


The influence of environmental variables on bdelloid rotifers of the genus *Rotaria* in Thailand

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Research Article

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Abstract

This research investigates the influence of environmental parameters that are known to affect bdelloid rotifer species richness and composition in temperate zones on the genus *Rotaria* in tropical zone. Our study analysed species richness and composition of the genus *Rotaria* from 390 samples collected from several types of aquatic habitats in Thailand. Coordinates, elevation, limnological parameters such as water temperature, conductivity, total dissolved solids, salinity, dissolved oxygen and pH were measured. A total of nine species was recorded. Of these, one species, *Rotaria macrura* (Ehrenberg, 1832), was a new record for Thailand and new to the oriental region, and was a yet undescribed species, *Rotaria* sp. The species diversity of this genus increased from eight to 10 species. The presence or absence of the genus *Rotaria* was significantly influenced by dissolved oxygen and habitat type. For the samples where the genus occurred, species richness was not affected by any of the limnological or bioclimatic variables. Differences in species composition were affected only by habitat type. The results support former suggestions that common abiotic parameters do not seem to strongly influence diversity in bdelloids, whereas major ecological differences between habitats influence bdelloid occurrence.

Introduction

The phylum Rotifera is one of the most common, abundant and widespread groups of microscopic animals in any continental aquatic environment (Fontaneto & De Smet 2015, Sa-ardrit *et al.* 2013). Rotifers are aquatic organisms, but they are also able to withstand periods without water such as drought and frost due to their desiccation capabilities (Ricci 2001). The two main groups of rotifers have differential adaptations to desiccation. For the Monogononta, generally the adult individuals die in the absence of water (García-Roger *et al.* 2014), but the product of sexual reproduction, called resting egg, survives (Pourriot & Snell 1983); for the Bdelloidea newborn, young stages and adult can survive, but for most species eggs are not desiccation-tolerant (Kaczmarek *et al.* 2019, Ricci 1987, 1992). Such different desiccation strategies, quicker for bdelloids and requiring more time for monogononts, have the consequence that bdelloids are better adapted to highly ephemeral waters and limno-terrestrial habitats such as mosses, lichens and soil (Donner 1965, Ricci 1987, 2001), whereas monogononts are more common and abundant in permanent water bodies and ephemeral water bodies that do not dry out too rapidly (Wallace *et al.* 2006).

Most ecological studies on rotifers in aquatic habitats deal with monogonont rotifers, especially in the plankton (Herzig 1987). The group Monogononta represents about 75% of the species diversity of the phylum, whereas Bdelloidea, with less than 500 known species (Segers 2007), is a group of rotifers that is rarely considered in ecological studies of aquatic habitats, and mostly in limno-terrestrial habitats (Fontaneto *et al.* 2006). In addition, identifying most bdelloid species can be reliably done only from living and active organisms (Wallace *et al.* 2006). Moreover, the only available complete identification key was written in German (Donner 1965), limiting its general use.

Here, we deal with an unusual survey, given that we focus on the genus *Rotaria* in several aquatic habitat types across one tropical country, Thailand, trying to disentangle the environmental correlates of biodiversity. The genus *Rotaria* is a common group of mostly freshwater taxa, with about 25 species. Many *Rotaria* species, as well as other rotifer species (Fontaneto *et al.* 2012), are well known from temperate and very little known from tropical areas. Thailand is potentially one of the best known areas for rotifers in tropical South-east Asia (Sa-ardrit *et al.* 2013), and previous studies on *Rotaria* revealed eight species, including one new to science and up to now still known only from Thailand (Jaturapruek *et al.* 2018).

Most of the data on the environmental correlates of the occurrence of species of *Rotaria* come from the temperate zones (Bērziņš & Pejler 1987, 1989; Pejler 1995, Pejler & Bērziņš 1993). Such a bias is common, not only in *Rotaria*, but also for rotifers in general (Fontaneto *et al.* 2012, 2015; Obertegger *et al.* 2010). The present study aims to address the knowledge gap in tropical

bdelloids living in aquatic habitats by analysing which environmental parameters may drive species diversity in hundreds of sampled communities across Thailand.

Materials and methods

Description of the study site

Thailand is a tropical country located in the middle of mainland South-east Asia between latitude 5°37'–20°27'N and longitude 97°22'–105°37'E (Setapan 1999). The geography of this country is diverse with high mountains, several watersheds, marine and freshwater habitats. Thus, a number of species are expected, given the heterogeneous ecosystems.

Aquatic habitats were chosen for this study because most *Rotaria* have been recorded in aquatic habitats around the world (Donner 1965). *Rotaria* were collected throughout Thailand covering six regions: North (N), Northeast (NE), West (W), Central (C), East (E) and South (S). Different type of water bodies, such as swamps, marshes, peat-swamps, ponds, rice fields, canals and rivers were analysed, in order to include the largest possible diversity of *Rotaria*.

Habitats were categorized in 15 types by their characteristics (Keddy 2010). A 'swamp' was defined as a wetland that is forested and occurs close to large rivers or lakes where it is critically dependent on natural water level fluctuations; some swamps were covered by aquatic vegetation, and were called 'vegetative swamp' or 'algae swamp' depending on the type of aquatic vegetation. A 'marsh' was defined as a wetland dominated by herbaceous vegetation such as grasses, rushes or reeds rather than woody plant species and often found at the edge of lakes and streams. A 'peat swamp' was defined as a wetland where waterlogged soil prevents dead leaves and wood to fully decompose; over time, this formed a thick layer of acidic peat. A 'pond' was defined as an area of standing water, either natural or artificial, which is smaller than a 'lake' (less than 80 000 m²), and could be covered by vegetation ('vegetative pond'). A 'rice field' was defined as a flooded parcel used for agricultural rice crops. A canal was defined as a waterway channel or artificial waterway of running ('canal RW') and standing water ('canal SW') smaller and shallower than a river; some standing water canals were covered by algae ('algae canal SW') or by vegetation ('vegetative canal'). A 'river' was defined as a natural flowing watercourse, usually freshwater, flowing towards an ocean, sea, lake or another river. A 'reservoir' was defined as a man-made body of standing water.

Sampling, sorting and examination

A total of 390 samples was taken by qualitative method among aquatic plants and water bodies with a plankton net of 20 µm mesh size. All samples were maintained alive in bottles without chemical preservation and observed in the laboratory within a few hours for sorting and identification. Sampling was conducted from May 2015 to June 2019. Some samples were taken from the same water body, but from different sites, different seasons and/or different habitats to cover their microhabitats as completely as possible. Several environmental measurements were taken in the field, including water temperature (°C), conductivity (µs cm⁻¹), total dissolved solids (mg L⁻¹), salinity (ppt), dissolved oxygen (mg L⁻¹) and pH, using a calibrated water analysis checker (YSI EXO1 Multiparameter Sonde and YSI EXO Handheld Display 599150). Coordinates (latitude and longitude in WGS84 system) and elevation were obtained by GPS tracker (Garmin eTrex Summit). The

detailed habitat information and sampling dates for each locality are listed in Supplementary Table S1 and Figure 1.

The active animals were sorted with a pipette under a stereomicroscope (Olympus SZ51) at magnifications up to 80×. The taxonomically relevant morphological characteristics, such as shape and length of whole body, rostrum, trunk, foot, spurs and toes were observed under compound light microscope (Olympus CH-2) at magnification up to 400×. The identification of species of *Rotaria* was performed following Donner (1965), Ricci & Melone (2000) and the recent update of Jaturapruek *et al.* (2018).

Species richness

Species richness of the genus *Rotaria* was counted as the total number of species in the genus for each sample. We also estimated how much of the potential diversity for the country was actually observed by using extrapolations from species accumulation curve. First-order Chao (Chao 1), first-order jackknife (jackknife 1), and bootstrap estimates were calculated to obtain a potential estimated number of species (Colwell & Coddington 1994). The analyses were performed in R v3.3.3 (R Core Team 2017) using the package *vegan* v2.4-5 (Oksanen *et al.* 2017).

We searched for potential environmental drivers of species richness. First we analysed which explanatory variables could affect the occurrence of the genus, using presence/absence data as a response variable in a generalized linear model (GLM) with binomial error structure; then we performed a similar analysis with species richness as a response variable in a GLM with Poisson error structure. As explanatory variables we included: a set of measurements obtained directly in the field, namely habitat type, water temperature, conductivity, salinity, total dissolved solids (TDS), dissolved oxygen (DO), pH, a set of measurements of bioclimatic variables, namely actual evapotranspiration (AET), precipitation, mean air temperature (MAT) and solar radiation (SRAD). The bioclimatic variables were obtained from the WorldClim2 database with average data from the period 1970–2000 (Fick & Hijmans 2017), extracted using the geographic coordinates of the sites, from raster layers of 30 seconds resolution (~1 km²). Bioclimatic data were gathered, handled and averaged by year using the R packages *raster* v2.6-7 (Hijmans 2017) and *rgdal* v1.2-16 (Bivand *et al.* 2017).

Before performing any analysis, we checked for multicollinearity between explanatory variables, using the logarithmic transformation for three of them (conductivity, salinity and TDS). Three variables revealed Pearson's *r* correlation values higher than 0.6: one of them, conductivity, was retained, whereas the other two, salinity and TDS, were removed from the models. Model fit was assessed by visual inspection of normality of histograms of residuals and plots of residual versus fitted values, normal Q-Q, scale-location, and residual versus leverage values (Crawley 2012). All analyses were performed in R. The outputs of GLMs are reported as Analysis of Deviance Tables, using the package *car* v2.1.6 (Fox & Sanford 2011).

Species composition

Differences in species composition were assessed using a Jaccard index of community dissimilarity (Wolda 1981). In order to test for the effect of limnological and bioclimatic variables on differences in species composition, a Permutational Analysis of Variance (PERMANOVA) was performed using the same explanatory variables used in the analyses of species richness. Such analysis was performed with the function *adonis* in the R package *vegan*. To

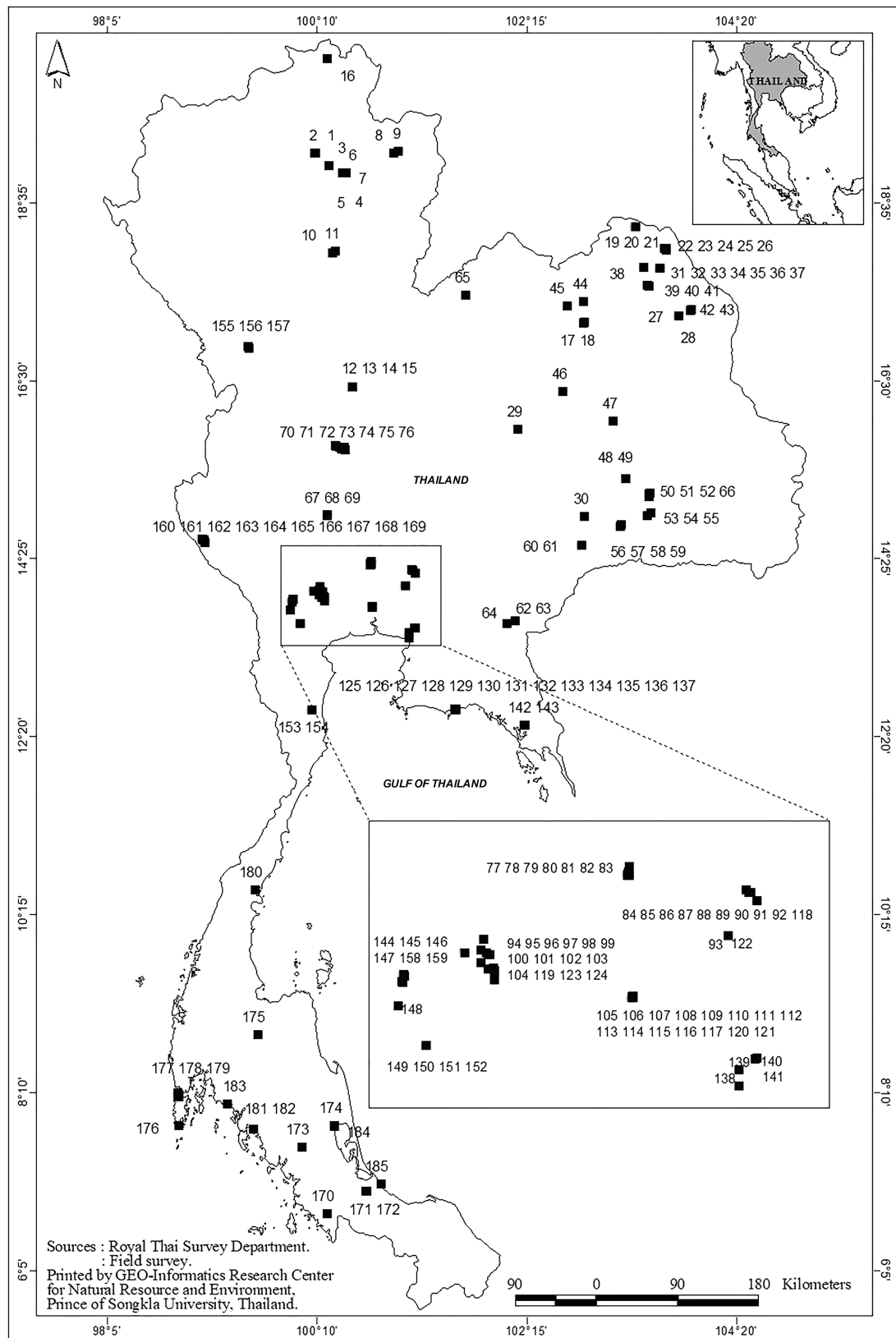


Figure 1. Map of sampling sites in Thailand. The north direction and a geographic scale in kilometres are reported. Code numbers are explained in Supplementary Table S1.

Table 1. Occurrence of bdelloid rotifers of the genus *Rotaria* in the 390 samples from Thailand from this study. The number identifying each sampling site refers to Figure 1 and Supplementary Table S1. * New record for Thailand and new to Oriental region.

Species	Sample number
<i>Rotaria macrura</i> (Ehrenberg, 1832)*	17, 264, 266
<i>Rotaria megarostris</i> Jaturaprupek, Fontaneto, Meksuwan, Pholpunthin & Maiphae, 2018	38, 85, 159, 216, 263, 264, 265, 267, 269, 270, 282, 286, 355, 367, 369, 375, 376, 377, 385, 390
<i>Rotaria mento</i> (Anderson, 1889)	22, 26, 32, 38, 60, 71, 72, 75, 76, 77, 78, 81, 88, 92, 96, 100, 103, 109, 140, 143, 171, 179, 181, 183, 184, 185, 186, 187, 188, 189, 191, 192, 197, 201, 216, 262, 263, 265, 268, 277, 284, 286, 289, 291, 292, 294, 322, 328, 332, 369, 371, 373, 378, 390
<i>Rotaria neptunia</i> (Ehrenberg, 1830)	4, 9, 14, 21, 22, 26, 32, 34, 36, 38, 66, 72, 81, 86, 88, 89, 91, 94, 95, 103, 118, 129, 149, 151, 153, 157, 158, 159, 161, 165, 178, 179, 180, 181, 182, 184, 185, 187, 190, 192, 229, 234, 244, 263, 264, 266, 267, 268, 269, 270, 272, 274, 275, 276, 277, 278, 279, 281, 282, 286, 287, 290, 309, 314, 322, 324, 330, 336, 343, 346, 348, 354, 365, 367, 387, 390
<i>Rotaria neptunoida</i> Harring, 1913	17, 71, 81, 111, 113, 140, 155, 159, 193, 196, 215, 229, 252, 268, 269, 271, 279, 281, 282, 287, 323, 332, 337, 366, 375, 376, 377, 385
<i>Rotaria ovata</i> (Anderson, 1889)	17, 21, 38, 108, 117, 207, 286, 323, 366, 371, 376, 377, 378, 385, 390
<i>Rotaria rotatoria</i> (Pallas, 1766)	21, 30, 32, 60, 69, 76, 78, 86, 88, 89, 94, 111, 113, 118, 131, 140, 151, 153, 155, 157, 158, 168, 171, 172, 173, 184, 185, 187, 189, 191, 203, 206, 215, 244, 247, 255, 257, 259, 262, 265, 268, 272, 274, 277, 278, 281, 282, 284, 290, 293, 322, 328, 330, 332, 337, 348, 352, 365, 367, 370, 375, 390
<i>Rotaria tardigrada</i> (Ehrenberg, 1830)	17, 21, 40, 43, 60, 81, 94, 108, 109, 151, 153, 155, 156, 158, 167, 171, 172, 173, 183, 185, 187, 190, 192, 200, 206, 229, 252, 253, 255, 264, 265, 266, 267, 270, 272, 286, 287, 314, 322, 323, 328, 332, 333, 365, 367, 369, 372, 373, 390
<i>Rotaria</i> sp.	273

visualize the relationship between species composition and habitats, a Two-Way Cluster Analysis (TWCA) was performed using PC-ORD program version 7 (McCune & Mefford 2016). However, as this approach can deal only with data where the genus was present, three habitat types including lake, reservoir and vegetative canal were excluded from this analysis.

Results

Species richness

A total of nine species, namely *Rotaria macrura* (Ehrenberg, 1832), *R. megarostris* Jaturaprupek, Fontaneto, Meksuwan, Pholpunthin & Maiphae, 2018, *R. mento* (Anderson, 1889), *R. neptunia* (Ehrenberg, 1830), *R. neptunoida* Harring, 1913, *R. ovata* (Anderson, 1889), *R. rotatoria* (Pallas, 1766), *R. tardigrada* (Ehrenberg, 1830), and a yet undescribed species, here called *Rotaria* sp., were identified in this study (Table 1). The highest number of species was recorded from Na Tub vegetative swamp, Songkhla (sample number 390 in Supplementary Table S1), with six species, followed by the vegetative swamp opposite dorm 13, Kasetsart University, Bangkok (sample number 286), with five species. *Rotaria neptunia* was the most common species (found in 76 of the 390 samples), followed by *R. rotatoria* (62 samples); *R. macrura* and *Rotaria* sp. were rare species, *Rotaria* sp. occurring in only one sample (Table 1).

One of the nine species, *R. macrura*, was a new record for Thailand and new to the oriental region. *R. macrura* was recorded in two parts of Thailand: KM. 62 Chiang Muan district vegetative swamp, Phayao (sample number 17) and NP12 vegetative swamp, Nakhon Pathom in different seasons (sample number 264 and 266). Most of the species have a wide range of distribution in the country: *R. megarostris*, *R. mento*, *R. neptunia*, *R. ovata* and *R. tardigrada* were found in every part of Thailand; *R. neptunoida* and *R. rotatoria* were reported in five parts, except in the eastern

Table 2. Results of the Generalized Linear Model with binomial error structure to explain the presence/absence of the genus *Rotaria* in the 390 samples from Thailand. The output of an Analysis of Deviance Table is reported, with Likelihood Ratio Chi-squared test (LR), degrees of freedom (df), P values (P) and adjusted R² values.

	LR	df	P	adjusted R ²
Habitat	27.10	14	0.02	2.7%
Temperature	2.58	1	0.11	0.7%
Conductivity	0.00	1	0.96	1.0%
Dissolved oxygen	7.75	1	0.01	2.3%
pH	0.94	1	0.33	0.1%
Actual evapotranspiration	0.38	1	0.54	0.0%
Precipitation	0.09	1	0.76	0.3%
Mean air temperature	1.02	1	0.31	0.0%
Solar radiation	1.40	1	0.24	0.2%

part of the country, whereas *Rotaria* sp. was recorded only in one specimen and restricted to a standing water canal opposite the Faculty of Engineering, Kasetsart University (sample number 273).

The distribution of the nine species in the 390 samples provided estimates of total species for the genus in Thailand to range from 9.0 ± 0.5 (Chao 1 estimator \pm standard error), to 9.4 ± 0.5 (bootstrap) and 9.9 ± 0.99 (jackknife 1).

The occurrence of the genus *Rotaria* in the 390 samples in Thailand was significantly related to dissolved oxygen (Table 2), even if this variable had an adjusted R² of only 2.3%: on average, samples without *Rotaria* had higher oxygen levels than the ones where *Rotaria* was present (Figure 2). Although pH did not significantly affect the presence of *Rotaria*, we noted that *Rotaria* was absent at sites with high pH, which were also the ones at high values

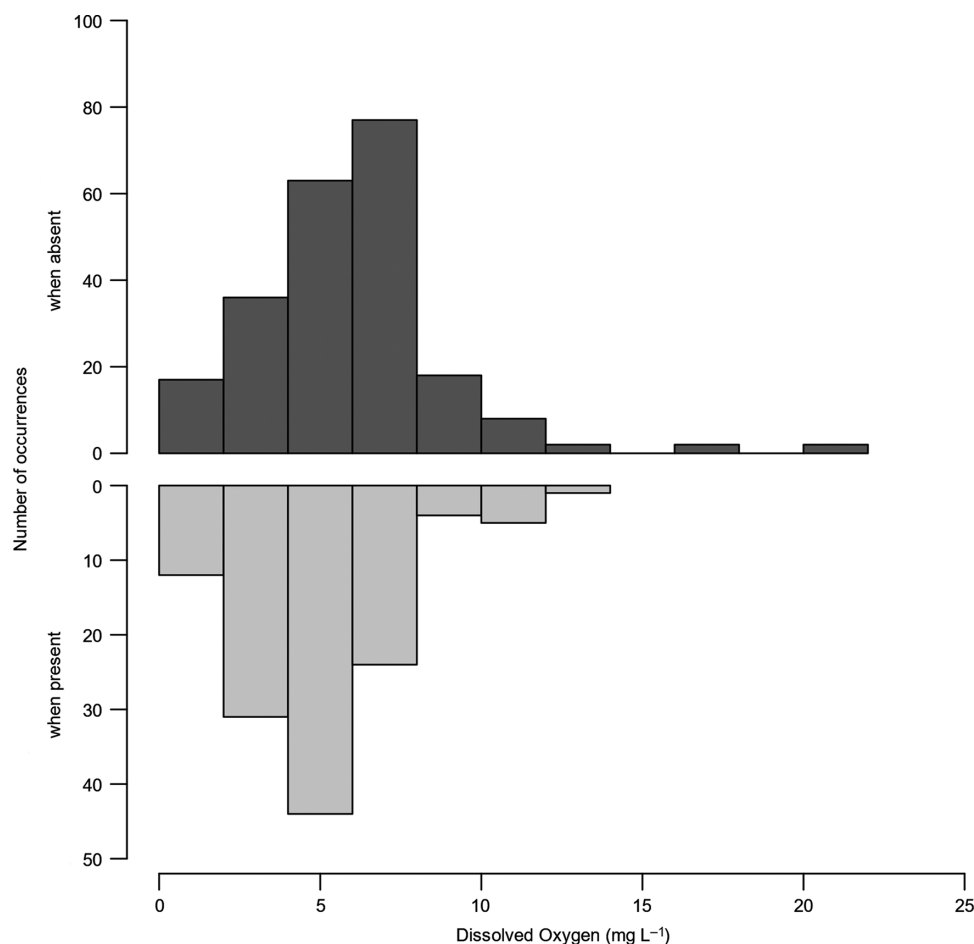


Figure 2. Histograms of the frequency distribution of the values of dissolved oxygen in sampling sites where *Rotaria* were absent or present. *Rotaria* had a tendency to disappear at higher values of dissolved oxygen.

of dissolved oxygen. Habitat type significantly affected occurrence of *Rotaria* as a genus (Table 2), which were absent from lakes, reservoirs and vegetative canals. None of the other variables revealed any significant effect at the genus level (Table 2).

Species richness of *Rotaria*, including only the 155 samples where the genus was present, was not affected by any of the limnological or bioclimatic variables that we analysed (Supplementary Table S2), even if differences in habitat type explained 4.4% of the variability and actual evapotranspiration 2.3%.

Species composition

The only significant variable explaining differences in species composition was habitat type (Table 3), with 13.3% of explained variance. None of the other limnological and bioclimatic variables reached 1.5% of explained variance (Table 3). The most dissimilar habitat from the others was river (Figure 3), whereas no clear further subdivision in groups could be highlighted (Figure 3).

Discussion

Most species of the genus *Rotaria* found in this study were already known in Thailand, except for *R. macrura*, which was encountered for the first time in South-east Asia after it was recorded from Korea (Song & Lee 2019), extending its distribution in the tropical region. One other species, *Rotaria* sp., was found only once and there are some characters that make this species different from any other previously known species. However, the detailed

characters were insufficient to clarify its status. The species diversity of *Rotaria* in Thailand increased from eight (Jaturapruerk *et al.* 2018) to 10 species. This confirmed that the effect of sampling effort still exists on faunistic studies in rotifers, even in relatively well known areas (Fontaneto *et al.* 2012, Jaturapruerk *et al.* 2018). However, the nine species found in our survey matched the expected richness by the estimators, which yielded an estimate of 9–10 species. Such estimates indicate that our sample collections seem to cover the actual diversity of this genus in Thailand for the habitat types we sampled, making our dataset reliable for ecological inference.

The limnological factors that significantly affected the occurrence of the genus *Rotaria*, expressed as presence/absence, were dissolved oxygen and habitat type (Table 2). Individuals of *Rotaria* tended to be present at lower values of dissolved oxygen at 0.2–13 (mg L⁻¹), most preferred sites at ~4.5–6.8 mg L⁻¹ (Figure 2). A lack of effect of other limnological and bioclimatic variables, confirmed for the occurrence of the genus and for species richness, might be the actual pattern. One explanation for the absence of effects on species richness could be related to compensatory effects, with different species displacing each other under different environmental regimes (Fischer *et al.* 2001). Another explanation could be that other unmeasured limnological variables could be more essential in driving species diversity in *Rotaria*, for example chlorophyll, as algae are known as the preferential food for some species of *Rotaria* (Donner 1965).

Differences in species composition of *Rotaria* seem to be related to habitat types (Table 3). Although bdelloid rotifers, including

Table 3. Output of the permutational analysis of variance to explain differences in species composition (Jaccard index) in relationship to limnological and bioclimatic variables. R² values and P values are reported.

	R ²	P
Habitat	13.3%	0.01
Temperature	1.3%	0.14
Conductivity	1.0%	0.28
Dissolved oxygen	0.3%	0.89
pH	1.4%	0.10
Actual evapotranspiration	1.1%	0.23
Precipitation	1.1%	0.24
Mean air temperature	0.6%	0.58
Solar radiation	0.5%	0.69

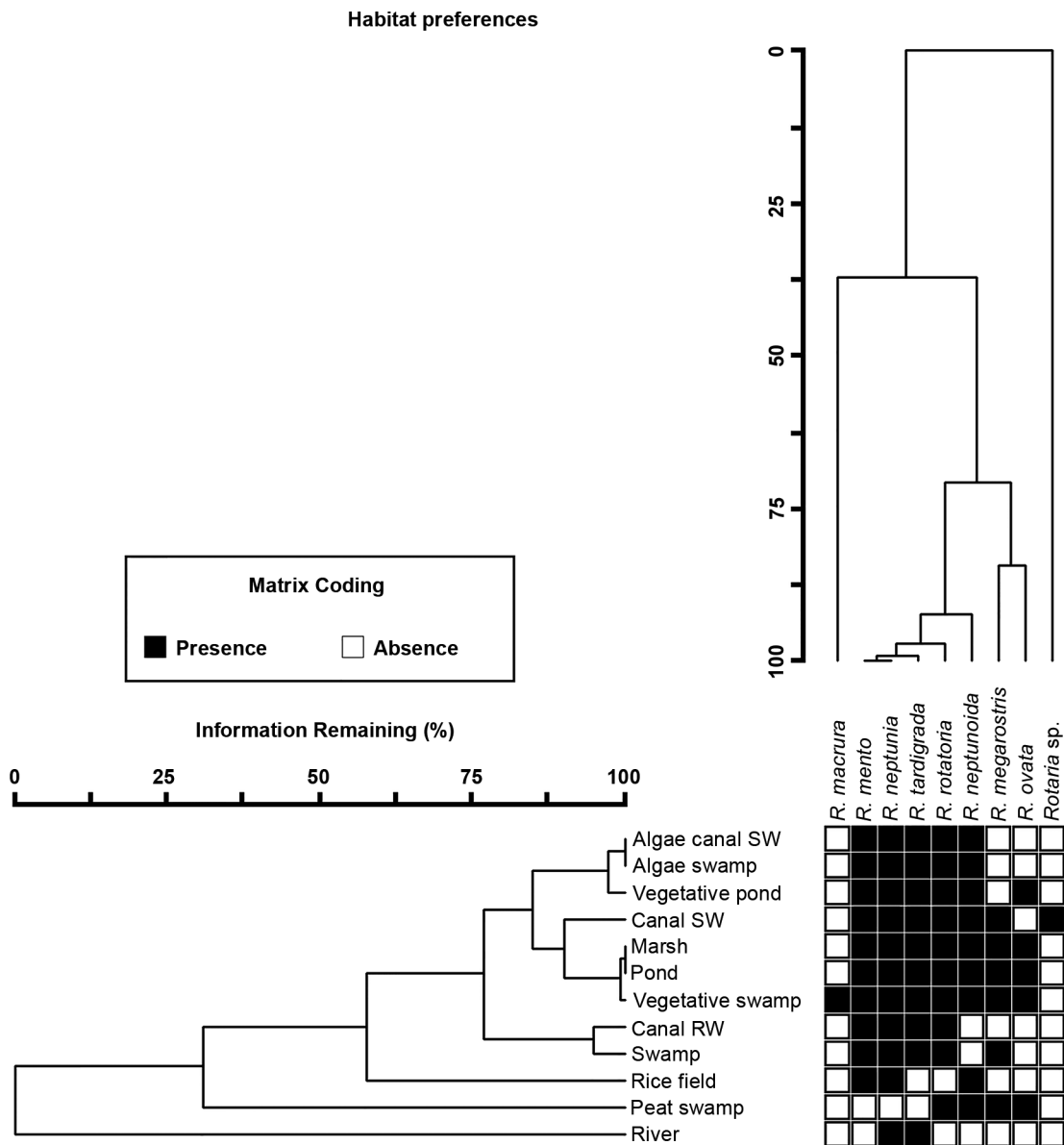


Figure 3. Habitat preferences of *Rotaria* reported as a two-way cluster plot with species and habitats ordered according to their similarity in species occurrence.

Rotaria, are small, disperse easily, and are able to withstand harsh conditions, allowing most species to become nearly cosmopolitan (Fontaneto 2019), habitat filtering still exists (Fontaneto *et al.* 2011). Habitat type, with all the inherent ecological differences between habitats, is indeed a strong selecting factor not only in bdelloid rotifers (Fenchel & Finlay 2004, Fontaneto *et al.* 2011), but also in other freshwater organisms (e.g. Bohonak & Jenkins 2003, Cohen & Shurin 2003). In the case of *Rotaria* in Thailand, species composition between rivers and other habitats was highly different; moreover, more *Rotaria* species live in vegetated than in non-vegetated habitats (GLM: $z = 4.4$, $P < 0.001$). The existence of vegetation in water bodies brings more microhabitats (Maiphae *et al.* 2005), increasing complexity of aquatic ecosystems (Kuczyńska-Kippen 2018). The importance of vegetation may be linked to the fact that only three species, namely *R. neptunia*, *R. rotatoria* and *R. tardigrada*, occurred in both standing and running water. If we assume that they occur in the littoral vegetated areas also of open running waters, their behavioural traits may explain their occurrence. *Rotaria neptunia* and *R. rotatoria* are considered good swimmers (Kuczyńska-Kippen 2018, Ricci & Melone 2000) and move frequently also in the open plankton, even if they were never found in lakes in our survey, where the distance to the littoral habitats is excessive for them. On the other hand, *R. tardigrada* commonly attaches to the substrate but has been reported also from open waters (Koste & Shiel 1986). Thus, even a small propensity for swimming in this species could have allowed it to be found in running waters, displaced from aquatic plants along the canal and river while swimming. In addition, we note that from the only 97 samples where we measured chlorophyll *a* content, *R. neptunia* was mostly recorded from meso-eutrophic habitats (chlorophyll *a* at 3.9–24.1 $\mu\text{g L}^{-1}$) (Supplementary Table S1). Our results support the suggestions from previous studies, reporting *R. neptunia* from eutrophic habitats (Sládeček 1983).

Overall, our research enhanced knowledge of bdelloid rotifers of the genus *Rotaria* in a tropical region by increasing the species diversity from eight to 10 species, of which one was a new record for Thailand and another one was a yet unidentified species. Such diversity could reliably represent the actual number of species for this genus in Thailand. In addition, we also tested the effect of limnological and bioclimatic variables on species richness and species composition. The results supported previous suggestions that parameters such as habitat type, potentially related to the level of habitat complexity, play a major role in driving species diversity of bdelloid species (Fontaneto *et al.* 2006, 2011; Kuczyńska-Kippen 2018).

Supplementary materials. For supplementary material for this article, please visit <https://doi.org/10.1017/S0266467421000018>

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Ethical statement. The present study was approved by the ethics committee of Kasetsart University (approval no. ACKU 59-SCI-004) for collecting *Rotaria* specimens.

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