integrates intrinsic and environmental signals. *Nature Genetics*, 33(Suppl.), 245–254. doi:10.1038/ng1089
- Jaffee, S. R., Caspi, A., Moffitt, T. E., Dodge, K. A., Rutter, M., Taylor, A., & Tully, L. A. (2005). Nature × Nurture: Genetic vulnerabilities interact with physical maltreatment to promote conduct problems. *Development* and Psychopathology, 17, 67–84.
- Johnson, W. E., Li, C., & Rabinovic, A. (2007). Adjusting batch effects in microarray expression data using empirical Bayes methods. *Biostatistics*, 8, 118–127. doi:10.1093/biostatistics/kxj037
- Jones, M. J., Fejes, A. P., & Kobor, M. S. (2013). DNA methylation, genotype and gene expression: Who is driving and who is along for the ride? *Genome Biology*, 14, 126. doi:10.1186/gb-2013-14-7-126
- Kent, W. J., Sugnet, C. W., Furey, T. S., Roskin, K. M., Pringle, T. H., Zahler, A. M., & Haussler, D. (2002). The human genome browser at UCSC. *Genome Research*, 12, 996–1006.
- Kim-Cohen, J., Caspi, A., Moffitt, T. E., Harrington, H., Milne, B. J., & Poulton, R. (2003). Prior juvenile diagnoses in adults with mental disorder: Developmental follow-back of a prospective-longitudinal cohort. Archives of General Psychiatry, 60, 709–717.
- Klengel, T., Pape, J., Binder, E. B., & Mehta, D. (2014). The role of DNA methylation in stress-related psychiatric disorders. *Neuropharmacology*, 80, 115–132. doi:10.1016/j.neuropharm.2014.01.013
- Kofink, D., Boks, M. P., Timmers, H. T., & Kas, M. J. (2013). Epigenetic dynamics in psychiatric disorders: Environmental programming of neurodevelopmental processes. *Neuroscience Biobehavioral Review*, 37, 831–845. doi:10.1016/j.neubiorev.2013.03.020
- Lewis, C. R., & Olive, M. F. (2014). Early-life stress interactions with the epigenome: Potential mechanisms driving vulnerability toward psychiatric illness. *Behavioural Pharmacology*, 25, 341–351. doi:10.1097/fbp. 0000000000000057
- Liao, J. C., Yang, T. T., Weng, R. R., Kuo, C. T., & Chang, C. W. (2015). TTBK2: A tau protein kinase beyond tau phosphorylation. *BioMed Research International*, 2015, 575170. doi:10.1155/2015/575170
- Lutz, P. E., & Turecki, G. (2014). DNA methylation and childhood maltreatment: From animal models to human studies. *Neuroscience*, 264, 142– 156. doi:10.1016/j.neuroscience.2013.07.069
- McAdams, T. A., Salekin, R. D., Marti, C. N., Lester, W. S., & Barker, E. D. (2014). Co-occurrence of antisocial behavior and substance use: Testing for sex differences in the impact of older male friends, low parental knowledge and friends' delinquency. *Journal of Adolescence*, 37, 247–256.
- McGowan, P. O., & Roth, T. L. (2015). Epigenetic pathways through which experiences become linked with biology. *Development and Psychopa*thology, 27, 637–648. doi:10.1017/s0954579415000206
- Moffitt, T. E. (2006). Life-course persistent versus adolescence-limited antisocial behaviour. In C. Cicchetti & D. J. Cohen (Eds.), *Developmental*

psychopathology: Vol. 3. Risk, disorder, and adaptation (pp. 570–598). Hoboken, NJ: Wiley.

- Moffitt, T. E., Arseneault, L., Jaffee, S. R., Kim-Cohen, J., Koenen, K. C., Odgers, C. L., . . . Viding, E. (2008). Research review: DSM-V conduct disorder: Research needs for an evidence base. *Journal of Child Psychol*ogy and Psychiatry, 49, 3–33.
- Moffitt, T. E., Caspi, A., Harrington, H., & Milne, B. J. (2002). Males on the life-course-persistent and adolescence-limited antisocial pathways: Follow-up at age 26 years. *Development and Psychopathology*, 14, 179–207.
- Monk, C., Spicer, J., & Champagne, F. A. (2012). Linking prenatal maternal adversity to developmental outcomes in infants: The role of epigenetic pathways. *Development and Psychopathology*, 24, 1361.
- Moul, C., Dobson-Stone, C., Brennan, J., Hawes, D. J., & Dadds, M. R. (2015). Serotonin 1B receptor gene (HTR1B) methylation as a risk factor for callous-unemotional traits in antisocial boys. *PLOS ONE*, 10, e0126903.
- Muldoon, P. P., Chen, J., Harenza, J. L., Abdullah, R. A., Sim-Selley, L. J., Cravatt, B. F., . . . Damaj, M. I. (2015). Inhibition of monoacylglycerol lipase reduces nicotine withdrawal. *British Journal of Pharmacology*, *172*, 869–882. doi:10.1111/bph.12948
- Murgatroyd, C., & Spengler, D. (2011). Epigenetics of early child development. *Frontiers in Psychiatry*, 2, 16.
- Nelson, R. J., & Trainor, B. C. (2007). Neural mechanisms of aggression. Nature Reviews Neuroscience, 8, 536–546.
- Nigg, J. T. (2016). Where do epigenetics and developmental origins take the field of developmental psychopathology? *Journal of Abnormal Child Psychology*, 44, 405–419.
- Odgers, C. L., Caspi, A., Broadbent, J. M., Dickson, N., Hancox, R. J., Harrington, H., . . . Moffitt, T. E. (2007). Prediction of differential adult health burden by conduct problem subtypes in males. *Archives of General Psychiatry*, 64, 476–484. doi:10.1001/archpsyc.64.4.476
- Odgers, C. L., Moffitt, T. E., Broadbent, J. M., Dickson, N., Hancox, R. J., Harrington, H., . . . Caspi, A. (2008). Female and male antisocial trajectories: From childhood origins to adult outcomes. *Development and Psychopathology*, 20, 673–716. doi:10.1017/S0954579408000333
- Pan, Z., Shen, Y., Ge, B., Du, C., McKeithan, T., & Chan, W. C. (2007). Studies of a germinal centre B-cell expressed gene, GCET2, suggest its role as a membrane associated adapter protein. *British Journal of Haematology*, 137, 578–590. doi:10.1111/j.1365-2141.2007.06597.x
- Pappa, I., St. Pourcain, B., Benke, K., Cavadino, A., Hakulinen, C., Nivard, M. G., . . . Tiemeier, H. (2015). A genome-wide approach to children's aggressive behavior: The EAGLE consortium. *American Journal of Medical Genetics B: Neuropsychiatric Genetics*. Advance online publication. doi:10.1002/ajmg.b.32333
- Pidsley, R., Wong, C. C. Y., Volta, M., Lunnon, K., Mill, J., & Schalkwyk, L. C. (2013). A data-driven approach to preprocessing Illumina 450K methylation array data. *BMC Genomics*, 14, 293. doi:10.1186/1471-2164-14-293
- Pingault, J. B., Cecil, C. A. M., Murray, J., Munafò, M. R., & Viding, E. (2016). Causal inference in psychopathology: A systematic review of Mendelian randomisation studies aiming to identify environmental risk factors for psychopathology. *Psychopathology Review*. Advance online publication.
- Pingault, J.-B., Rijsdijk, F., Zheng, Y., Plomin, R., & Viding, E. (2015). Developmentally dynamic genome: Evidence of genetic influences on increases and decreases in conduct problems from early childhood to adolescence. *Scientific Reports*, 5, 10053.
- Price, M. E., Cotton, A. M., Lam, L. L., Farre, P., Emberly, E., Brown, C. J., . . . Kobor, M. S. (2013). Additional annotation enhances potential for biologically-relevant analysis of the Illumina Infinium HumanMethylation450 BeadChip array. *Epigenetics Chromatin*, 6, 4. doi:10.1186/ 1756-8935-6-4
- Provençal, N., Suderman, M. J., Caramaschi, D., Wang, D., Hallett, M., Vitaro, F., . . . Szyf, M. (2013). Differential DNA methylation regions in cytokine and transcription factor genomic loci associate with childhood physical aggression. *PLOS ONE*, 8, e71691. doi:10.1371/journal.pone. 0071691
- Provençal, N., Suderman, M. J., Guillemin, C., Vitaro, F., Côté, S. M., Hallett, M., . . . Szyf, M. (2014). Association of childhood chronic physical aggression with a DNA methylation signature in adult human T cells. *PLOS ONE*, 9, e89839. doi:10.1371/journal.pone.0089839
- R Core Team. (2014). R: A language and environment for statistical computing [Computer software]. Retrieved from http://www.R-project.org/

- Relton, C. L., & Davey Smith, G. (2012). Two-step epigenetic Mendelian randomization: A strategy for establishing the causal role of epigenetic processes in pathways to disease. *International Journal of Epidemiology*, 41, 161–176. doi:10.1093/ije/dyr233
- Relton, C. L., Gaunt, T., McArdle, W., Ho, K., Duggirala, A., Shihab, H., . . . Davey Smith, G. (2015). Data resource profile: Accessible Resource for Integrated Epigenomic Studies (ARIES). *International Journal of Epidemiology*. Advance online publication. doi:10.1093/ije/dyv072
- Richmond, R. C., Simpkin, A. J., Woodward, G., Gaunt, T. R., Lyttleton, O., McArdle, W. L., . . . Tilling, K. (2015). Prenatal exposure to maternal smoking and offspring DNA methylation across the lifecourse: Findings from the Avon Longitudinal Study of Parents and Children (ALSPAC). *Human Molecular Genetics*, 24, 2201–2217.
- Rijlaarsdam, J., Cecil, C., Walton, E., Chapman Mesirow, M. S., Relton, C., Gaunt, T. R., . . . Barker, E. (2016). Prenatal unhealthy diet, insulin-like growth factor 2 gene (IGF2) methylation and attention deficit hyperactivity disorder (ADHD) symptoms for early-onset conduct problem youth: Prenatal unhealthy diet, IGF2 methylation and ADHD. *Journal of Child Psychology and Psychiatry*. Advance online publication.
- Rijlaarsdam, J., Pappa, I., Walton, E., Bakermans-Kranenburg, M. J., Mileva-Seitz, V. R., Rippe, R. C., . . . van IJzendoorn, M. H. (2016). An epigenome-wide association meta-analysis of prenatal maternal stress in neonates: A model approach for replication. *Epigenetics*. Advance online publication. doi:10.1080/15592294.2016.1145329
- Rodgers, A. B., & Bale, T. L. (2015). Germ cell origins of posttraumatic stress disorder risk: The transgenerational impact of parental stress experience. *Biological Psychiatry*, 78, 307–314.
- Rodriguez-Arias, M., Navarrete, F., Daza-Losada, M., Navarro, D., Aguilar, M. A., Berbel, P., . . . Manzanares, J. (2013). CB1 cannabinoid receptormediated aggressive behavior. *Neuropharmacology*, 75, 172–180. doi:10.1016/j.neuropharm.2013.07.013
- Salvatore, J. E., & Dick, D. M. (2016). Genetic influences on conduct isorder. Neuroscience & Biobehavioral Reviews. Advance online publication.
- Shumay, E., Logan, J., Volkow, N. D., & Fowler, J. S. (2012). Evidence that the methylation state of the monoamine oxidase A (MAOA) gene predicts brain activity of MAO A enzyme in healthy men. *Epigenetics*, 7, 1151– 1160. doi:10.4161/epi.21976
- Szyf, M., & Bick, J. (2013). DNA methylation: A mechanism for embedding early life experiences in the genome. *Child Development*, 84, 49–57.
- Tobi, E. W., Goeman, J. J., Monajemi, R., Gu, H., Putter, H., Zhang, Y., . . . Müller, F. (2014). DNA methylation signatures link prenatal famine exposure to growth and metabolism. *Nature Communications*, 5, 5592.
- Tremblay, R. E., & Szyf, M. (2010). Developmental origins of chronic physical aggression and epigenetics. *Epigenomics*, 2, 495–499.
- Tsai, P. C., & Bell, J. T. (2015). Power and sample size estimation for epigenome-wide association scans to detect differential DNA methylation. *International Journal of Epidemiology*. Advance online publication. doi:10.1093/ije/dyv041
- Turecki, G., & Meaney, M. J. (2016). Effects of the social environment and stress on glucocorticoid receptor gene methylation: A systematic review. *Biological Psychiatry*, 79, 87–96.
- Tyrka, A. R., Parade, S. H., Eslinger, N. M., Marsit, C. J., Lesseur, C., Armstrong, D. A., . . . Seifer, R. (2015). Methylation of exons 1 D, 1 F, and 1 H of the glucocorticoid receptor gene promoter and exposure to adversity in preschool-aged children. *Development and Psychopathology*, 27, 577–585.
- van Mil, N. H., Steegers-Theunissen, R. P., Bouwland-Both, M. I., Verbiest, M. M., Rijlaarsdam, J., Hofman, A., . . . Verhulst, F. C. (2014). DNA methylation profiles at birth and child ADHD symptoms. *Journal of Psychiatric Reserch*, 49, 51–59.
- Waltes, R., Chiocchetti, A. G., & Freitag, C. M. (2015). The neurobiological basis of human aggression: A review on genetic and epigenetic mechanisms. *American Journal of Medical Genetics B: Neuropsychiatric Genetics*. Advance online publication. doi:10.1002/ajmg.b.32388
- Walton, E., Pingault, J.-B., Cecil, C., Gaunt, T., Relton, C., Mill, J., & Barker, E. (2016). Epigenetic profiling of ADHD symptoms trajectories: A prospective, methylome-wide study. *Molecular Psychiatry*. Advance online publication.
- Wang, D., Szyf, M., Benkelfat, C., Provençal, N., Turecki, G., Caramaschi, D., . . . Booij, L. (2012). Peripheral SLC6A4 DNA methylation is associated with in vivo measures of human brain serotonin synthesis and childhood physical aggression. *PLOS ONE*, 7, e39501.
- Weaver, I. C., Cervoni, N., Champagne, F. A., D'Alessio, A. C., Sharma, S., Seckl, J. R., . . . Meaney, M. J. (2004). Epigenetic programming by maternal behavior. *Nature Neuroscience*, 7, 847–854.

- Weder, N., Zhang, H., Jensen, K., Yang, B. Z., Simen, A., Jackowski, A., ... Kaufman, J. (2014). Child abuse, depression, and methylation in genes involved with stress, neural plasticity, and brain circuitry. *Journal of the American Academy of Child & Adolescent Psychiatry*, 53, 417– 424. doi:10.1016/j.jaac.2013.12.025
- Wolke, D., Steer, C., & Bowen, E. (2004). The ALSPAC Family Adversity Index (FAI): Background and development. Unpublished manuscript, University of Bristol.
- Zannas, A., & Binder, E. (2014). Gene–environment interactions at the FKBP5 locus: Sensitive periods, mechanisms and pleiotropism. *Genes, Brain and Behavior*, 13, 25–37.
- Zannas, A. S., & West, A. E. (2014). Epigenetics and the regulation of stress vulnerability and resilience. *Neuroscience*, 264, 157–170.
- Zannas, A. S., Wiechmann, T., Gassen, N. C., & Binder, E. B. (2015). Gene– stress–epigenetic regulation of FKBP5: Clinical and translational implications. *Neuropsychopharmacology*. Advance online publication.