Survival and behavioural responses of the predatory ladybird beetle, *Eriopis connexa* populations susceptible and resistant to a pyrethroid insecticide

A.F. Spíndola, C.S.A. Silva-Torres*, A.R.S. Rodrigues and J.B. Torres

Departamento de Agronomia–Entomologia, Universidade Federal Rural de Pernambuco (UFRPE), Rua Dom Manoel de Medeiros, s/n, Dois Irmãos, Recife – PE, 52171-900, Brazil

Abstract

The ladybird beetle, Eriopis connexa (Germar) (Coleoptera: Coccinellidae), is one of the commonest predators of aphids (Hemiptera: Aphididae) in the cotton agroecosystem and in many other row and fruit crops in Brazil, and has been introduced into other countries such as the USA for purposes of aphid control. In addition, the boll weevil, Anthonomus grandis Boheman (Coleoptera: Curculionidae) is the most serious cotton pest where it occurs, including Brazil. Controlling boll weevils and other pests such as cotton defoliators still tends to involve the intense application of insecticides to secure cotton production. The pyrethroid insecticide lambda-cyhalothrin (LCT) is commonly used, but this compound is not effective against aphids; hence, a desirable strategy would be to maintain E. connexa populations in cotton fields where LCT is applied. Using populations of E. connexa resistant (Res) and susceptible (Sus) to LCT, we compared behavioural responses on treated cotton plants and under confinement on partially and fully treated surfaces, and assessed the insects' survival on treated plants compared with that of the boll weevil. The E. connexa resistant population caged on treated plants with 15 and 75 g a.i. ha^{-1} exhibited \gg 82% survival for both insecticide concentrations compared with $\ll 3\%$ and $\ll 17\%$ survival for susceptible *E. connexa* populations and boll weevils, respectively. The response of E. connexa Res and Sus populations when released, either on the soil or on the plant canopy, indicated avoidance towards treated plants, as measured by elapsed time to assess the plant. When compared with susceptible individuals, resistant ones took longer time to suffer insecticide knockdown, had a higher recovery rate after suffering knockdown, and spent more time in the plant canopy. Based on behavioural parameters evaluated in treated arenas, no ladybird beetles exhibited repellency. However, irritability was evident, with the susceptible population exhibiting greater irritability compared with the resistant population and a subgroup comprising resistant individuals that had recovered from knockdown. The outcomes for the E. connexa Res population indicate a promising strategy for its maintenance when using the insecticide LCT in integrated pest management schemes to control boll weevil or other non-target pest of ladybird beetles in cotton fields.

*Author for correspondence Phone: +55 81 3320 6218 Fax: +55 81 3320 6205 E-mail: christian@depa.ufrpe.br **Keywords:** Coccinellidae, lambda-cyhalothrin, insecticide resistance, behaviour, repellency, irritability

(Accepted 25 January 2013; First published online 22 March 2013)

Introduction

The control of pest insects through joint use of synthetic insecticides and natural enemies (integrated pest management (IPM) schemes) is impossible and/or impractical in most agroecosystems (e.g. Tabashink & Johnson, 1999; Torres, 2012). However, by exploiting the general ability of insect populations to evolve resistance to insecticides, insect natural enemies might survive applications of certain insecticide at rates capable of killing the target insect pest. Such resistance has been detected in populations of predatory mites, hymenopteran parasitoids, chrysopids (Neuroptera) and predatory ladybird beetles (Head *et al.*, 1977; Croft, 1990; Kumral *et al.*, 2011; Whalon *et al.*, 2011; Rodrigues, 2012).

Resistance can take many forms because, in addition to being toxic to the target insect pest, insecticides can have other effects on natural enemies of one form and another. For example, in the case of insect natural control agents, such animals can survive insecticide applications by metabolic detoxification, as well as by behavioural mechanisms including repellency and irritability that reduce their exposure to the poisons (Gould, 1984; Hoy et al., 1998; Jallow & Hoy, 2005). Repellency is generally associated with sensory perceptions that allow an insect to recognize and to avoid insecticidetreated areas, whereas irritability is associated with the insect's neurotoxic response to direct exposure to the insecticide (Haynes, 1988; Soderlund & Bloomquist, 1989). Nevertheless, the associated behaviours differ primarily as to whether they appear before or after insecticide contact, and thus irritability could be considered as repellency in a broader sense (Georghiou, 1972).

An insecticide of particular concern is the synthetic pyrethroid lambda-cyhalothrin (LCT), a product largely recommended for use to control Lepidopteran and Coleopteran pests in several crops, including cotton, and with field application rates ranging from 5 to 20 g a.i. ha⁻¹ to control cotton pests such as staining bugs, cotton leafworms, bollworms, pink bollworms, and specifically in terms of the present study, the boll weevil, Anthonomus grandis Boheman (Coleoptera: Curculionidae) (MAPA, 2012). Sprayed on the plant canopy, LCT may well also reach non-target insect pests such as insect predators and parasitoids. Several studies have demonstrated that this compound has low selectivity to insect natural enemies, often causing lethal and sublethal effects (Tillman & Mulrooney, 2000; Torres et al., 2002; Wang et al., 2003; Liu & Stansly, 2004; Rocha et al., 2010). In addition, since LCT is a pyrethroid, it is known that this class of insecticides have repellent properties to phytophagous and predatory mites (Penman & Chapman, 1983; Riedl & Hoying, 1983), Coleoptera (Moore, 1980; Riedl & Hoying, 1983), Lepidoptera (Ruscoe, 1977; Gist & Pless, 1985) and irritability effects to various arthropods (Soderlund & Bloomquist, 1989; Alzogaray et al., 2005).

A natural enemy of particular interest is the ladybird beetle, *Eriopis connexa* (Germar) (Coleoptera: Coccinellidae). Globally, it plays an important role in the natural control of aphids and mites, characterized by its polyphagy, voracity and natural occurrence in various crops of economic importance, including cotton (Torres *et al.*, 2009).

The potential of *E. connexa* as an aphid predator resulted in its introduction into the USA in order to control the Russian wheat aphid, Diuraphis noxia (Mordvilko) (Reed & Pike, 1991). In contrast one of the most important obstacles to increased use of predatory ladybird beetles as biological control agents is the constant need for insecticide sprays to control many pests that are not controlled by the ladybird beetles themselves, such as several lepidopteran pest species, and especially the boll weevil in cotton. Worldwide, the latter is the most important cotton pest where it occurs, and alone it is responsible for about 50% of all insecticide sprays used during the cropping season in Brazil (Richetti et al., 2004; Haney et al., 2009). Such an elevated number of sprays can directly compromise the survival of predatory ladybird beetles in cotton fields and could also result in secondary pest resurgence, especially of aphids (Kidd & Rummel, 1997; Longley, 1999), including escalating aphid numbers in LCT treated areas (Hardin et al., 1995; Deguine et al., 2000; Obrycki et al., 2009), as well as having indirect effects on E. connexa through changes in its behaviour.

Recently, studies have found and characterized resistance to LCT in some populations of *E. connexa* (Rodrigues, 2012), and have documented the occurrence of a fitness cost along with the predatory potential of resistant beetles after exposure to this insecticide (Ferreira *et al.*, 2012). This is of considerable interest because the presence of LCT resistant predatory ladybirds in cotton fields might allow growers to maintain the beetles for aphid control, while at the same time maintaining insecticide action against other target pests such as boll weevils. In order to build on these discoveries, it is important to obtain information about possible direct and indirect effects of LCT on *E. connexa* adult survival and behaviour, respectively.

Since natural enemies could be exposed to insecticides, both during spraying and afterwards through residual effects on the plant (Croft, 1990), two hypotheses were tested, namely that: (i) insecticide resistance allows the survival of E. connexa adults after LCT sprays on cotton plants at concentrations that effectively control the target pest, the boll weevil, A. grandis; and (ii) the resistance of E. connexa adults to LCT includes behavioural responses that favour their survival after insecticide exposure. To investigate these possibilities, we evaluated the effects of LCT on the survival of two populations of E. connexa (resistant, Res, and susceptible, Sus, to the insecticide) in comparison with that of the boll weevil pest population. By using a computer behaviour tracking system, we also studied the behavioural patterns, particularly repellency and irritability, of these two E. connexa populations when exposed to the insecticide.

Materials and methods

Experiments were performed at the Laboratory of Biological Control and Insect Ecology of the Universidade

Federal Rural de Pernambuco (UFRPE), Recife-PE, Brazil, behavioural in the Laboratory of Insect Behaviour of the same institute.

Collection and rearing of the boll weevil

Cotton squares and bolls showing some signs of insect attack (feeding and oviposition) were collected from a conventional cotton field located in Frei Miguelinho County, Pernambuco State (07°55'90.1"S and 35°51'45.6"W). The collected material was placed in plastic trays and kept in Plexiglas cages (45 cm wide×45 cm length×30 cm height) in the laboratory at $25\pm1.5^{\circ}$ C and 12h daylight until adult emergence. After emergence, adults were transferred to plastic containers (500 ml) and fed cotton flower buds and cotyledon leaves until the start of experiments.

Collection and rearing of E. connexa *populations*

One population of *E. connexa* was collected in the same cotton field location as the boll weevil population. Initially, this *E. connexa* population was reared in the laboratory, and its response to the commercial insecticide Karate Zeon 50 CS (LCT 5% w/v – 50 g l^{-1} , Syngenta) determined. Individuals of the F₂ generation exhibited LD₅₀ = 0.038 g of a.i. l^{-1} . These were designated as the susceptible reference population (Sus).

The second *E. connexa* population was collected in a conventional cabbage field in Viçosa County, Minas Gerais State (20°45′S and 42°51′W). This population exhibited $LD_{50}=1.45$ g a.i. 1^{-1} as determined through the dose–mortality curve in F_1 generation. Therefore, these two populations showed an initial resistance ratio of 38.1 times (ratio between the LD_{50s}). This second *E. connexa* population was kept under constant selection with increasing doses of the commercial insecticide, Karate Zeon 50 CS. By F_9 generation, its LD_{50} was 2.16 g a.i. 1^{-1} . This group was designated as the resistant population (Res).

Colonies of the two ladybird populations were kept separated in the laboratory, at a temperature of 25°C and 12h light phase. Eggs of *Anagasta kuehniella* (Zeller) (Lepidoptera: Pyralidae), obtained as described in Torres *et al.* (1995), were provided as food, with ladybird rearing procedures conducted according to Rodrigues (2012).

Survival of E. connexa and boll weevil to LCT

Cotton plants (cv. BRS Rubi) were grown in microplots (1 m diameter) up to ~70 days old, whereupon they were sprayed with two different concentrations of LCT, 15 and 75 g a.i. ha⁻¹, corresponding to the lower recommended field rate to control boll weevils and five times that field rate, respectively (MAPA, 2012). The LCT used corresponded to the commercial product Karate Zeon 50 CS (5% w/v – 50 g l⁻¹ SC (soluble concentrated); Syngenta, Brazil). The plants of the control group were sprayed only with water. Insecticide applications were made by using a coastal-manual sprayer Jacto PJH 20L (Jacto, Pompéia, SP) with a hollow cone nozzle obtaining complete plant coverage.

Two hours after insecticide application, a food diet for the adult ladybirds, comprising honey and yeast (50:50%), was applied on the top four leaves of the cotton plants. Next, the upper plant stratum (5–6 nodes) was enclosed within a sleeve cage made of voile fabric (60 cm length × 40 cm width). The lower end of each cage was fastened by a string around the

plant's main stem, while the upper end was closed with a zipper. This allowed access to the plant top for insect release and later evaluation.

Adult males and females, 8–10 days old, of the boll weevil and of *E. connexa* Sus and Res populations were introduced separately into each caged plant top. These three insect groups represented the treatments in a 3×3 randomized design that encompassed three LCT concentrations (0, 15 and 75 g a.i. ha⁻¹). The experiment was set up with different numbers of individuals owing to the different numbers of insects available (180 *E. connexa* Sus, 138 R, and 281 boll weevils). These insect numbers allowed for 3, 4 and 5 replications for *E. connexa* Sus; 4, 4 and 4 replications for Res; and 4, 5 and 5 replications for boll weevil, for the 0, 15 and 75 g a.i. ha⁻¹ concentrations, respectively.

A day after the insects had been released onto plants, the cages were collected and brought to the laboratory to evaluate insect survival by counting the number of dead and alive in each cage. The data for percentage survival of ladybirds and boll weevils were tested by Kolmogorov–Smirnov and Bartlett tests for normality and homogeneity, respectively, (with subsequent arcsine transformation of square roots (x/100) after correction for natural mortality observed in the controls (Abbott, 1925). These data were submitted to analysis of variance through the PROC GLM of SAS (SAS Institute, 2001).

Behavioural response of E. connexa to LCT treated plants

The behavioural responses of ladybird beetles in relation to their position on treated plants were studied using cotton plants cultivated in 5 L-plastic pots and with four to five expanded leaves. The plants used were infested with cotton aphids. Infestation was achieved by allowing the aphids contact over 24h with old, previously aphid-infested cotton plants. These plants naturally infested with aphids were randomly assigned for each treatment. We did not estimate the number of aphids per plant, but the plants were equally exposed to the possibility of aphid invasion (by walking) and infestation and randomly assigned to different treatments.

The test cotton plants were either treated via a 1-L hand sprayer until full coverage with Karate Zeon 50 CS at the field rate 15g a.i. ha⁻¹, recommended to control boll weevil and lepidopteran larvae or treated with distilled water only. Before spraying, plastic pots holding the plants were completely covered with aluminium foil, including the soil surface, leaving only the plant canopy exposed to the chemical application. Plants were treated and kept in a greenhouse about 2–3 h after insecticide spray for drying. After spraying, aphids were still present and alive on the plants. Next, the plants were taken to the laboratory under a temperature regime of 25°C, and were placed on benches for the beginning of observations.

Adult ladybird beetles, 8–10 days old, from the *E. connexa* Sus and Res populations earlier reared in the laboratory were used in these experiments. They were fed as described for the previous experiments until 24 h before tests; at this point they were deprived of food and the distal 1/3 of their membranous wings was cut-off with scissors to prevent beetles escaping during the observations. Previous tests indicated that this procedure did not impair any behaviour except flight.

Ladybirds were released on untreated plants (controls – sprayed with distilled water) and those treated with LCT (sprayed with 15 g a.i. ha^{-1}), in one of two different releasing sites: onto the soil next to the plant's main stem or in the

canopy on the top leaf of the plant. This experiment was conducted in a complete randomized $2 \times 2 \times 2$ factorial design, with two plant treatments (untreated and insecticide treated plants), two ladybird populations (Res and Sus) and two release sites (soil and canopy). Twenty-five adult beetles were observed individually per treatment level with each individual representing one replicate, with a total of 200 observed ladybirds from each of the two populations.

The behaviour and position of each released beetle was tracked continuously for 30 min. The following parameters were measured for beetles released on the soil: elapsed time to reach and start climbing on a test plant's main stem (i.e. time on soil surface); time walking on the plant stem; time on plant leaves; and time to suffer insecticide knockdown, which was only measured for beetles released on treated plants. The parameters measured for beetles released on the plant canopy were: time on plant leaves; time walking on the stem; time spent out of the plant (i.e. on the soil surface); and time to suffer knockdown. Ladybirds that had suffered knockdown were collected and kept individually in Petri dishes (9 cm diameter), to evaluate recovery rate after 24 h of insecticide exposure on treated plants.

Based on a null hypothesis of similarity between populations (*E. connexa* Sus and Res) and both releasing sites (soil and canopy), recovery rates were examined through the PROC FREQ of SAS (SAS Institute, 2001), and statistically significant differences were tested using a chi-square test at 5% probability.

Time to suffer knockdown was submitted to analysis of variance through the PROC ANOVA of SAS (SAS Institute, 2001), in a complete randomized factorial design, comparing populations Sus and Res as treatments with releasing sites (soil and canopy) as main factors only for insecticide-treated plants. Elapsed time to reach and start climbing on the plant's main stem was considered only for beetles released on the soil surface, but the analysis included insecticide-treated and untreated plants and populations Sus and Res.

Behavioural responses of E. connexa to an LCT treated area

The effects of insecticide repellency and irritability were investigated using a dry residue of the insecticide and three groups of adult beetles: E. connexa Sus (n = 60), E. connexa Res (n=60) and E. connexa Res recovered (n=60). The E. connexa resistant-recovered (R-rec) subgroup was represented by those adults of the resistant population that recovered from knockdown after being in contact with the dry residue of the insecticide, and hence that had previously experienced direct contact with it. Filter papers (9 cm diameter, Whatman No. 1) were treated with 1 ml of the formulated product Karate Zeon 50 SC in the concentration 435 mg a.i. l^{-1} (ca. LD_{90} for the susceptible population). This concentration resulted in a knockdown effect for resistant individuals when exposed to the insecticide residue for a period $\gg 10$ min, and it allowed for a high rate of recovery. Residual contamination consisted of releasing resistant adults inside the Petri dishes with treated filter paper lined in the bottom for a period of 30 min. After contact with the insecticide treated paper, all insects were collected and transferred to another Petri dish lined with clean filter paper. At one day following insecticide contact, the recovering adults were placed in individual containers and fed for 72 h, at which time they were used as a resistant-recovered (R-rec) subgroup for behavioural tests.

Insecticide repellency and irritability were evaluated as residual effects of LCT on a filter paper by using an arena made of a glass Petri dish (9 cm diam.) as described in Cordeiro *et al.* (2010). Test conditions consisted of (i) non-choice on treated area; (ii) choice between treated and untreated areas; and (iii) non-choice on untreated area.

The first of these conditions, non-choice on treated area, used arenas that received a filter paper (9cm diameter) previously treated with 1 ml of LCT at a concentration of 435 mg a.i. 1^{-1} (LD₉₀ for S population). Prior to insect exposure, the insecticide was evenly applied on paper by using a 1000 µl automated pipette (HTL Labmate[®]). Treated papers were allowed to dry for 60 min at 25°C before introduction into the arenas. To avoid ladybirds escaping from the arenas, the Petri dish inner walls were coated with Teflon[®] (polytetrafluoroethylene – PTFE) solution and allowed to dry for 30 min prior to experiments.

The second condition, allowing the beetles to choose between insecticide treated and untreated areas, used areass partially treated with the insecticide. Filter papers (9 cm diameter) were treated either with insecticide in the same concentration of 435 mg a.i. ha⁻¹ or with distilled water, then dried and cut into two symmetrical halves. Pairs of treated and untreated half-discs were joined with adhesive tape on the underside of the paper. Therefore, these connected paper halves (one treated, the other untreated) were introduced into Petri dish arenas that had been previously marked on the bottom to indicate treated and untreated sides. For the third variation, non-choice on a non-treated arena, beetles were introduced to arenas that had received filter paper evenly treated only with distilled water.

These experiments included nine treatments (3×3) in total corresponding to three ladybird groups (Res, Sus and R-rec) observed under three different conditions (i.e. arenas): full insecticide coverage ('full'), partially treated ('partially'), and devoid of insecticide ('empty'). Twenty individual adult replicates were observed for each treatment and all replicates were observed under the same laboratory conditions (25–27°C), during the light period of the day from 10.00 to 17.00 h. Arenas were replaced after each four consecutive observations, but filter papers were substituted after each trial to avoid any traces of tested ladybirds.

After the arenas were established, adult ladybirds, regardless of gender, were singly introduced in the arena. Observations were conducted for 10 min with the help of the computer software ViewPoint™ (ViewPoint Life Sciences Inc., Montreal, Canada). This system consisted of a video camera attached to a vertical support and positioned above the arena, which captured the insect behaviour, and which was directed and saved as a computer file. Before each 10-min observation, the insect was allowed to acclimatize inside the arena for 60s. Behaviour parameters observed for full and empty arenas included walking distance, walking time, walking speed, and number of stops. Meanwhile, for partially treated arenas, the proportion of time spent on each half of the arena was also measured. Insecticide repellency was evident when the insect did not enter the treated half of the arena, whereas insecticide irritability when the insect stayed in the treated half of the arena for $\ll 50\%$ of the total 10-min observation period (Cordeiro et al., 2010).

For the *E. connexa* Res, Sus and R-rec groups in fully, empty, and partially treated arenas, observational data of walking distance, walking time, walking speed and number of stops were analyzed by MANOVA using PROC GLM of SAS Table 1. Average survival (95% confidence interval) for boll weevil adults, *A. grandis*, and for ladybird beetles, *E. connexa*, from resistant and susceptible populations, caged on cotton plants in the field treated with lower and $5 \times$ higher LCT recommended field rate for cotton pest control.

| Insects | Lambda-cyhalothrin concentrations ¹ | | | | |
|--|---|--|--|--|--|
| | 15 g a.i. ha ⁻¹ | 75 g a.i. ha $^{-1}$ | | | |
| A. grandis E. connexa resistant E. connexa susceptible | 17.4 ^b (7.0–27.8) 84.0 ^a (71.6–96.4) 3.3 ^b (0.3–7.6) | 15.8 ^b (13.4–18.2) 82.5 ^a (75.2–89.7) 0 ^c | | | |

¹ Means followed by the same letter within column do not differ statistically (One-way ANOVA; Tukey HSD test; P > 0.05).

(SAS Institute, 2001). In addition, to determine the occurrence of insecticide repellency or irritability on ladybirds in partially treated arenas, data were analyzed through the Wilcoxon rank sum test using PROC NPAR1WAY of SAS (SAS Institute, 2001). Meanwhile, comparison among all three groups regarding insecticide irritability and repellency was analyzed using a Kruskal–Wallis test using PROC NPAR1WAY of SAS (SAS Institute, 2001).

Results

Survival of E. connexa and boll weevils to LCT

For Res and Sus ladybirds and boll weevils confined on untreated cotton plants (controls), survival rates were 90.2, 95.5 and 100%, respectively. Considering the insecticide concentrations used of 15 and 75 g of a.i. ha⁻¹, respectively, the results revealed that there was a significant variation on insect survival of the two *E. connexa* populations, Sus and Res, and of boll weevil (P<0.0001), but between the two tested insecticide concentrations, these populations exhibited the same survival pattern ($F_{1,24}$ =0.31, P=0.5846). Furthermore, there was no interaction across insect populations and insecticide concentrations tested (P=0.6474).

Comparing the lower LCT concentration used $(15 \text{ g a.i. ha}^{-1})$ and a concentration $5 \times$ higher (75 g a.i. ha⁻¹), there was no difference between insecticide concentrations within insect populations (table 1). In contrast, there was a significant difference in survival of the insects within the same insecticide concentration. Overall, resistant ladybirds had the highest survival rates, at 84 and 82.5% at the lowest and highest LCT concentrations applied, respectively (table 1). Susceptible ladybirds and boll weevils shown at the lowest and highest LCT dose applied 3.3% and 17.4%, and 0% and 15.8% survival rates, respectively.

Behavioural response of E. connexa to LCT treated plants

Regarding the elapsed time to reach and start climbing on the plant's main stem, resistant and susceptible ladybirds released on the soil showed variable responses to LCT treated plants (P=0.012) (table 2). Resistant beetles spent approximately twice as much time on the soil surface (15.95 min) before accessing the stem of treated plants than untreated plants (8.81 min), whereas the susceptible beetles spent a statistically similar amount of time on the soil surface before they all climbed onto the stems of insecticide-treated (9.41 min) and untreated plants (14.70 min) (P=0.172) (table 2). Therefore, owing to this difference between populations of *E. connexa* Res and Sus functions the type of plant (treated and untreated), there was a significant interaction between these treatments (P=0.005).

With regard to the time spent on plants and walking on the stem, when insects were on the plant canopy they were easily found on leaves or walking between leaves, and on the plant main stem. Upon comparing individuals released on the plant canopy with those released on the soil and which later climbed onto the plant, there were statistically significant differences in the time spent on the plant (P < 0.0001), and whether the plants were insecticide-treated or untreated (P < 0.0001). However, these differences were not related to whether the *E. connexa* populations were Res or Sus (P = 0.174).

The time spent on plants when beetles were released on the soil was 65.8% compared with 83.5% when released on the plant canopy. In addition, irrespective of their population type, beetles stayed two to three times longer on the canopy of untreated plants (94%) compared with treated plants (58.7%) (table 2). When ladybird beetles were not on the plant canopy of treated plants, they were found on the soil surface (before or after suffering knockdown) or on the inner border of the plastic pots. Moreover, the time that *E. connexa* Res and Sus populations spent on the plant canopy was similar (P=0.1181), at 76.7 and 72.6%, respectively.

After insecticide contact on treated plants, significant differences in the time to knockdown (P < 0.0001) were evident between the two populations and between beetles at the two release sites (P = 0.0166). Resistant ladybirds were seen to show a delayed response to knockdown when released either on soil (average difference of 4.6 min longer) or on plant canopy (7.0 min longer) compared with susceptible ladybird beetles (table 2). In contrast, there was no interaction between insect populations and release site (P = 0.2584).

Disregarding the releasing sites, the time to suffer knockdown was higher for resistant ladybird beetles (19.94 min) than for susceptible ladybird beetles (13.87 min). Seven *E. connexa* Res individuals completely avoided treated plants, not even climbing on them. Therefore, these were not considered in the analyses of knockdown.

Regarding the releasing site, only individuals from the resistant population showed a significant variation in the time to knockdown (P=0.0003). Beetles from the resistant population took longer time to suffer knockdown when released on the plant canopy (21.35 min) compared with beetles released on the soil (17.9 min). In contrast, for susceptible ladybird beetles, the time to suffer knockdown was similar whether they were released on the canopy (14.35 min) or on the soil (13.39 min) (table 2).

After suffering knockdown, ladybirds were kept singly and their recovery/survival rate after 24h measured. There was a significant difference in the survival rate of both Res and Sus *E. connexa*, irrespective of releasing site (soil, P < 0.0001; canopy, P < 0.0001). For resistant beetles that had accessed the treated plants from the soil, 85.7% of individuals that suffered knockdown recovered after 24h, in comparison with only 14.3% of susceptibles. Among those individuals released on the plant canopy, 100% of resistant beetles recovered after 24h, whereas no susceptibles survived after knockdown.

Behavioural response of E. connexa to an LCT treated area

For all behavioural parameters measured, the three *E. connexa* groups exposed to LCT in a full-insecticide arena

| | | 1, 1 | | e · | | |
|-------------|----------------|--|--|--|--|--|
| Populations | Releasing site | Time on treated plant ¹ | | Time on untreated plant ¹ | | Time to suffer |
| | | Canopy | Main stem | Canopy | Main stem | knockdown ² |
| Resistant | Soil Canopy | $6.26^{a} (\pm 1.06)$ 11.5 ^a (±0.92) | $3.06^{b} (\pm 0.59)$ $3.64^{b} (\pm 0.72)$ | 17.79 ^a (±1.91) 27.57 ^a (±1.05) | $1.93^{b} (\pm 0.48)$ $0.42^{b} (\pm 0.17)$ | $17.99^{b} (\pm 1.5)$ 21.34 ^a (±1.34) |
| Susceptible | Soil Canopy | $3.9^{a} (\pm 0.70)$ $6.73^{a} (\pm 0.88)$ | $3.54^{a} (\pm 0.42)$ $1.53^{b} (\pm 0.27)$ | 15.21 ^a (±2.0) 26.21 ^a (±1.38) | $2.09^{\rm b}$ (±0.44) $0.77^{\rm b}$ (±0.25) | 13.39 ^a (±1.51) 14.35 ^a (±0.94) |

Table 2. Average time in minutes (\pm SE, standard error) for *E. connexa* from resistant and susceptible populations spent on cotton plants when released on soil surface or in the canopy of plants treated and untreated with LCT (15g a.i. ha⁻¹), and time to suffer knockdown.

¹ Means followed by the same letter within row do not differ between the canopy and the main stem of the plant when comparing treated or untreated cotton plants for each release site and population (χ^2 -test; *P*>0.05).

² Means followed by the same letter within column for knockdown effect are not statistically significant upon comparing release sites within the same population (χ^2 -test; *P* > 0.05).

Table 3. Behaviour parameters evaluated for *E. connexa* from susceptible (Sus) and resistant (Res) populations and a resistant-recovered (R-rec) subgroup during 10 min of continuous observation in fully and partially treated, and untreated arenas.

| Populations | Behavioural parameters (mean±SE) | | | | | |
|--|---|--|--|---|--|--|
| | Distance walked (cm) | Walking time (min) | Number of stops | Walking speed $(10^{-2} \text{ cm s}^{-1})$ | | |
| Fully treated arena Recovered Resistant Susceptible Statistics | 163.7 ± 21.1^{b} 263.0 ± 27.6^{a} 297.9 ± 25.3^{a} F = 6.97; P = 0.002 | 7.14 ± 0.66^{b} 8.32 ± 0.34 ^a 9.08 ± 0.21 ^a F=5.19; P=0.008 | 185.6 ± 48.6^{ab} 292.3 ± 55.6^{a} 125.0 ± 37.6^{b} F = 3.27; P = 0.044 | 40.1 ± 3.3 50.9 ± 4.3 54.1 ± 4.0 F=3.12; P=0.051 | | |
| Partially treated arena Recovered Resistant Susceptible Statistics | 117.8 ± 15.2^{b} 200.7 ± 32.3^{ab} 229.9 ± 25.6^{a} F=5.25; P=0.008 | 7.56 ± 0.58 8.14 ± 0.36 8.27 ± 0.43 F = 0.65; P = 0.526 | 198.1 ± 62.7 183.1 ± 49.4 143.0 ± 56.2 F = 0.26; P = 0.776 | 30.9 ± 4.8 42.5 ± 6.8 47.1 ± 4.6 F=2.31; P=0.109 | | |
| <i>Untreated arena</i> Recovered Resistant Susceptible Statistics | $143.6 \pm 27.8 \\ 141.2 \pm 15.0 \\ 138.7 \pm 32.1 \\ F = 0.01; P = 0.991$ | $7.32 \pm 0.61 7.74 \pm 0.51 6.03 \pm 0.65 F=2.26; P=0.113$ | 306.2 ± 66.1 298.4 ± 64.6 428.2 ± 66.5 F = 1.23; P = 0.301 | 34.2 ± 5.4 32.2 ± 3.3 36.9 ± 5.8 F=0.22; P=0.799 | | |

¹ Means followed by different letters within the column differ in a statistically significant manner among populations (Waller–Duncan test; P < 0.05).

showed statistically significant differences in their responses: walking distance (P=0.002); walking time (P=0.008); and number of stops (P=0.044) and a tendency for differences in walking speed (P=0.051) (table 3).

Resistant and susceptible *E. connexa* showed the longest walking distances and walking time in comparison with resistant-recovered (R-rec) individuals (table 3). Beetles from the Sus population made the fewest stops, those from the Res population the most; R-rec group individuals showed intermediate values (table 3). The opposite trend was found for walking speed. *E. connexa* Sus individuals showed the greatest speed, members of the R-rec group the slowest; *E. connexa* Res individuals again intermediate values (table 3). Meanwhile, when beetles were in untreated arenas, no behavioural differences among groups/populations were observed (table 3).

For those behavioural parameters measured for beetles in partially treated arenas, a significant difference was found only in terms of the total walking distance. It was significantly lower for *E. connexa* R-rec and higher for *E. connexa* Sus individuals (P=0.0081) (table 3). Beetles of all three groups were not behaviourally repelled by LCT (Kruskal–Wallis, χ^2 =0.6705, P=0.7152). In contrast, regarding insecticide irritability, variation in responses was more evident

across groups (Kruskal–Wallis, χ^2 =5.68, *P*=0.0584; fig. 1). Susceptibles exhibited the highest irritability rate (Wilcoxon, *Z*=5.60, *P*<0.0001), followed by resistant recovered individuals (Wilcoxon, *Z*=2.48, *P*=0.0131); *E. connexa* Res individuals exhibited only a partial and statistical non-significant difference (Wilcoxon, *Z*=1.85, *P*=0.0632).

Given a choice, susceptible *E. connexa* and R-rec subgroup individuals spent more time in the untreated area of the partially treated arean than resistant individuals (Kruskal-Wallis, χ^2 =6.88, *P*=0.0321; fig. 2). Between treated and untreated areas of the partially treated arena, within the same group, a significant difference was recorded in the time spent on the untreated area of the arena only for resistant recovered individuals (Wilcoxon, *Z*=-2.96, *P*=0.003) and susceptibles (Wilcoxon, *Z*=-5.29, *P*<0.0001) (fig. 2).

Discussion

In this study, we evaluated the survival of two *E. connexa* ladybird populations (Res and Sus) with that of the boll weevil, *A. grandis*, and observed the behavioural responses of the two *E. connexa* populations towards LCT insecticide treatment. The *E. connexa* resistant population showed the highest survival rates at the two insecticide concentrations

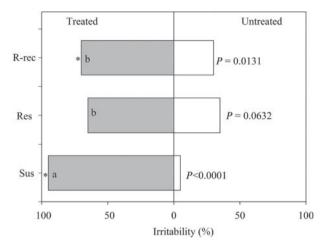


Fig. 1. Irritability of *E. connexa* from susceptible (Sus) and resistant (Res) populations and a resistant-recovered (R-rec) subgroup in partially treated arenas with dried LCT residues. Bars with an asterisk indicate a statistically significant difference between treated and untreated halves of the arena (Wilcoxon rank sum test; P < 0.05), whereas bars with different letters indicate statistically significant differences among the groups tested (Kruskal–Wallis test; Sus × Res: P = 0.0192; Sus × R-rec: P = 0.0399; Res × R-rec: P = 7389).

used in laboratory and field cage experiments. Previous results by Rodrigues (2012) had shown that four out of seven *E. connexa* populations collected in different crop fields and localities, and submitted to topical LCT application, had resistance ratios (ratio between LD_{50s}) varying from 10.5- to 37.5-fold, including the resistant population as studied here.

Our results support the expected response of *E. connexa* Res and Sus populations to lambda-cyahlothrin; the variation in response found for both populations is compatible with the mortality estimates through dose-mortality curves determined by Rodrigues (2012). Based on the estimated LDs determined by Rodrigues (2012), we expected susceptibles to show an approximately 85% mortality rate for the highest LCT concentration here tested (75 g a.i. ha^{-1}) and resistant individuals mortality rates varying from 10 to 20%. Instead, our results of field exposure of ladybirds to LCT on cotton plants revealed that Res E. connexa individuals had an even higher survival rate than expected from topically treated individuals in the laboratory. In fact in the field, other factors such as humidity on leaves from dew and the abundance of hiding places can lower insect contamination by insecticide. In contrast, the knockdown effect shown by ladybirds caged on treated cotton plants confirmed the insect's response to residual contact with the insecticide, but with substantial recovery after 24 h. Therefore, these results suggest the ability of resistant individuals to detoxify the insecticide, resulting in physiological selectivity in favour of resistant E. connexa, since such individuals had an elevated resistance ratio compared with susceptibles.

For ladybirds, over 210 laboratory-based bioassays for lethal acute toxicity at different developmental stages and for adult reproductive and survival output have been published for several insecticides, including 20 papers based on fieldrecommended rates or sub-lethal doses of pyrethroids (Web of Science[®], research topic 'Coccinellidae and insecticide' from 1970 to May 2012). Such tests are important in defining the

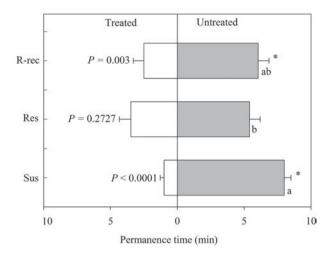


Fig. 2. Average time (+SE) spent on treated and untreated halves of the partially treated arena by *E. connexa* from susceptible (Sus) and resistant (Res) populations and a resistant-recovered (R-rec) subgroup. Bars with an asterisk indicate statistically significant differences between times in the treated and untreated halves of the arena (Wilcoxon rank sum test; P < 0.05), whereas bars with different letters indicate statistically significant differences among the groups tested (Kruskal–Wallis test; P < 0.05).

impact of insecticides on ladybird beetles in terms of designing IPM programmes. With the exception of a possible resistance to the pyrethroid bifenthrin mentioned by Kumral *et al.* (2011) in the coccinelid *Stethorus gilvifrons* (Mulsant), all other reports show incompatibility of the studied ladybird species with pyrethroid insecticides. In fact, there are very few examples of organic synthetic insecticides being compatible with ladybirds that can be safely recommended within an IPM approach; these include chemicals such as pymetrozine, pirimicarb, and some growth regulators for ladybird adults (Torres *et al.*, 2003; Cabral *et al.*, 2008).

Previous studies have detected pesticide resistance in populations of many arthropod pests, while only a few have shown resistance in their arthropod natural enemies (Tabashink & Johnson, 1999). Among the hypotheses to explain such a discrepancy, one states that phytophagous arthropods have biochemical mechanisms with a higher capacity to detoxify toxic compounds, because they are frequently challenged by plant secondary compounds (Plapp & Bull, 1978; Croft & Morse, 1979). Alternatively, according to Roush & Daly (1990), despite the ecological differences between pests and predators, the latter could develop resistance to insecticides through selection pressure caused by repetitive applications of insecticides to control arthropod pests in the field.

Studies related to the survival of lacewings such as *Chrysoperla externa* (Hagen) and *Ceraeochrysa cubana* (Hagen) (both Neuroptera: Chrysopidae) following exposure to the pyrethroid permethrin, indicate that these insects have a high level of detoxification, in addition to the low repellency rate of *C. externa* to permethrin-treated areas (Cordeiro *et al.*, 2010). In addition, research conducted in Pakistan with different populations of *Chrysoperla carnea* (Stephens) has shown resistance to both organophosphates and pyrethroids (Pathan *et al.*, 2008), involving a biochemical resistance mechanism (Sayyed *et al.*, 2010).

When searching for potential natural enemies to use in IPM schemes, behavioural responses after exposure to insecticides are important aspects to be investigated. In light of this, the present results may have a major and important influence upon decisions related to pest control. For example, insecticide repellency on a predator's part may favour the avoidance of treated areas, but at the same time the biological control agent would spend less time on treated infested areas. Behavioural alterations could be detected by changes in time spent on the treated areas, walking pattern and speed, irritability and even repellency (Chareonviriyaphap *et al.*, 1997; Pothikasikorn *et al.*, 2007).

As previously expected, when ladybirds of the two different populations were placed in a confined environment, but were not exposed to the insecticide, there were no differences in behaviour, since no toxic compound was present to elicit escape or avoidance. In contrast, when insects from the Sus, Res and R-rec groups were exposed to insecticide in fully treated arenas or partially treated arenas, behavioural differences were noted even though in most cases they were subtle. Probably these responses are directly linked to insecticide metabolism in *E. connexa* Res individuals.

The *E. connexa* Res and Sus populations exhibited overall similar trends among the behaviour responses to treated and untreated areas (table 3). Even so, susceptible *E. connexa* showed numerically greater walking speed, slightly greater walking distance, and a lower number of stops when exposed to treated areas. Hence, direct contact with insecticide residue in treated areas induced a more pronounced response in susceptibles than in resistant insects. In both fully treated and partially treated areas, *E. connexa* R-rec insects showed a slightly reduced walking distance and walking speed than resistant individuals, suggesting that the experience of recovering from knockdown had some lingering behavioural effects on them. They could also have some learning ability to avoid contact with the toxic compound, but this possibility needs further investigation.

A highest avoidance rate to insecticide-treated areas could be related to behavioural and physiological traits in the insects because of continuous selection pressure, generation after generation. However, this was not clear with E. connexa Res or R-rec populations caused by insecticide exposure as expected by our hypothesis. Chareonviriyaphap et al. (1997) found that permethrin-resistant mosquitoes, Anopheles albimanus Wiedemann (Diptera: Culicidae) tended to avoid treated areas more than susceptible individuals. Similarly, in our study, some behavioural changes were found in the ladybirds of the three groups, leading us to investigate whether or not the resistant insects would show a higher avoidance rate in a more confined and insecticide-treated environment, as well as differences in repellency and irritability. Nevertheless, all three groups exhibited similar trends for permanence and irritability when exposed to treated areas, differing only in the degree of response.

A combination of insecticide irritability and repellency as an escape mechanism driven by behaviour stimulation was previously described by Chareonviriyaphap *et al.* (1997). Irritability is a result of direct contact with insecticide treated areas, causing a physiological hypersensitivity in individuals to the toxic compound, such that they avoid further contact with the treated area. On the other hand, repellency prevents direct contact with the toxic compound because of various physiological (Klowden, 2007) and sensorial (Haynes, 1988; Soderlund & Bloomquist, 1989) mechanisms related to detection of the insecticide, such that the insect avoids entering the treated area. Regardless of their susceptibility status, the ladybird beetles in our study were not repelled by LCT, because none of them avoided direct contact with treated areas. In contrast, LCT caused irritability, as shown by a significant reduction in the amount of time spent on treated areas of the arena, with insects spending more time on untreated areas, although this was less pronounced. Susceptibles exhibited the highest irritability response, with the shortest amount of time spent on treated areas in comparison with the other two populations, an outcome that supports an irritability hypothesis predicting greater speed for such individuals. Susceptible individuals do not have the same detoxifying ability as resistant ones, and could try to find a refuge area comparatively faster in order to avoid the negative effects of the toxic compound. This was observed in full or partially treated arenas.

Studies investigating the behavioural response of natural enemies to insecticides are very few. Recently, Campos *et al.* (2011) found that the earwig, *Doru luteipes* (Scudder) (Dermaptera: Forficulidae) showed changes in motility and other behaviours, after contacting a surface treated with LCT. Escape behaviour from treated areas was also found for the green lacewings *C. externa* and *C. cubana* when exposed to permethrin (Cordeiro *et al.*, 2010). Regarding ladybirds, our study demonstrated that *E. connexa* showed variations in the elapsed time required to respond to the insecticide, both in its walking behaviour and in the time spent on treated plants.

More detailed information about insecticide repellency and irritability in populations of natural enemies would help inform decisions regarding the potential of these predators as biological control agents in IPM scenarios that involve the concurrent use of pesticides. In addition, behavioural studies of natural enemies would help in the design of appropriate insecticide application methods that would not compromise the survival of the natural enemies in the crop. For example, localized or systemic application might be necessary to favour the survival of natural enemies by ecological selectivity in situations in which they cannot be repelled or irritated by the toxic compound.

Based on our results, resistant *E. connexa* do not depend on their behaviour in terms of resistance to LCT, but instead rely upon some major physiological resistance mechanism. According to Rodrigues (2012), when the synergist piperonyl butoxide (PBO) was added to LCT, the mortality of resistant *E. connexa* tested increased over 1400-fold, and attained levels similar to that of susceptibles. Bioassays performed *in vitro* showed a differentiated action of esterases between the *E. connexa* populations studied, indicating carboxylesterase type B resistance, a phenomenon associated with the metabolism of LCT.

The fact that resistant *E. connexa* had a higher survival rate than even the boll weevil populations (these results) has practical implications to the use of LCT in the cotton pest management programme, as well as in other agroecosystems where *E. connexa* appears as an important predator of aphids. Generally, the use of broad spectrum insecticides such as LCT to control pests such as coleopteran and lepidopteran larvae and adults that are non-targets of ladybird beetles could result in negative effects for these beetles as biological control agents (Tillman & Mulrooney, 2000; Torres *et al.*, 2002).

Owing to seasonal growth patterns in cotton plants and their pest populations, mid- to late-season sprays of LCT are generally recommended to control lepidopteran larvae and boll weevils. However, outbreaks of cotton aphids commonly occur after LCT application (Hardin *et al.*, 1995; Deguine *et al.*, 2000; Obrycki *et al.*, 2009), and significant crop loss can take place as a result of sooty mould that grows on opening bolls in response to aphid honeydew excreta. Even with a low population density, resistant *E. connexa* that survive such pesticide application would be expected to help minimize aphid numbers and thus reduce these losses.

Cotton aphids may also infest cotton plants at an earlier stage. To manage these early-season infestations using insecticides, two practices could be usefully adopted: (i) applying systemic insecticide seed treatments, especially at sites that have a history of aphid infestation; (ii) spraying plants with more selective insecticides, such as pymetrozine (Torres et al., 2003). As an additional biological control measure, we recommend inoculative releases of resistant E. connexa at this stage, when the cotton plants have a relatively small leaf area for the ladybird beetles to search, and aphid infestations are low, allowing the predator population to increase as the plants grow. Such an approach would seem both promising and feasible. Although little information has been published on mass production of ladybirds, we have found that large numbers of resistant E. connexa can be reliably raised in the laboratory, using A. kuehniella eggs to rear the larvae and providing a honey-yeast mixture plus A. kuehniella eggs as diet for the ladybird beetle adults, as we have done here (see section 'Collection and rearing of E. connexa populations').

Lastly, the fact that E. connexa was not repelled by LCT but instead showed irritability is another plus for IPM, because resistant ladybirds do not avoid and presumably would still search for prev in areas treated with LCT. Using resistant ladybirds in conjunction with recommended rates of LCT in pest control also might be expected to reduce some of the negative effects of insecticide sprays. For example, the potential for pest resurgence as a result of reduction in natural enemy populations (Kidd & Rummel, 1997; Longley, 1999) such as escalating aphid populations in LCT treated areas (Hardin et al., 1995; Deguine et al., 2000; Obrycki et al., 2009). Aphid outbreaks after LCT sprays strongly suggest mortality of ladybird beetles and its inefficacy against aphids. Therefore, the combined use of resistant populations of ladybird beetles and rational use of LCT might reduce the number of insecticide sprays in an area, saving costs for the grower, as well as at the same time reducing environmental pollution, thereby improving environmental protection.

Acknowledgments

This research was supported in part by 'Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) Edital Universal' and by Fundação de Amparo à Ciência e Tecnologia do Estado de Pernambuco (FACEPE) through BFT, BCT and IBPG grants to the authors and financial aid (APQ). We further thank the two anonymous reviewers for their valuable comments and suggestions on the manuscript, and Professor Hugh D. Loxdale for his helpful editorial comments on the manuscript.

References

Abbott, W.S. (1925) A method of computing the effectiveness of an insecticide. *Journal of Economic Entomology* 18, 265–267.

- Alzogaray, R.A., Fontán, A. & Zerba, E.N. (2005) Evaluation of hyperactivity produced by pyrethroid treatment on third instar nymphs of *Triatoma infestans* (Hemiptera: Reduviidae). Archives of Insect Biochemistry and Physiology 35, 323–333.
- Cabral, S., Garcia, P. & Soares, A.O. (2008) Effects of pirimicarb, buprofezin and pymetrozine on survival, development and reproduction of *Coccinella undecimpunctata* (Coleoptera: Coccinellidae). *Biocontrol Science and Technology* 18, 307– 318.
- Campos, M.R., Picanço, M.C., Martins, J.C., Tomaz, A.C. & Guedes, R.N.C. (2011) Insecticide selectivity and behavioral response of the earwig *Doru luteipes*. Crop Protection 30, 1535–1540.
- Chareonviriyaphap, T., Roberts, D.R., Andre, R.G., Harlan, H. & Bangs, M.J. (1997) Pesticide avoidance behaviour in Anopheles albimanus Wiedemann. Journal of the American Mosquito Control Association 13, 171–183.
- Cordeiro, E.M.G., Corrêa, A.S., Venzon, M. & Guedes, R.N.C. (2010) Insecticide survival and behavioural avoidance in the lacewings *Chrysoperla externa* and *Ceraeochrysa cubana*. *Chemosphera* 81, 1352–1357.
- **Croft, B.A.** (1990) Arthropod Biological Control Agents and Pesticides. New York, John Wiley & Sons.
- Croft, B.A. & Morse, J.G. (1979) Recent advances in natural enemy-pesticide research. *Entomophaga* 24, 3–11.
- Deguine, J.P., Gozé, E. & Leclant, F. (2000) The consequences of late outbreaks of the aphid *Aphis gossypii* in cotton growing in Central Africa: towards a possible method for the prevention of cotton stickness. *International Journal of Pest Management* 46, 86–89.
- Ferreira, E.S., Rodrigues, A.R., Silva-Torres, C.S.A. & Torres, J.B. (2012) Life-history costs associated with resistance to lambda-cyhalothrin in the predatory ladybird beetle *Eriopis connexa*. Agriculture and Forest Entomology in press (http:// onlinelibrary.wiley.com/doi/10.1111/j.1461-9563.2012.00599. x/abstract).
- Georghiou, G.P. (1972) The evolution of resistance to pesticides. Annual Review of Ecology and Systematics 3, 133–68.
- Gist, G.L. & Pless, C.D. (1985) Ovicidal activity and ovipositional repellent properties of synthetic pyrethroids to the fall armyworm *Spodoptera frugiperda*. Florida Entomologist 68, 462–466.
- Gould, F. (1984) Role of behaviour in the evolution of insect adaptation to insecticides and resistant host plants. Bulletin of the Entomological Society of America 30, 34–41.
- Haney, P.B., Lewis, W.J. & Lambert, W.R. (2009) Cotton Production and The Boll Weevil in Georgia: History, Cost of Control, and Benefits of Eradication. Athens, GA, College of Agricultural and Environmental Sciences, The University of Georgia. Research Bulletin 428.
- Hardin, M.R., Benrey, B., Coll, M., Lamp, W.O., Roderick, G.K. & Barbosa, P. (1995) Arthropod pest resurgence: an overview of potential mechanisms. *Crop Protection* 14, 3–18.
- Haynes, K.F. (1988) Sublethal effects of neurotoxic insecticides on insect behavior. Annual Review of Entomology 33, 49–168.
- Head, R., Neel, W.W., Sartor, C.R. & Chambers, H. (1977) Methyl parathion and carbaryl resistance in *Chrysomela scripta* and *Coleomegilla maculata. Bulletin of Environmental Contamination* and Toxicology 17, 163–164.
- Hoy, C.W., Head, G.P. & Hall, F.R. (1998) Spatial heterogeneity and insect adaptation to toxins. *Annual Review of Entomology* 43, 571–594.

- Jallow, M.F.A. & Hoy, C.W. (2005) Phenotypic variation in adult behavioral response and offspring fitness in *Plutella xylostella* (Lepidoptera: Plutellidae) in response to permethrin. *Journal* of Economic Entomology 98, 2195–2202.
- Kidd, P.W. & Rummel, D.R. (1997) Effect of insect predators and a pyrethroid insecticide on cotton aphid, *Aphis gossypii* Glover, population density. *Southwestern Entomologist* 22, 381–393.
- Klowden, M.J. (2007) Physiological Systems In Insects. 2nd edn. San Diego, Academy Press.
- Kumral, N.A., Gencer, N.S., Susurluk, H. & Yalcin, C. (2011) A comparative evaluation of the susceptibility to insecticides and detoxifying enzyme activities in *Stethorus gilvifrons* (Coleoptera: Coccinellidae) and *Panonychus ulmi* (Acarina: Tetranychidae). *International Journal of Acarology* 37, 255–268.
- Liu, T.X. & Stansly, P.A. (2004) Lethal and sublethal effects of two insect growth regulators on adult *Delphastus catalinae* (Coleoptera: Coccinellidae), a predator of whiteflies (Homoptera: Aleyrodidae). *Biological Control* **30**, 298–305.
- Longley, M. (1999) A review of pesticide effects upon immature aphid parasitoids within mummified hosts. *International Journal of Pest Management* 45, 139–145.
- MAPA (2012) Ministério da Agricultura, Pecuária e Abastecimento. AGROFIT: sistema de agrotóxicos fitossanitários. Available online at http://extranet.agricultura.gov.br/agrofit_cons/ principal_agrofit_cons (accessed on 1 March 2012).
- Moore, R.F. (1980) Behavioral and biological effects of NRDC-161 factors in control of the boll weevil. *Journal of Economic Entomology* 73, 265–267.
- Obrycki, J.J., Harwood, J.D., Kring, T.J. & O'neil, R.J. (2009) Aphidophagy by Coccinellidae: application of biological control in agroecosystems. *Biological Control* 51, 244–254.
- Pathan, A.K., Sayyed, A.H., Aslam, M., Razaq, M., Jilani, G. & Saleem, M.A. (2008) Evidence of field-evolved resistance to organophosphates and pyrethroids in *Chrysoperla carnea* (Neuroptera: Chrysopidae). *Journal of Economic Entomology* 101, 1676–1684.
- Penman, D.R. & Chapman, R.B. (1983) Fenvalerate induced distributional imbalances of two-spotted spider mite on bean plants. *Entomologia Experimentalis et Applicata* 33, 71–78.
- Plapp, F.W. Jr & Bull, D.L. (1978) Toxicity and selectivity of some insecticides to *Chrysopa carnea*, a predator of the tobacco budworm. *Environmental Entomology* 7, 431–434.
- Pothikasikorn, J., Overgard, H., Ketavan, C., Visetson, S., Bangs, M.J. & Chareonviriyaphap, T. (2007) Behavioral responses of malaria vectors, *Anopheles harrisoni complex*, to three classes of agrochemical in Thailand. *Journal of Economic Entomology* 44, 1032–1039.
- Reed, D.K. & Pike, K.S. (1991) Summary of an exploration trip to South America. *IOBC/Neartic Region Newsletter* 36, 16–17.
- Richetti, A., Melo Filho, G.A., Lamas, F.M., Staut, L.A. & Fabrício, A.C. (2004) Estimativa do custo de produção de algodão, safra 2004/05, para Mato Grosso do Sul e Mato Grosso. Dourados, Embrapa Pecuária Oeste (Embrapa, Comunicado Técnico, 91).
- Riedl, H. & Hoying, S.A. (1983) Toxicity and residual activity of fenvalerate to *Typhlodromus occidentalis* (Acari: Phytoseiidae) and its prey, *Tetranychus urticae* (Acari: Tetranychidae), on pear. *Canadian Entomologist* **115**, 807–813.
- Rocha, L.C.D., Carvalho, G.A., Moura, A.P., Moscardini, V.F., Rezende, D.T. & Santos, O.M. (2010) Seletividade fisiológica

de inseticidas utilizados em cultura cafeeira sobre ovos e adultos de *Cryptolaemus montrouzieri* Mulsant. *Arquivos do Instituto Biológico* **77**, 119–127.

- Rodrigues, A.R.S. (2012) Caracterização da resistência de joaninhas predadoras ao lambda-cialotrina. DSc. Thesis, Universidade Federal Rural de Pernambuco, Recife, Brazil.
- Roush, R.T. & Daly, J.C. (1990) The role of population genetics in resistance research and management. pp. 97–152 *in* Roush, R.T. & Tabashink, B.E. (*Eds*) *Pesticide Resistance in Arthropods*. New York, NY, USA, Chapman and Hall.
- Ruscoe, C.N.E. (1977) The new NRDC pyrethroids as agricultural insecticides. *Pesticide Science* 8, 236–242.
- SAS Institute (2001) SAS/STAT User's Guide, version 8.02, TS level 2MO. Cary, NC, SAS Institute Inc.
- Sayyed, A.H., Pathan, A.K. & Faheem, U. (2010) Cross-resistance, genetics and stability of resistance to deltamethrin in a population of *Chrysoperla carnea* from Multan, Pakistan. *Pesticide Biochemistry and Physiology* 98, 325–332.
- Soderlund, D.M. & Bloomquist, J.R. (1989) Neurotoxic actions of pyrethroid insecticides. *Annual Review of Entomology* 34, 77–96.
- Tabashink, B.E. & Johnson, M.W. (1999) Evolution of Pesticide Resistance in Natural Enemies. New York, NY, USA, Academic Press.
- Tillman, P.G. & Mulrooney, J.E. (2000) Effect of selected insecticides on the natural enemies Coleomegilla maculata and Hippodamia convergens (Coleoptera: Coccinellidae), Geocoris punctipes (Hemiptera: Lygaeidae), and Bracon mellitor, Cardiochiles nigriceps, and Cotesia marginiventris (Hymenoptera: Braconidae) in cotton. Journal of Economic Entomology 93, 1638–1643.
- Torres, J.B. (2012) Insecticide resistance in natural enemies seeking for integration of chemical and biological controls. *Journal of Biofertilizers and Biopesticides* 3, e104.
- Torres, J.B., Freitas, F.S. & Pratissoli, D. (1995) Avaliação de diferentes porcentagens da mistura de farinha de milho com farinha de trigo integral e levedura-de-cerveja na criação de *Anagasta kuheniella* (Zeller, 1879). *Revista Ciência e Prática* 19, 365–368.
- Torres, J.B., Silva-Torres, C.S.A., Silva, M.R. & Ferreira, J.F. (2002) Compatibilidade de inseticidas e acaricidas com o percevejo predador *Podisus nigrispinus* (Dallas) (Heteroptera: Pentatomidae) em algodoeiro. *Neotropical Entomology* 31, 311–317.
- Torres, J.B., Silva-Torres, C.S.A. & Oliveira, J.V. (2003) Toxicity of pymetrozine and thiamethoxam to Aphelinus gossypii and Delphastus pusillus. Pesquisa Agropecuária Brasileira 38, 459–466.
- Torres, J.B., Schetino, C.B. & Pratissoli, D. (2009) Controle biológico de pragas com uso de insetos predadores. *Informe Agropecuário* 30, 17–32.
- Wang, X., Shen, Z., Xu, W. & Lu, J. (2003) Sublethal effects of insecticides on fecundity of multicolored Asian ladybird Harmonia axyridis. Journal of Applied Ecology 14, 1345–1348.
- Whalon, M.E., Mota–Sanchez, D., Hollingworth, R.M. & Duynslager, L. (2011) Arthropod Pesticide Resistance Database. Available online at http://www.pesticideresistance. org/search/1 (accessed on 25 May 2012).