

Efficiency and kinetics of the *in vitro* fertilization process in bovine oocytes with different meiotic competence

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Summary

The aim of the study was to investigate the efficiency and kinetics of fertilization in oocytes with different meiotic competence, as defined by the phase of the follicular wave and follicle size. Oocytes were recovered from cows with synchronized estrus cycles, slaughtered in either the growth (day 3) or the dominant (day 7) phase, separately from large, medium and small follicles. The oocytes were matured and fertilized by a standard protocol. Twenty-four hours after fertilization, the oocytes were denuded from cumulus cells, fixed and stained with bisbensimid Hoechst–PBS. Fertilization was more efficient and the first cleavage was accelerated in growth phase-derived oocytes, as shown by significantly higher ($p \leq 0.01$) proportions of both normally fertilized and cleaved oocytes (68.8 and 25.1%), in comparison with dominant phase-derived oocytes (44.2 and 10.3%). In the growth-phase derived oocytes, proportions of normally fertilized and cleaved oocytes were significantly higher ($p \leq 0.01$) in oocytes from large (100.0 and 36.4%) and medium (83.3 and 36.5%) follicles than in those from small (54.8 and 14.6%) follicles. The dominant phase-derived oocytes showed higher proportions of normally fertilized and cleaved oocytes in the populations recovered from small (51.5 and 10.0%) and medium (43.1 and 12.0%) follicles than in those from large (25.0 and 0%) follicles; however, the differences were not significant. It can be concluded that: (i) efficiency and kinetics of fertilization differ in relation to oocyte's meiotic competence; (ii) improved development of embryos from oocytes with greater meiotic competence is associated with a more effective fertilization process.

Keywords: Bovine, Efficiency, *In vitro* fertilization, Kinetics, Oocyte

Introduction

Although considerable improvements have been achieved for *in vitro* embryo technologies over the last years, the embryo yields still remain much lower than those of *in vivo* produced embryos (Farin & Farin, 1995). This is caused by differences between *in vitro* and *in vivo* environments for oocyte maturation. The maturation potential of an oocyte, which plays a key role not only in its maturation but also in its fertilization and early embryo development, is changing during follicular development.

The estrus cycle of cows is usually characterized by two or three follicular waves. Each wave involves

follicular growth, stagnation, dominance and regression phases. In the growth phase a cohort of 20–24 follicles, at least 4 mm in diameter, begins to grow and one of these is selected to be the dominant follicle. The dominant follicle produces factors that inhibit its counterparts in the same wave and suppress their growth (Ginther *et al.*, 1996). This effect is more apparent in 6–8 mm follicles than in smaller follicles (Hendriksen *et al.*, 2000). In non-ovulatory waves, all follicles, including dominant ones, become atretic and undergo apoptosis (Hendriksen *et al.*, 2000, Mihm & Austin, 2002, Mihm *et al.*, 2002) and in the last wave the dominant follicle ovulates.

The size of a follicle and the level of its atresia determine the quality of oocyte cytoplasm and, consequently, the ability of this oocyte to resume meiosis, to be fertilized and cleave *in vitro* (Hyttel *et al.*, 1997). During follicular growth, RNA molecules and proteins are synthesized and accumulated in oocyte

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cytoplasm and stored for early embryo development (Gandolfi & Gandolfi, 2001, Lonergan *et al.*, 2006, Sirard *et al.*, 2006).

As concerns meiotic competence, bovine embryos for *in vitro* production are generally prepared from heterogeneous populations of oocytes. They are selected only on the basis of their morphological evaluation. Recently, a selection by the metabolic activity of oocytes has been described (Alm *et al.*, 2005).

Some authors have studied the developmental competence of oocytes in relation to different sizes of follicles (Pavlok *et al.*, 1992, Lonergan *et al.*, 1994,) and ovarian cycle status (Varisanga *et al.*, 1998, Hagemann *et al.*, 1999, Machatkova *et al.*, 2000, de Wit *et al.*, 2000). In agreement with this, it has been shown in our previous studies that the developmental competence of oocytes increases with an increasing follicle size and is also influenced by the phase of follicular wave. More oocytes were able to develop to the blastocyst stage, when they were recovered in the growth phase, as compared with the dominant phase (Machatkova *et al.*, 2004).

The aim of this study was to characterize fertilization in the populations of oocytes with different meiotic competence derived from large, medium and small follicles collected in the growth and the dominant phase of the first follicular wave. This was based on a comparison of the efficiency and kinetics of the oocyte fertilization process.

Materials and methods

Synchronization of donors

A total of 36 cows were used as oocyte donors. The estrus cycles of these donors were synchronized by two doses of PGF₂α at an interval of 11 days. Ovulation was stimulated by one dose of hCG (1500 IU, Organon Co, Holland) on the day of estrus onset (Day 0). Animals were slaughtered in the growth (day 3, *n* = 18) or in the dominant (day 7, *n* = 18) phase of the first follicular wave. A pair of ovaries from each donor was examined for their ovarian status and transported, at 27 °C, to the laboratory.

Evaluation criteria for the growth and dominant phases included the presence of a hemorrhagic corpus luteum with signs of ovulation and no follicle larger than 11 mm in diameter and the presence of an advanced corpus luteum and two large follicles, 14–15 and 8–11 mm in diameter, respectively. The experiment was carried out in three replicate procedures.

Oocyte isolation

Oocytes in either the growth or the dominant phase were collected from large (11–15 mm) and medium-

sized (6–10 mm) follicles by aspiration and from small (2–5 mm) follicles by total slicing of the ovarian cortex.

The oocytes were evaluated morphologically and all collected oocytes except for heavy atretic ones without cumulus cells were used.

Oocyte maturation and fertilization

Procedures were carried out according to the standard protocol (Machatkova *et al.*, 2004). Oocytes were matured in TCM-199 medium (Earle's salt), supplemented with 50 IU/ml penicillin, 50 µg/ml streptomycin, 0.20 mmol/l sodium pyruvate (Sigma), gonadotropins (P.G. 600, 15 IU/ml, Intervet) and 5% estrus cow serum (ECS) in four-well plates (Nunclon Intermed) for 24 h.

Spermatozoa of the same bull were used for oocyte fertilization in all experiments. Motile spermatozoa were isolated by the swim-up method using modified Tyrode's medium (SP-TALP) from frozen-thawed sperm. Oocytes were fertilized for 24 h in modified Tyrode's medium (IVF-TALP), supplemented with 10 µg/ml heparin and 75 µg/ml kanamycin (Sigma), containing 1 × 10⁶ spermatozoa /ml. All procedures were carried out at 39 °C in a humidified atmosphere and 5% CO₂.

Evaluation of oocyte fertilization

Twenty-four hours after fertilization, oocytes were denuded from cumulus cells by vortex, fixed in aqueous glutaraldehyde solution, 2.5 % (v/v) and stained with bisbenzimidazole 332 58 Hoechst solution (10 µg/ml in 0.154 mol/l sodium chloride and 0.015 mol/l trisodium citrate buffer). The oocytes were rinsed once in Dulbecco-PBS, stained at room temperature for 10 min and rinsed again three times in Dulbecco-PBS. Wet slides with oocytes in 5 µl glycerin buffer solution were prepared and examined by epifluorescence (×400; L 420 barrier filter).

The efficiency of fertilization was evaluated by the percentages of normally fertilized, abnormally fertilized (polyspermy or pronuclei fragmentation) and unfertilized oocytes from all inseminated oocytes.

The kinetics of fertilization was assessed by percentages of oocytes developing to pronuclei, syngamy or first-cleavage stages from normally fertilized oocytes.

Statistical analysis

The data were analysed by the chi-squared test, using the SPSS, Version for Windows software (SPSS Inc.).

Table 1 Fertilization efficiency of oocytes derived from the growth and the dominant phase of follicular wave

Phase of follicular wave	Inseminated oocytes <i>n</i>	Fertilized					
		Normally		Abnormally		Unfertilized	
		<i>n</i>	%	<i>n</i>	%	<i>n</i>	%
Growth	446	307	68.8 ^a	65	14.6 ^a	74	16.6 ^a
Dominant	353	156	44.2 ^b	104	29.5 ^b	93	26.3 ^b

Values with different superscripts within the column are significantly different (^{a,b} $p \leq 0.01$).

Results

Efficiency of oocyte fertilization

There was a difference in fertilization efficiency between oocytes derived from growth-phase (GP) oocytes and those derived from dominant-phase (DP) oocytes. In the GP oocytes, the proportion of normally fertilized oocytes was significantly higher ($p \leq 0.01$) and the proportions of abnormally fertilized and unfertilized oocytes were significantly lower ($p \leq 0.01$) than in the DP oocytes (Table 1).

The differences in fertilization efficiency between the GP and DP oocytes were also found in relation to follicular size. In the GP oocytes, the proportions of normally fertilized oocytes in the oocytes from large and medium follicles were significantly higher ($p \leq 0.01$) than the proportion of normally fertilized oocytes from small follicles. A significant difference

($p \leq 0.01$) in the percentage of unfertilized oocytes was only between oocytes from large and those from small follicles (Table 2). Contrary to the GP oocytes, in the DP oocytes the percentage of normally fertilized oocytes was higher in oocytes from small follicles than in those from medium-sized and large follicles, but the differences were not significant. The percentage of unfertilized oocytes was lower in oocytes from small follicles than in those from large follicles and this difference was significant ($p \leq 0.01$; Table 3).

Kinetics of oocyte fertilization

The kinetics of oocyte fertilization differed between GP and DP oocytes. In the GP oocytes, the proportion of cleaved oocytes was significantly higher ($p \leq 0.01$) and the proportion of pronuclei-stage oocytes was significantly lower ($p \leq 0.01$), as compared with the DP oocytes (Table 4).

Table 2 Fertilization efficiency of oocytes derived from different sized follicles collected in the growth phase

Follicles	Inseminated oocytes <i>n</i>	Fertilized					
		Normally		Abnormally		Unfertilized	
		<i>n</i>	%	<i>n</i>	%	<i>n</i>	%
Large	11	11	100.0 ^a	0	0.0 ^a	0	0.0 ^a
Medium	102	85	83.3 ^a	5	4.9 ^a	12	11.8 ^{a,b}
Small	188	103	54.8 ^b	49	26.1 ^b	36	19.1 ^b

Values with different superscripts within the column are significantly different (^{a,b} $p \leq 0.01$).

Table 3 Fertilization efficiency of oocytes derived from different sized follicles collected in the dominant phase

Follicles	Inseminated oocytes <i>n</i>	Fertilized					
		Normally		Abnormally		Unfertilized	
		<i>n</i>	%	<i>n</i>	%	<i>n</i>	%
Large	12	3	25.0 ^a	2	16.7 ^a	7	58.3 ^a
Medium	58	25	43.1 ^a	19	32.7 ^a	14	24.1 ^{a,b}
Small	134	69	51.5 ^a	41	30.6 ^a	24	17.9 ^b

Values with different superscripts within the column are significantly different (^{a,b} $p \leq 0.01$).

Table 4 Fertilization kinetics of oocytes derived from the growth and the dominant phase of follicular wave

Phase of follicular wave	Normally fertilized/inseminated oocytes <i>n</i>	Oocytes stage					
		Pronuclei		Syngamy		Cleavage	
		<i>n</i>	%	<i>n</i>	%	<i>n</i>	%
Growth	199/301	87	43.7 ^a	62	31.2 ^a	50	25.1 ^a
Dominant	97/204	54	55.7 ^b	33	34.0 ^a	10	10.3 ^b

Values with different superscripts within the columns are significantly different (^{a,b} $p \leq 0.01$).

The differences in fertilization kinetics between the GP and DP oocytes were also found in relation to follicular size. In the GP oocytes, the proportion of pronuclei-stage oocytes was significantly lower ($p \leq 0.01$) and the proportion of cleaved oocytes was significantly higher ($p \leq 0.01$) in oocytes from large follicles in comparison with these proportions in oocytes from small follicles (Table 5). In the DP oocytes, no significant differences in the proportions of pronuclei- and cleavage-stage oocytes were found amongst oocytes from all three categories of follicles (Table 6).

Discussion

Oocytes obtained from ovaries of cows slaughtered without regard to their ovarian status and follicle size represent a widely heterogeneous population subsequently used for *in vitro* embryo production.

This population includes oocytes at various stages of follicular development, i.e., growth, stagnation, dominance and regression. It is well known that 90% of oocytes in this population complete meiosis when matured *in vitro*, but only 30% of the oocytes are able to continue their development to the blastocyst stage after *in vitro* fertilization. This low embryo production is due to the inability of some oocytes to accomplish cytoplasmic maturation and undergo successful fertilization and subsequent development (Leibfried-Rutledge, 1999). It has been suggested that low meiotic competence and incomplete cytoplasmic maturation are associated with embryo losses (Arlotto *et al.* 1996, Sirard, 2001).

Our previous studies have shown, in accordance with other reports (Hagemann 1999, Machatkova *et al.* 2004), that embryo development is more efficient when oocytes with greater meiotic competence are derived from follicles larger than 6 mm collected in the growth phase of the first follicular wave. On the basis of

Table 5 Fertilization kinetics of oocytes derived from different sized follicles collected in the growth phase

Follicles	Normally fertilized/inseminated oocytes <i>n</i>	Oocyte stage					
		Pronuclei		Syngamy		Cleavage	
		<i>n</i>	%	<i>n</i>	%	<i>n</i>	%
Large	11/11	1	9.1 ^a	6	54.5 ^a	4	36.4 ^a
Medium	85/102	33	38.8 ^b	21	24.7 ^a	31	36.5 ^a
Small	103/188	53	51.4 ^b	35	34.0 ^a	15	14.6 ^b

Values with different superscripts within the columns are significantly different (^{a,b} $p \leq .01$).

Table 6 Fertilization kinetics of oocytes derived from different sized follicles collected in the dominant phase

Follicles	Normally fertilized/ inseminated oocytes <i>n</i>	Oocyte stage					
		Pronuclei		Syngamy		Cleavage	
		<i>n</i>	%	<i>n</i>	%	<i>n</i>	%
Large	3/12	1	33.3 ^a	2	66.7 ^a	0	0.0 ^a
Medium	25/58	11	44.0 ^a	11	44.0 ^a	3	12.0 ^a
Small	69/134	42	60.9 ^a	20	29.0 ^a	7	10.1 ^a

^aNo significant differences were found.

these results we assumed that also the efficiency of fertilization might be improved for these oocytes.

In this study we examined the efficiency and kinetics of fertilization in bovine oocytes isolated from three different follicle-size categories of the growth and the dominant phase. As donors of oocytes and spermatozoa, we used cows with defined follicular development and in in vitro system tested bull in order to standardize the paternal influence on the course of fertilization (Eid *et al.*, 1994; Ward *et al.*, 2002).

Our results show that a higher efficiency of fertilization and accelerated kinetics of cleavage can be achieved in oocytes with greater meiotic competence collected in the growth phase, as compared with those with lesser meiotic competence collected in the dominant phase. They also confirm that a higher embryo yield is not related only to greater meiotic competence of an oocyte, but also to its greater capability of being fertilized. In our experiments, at 24 h after insemination, the rate of cleaved oocytes derived from the growth phase was higher more than twice in comparison with the rate of dominant phase-derived oocytes.

It is known that the meiotic and developmental competence of an oocyte increase with a growing size of the follicle (Hagemann *et al.*, 1999, Gandolfi & Gandolfi, 2001). An interference between follicle size and duration of fourth cell cycle in developing embryos has recently been described by Lequarre *et al.* (2005). However, no information has so far been available on the relationship between the size of a follicle and the ability of its oocyte to be fertilized.

In this study we observed that oocytes collected from the three defined size categories of follicles had different ability to be fertilized. In the growth phase-derived oocytes, the proportion of normally fertilized oocytes increased from small to large follicles while in the dominant phase-derived oocytes this trend was reversed. This can be explained by a negative influence of the dominant follicle, which seems to be selective, because we found that fertilization efficiency was less negatively affected in oocytes from small follicles than in those from medium and large follicles. These results are in agreement with the findings made by Hendriksen *et al.* (2000) who reported more apparent inhibitory effects of the dominant follicle on oocytes from 6–8 mm follicles than on those from small follicles.

Dominko & First (1992) and Iwata *et al.* (2004) demonstrated that embryo development was improved in oocytes that extruded the polar body earlier in the process of maturation. In our previous study we described that embryo development was faster in oocytes with greater meiotic competence (Machatkova *et al.*, 2006). Nevertheless, until now no data on the kinetics of fertilization in oocytes with different meiotic competence has been published.

In this study we found that, in growth phase-derived oocytes, fertilization was accelerated in oocytes from large follicles, as compared with those from small follicles. On the other hand, in dominant phase-derived oocytes, fertilization in oocytes from large follicles was delayed in comparison with oocytes from small follicles.

In conclusion: (i) efficiency and kinetics of fertilization process differ in relation to oocyte's meiotic competence; (ii) efficiency of fertilization is higher and first cleavage is accelerated in the greater-meiotic competent oocytes compared with the lesser-meiotic competent oocytes; and (iii) improved development of embryos from oocytes with greater meiotic competence is associated with a more effective fertilization process.

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