

Review

Cite this article: Spackman CN, Monaco TA, Stonecipher CA, and Villalba JJ (2020) Plant silicon as a factor in medusahead (*Taeniatherum caput-medusae*) invasion. *Invasive Plant Sci. Manag* **13**: 143–154. doi: [10.1017/inp.2020.20](https://doi.org/10.1017/inp.2020.20)

Received: 31 March 2020
Revised: 29 May 2020
Accepted: 3 July 2020
First published online: 14 July 2020

Associate Editor:

Edith Allen, University of California, Riverside


Keywords:

Plant physiology; plant fitness; weed control; invasion feedback

Author for correspondence:

Casey N. Spackman, New Mexico State University, MSC 3AE, P.O. Box 30001, Las Cruces, NM 88003-0031.
(Email: spackman@nmsu.edu)

Plant silicon as a factor in medusahead (*Taeniatherum caput-medusae*) invasion

Casey N. Spackman¹ , Thomas A. Monaco², Clinton A. Stonecipher³ and Juan J. Villalba⁴

¹Extension Range Management Specialist, New Mexico State University, Las Cruces, NM, USA; ²Research Ecologist, Forage and Range Research Laboratory, USDA-ARS, Logan, UT, USA; ³Rangeland Scientist, Poisonous Plants Research Laboratory, USDA-ARS, Logan UT, USA and ⁴Professor, Department of Wildland Resources, Utah State University, Logan, UT, USA

Abstract

Medusahead [*Taeniatherum caput-medusae* (L.) Nevski] is one of the most detrimental invasive annual grasses impacting the sustainability and function of rangeland in the western United States. This annual grass possesses high concentrations of tissue silicon (Si) that may facilitate invasion through key plant characteristics such as increased plant fitness, structure, and antinutritive qualities. These characteristics may affect known invasive processes such as increased plant productivity, slow litter decomposition, and decreased herbivory, facilitating a positive feedback cycle of invasion. However, Si is not considered an essential element and is often overlooked as a factor of *T. caput-medusae* invasion. Thus, this article provides a synthesis of plant Si, *T. caput-medusae*, and the self-reinforcing feedback cycle of invasion. We also discuss how current control strategies address plant characteristics determined by Si and suggest research avenues that may aid in novel or improved control strategies that target the *T. caput-medusae*–silica relationship.

Introduction

Medusahead [*Taeniatherum caput-medusae* (L.) Nevski] is one of the most detrimental invasive plants impacting rangeland sustainability and livestock operations (Johnson and Davies 2012; Mangla et al. 2011; Nafus and Davies 2014); it decreases wildlife habitat and plant diversity and increases the frequency of fires (Davies 2011). Recently, the integration of multiple control strategies has reestablished desirable plant species (Davies and Boyd 2018; Schantz et al. 2019), but the cost and uncertainty of success still exists for land managers (James et al. 2015). Consequently, there is a critical need to assess the underlying causes of *T. caput-medusae* invasion and how these causes impact traditional control strategies and to develop management strategies that decrease success uncertainty. These challenging questions can be partially addressed through a conceptual model that considers how tissue silicon (Si) concentration influences *T. caput-medusae* plant characteristics and the invasive processes.

High tissue Si concentration is a vital attribute of *T. caput-medusae*, as it forms mineral silica complexes within stems and an epidermal silica varnish on the leaves, awns, and glumes (Bovey et al. 1961; Epstein 1999; Swenson et al. 1964). The varnish hinders digestive processes within the rumen of herbivores (Montes-Sánchez and Villalba 2017) and contributes to a defense mechanism that facilitates aversive grazing behavior (Coskun et al. 2019; Hunt et al. 2008; McNaughton et al. 1985). The same varnish also limits leaf litter decomposition, causing a persistent litter or thatch layer (Torell et al. 1961; Young 1992). Accumulated litter inhibits establishment of non-*T. caput-medusae* seedlings and increases fine fuels, facilitating a grass-fire positive feedback cycle (Davies and Nafus 2013; Kyser et al. 2007; Young et al. 1999). Finally, high tissue Si concentrations may increase *T. caput-medusae* productivity directly through enhanced plant fitness and indirectly through accumulated litter, which may improve *T. caput-medusae* germination and root development (Evans and Young 1970). Consequently, *T. caput-medusae* Si concentrations are likely linked to increasing both the relative and overall abundance of the weed in plant communities, decreasing native species diversity, and reducing net primary productivity, and contribute to the creation of positive feedbacks that favor persistence of exotic species at the expense of native species (Elgersma et al. 2012; Inderjit and Cahill 2015; Suding et al. 2013).

The conceptual model we propose portrays linkages between tissue Si concentrations and a self-reinforcing positive feedback cycle of *T. caput-medusae* invasion. These linkages may enhance a more mechanistic understanding of the invasive processes and potentially refine *T. caput-medusae* control and restoration effort opportunities (Figure 1). This model illustrates how tissue Si concentrations directly influence critical plant characteristics (e.g., fitness, structure, and chemical composition), which subsequently impact fundamental invasive processes

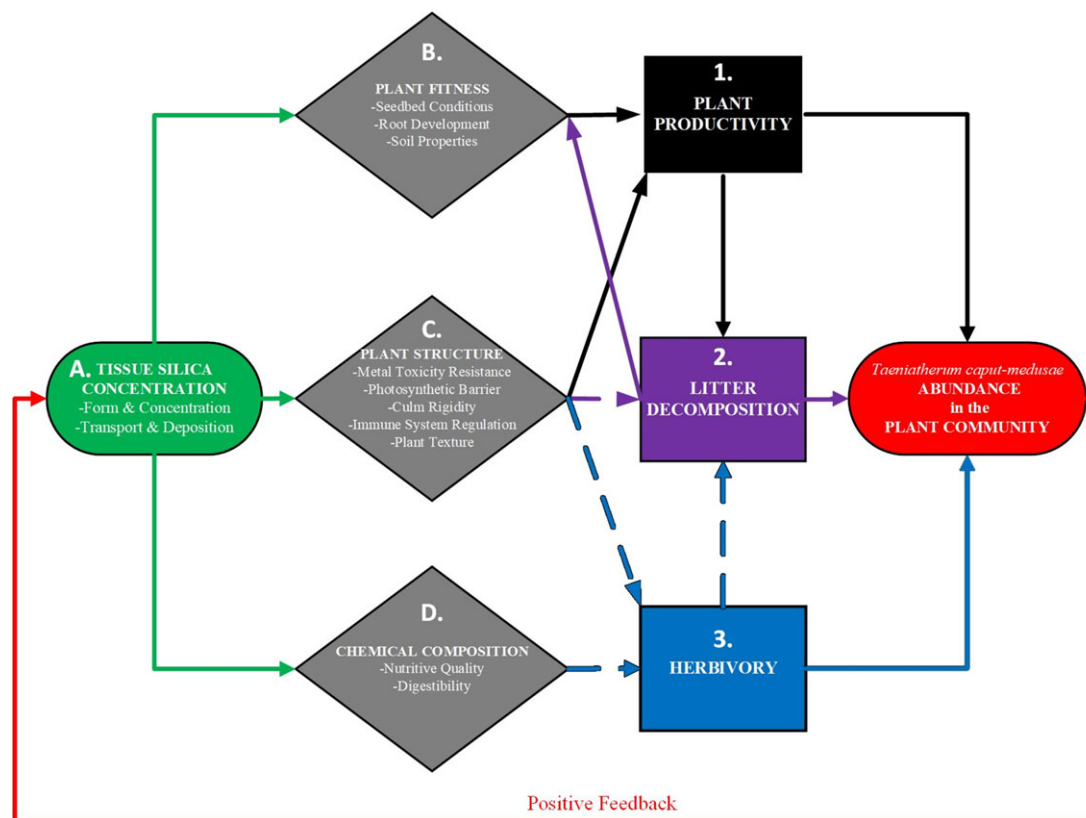


Figure 1. A model explaining the self-reinforcing positive cycle feedback of *Taeniatherum caput-medusae* invasion in relation to silicon (Si). Rhomboids represent plant characteristics; rectangles represent invasion processes; solid lines represent net positive effects and dashed lines net negative effects.

(e.g., plant productivity, litter decomposition, and herbivory). However, despite the connectivity between plant characteristics and invasive processes associated with *T. caput-medusae* abundance, close examination of the current knowledge base suggests that considerable uncertainty exists in how control strategies impact tissue Si concentrations.

Several categories of *T. caput-medusae* control (i.e., mechanical, cultural, and chemical) have been applied successfully to reduce and/or suppress the plant's abundance (Davies 2010; Kyser et al. 2007; Monaco et al. 2005; Stonecipher et al. 2016). However, tissue Si concentration is an underlying cause of invasion that is often overlooked, because Si is not considered an essential element of plant nutrition (Arnon and Stout 1939; Epstein 1999). Thus, the focus of this article will be to summarize the role of tissue Si in plants, identify *T. caput-medusae*'s characteristics and its invasive processes in relation to tissue Si using a conceptual model (Figure 1), address how control strategies address *T. caput-medusae* Si, and finally, suggest research avenues that may aid in novel or improved control strategies that target *T. caput-medusae*–silica relationships.

Silicon and Plants

Form and Concentration

Si is the second most abundant element found within the earth's crust (Epstein 1999), and the majority of plant Si exists as amorphous Si (i.e., phytoliths), which is insoluble in water. In contrast, its soluble form, orthosilicic acid, or simply silicic acid (Ma and Yamaji 2006), is the form taken up by plant roots and deposited

as the insoluble form, silica (Currie and Perry 2007; Epstein 1994, 1999). Often the literature does not distinguish between the two forms of Si, particularly with *T. caput-medusae*. Thus, unless soluble Si is directly stated herein, it is referring to the insoluble form, silica. Tissue silica varies widely among plant species (Hodson et al. 2005), and plants are generally classified as accumulators (1% to 10% silica on a dry matter basis) or nonaccumulators (<1%). Some monocots such as rice (*Oryza sativa* L.), sugarcane (*Saccharum officinarum* L.), and members of the Cyperaceae family (i.e., sedges) are considered hyperaccumulators, having greater than 10% silica in their tissues (Handreck and Jones 1967; Ma and Takahashi 2002; Neumann 2003). Consequently, many grasses are considered accumulators, but tissue silica varies widely depending on geographic location, phenological stage, and environmental conditions (McNaughton et al. 1985). For instance, tissue silica ranges between 1% and 4% in alpine tundra grasses, whereas African savanna grasses contain up to 20% (Johnston et al. 1968; McNaughton et al. 1985). Semiarid rangeland grasses are considered intermediate accumulators, with tissue silica ranging between 3% and 9% (Shewmaker et al. 1989), but *T. caput-medusae* stands out as a hyperaccumulator, with values ranging between 10% and 19% (Bovey et al. 1961; Epstein 1999; Swenson et al. 1964).

Transport and Deposition

The uptake of soluble Si varies based on species, genotype, and root structure, and its transport occurs via the transpiration stream (Ma and Yamaji 2006). Transport can be active, passive, and/or rejective (Ma et al. 2004; Mitani and Ma 2005; Takahashi et al. 1990).

For example, accumulation in grasses is believed to be a passive process, wherein these plants often display tissue silica concentrations reflective of soluble soil Si concentrations (Handreck and Jones 1967; Ma et al. 2001). In contrast, some dicots exhibit a rejective transport mechanism associated with a physical barrier in roots, allowing water and nutrient passage but limiting soluble Si uptake (Jones and Handreck 1969). Hyperaccumulators such as rice (Feng et al. 2011; Ma et al. 2007), barley (*Hordeum vulgare* L.) (Chiba et al. 2009), maize (*Zea mays* L.) (Mitani et al. 2008), wheat (*Triticum aestivum* L.) (Casey et al. 2004; Rains et al. 2006), and cucumber (*Cucumis sativus* L.) (Liang et al. 2005) display an active transport mechanism that facilitates soluble Si uptake and deposition (Chiba et al. 2009; Ma et al. 2006, 2007). In passive transport, soil moisture is one of the primary limiting factors of mineral uptake (Bloom et al. 1985; Hu and Schmidhalter 2005). In semiarid environments where soil moisture is limited, grass species display low to moderate levels of tissue Si concentrations (Shewmaker et al. 1989), likely due to passive transport. *Taeniatherum caput-medusae* has tissue silica concentrations similar to that of rice, although it is unknown whether it uses an active or passive transport mechanism. An active transport mechanism is more plausible, as concentrations of *T. caput-medusae* tissue silica are greater than those of other semiarid species (Bovey et al. 1961; Swenson et al. 1964).

As soluble Si is transported to various parts of a plant, it is dehydrated and polymerized into a di- or poly-silicic acid, which can be further polymerized within cellular structures to silica (Casey et al. 2004; Ma and Yamaji 2006; Mitani and Ma 2005). Cellular constituents such as hemicellulose, callose, pectin, and lignin have been shown to provide a framework for deposition (Guerriero et al. 2016). Tissue silica can also take on various forms, depending on deposition location, cell shape, and environmental conditions (Blackman 1971; Li et al. 2014). The location for silica deposition also varies greatly among plant species (Lewin and Reimann 1969; Lux et al. 2003). Tissue silica bodies have been observed in all major aboveground plant parts of *T. caput-medusae*, including sausage-shaped bodies beneath the epidermis of culms and a varnish-like structure on the awns as silicified barbs (Swenson et al. 1964).

Tissue Silicon and *Taeniatherum caput-medusae* Plant Characteristics

Plant Fitness

High tissue Si in *T. caput-medusae* may influence many factors associated with plant fitness that can be linked to plant productivity (Figure 1). For instance, tissue Si may increase root development, slow litter decomposition, alter soil conditions, and contribute to microsite conditions that favor *T. caput-medusae* germination. Overall, high tissue Si likely has a cascade of net positive effects on *T. caput-medusae* productivity and abundance within plant communities, which in turn strengthens the feedback cycle of invasion.

Seedbed Conditions

Successful seedling establishment in plants is often determined by the number of safe sites provided by the soil surface (Evans and Young 1970; Harper et al. 1965). Litter can create a favorable microenvironment for *T. caput-medusae* seeds that do not arrive at soil safe sites. For instance, as a consequence of high tissue Si concentrations, *T. caput-medusae* litter is slow to decompose (Figure 1), contributing to years of persistent decadent plant

material (Bovey et al. 1961; Young 1992). While many plant components such as lignin (Aerts 1997; Laishram and Yadava 1988; Stott et al. 1983), tannin-protein complexes (Palm 1988), and high C:N and C:P ratios (Goldman et al. 1987) reduce litter decomposition rates, Si is thought to be the primary contributor to slowed decay rates in *T. caput-medusae* (Torell et al. 1961; Young 1992). Evans and Young (1970) found that *T. caput-medusae* litter reduces seasonal and daily temperature fluctuations and water evaporation and increases humidity, which together contribute to a more favorable seedling microenvironment. The awns of *T. caput-medusae* seeds also get caught within the litter, limiting seed burial and intimate contact with the soil surface (Evans and Young 1970). For other plant species, this may pose a dire problem, as they require direct contact with a moisture-supplying substrate such as the soil, but *T. caput-medusae* seeds absorb moisture from the litter environment (i.e., they are hygroscopic) (Young 1992). In support of the favorable, self-induced seedbed conditions, *T. caput-medusae* was found to produce 47 times more seedlings within its litter environment than on bare ground conditions (Evans and Young 1970). This is not to say that *T. caput-medusae* cannot germinate without the presence of litter, but rather the accumulation of litter enhances *T. caput-medusae* establishment and persistence.

Root Development

Cheatgrass (*Bromus tectorum* L.) and *T. caput-medusae* roots develop at comparable rates and can grow to depths of 100 cm (Hironaka 1961). Both annual grasses primarily germinate in the autumn, continue slow root growth through the winter, and have greater root elongation into spring and summer than newly established native perennial grass species (Harris 1977; Hironaka 1961; Vasquez et al. 2008). Collectively, these unique root development traits enable *T. caput-medusae* and similar annual grasses to gain a competitive advantage over perennial plant species through greater resource acquisition (James et al. 2010; Vasquez et al. 2008; Young and Mangold 2008).

Taeniatherum caput-medusae diverges from other invasive annual grass species when it comes to root cell size and structure. For instance, Harris (1977) compared the root cellular composition of *T. caput-medusae*, *B. tectorum*, and a perennial grass through photomicrographs and showed that *T. caput-medusae* had thicker cell walls and overall larger root diameters than *B. tectorum*, but lower values than the perennial grass species. It was proposed that the thicker cellular root endodermis may allow for transpiration to occur even if the surrounding soil environment is drier, particularly in the upper soil horizons. When a Si-based fertilizer was applied to a variety of plant species (e.g., sorghum [*Sorghum bicolor* (L.) Moench], rice, wheat, and potatoes [*Solanum tuberosum* L.]), root endodermis cells were more rigid and efficient in water transpiration under moisture-lacking conditions (Chen et al. 2011; Crusciol et al. 2009; Gong et al. 2005; Hattori et al. 2003). In general, Si has been associated with increased plant tissue rigidity in several plant species (Ma et al. 2006; Namaganda et al. 2009; Raven 1983). Despite considerable information on Si and its beneficial role in other plants, there is very limited information pertaining to *T. caput-medusae* roots and how Si influences root development.

Soil Properties

Taeniatherum caput-medusae can be found on a variety of soil types, but more frequently in clayey soils as opposed to coarse-textured, sandy soils (Dahl and Tisdale 1975). Because clayey soils

are known for their high water-holding capacity and shrink–swell characteristics, which create large soil cracks and/or fissures when they dry, plant seedlings and their roots can experience increased desiccation as cracks develop (Young et al. 1999). However, *T. caput-medusae* roots are well adapted to this potential threat. For instance, if the primary root of *T. caput-medusae* is injured or dries out, an adventitious root emerges to replace the primary root (Young 1992). Vascular tissues have been found to be strengthened by Si accumulation and may be able to withstand the shrinkage-induced tissue stress from the drying of clayey soils (Hattori et al. 2003). With high tissue Si concentrations, *T. caput-medusae* likely responded more favorably (i.e., greater root growth) on clayey soils and large/infrequent watering treatments compared with other invasive annual grasses such as *B. tectorum* or *ventenata* [*Ventenata dubia* (Leers) Coss.] (Bansal et al. 2014).

Edaphic conditions, such as nutrient availability, may also be related to *T. caput-medusae* invasion. A transplant study between *T. caput-medusae* seeds from California and France showed that California soils produced larger plants than those from France (Blank and Sforza 2007). It was proposed that California soils have greater nutrient content than those from France. In a comparative growth study, nitrogen and phosphorus additions increased biomass production of *B. tectorum* more so than *T. caput-medusae* (Dakheel et al. 1993). In the same study, when soils were deficient in phosphorus, *B. tectorum* also outperformed *T. caput-medusae*. Similarly, *B. tectorum* produced more shoot mass than *T. caput-medusae* when grown in different nitrogen nutrient solutions (MacKown et al. 2009). Nevertheless, none of these studies reported soil or plant Si concentrations, likely due to the fact that Si is not considered an essential element of plant nutrition (Arnon and Stout 1939; Epstein 1972). Because Si has been shown to increase root and leaf elongation in hyperaccumulator plant species (i.e., sorghum and rice) (Hattori et al. 2003; Hossain et al. 2002), it is possible that the combination of Si dissolution in clayey soils (Tubaña and Heckman 2015) and the potentially high uptake capacity of soluble Si by *T. caput-medusae* may facilitate increased growth and offset the competitive superiority of *B. tectorum* described in the aforementioned studies.

Plant Structure

Tissue Si may influence many plant structural factors of *T. caput-medusae*, such as metal toxicity resistance, protection from ultraviolet-b (UV-B) radiation damage, increased culm rigidity, and upregulation of the innate immune system. These factors may directly decrease litter decomposition rate and herbivory, indirectly increase plant fitness and productivity, and contribute to the overall abundance of *T. caput-medusae* in the plant community (Figure 1).

Metal Toxicity Resistance

The accumulation of toxic metals can negatively impact plant fitness (Nagajyoti et al. 2010), and evidence suggests that tissue Si mitigates this toxicity through structural adaptations within the plant (Ma 2004). An apoplastic root membrane was shown to regulate the translocation of metals through plant cell walls (Emamverdian et al. 2018). Deposition of Si within the membrane was thought to decrease the porosity of the cell wall, consequently reducing the movement of toxic metals and salts into the plant (Coskun et al. 2019; Gong et al. 2006; Wu et al. 2013). Some toxic metals may pass through the membrane, but soluble Si can form mineral complexes to increase adsorption and reduce toxic metal

movement within plants (Keller et al. 2015; Wang et al. 2004; Ye et al. 2012). However, Adrees et al. (2015) suggested that changes in root structure were not the only mechanism of decreased toxic metal uptake. Modulation of the influx transporters by Si may also decrease toxic metal uptake. *Taeniatherum caput-medusae* was found to colonize waste mine sites high in toxic metals, although the aforementioned mechanisms of reduced metal toxicity were not addressed (Martinez-Ruiz et al. 2001; Reglero et al. 2008). Consequently, these mechanisms could reduce metal toxicity in *T. caput-medusae*, contributing to its invasion, but further exploration is needed.

Photosynthetic Barrier

Reductions in the atmospheric ozone layer have resulted in increased UV-B radiation reaching the earth's surface (Madronich et al. 1998). Increased UV-B radiation can damage leaves and reduce the photosynthetic capacity of plants (Kakani et al. 2003). Supplementing rice with soluble Si reduced UV-B damage through increased leaf rigidity and the production of phenolic compounds (Li et al. 2004; Tamai and Ma 2008). The presence of tissue Si within the cellular membranes also mitigated UV-B–induced reactive oxygen species, which are known to cause membrane damage, decrease enzyme activity, increase electrolyte leakage, and alter gene expression (Coskun et al. 2019; Shen et al. 2010). Despite these studies, the relationship between photosynthesis and tissue Si in *T. caput-medusae* has not been evaluated.

Culm Rigidity

Taeniatherum caput-medusae plants can reach a height of more than 60 cm, which is impressive, given that its culms are typically thin (e.g., <1-mm diameter) and inflorescences are relatively large and “wispy” (McKell et al. 1962), possibly making them susceptible to lodging and breakage (Savant et al. 1996). Culm integrity of *T. caput-medusae* is potentially enhanced due to the accumulation of silica, which increases the rigidity of the stem and leaves of certain plants (Ma et al. 2006; Namaganda et al. 2009). For instance, rice lodging was reduced with increasing tissue Si concentrations (Lee et al. 1990; Savant et al. 1996). However, no studies have examined the associations between culm rigidity in *T. caput-medusae*, tissue Si, and lodging.

Immune System Regulation

Wagner (1940) was the first to suggest that tissue Si increases plant defenses against stem-boring pathogens; this is known as the mechanical barrier hypothesis. However, studies in rice (Rodrigues et al. 2003, 2004) and wheat (Bélanger et al. 2003; Rémus-Borel et al. 2009) have shown that there is an upregulation of the innate immune system (e.g., chitinases, peroxidase, polyphenol oxidases, phytoalexins, and phenolic compounds) following predation rather than the aforementioned physical defense. Tissue Si is known to act as a modulator of these plant defenses. For instance, tissue Si–associated defenses increase the binding affinity of plant proteins within the defense signaling pathway, preventing these pathogenic enzymes from reaching their target (Datnoff et al. 2007; Fautoux et al. 2005). Recent work on pathogens and stem-boring insects demonstrated that these predators release effector proteins that interfere with the defense signaling pathway (Giraldo and Valent 2013; Mugford et al. 2016). Soluble Si deposition strengthens the apoplastic membrane and prevents the effector proteins from reaching their target (Holub and Cooper 2004; Nuernberger and Lipka 2005). Another postulation is that tissue Si allows for the redistribution of energy stores to upregulate

defense responses. For instance, Si-accumulating plants may replace energy-dependent processes involved in building structural carbohydrates for non-energy dependent ones (McNaughton et al. 1985; Van Soest 2006). Energy stores that would otherwise be used for growth and development of plant structural components are redistributed to the innate immune system (Coskun et al. 2019). Overall, the redistribution of energy stores and production of a tissue Si physical barrier allow for a stronger defense response upon predation. However, these theories on tissue Si and the upregulation of the innate immune response have not been explored in *T. caput-medusae*.

Plant Texture

Invertebrate insects (Massey et al. 2006), voles (Massey and Hartley 2006), rabbits (Cotterill et al. 2007), livestock (Massey et al. 2009; Shewmaker et al. 1989), small granivores, and birds (Longland 1994; Savage et al. 1969) all show decreased preference for Si-rich grasses. This avoidance may be a consequence of an undesirable oral texture. For instance, *T. caput-medusae* inflorescences have a large vertical awn attached to the seed, with shorter lateral spiked glumes (Miller et al. 1999). Microscopic examination of *T. caput-medusae* awns revealed that they contain a Si varnish with barbs (Swenson et al. 1964), which may contribute to the undesirable texture causing irritation to mouthparts (Massey and Hartley 2009). In addition, tissue Si in grasses has been associated with increased tooth wear (Baker et al. 1959), gastrointestinal urolithiasis (Bailey 1981), and esophageal tumors (O'Neill et al. 1982, 1980), possibly causing further *T. caput-medusae* avoidance.

Disarticulation of seeds from the seed head occurs from July to October, with the majority being dropped in August (Davies 2008), providing an opportunity for herbivory without the consequence of oral irritation. However, Si bodies are still present in the stem (Swenson et al. 1964), and likely in the spiked glume after seed disarticulation, and continue to provide an abrasive texture. In contrast, other annual grasses such as *B. tectorum* lack these short spiked glumes, and when seed drop occurs, the plants become susceptible to herbivory (Vallentine and Stevens 1994). Overall, the texture caused by *T. caput-medusae* tissue Si may deter herbivory of *T. caput-medusae*, facilitating a self-reinforcing positive feedback cycle.

Chemical Composition

Tissue Si concentrations have a direct effect on the nutrient content and digestibility of *T. caput-medusae* (Figure 1). These effects negatively impact herbivory and increase litter accumulation, which both directly and indirectly increase *T. caput-medusae* abundance within the plant community.

Nutritive Quality

Tissue nutrient concentrations and their association to plant secondary compounds or toxins largely influence the likelihood of a plant being consumed by an herbivore (Provenza et al. 2002). Bovey et al. (1961) showed that *T. caput-medusae* contains 10.4% crude protein (CP), 2.6% fat, 26.8% crude fiber, 6.1% lignin, and 13.9% ash during the vegetative phenological stage. Subsequently, rapid maturation causes a decline in CP and fat content, while fiber, lignin, and ash concentrations increase. Other more recent studies show similar trends for the nutritional composition of *T. caput-medusae* over time (Montes-Sánchez and Villalba 2017; Villalba and Burritt 2015). Plant ash is an indicator of tissue silica content, and silica in ash comprises values greater

than 90% in rice plants (Charca et al. 2007; Yalçın and Sevinç 2001). These values can be lower, depending on plant species and tissue location (Lanning et al. 1980; Lanning and Eleuterius 1983). *Taeniatherum caput-medusae* ash comprises more than 70% tissue silica, whereas *B. tectorum* comprises 47% (Bovey et al. 1961). In the early phenological stages, *T. caput-medusae* has similar nutritional value to that of other desirable grass, and livestock grazing can be used as a method of control (Bovey et al. 1961; DiTomaso et al. 2008; Hamilton et al. 2015; Villalba and Burritt 2015). Because of its rapid maturation, decline in nutritional value, and high silica concentrations, the window of palatability is limited to 2 to 4 wk (Brownsey et al. 2017). After this period, consumption of *T. caput-medusae* is reduced, and grazing pressure on more nutritious plant species may occur and be deleterious (DiTomaso and Smith 2012; Sheley et al. 2008). Even if *T. caput-medusae* was palatable after seed drop, the nutritional quality of the plant is inadequate to sustain grazers. Domestic ungulates require a minimum of ~7.5% CP for body maintenance (National Research Council 2000, 2007a, 2007b), and supplementation is required for animals grazing most grasses at later phenological stages (Ganskopp and Bohnert 2001). Animal supplementation of nutrient-rich forages in combination with *T. caput-medusae* were shown to increase consumption of the troublesome grass (Hamilton et al. 2015; Montes-Sánchez et al. 2017; Stonecipher et al. 2016). Nevertheless, in these studies, *T. caput-medusae* consumption was low and would not likely discourage *T. caput-medusae* abundance within the plant community. Additionally, tissue Si was unaltered when fed to grazers, and thus the defense mechanism of the plant still existed, possibly limiting utilization of the chemical constituents within. Supplemental grazing of *T. caput-medusae* in the autumn also does not reduce the soil seedbank, as seed drop has already occurred, and may actually increase *T. caput-medusae* abundance in subsequent years (DiTomaso et al. 2008; Lusk et al. 1961). These studies indicate that similar to plant texture (discussed earlier), chemical composition alone does not explain the low intake and palatability of *T. caput-medusae*.

Digestibility

Tissue Si within plants has been proposed as a defense mechanism against herbivory (Hunt et al. 2008), similar to those induced by plant secondary compounds or toxins (Provenza et al. 2002). An epidermal silica varnish in *T. caput-medusae* acts as a physical barrier limiting the degradation of the cell wall by rumen microorganisms (Hunt et al. 2008; Montes-Sánchez and Villalba 2017). This barrier prevents utilization of the organic constituents beneath such a layer (Mayland and Shewmaker 2001; Van Soest 1994; Van Soest and Jones 1968). Support for this reduced digestion mechanism was shown by Montes-Sánchez and Villalba (2017), who reported declines in digestibility of *T. caput-medusae* with increments in the plant's particle size; with larger particles conserving the tissue Si barrier to a greater extent than particles of a smaller size. Although lignin content has been associated with decreased digestibility of certain plants, differences in *T. caput-medusae* digestion rates were not influenced by this factor, as alfalfa (*Medicago sativa* L.) and tall fescue (*Schedonorus arundinaceus* (Schreb.) Dumort., nom. cons.) hay had lignin concentrations similar to those present in *T. caput-medusae*. In addition, soluble Si limits enzymatic activity (Kind et al. 1954), thus decreasing plant digestibility in the rumen (Smith et al. 1971). Finally, for every percentile unit increase in tissue silica, there are between 1 and 3 percentile units of reduction in forage digestibility (Smith et al. 1971;

Van Soest and Jones 1968). In avian species, such as chukar partridges (*Alectoris graeca* Meisner), *B. tectorum* seeds were preferred over *T. caput-medusae* seeds (Savage et al. 1969), likely due to undesirable high tissue Si in *T. caput-medusae* and a concomitant lower digestibility, but other factors such as seed size may influence ingestion.

Traditional Control Strategies

Commonly used *T. caput-medusae* control strategies can be categorized into mechanical, cultural, and chemical. Each addresses certain aspects of the Si positive feedback cycle (Figure 1). Integrating multiple strategies is often more successful than using one strategy by itself (Davies 2010; Davies and Sheley 2011; Kyser et al. 2007; Monaco et al. 2005). This section will address individual strategies and their roles in the Si positive feedback cycle, potentially giving insight into why integrated control is often more successful.

Mechanical Control

Mowing

A window of opportunity for mowing *T. caput-medusae* was proposed, which was ~35 d from late vegetative to early reproductive phenological stage (Brownsey et al. 2017). The same study estimated that mowing before seed head emergence decreased *T. caput-medusae* seed production by ~50%. However, early spring mowing may miss younger plants and allow recovery of damaged plants, enabling *T. caput-medusae* to produce seeds and only temporarily reduce plant productivity. The optimal timing of mowing and control should be tailored to occur when the majority of plants are in the boot phenological stage, thus limiting seed production and additional late-maturing plants. Plant Si has been shown to increase plant tissue rigidity (Ma et al. 2006; Namaganda et al. 2009; Raven 1983) and potentially decrease microbial degradation, increasing litter accumulation (Torell et al. 1961; Young 1992). Mowing mulches the standing biomass and may artificially accelerate litter decomposition, but it may also create favorable microsites for *T. caput-medusae* seedlings (Evans and Young 1970). *Taeniatherum caput-medusae* seeds remain viable for up to 3 yr (Nelson and Wilson 1969); thus, repeated mowing treatments must be applied, which may lead to deleterious impacts on native plant species (Davies et al. 2012). The small window of opportunity for mowing may also be impractical for repeated application over large-scale infestations. Finally, *T. caput-medusae* infestations often occur on landscapes that are inaccessible to machinery due to slope and terrain, as well as being on uneven, rocky soils that can damage equipment (Young 1992). Overall, mowing may partially address the plant fitness and structural characteristics of *T. caput-medusae* within the Si positive feedback cycle (Figure 1), but it does not address the chemical composition of the plant.

Tillage

Tilling can break up litter, incorporating it into the soil, and create favorable seedbed conditions for revegetation efforts (Kaltenecker 1997). In addition, tilling can damage *T. caput-medusae* roots, bury germinable seeds, and reduce *T. caput-medusae* cover by ~50% (Kyser et al. 2007). These impacts on *T. caput-medusae* address both plant fitness and structure in the Si positive feedback cycle (Figure 1). It has been shown that *T. caput-medusae* seeds emerge poorly at depths greater than 5 cm (Young et al. 1969). However,

tillage may also increase erosion potential, facilitate soil moisture and organic matter losses, and have deleterious effects on native vegetation (Nafus and Davies 2014; Pierson et al. 2007). Soil disturbance has been shown to favor *T. caput-medusae* invasion through increased standing biomass and seed production in the subsequent year (Miller 1996). As with mowing, tillage may not be feasible, due to topographic features and surface obstacles that could damage equipment. Finally, tillage may negatively impact fragile biological crusts and interrupt nutrient cycling, all while providing a disturbance regime that favors additional undesirable exotic plant species (Kaltenecker 1997; Locke and Bryson 1997; Young 1992).

Cultural Control

Prescribed Fire

Prescribed fire can consume standing *T. caput-medusae* biomass, seeds, and litter (Kyser et al. 2008), while potentially interrupting aspects of the Si positive feedback cycle (Figure 1). Fire may also provide an opportunity for revegetation and additional treatments (Davies 2010; Davies et al. 2015). *Taeniatherum caput-medusae* matures 2 to 4 wk later than other annual grasses, including *B. tectorum* (Dahl and Tisdale 1975; Hironaka 1961; Young 1992), and retains its seeds within inflorescences until August (Davies 2008). Thus, timing a prescribed fire to coincide with viable seed retention can destroy these seeds. Coordinating the ideal timing and achieving suitable conditions for a prescribed fire may be difficult in the Intermountain Region of the western United States due to low air temperatures in the autumn/winter and insufficient amounts of combustible material in the spring and early summer (Kyser et al. 2008). Delaying prescribed fires to late summer or early autumn raises the risk of escape due to the contribution of other community vegetation to the overall fuel load. After seed drop, fires must reach temperatures greater than 250 C to consume seeds located on the soil surface (DiTomaso et al. 1999; Sweet et al. 2008). Slow-burning, high-temperature fires may result in unintended damage to desirable plant species, increasing the dominance of fire-adapted invasive annual grasses (Billings 1994; Whisenant 1990), possibly perpetuating the grass-fire feedback cycle (D'Antonio and Vitousek 1992). Uncontrolled fires have been shown to create opportunities for exotic plant establishment and contribute to air pollution and atmospheric CO₂ that impact the climate (Campbell and Cahill 1996; D'Antonio and Vitousek 1992; Peters and Bunting 1994; Young 1992). Fire also alters soil infiltration rates, soil porosity, soil hydraulic conductivity, and soil water storage capacity (Neary et al. 1999). Despite these difficulties and risks, under optimal timing and conditions, prescribed fire has been shown to successfully reduce *T. caput-medusae* populations in the California annual grasslands (Kyser et al. 2008; Meyer and Schiffman 1999).

Grazing

Livestock grazing has been shown to be one of the preferred methods of *T. caput-medusae* control due to its low cost and practicality (Hamilton et al. 2015; James et al. 2015; Johnson et al. 2011), positive effects on nutrient cycling (Davies et al. 2010; Hobbs 1996), and limited disturbance compared with other control methods (e.g., mowing, tillage, and fire). For instance, livestock grazing removes standing vegetation and litter, and the deposition of animal wastes (i.e., feces and urine), which are high in nitrogen (N), can directly influence soil N mineralization (Davies et al. 2010; Hobbs 1996). However, livestock consumption

of *T. caput-medusae* has a narrow time frame of palatability before seed head emergence; livestock tend to avoid the plant after this time (DiTomaso et al. 2008; Young 1992). Livestock avoidance may be due to high tissue Si concentration, which presents an abrasive oral texture (McNaughton et al. 1985) and limits rumen digestibility (Montes-Sánchez and Villalba 2017). The plant's structure and chemical composition may directly and indirectly increase *T. caput-medusae* abundance in the plant community (Figure 1) through reduced livestock consumption and repeated defoliation of other, more palatable grasses (Belsky and Gelbard 2000; Heady 1961; Hunt et al. 2008; Mueggler 1972). Grazing before seed head emergence has been shown to overcome these obstacles, but the window of opportunity is narrow (Brownsey et al. 2017) and high stocking rates when *T. caput-medusae* is most palatable may not be feasible (DiTomaso et al. 2008). Grazing has been shown to be successful in the California annual grasslands when proper timing and stocking rates are considered (DiTomaso et al. 2008; Lusk et al. 1961). In contrast, year-round grazing and early spring grazing saw increases in *T. caput-medusae* abundance (DiTomaso et al. 2008; Harrison et al. 2003). Overall, grazing can be used as a method of control to address the Si positive feedback cycle (Figure 1) if high-intensity grazing is timed to occur when *T. caput-medusae* is in the boot phenological stage.

Chemical Control

Herbicides are the primary form of chemical control of *T. caput-medusae* and have the potential to reduce plant fitness, alter plant structural components, and increase nutritive quality, consequently interrupting the Si positive feedback cycle (Figure 1). There are many herbicides labeled for the control of *T. caput-medusae*, including PRE, POST, broad-spectrum, grass-selective, and growth-regulator herbicides. Imazapic and rimsulfuron are two PRE herbicides that have been used to control *T. caput-medusae* (Davies 2010; Davies and Sheley 2011; Kyser et al. 2007, 2012b; Monaco et al. 2005; Sheley et al. 2007), but they are highly variable in their efficacy (Kyser et al. 2007, 2012b). This may in part be due to the accumulated litter inhibiting penetration of the herbicide to the soil surface (Kyser et al. 2007) or rapid degradation in warmer soils (Kyser et al. 2012b). A new herbicide, indazaflam, has been shown to effectively control annual grasses through its PRE residual capacity (Sebastian et al. 2016a, 2016b, 2017a, 2017b), but it is not yet labeled for areas grazed by livestock. This poses a problem, as most areas heavily infested with *T. caput-medusae* are also used for livestock production. Aminopyralid, a PRE growth-regulator herbicide, was shown to suppress *T. caput-medusae* seed production >95% in a greenhouse setting (Rinella et al. 2014), with similar success in the field (Rinella et al. 2018). Glyphosate, a broad-spectrum POST herbicide, has been used at low rates to decrease *T. caput-medusae* abundance by ~95% with limited damage to shrubs and other native vegetation in big sagebrush (*Artemisia tridentata* Nutt.)-scrub ecosystems (Kyser et al. 2012a). Glyphosate application before seed set may also temporarily decrease plant fitness by reducing seed production and viability and arresting root development (Figure 1). However, multiple applications are needed, as glyphosate is a foliar contact herbicide and does not directly influence the soil seedbank (Kyser et al. 2012a). Even with the potential of *T. caput-medusae* herbicide control, there are risks associated with herbicide application. For instance, spray drift due to wind, volatilization, and different herbicide formulations may cause nonuniform application and injury to non-target species (DiTomaso 1997). Repetitive herbicide

application selects for herbicide-tolerant plants and may decrease desirable plant species within the population (DiTomaso 1997). Chemical application is often temporary, with multiple applications needed, but has shown potential for *T. caput-medusae* control and interrupting the Si positive feedback cycle.

Research Needs

Although we know orthosilicic acid is the soluble form taken up by many plants (Ma and Yamaji 2006) and deposited as amorphous mineral silica in *T. caput-medusae* tissues (Bovey et al. 1961; Epstein 1999; Swenson et al. 1964), critical knowledge gaps exist. In particular, our understanding of how Si is transported through the soil to the root of *T. caput-medusae*, how roots respond to available Si, what mechanisms within the root are responsible for uptake, and how the Si is transported through the plant are largely unknown. In addition, addressing ways to constrain tissue Si deposition within *T. caput-medusae* may be key to interrupting the positive feedback cycle of invasion and facilitate control of *T. caput-medusae*. Consequently, there are critical research needs in relation to plant Si and *T. caput-medusae* that may lead to alternative methods of control and more successful management strategies.

The Soil

The normal range of soluble Si in soil is between 0.6 to 1.0 mM but can be higher or lower depending on soil weathering processes (Epstein 1994). When soluble Si concentrations exceed 2 mM, the Si polymerizes into a gel and becomes unavailable for plant uptake (Ma et al. 2001). Rice and an *Equisetum* species began to wilt and leaves became necrotic when grown in deficient soluble Si conditions (Chen and Lewin 1969; Yoshida et al. 1962). Reduced growth and production yields were also observed in Gramineae species when grown in similar deficient conditions (Vlamis and Williams 1967). Thus, soil concentrations of soluble Si play a fundamental role in plant uptake and fitness for these species.

Understanding the role of soil soluble Si in *T. caput-medusae*-invaded landscapes could help in improving and/or developing better control strategies. Similar to the aforementioned Si hyper-accumulators, Si-deficient soils may reduce *T. caput-medusae* fitness (Figure 1). Nevertheless, it is unknown how low soil values of soluble Si impact *T. caput-medusae*. If the soil soluble Si deficiency can reduce *T. caput-medusae* fitness, novel studies could develop control strategies that decrease available soil Si. Because soluble Si polymerizes at concentrations greater than 2 mM (Ma et al. 2001), future studies could help develop strategies that target reductions or increases in soil soluble Si concentrations, limiting uptake and consequently decreasing *T. caput-medusae* fitness.

The Root

Fitness of rice plants has been associated with the ability to take up soluble Si through the root (Takahashi et al. 1990) and uptake regulation by the *Ls1* gene (Ma et al. 2004). Modification of the active transport mechanisms in rice has led to increased plant production and disease resistance (Ma et al. 2006). If increased plant fitness can be achieved by identifying the uptake mechanism of soluble Si in rice, it is reasonable to conclude that identifying the transport mechanism in *T. caput-medusae* might enable modifications to be made to reduce *T. caput-medusae* fitness. Nevertheless, the practicality of applying these modifications to *T. caput-medusae*

in highly variable ecological system will require considerable research before implementation.

Chemical application, rather than genetic modification, may provide a more viable means of interrupting the uptake mechanism. For instance, in a laboratory setting, the active transporter in rice was inhibited through the use of metabolic inhibitors such as NaCN and 2,4-dinitrophenol at low temperatures (Ma et al. 2004). Although these chemicals would be hazardous if applied in a landscape-type setting, this research sets the stage for exploring environmentally friendly chemicals that could have similar beneficial results. While previous methods of restricting Si uptake rely on an active transport mechanism, if a passive transport mechanism exists, the addition of specific soil minerals may provide a form of competitive adsorption, reducing soluble Si uptake into the plant. However, identifying the uptake mechanism is critical before exploring soluble Si inhibition techniques.

When compared with roots of other grasses, the roots of *T. caput-medusae* were found to have thicker cell walls and overall larger root diameters than an annual grass, but values were lower than those of a perennial grass species (Harris 1977). A thicker root endodermis may allow for transpiration to occur even if the surrounding soil environment is drier, particularly in the upper soil horizons; however (Harris 1977), root Si concentrations were not reported. Evaluating root Si concentrations may give insight into why *T. caput-medusae* is superior in acquiring soil resources and tolerating soil environmental fluctuations (e.g., drought, temperature extremes). If root tissue Si plays a role in *T. caput-medusae* fitness, opportunities for research become available for discovering how deposition of soluble Si occurs in roots. This may lead to control strategies that manipulate soluble Si deposition processes within roots, decrease *T. caput-medusae* fitness, and reduce invasive superiority. Overall, little is known about *T. caput-medusae* roots in relation to Si uptake and tissue silica concentrations; thus, further examination is needed.

The Shoot

Little information is available about soluble Si transport and deposition as amorphous silica within tissues. Manipulation and reduction of Si deposition would be key to increased herbivore preference and digestibility and would make the plant texture less abrasive for herbivores. For instance, sublethal doses of a glyphosate-containing herbicide with a surfactant were shown to reduce tissue silica concentrations and increase tillering in quackgrass [*Elymus repens* (L.) Gould] (Coupland and Caseley 1975). Similarly, low rates of glyphosate increased the nutritional quality of annual ryegrass (*Lolium rigidum* Gaudin) (Armstrong et al. 1992; Gatford et al. 1999) and increased cattle preference for glyphosate-treated fescue pastures (Kisseberth et al. 1986). However, tissue Si concentrations were not reported in these last three studies. Understanding how glyphosate alters the soluble Si deposition process and increases nutritional composition may give insight into other chemical treatments with similar results.

Aspects of plant structure (e.g., toxicity resistance, photosynthetic barrier, culm rigidity, immune system response) are possibly linked to *T. caput-medusae* tissue Si, but these aspects need further study. For instance, toxic metal accumulation in plants may affect plant productivity (Adrees et al. 2015; Nagajyoti et al. 2010), increased UV-B radiation could damage leaves and reduce the photosynthetic capacity of plants (Kakani et al. 2003), low tissue Si can increase susceptibility of culms to breakage in rice (Lee et al. 1990; Savant et al. 1996), and a weak immune system response

to predation has detrimental consequences to fitness (Holub and Cooper 2004; Nuernberger and Lipka 2005), whereas tissue Si mitigates these constraints. These principles and processes are known for crops; however, they are unexplored for *T. caput-medusae*. Elucidation of the influence of Si on *T. caput-medusae* fitness could lead to more efficient and/or novel control strategies for *T. caput-medusae*, interrupting its invasive superiority.

Conclusion

Taeniatherum caput-medusae is a superior invasive annual grass overtaking rangeland ecosystems and leading to drastic changes in ecosystem services. While control strategies exist for invasive annual grasses, these strategies do not take into consideration the role of high tissue Si in *T. caput-medusae* invasion. High tissue concentration of Si in *T. caput-medusae* creates a self-reinforcing feedback cycle that contributes to its invasive success and difficulty of control. The proposed conceptual model in this article summarizes *T. caput-medusae* characteristics associated with Si content (e.g., plant fitness, structure, and chemical composition) and the invasive processes associated with these characteristics (e.g., plant productivity, litter decomposition, and herbivory). These characteristics and processes help to identify critical research needs that can be used to develop improved and/or novel management practices of control, strengthening current strategies. Interrupting the Si positive feedback cycle of *T. caput-medusae* invasion should be considered when researching and developing future management strategies.

Acknowledgments. This research was funded by a Western SARE grant (SW15-003) in collaboration with Utah State University Department of Wildland Resources and USDA Agriculture Research Service, Logan, UT. No conflicts of interest have been declared.

References

- Adrees M, Ali S, Rizwan M, Zia-ur-Rehman M, Ibrahim M, Abbas F, Farid M, Qayyum MF, Irshad MK (2015) Mechanisms of silicon-mediated alleviation of heavy metal toxicity in plants: a review. *Ecotoxicol Environ Saf* 119:186–197
- Aerts R (1997) Climate, leaf litter chemistry and leaf litter decomposition in terrestrial ecosystems. *Oikos* 79:439–449
- Armstrong RD, Simpson RJ, Pearce GR, Radojevic I (1992) Digestibility of senescing annual ryegrass following application of glyphosate. *Aust J Agric Res* 43:871–885
- Arnon DI, Stout PR (1939) The essentiality of certain elements in minute quantity for plants with special reference to copper. *Plant Physiol* 14:371
- Bailey CB (1981) Silica metabolism and silica urolithiasis in ruminants: a review. *Can J Anim Sci* 61:219–235
- Baker G, Jones LHP, Wardrop ID (1959) Cause of wear in sheep's teeth. *Nature* 184:1583–1584
- Bansal S, James JJ, Sheley RL (2014) The effects of precipitation and soil type on three invasive annual grasses in the western United States. *J Arid Environ* 104:38–42
- Bélanger RR, Benhamou N, Menzies JG (2003) Cytological evidence of an active role of silicon in wheat resistance to powdery mildew (*Blumeria graminis* f. sp. *tritici*). *Phytopathology* 93:402–412
- Belsky AJ, Gelbard JL (2000) *Livestock Grazing and Weed Invasions in the Arid West*. Portland: Oregon Natural Desert Association. 31 p
- Billings WD (1994) Ecological impacts of cheatgrass and resultant fire on ecosystems in the western Great Basin. Pages 22–30 in Mosen SB, Kitchen SG, eds. *Proceedings—Ecology and Management of Annual Rangelands*. Gen. Tech. Rep INT-GTR-313. Ogden, UT: U.S. Department of Agriculture, Forest Service

- Blackman E (1971) Opaline silica bodies in the range grasses of southern Alberta. *Can J Bot* 49:769–781
- Blank RR, Sforza R (2007) Plant–soil relationships of the invasive annual grass *Taeniatherum caput-medusae*: a reciprocal transplant experiment. *Plant Soil* 298:7–19
- Bloom AJ, Chapin FS, Mooney HA (1985) Resource limitation in plants—an economic analogy. *Annu Rev Ecol Syst* 16:363–392
- Bovey RW, Tourneau DL, Erickson LC (1961) The chemical composition of medusahead and downy brome. *Weeds* 9:307–311
- Brownsey P, James JJ, Barry SJ, Becchetti TA, Davy JS, Doran MP, Forero LC, Harper JM, Larsen RE, Larson-Praplan SR, Zhang J, Laca EA (2017) Using phenology to optimize timing of mowing and grazing treatments for medusahead (*Taeniatherum caput-medusae*). *Rangeland Ecol Manag* 70:210–218
- Campbell D, Cahill TA (1996) Air quality impact of forest burning in the Sierra Nevada. Proceedings of the Forest Vegetation Management Conference (USA), Redding, CA, January 16–18, 1996
- Casey WH, Kinrade SD, Knight CTG, Rains DW, Epstein E (2004) Aqueous silicate complexes in wheat, *Triticum aestivum* L. *Plant Cell Environ* 27:51–54
- Charca G, Guzman B, Flora R (2007) Estudio para la obtencion de silice amorfa a partir de la cascara de arroz. *Acta Microscopica* 16:212–213
- Chen C, Lewin J (1969) Silicon as a nutrient element for *Equisetum arvense*. *Can J Bot* 47:125–131
- Chen W, Yao X, Cai K, Chen J (2011) Silicon alleviates drought stress of rice plants by improving plant water status, photosynthesis and mineral nutrient absorption. *Biol Trace Elem Res* 142:67–76
- Chiba Y, Mitani N, Yamaji N, Ma JF (2009) HvLsi1 is a silicon influx transporter in barley. *Plant J* 57:810–818
- Coskun D, Deshmukh R, Sonah H, Menzies JG, Reynolds O, Ma JF, Kronzucker HJ, Bélanger RR (2019) The controversies of silicon's role in plant biology. *New Phytol* 221:67–85
- Cotterill JV, Watkins RW, Brennon CB, Cowan DP (2007) Boosting silica levels in wheat leaves reduces grazing by rabbits. *Pest Manag Sci* 63:247–253
- Coupland D, Caseley JC (1975) Reduction of silica and increase in tillering induced in *Agropyron repens* by glyphosate. *J Exp Bot* 26:138–144
- Crusciol CAC, Pulz AL, Lemos LB, Soratto RP, Lima GPP (2009) Effects of silicon and drought stress on tuber yield and leaf biochemical characteristics in potato. *Crop Sci* 49:949–954
- Currie HA, Perry CC (2007) Silica in plants: biological, biochemical and chemical studies. *Ann Bot* 100:1383–1389
- Dahl BE, Tisdale EW (1975) Environmental factors related to medusahead distribution. *J Range Manage* 28:463–468
- Dakheel AJ, Radosevich SR, Barbour MG (1993) Effect of nitrogen and phosphorus on growth and interference between *Bromus tectorum* and *Taeniatherum asperum*. *Weed Res* 33:415–422
- D'Antonio CM, Vitousek PM (1992) Biological invasions by exotic grasses, the grass/fire cycle, and global change. *Annu Rev Ecol Syst* 23:63–87
- Datnoff LE, Elmer WH, Huber DM (2007) Mineral Nutrition and Plant Disease. St Paul, MN: American Phytopathological Society
- Davies KW (2008) Medusahead dispersal and establishment in sagebrush steppe plant communities. *Rangeland Ecol Manag* 61:110–115
- Davies KW (2010) Revegetation of medusahead-invaded sagebrush steppe. *Rangeland Ecol Manag* 63:564–571
- Davies KW (2011) Plant community diversity and native plant abundance decline with increasing abundance of an exotic annual grass. *Oecologia* 167:481–491
- Davies KW, Bates JD, Nafus AM (2012) Mowing Wyoming big sagebrush communities with degraded herbaceous understories: has a threshold been crossed? *Rangeland Ecol Manag* 65:498–505
- Davies KW, Bates JD, Svejcar TJ, Boyd CS (2010) Effects of long-term livestock grazing on fuel characteristics in rangelands: an example from the sagebrush steppe. *Rangeland Ecol Manag* 63:662–669
- Davies KW, Boyd CS (2018) Longer-term evaluation of revegetation of medusahead-invaded sagebrush steppe. *Rangeland Ecol Manag* 71:292–297
- Davies KW, Boyd CS, Johnson DD, Nafus AM, Madsen MD (2015) Success of seeding native compared with introduced perennial vegetation for revegetating medusahead-invaded sagebrush rangeland. *Rangeland Ecol Manag* 68:224–230
- Davies KW, Nafus AM (2013) Exotic annual grass invasion alters fuel amounts, continuity and moisture content. *Int J Wildland Fire* 22:353–358
- Davies KW, Sheley RL (2011) Promoting native vegetation and diversity in exotic annual grass infestations. *Restor Ecol* 19:159–165
- DiTomaso JM (1997) Risk analysis of various weed control methods. *Proc. Calif. Exotic Pest Plant Counc. Symp* 3:34–39
- DiTomaso JM, Kyser GB, George MR, Doran MP, Laca EA (2008) Control of medusahead (*Taeniatherum caput-medusae*) using timely sheep grazing. *Invasive Plant Sci Manag* 1:241–247
- DiTomaso JM, Kyser GB, Hastings MS (1999) Prescribed burning for control of yellow starthistle (*Centaurea solstitialis*) and enhanced native plant diversity. *Weed Sci* 47:233–242
- DiTomaso JM, Smith BS (2012) Linking ecological principles to tools and strategies in an EBIPM program. *Rangelands* 34:30–34
- Elgersma KJ, Yu S, Vor T, Ehrenfeld JG (2012) Microbial-mediated feedbacks of leaf litter on invasive plant growth and interspecific competition. *Plant Soil* 356:341–355
- Emamverdian A, Ding Y, Xie Y, Sangari S (2018) Silicon mechanisms to ameliorate heavy metal stress in plants. *BioMed Research International* 2018:8492898. <https://www.hindawi.com/journals/bmri/2018/8492898>. Accessed: May 15, 2019
- Epstein E (1972) Mineral nutrition of plants: principles and perspectives. Oxford: Wiley. 412 p
- Epstein E (1994) The anomaly of silicon in plant biology. *Proc Natl Acad Sci USA* 91:11–17
- Epstein E (1999) Silicon. *Annu Rev Plant Physiol Plant Mol Biol* 50:641–664
- Evans RA, Young JA (1970) Plant litter and establishment of alien annual weed species in rangeland communities. *Weed Sci* 18:697–703
- Fauteux F, Rémus-Borel W, Menzies JG, Bélanger RR (2005) Silicon and plant disease resistance against pathogenic fungi. *FEMS Microbiol Lett* 249:1–6
- Feng J, Yamaji N, Mitani-Ueno N (2011) Transport of silicon from roots to panicles in plants. *Proc Jpn Acad Ser B Phys Biol Sci* 87:377–385
- Ganskopp D, Bohnert D (2001) Nutritional dynamics of 7 northern Great Basin grasses. *Rangeland Ecol Manag/J Range Manage Archives* 54:640–647
- Gatford KL, Simpson RJ, Siever-Kelly C, Leury BJ, Dove H, Ciavarella TA (1999) Spray-topping annual grass pasture with glyphosate to delay loss of feeding value during summer. I. Effects on pasture yield and nutritive value. *Aust J Agric Res* 50:453–464
- Giraldo MC, Valent B (2013) Filamentous plant pathogen effectors in action. *Nature Rev Microbiol* 11:800
- Goldman JC, Caron DA, Dennett MR (1987) Regulation of gross growth efficiency and ammonium regeneration in bacteria by substrate C: N ratio. *Limnol Oceanogr* 32:1239–1252
- Gong H, Zhu X, Chen K, Wang S, Zhang C (2005) Silicon alleviates oxidative damage of wheat plants in pots under drought. *Plant Sci* 169:313–321
- Gong HJ, Randall DP, Flowers TJ (2006) Silicon deposition in the root reduces sodium uptake in rice (*Oryza sativa* L.) seedlings by reducing bypass flow. *Plant Cell Environ* 29:1970–1979
- Guerriero G, Hausman J-F, Legay S (2016) Silicon and the plant extracellular matrix. *Front Plant Sci* 7:463
- Hamilton T, Burritt EA, Villalba JJ (2015) Assessing the impact of supplements, food aversions, and silica on medusahead (*Taeniatherum caput-medusae* (L.) Nevski) use by sheep. *Small Ruminant Res* 124:45–54
- Handreck KA, Jones LP (1967) Uptake of monosilicic acid by *Trifolium incarnatum* (L.). *Aust J Bio Sci* 20:483–486
- Harper JL, Williams JT, Sagar GR (1965) The behaviour of seeds in soil: I. The heterogeneity of soil surfaces and its role in determining the establishment of plants from seed. *J Ecol* 53:273–286
- Harris GA (1977) Root phenology as a factor of competition among grass seedlings. *J Range Manage* 30:172–177
- Harrison S, Inouye BD, Safford HD (2003) Ecological heterogeneity in the effects of grazing and fire on grassland diversity. *Conserv Biol* 17:837–845
- Hattori T, Inanaga S, Tanimoto E, Lux A, Luxová M, Sugimoto Y (2003) Silicon-induced changes in viscoelastic properties of sorghum root cell walls. *Plant Cell Physiol* 44:743–749
- Heady HF (1961) Continuous vs. specialized grazing systems: a review and application to the California annual type. *J Range Manage* 14:182–193

- Hironaka M (1961) The relative rate of root development of cheatgrass and medusahead. *J Range Manage* 14:263–267
- Hobbs NT (1996) Modification of ecosystems by ungulates. *J Wildl Manag* 60:695–713
- Hodson MJ, White PJ, Mead A, Broadley MR (2005) Phylogenetic variation in the silicon composition of plants. *Ann Bot* 96:1027–1046
- Holub EB, Cooper A (2004) Matrix, reinvention in plants: how genetics is unveiling secrets of non-host disease resistance. *Trends Plant Sci* 9:211–214
- Hossain MT, Mori R, Soga K, Wakabayashi K, Kamisaka S, Fujii S, Yamamoto R, Hoson T (2002) Growth promotion and an increase in cell wall extensibility by silicon in rice and some other Poaceae seedlings. *J Plant Res* 115:23–27
- Hu Y, Schmidhalter U (2005) Drought and salinity: a comparison of their effects on mineral nutrition of plants. *Z Pflanzenernähr Bodenk* 168:541–549
- Hunt JW, Dean AP, Webster RE, Johnson GN, Ennos AR (2008) A novel mechanism by which silica defends grasses against herbivory. *Ann Bot* 102:653–656
- Inderjit, Cahill JF (2015) Linkages of plant–soil feedbacks and underlying invasion mechanisms. *AoB Plants* 7:plv022
- James JJ, Gornish ES, DiTomaso JM, Davy J, Doran MP, Becchetti T, Lile D, Brownsey P, Laca EA (2015) Managing medusahead (*Taeniatherum caput-medusae*) on rangeland: a meta-analysis of control effects and assessment of stakeholder needs. *Rangeland Ecol Manag* 68:215–223
- James JJ, Ziegenhagen L, Aanderud ZT (2010) Exploitation of nutrient-rich soil patches by invasive annual and native perennial grasses. *Invasive Plant Sci Manag* 3:169–177
- Johnson DD, Davies KW (2012) Medusahead management in sagebrush-steppe rangelands: prevention, control, and revegetation. *Rangelands* 34:32–38
- Johnson DD, Davies KW, Schreder PT, Chamberlain A-M (2011) Perceptions of ranchers about medusahead (*Taeniatherum caput-medusae* (L.) Nevski) management on sagebrush steppe rangelands. *Environ Manag* 48:400–417
- Johnston A, Bezeau LM, Smoliak S (1968) Chemical composition and in vitro digestibility of alpine tundra plants. *J Wildl Manag* 32:773–777
- Jones LHP, Handreck KA (1969) Uptake of silica by *Trifolium incarnatum* in relation to the concentration in the external solution and to transpiration. *Plant Soil* 30:71–80
- Kakani VG, Reddy KR, Zhao D, Sailaja K (2003) Field crop responses to ultraviolet-B radiation: a review. *Agric For Meteorol* 120:191–218
- Kaltenecker JH (1997) The Recovery of Microbiotic Crusts following Post-fire Rehabilitation on Rangelands of the Western Snake River Plain. M.S. thesis. Boise, ID: Boise State University. 99 p
- Keller C, Rizwan M, Davidian J-C, Pokrovsky OS, Bovet N, Chaurand P, Meunier J-D (2015) Effect of silicon on wheat seedlings (*Triticum turgidum* L.) grown in hydroponics and exposed to 0 to 30 μM Cu. *Planta* 241:847–860
- Kind P, King EJ, Pash V, Roman W, Schmidt E (1954) Inhibition of enzymes by silicic acid. *Biochem J* 56:xliv–xlv
- Kisseberth WC, Buck WB, Mansfield ME, Manuel RK (1986) Preferential grazing by cattle on glyphosate-treated fescue pastures. *Am J Vet Res* 47:696–698
- Kyser GB, Creech JE, Zhang J, DiTomaso JM (2012a) Selective control of medusahead (*Taeniatherum caput-medusae*) in California sagebrush scrub using low rates of glyphosate. *Invasive Plant Sci Manag* 5:1–8
- Kyser GB, DiTomaso JM, Doran MP, Orloff SB, Wilson RG, Lancaster DL, Lile DF, Porath ML (2007) Control of medusahead (*Taeniatherum caput-medusae*) and other annual grasses with imazapic. *Weed Technol* 21:66–75
- Kyser GB, Doran MP, McDougald NK, Orloff SB, Vargas RN, Wilson RG, DiTomaso JM (2008) Site characteristics determine the success of prescribed burning for medusahead (*Taeniatherum caput-medusae*) control. *Invasive Plant Sci Manag* 1:376–384
- Kyser GB, Peterson VF, Davy JS, DiTomaso JM (2012b) Preemergent control of medusahead on California annual rangelands with aminopyralid. *Rangeland Ecol Manag* 65:418–425
- Laishram ID, Yadava PS (1988) Lignin and nitrogen in the decomposition of leaf litter in a sub-tropical forest ecosystem at Shiroy hills in north-eastern India. *Plant Soil* 106:59–64
- Lanning FC, Eleuterius LN (1983) Silica and ash in tissues of some coastal plants. *Ann Bot* 51:835–850
- Lanning FC, Hopkins TL, Loera JC (1980) Silica and ash content and depositional patterns in tissues of mature *Zea mays* L. plants. *Ann Bot* 45:549–554
- Lee DB, Kwon TO, Park KH (1990) Influence of nitrogen and silica on the yield and the lodging related traits of paddy rice. *Research Reports of the Rural Development Administration, Soil & Fertilizer* 32:15–23. Korean
- Lewin J, Reimann BE (1969) Silicon and plant growth. *Annu Rev Plant Physiol* 20:289–304
- Li W-B, Shi X-H, Wang H, Zhang F-S (2004) Effects of silicon on rice leaves resistance to ultraviolet-B. *Acta Botanica Sinica (English edition)* 46:691–697
- Li Z, Song Z, Cornelis J-T (2014) Impact of rice cultivar and organ on elemental composition of phytoliths and the release of bio-available silicon. *Front Plant Sci* 5:529
- Liang Y, Si J, Römheld V (2005) Silicon uptake and transport is an active process in *Cucumis sativus*. *New Phytol* 167:797–804
- Locke MA, Bryson CT (1997) Herbicide-soil interactions in reduced tillage and plant residue management systems. *Weed Sci* 45:307–320
- Longland WS (1994) Seed use by desert granivores. *Desert Plants* 30:233–237
- Lusk WC, Jones MB, Torell DT, Kell CMM (1961) Medusahead Palatability. *J Range Manage* 14:248–251
- Lux A, Luxová M, Abe J, Tanimoto E, Hattori T, Inanaga S (2003) The dynamics of silicon deposition in the sorghum root endodermis. *New Phytol* 158:437–441
- Ma JF (2004) Role of silicon in enhancing the resistance of plants to biotic and abiotic stresses. *Soil Sci Plant Nutr* 50:11–18
- Ma JF, Mitani N, Nagao S, Konishi S, Tamai K, Iwashita T, Yano M (2004) Characterization of the silicon uptake system and molecular mapping of the silicon transporter gene in rice. *Plant Physiol* 136:3284–3289
- Ma JF, Miyake Y, Takahashi E (2001) Silicon as a beneficial element for crop plants. Pages 17–39 in LE Datnoff, GH Snyder, GH Korndörfer, eds. *Studies in Plant Science*. Amsterdam: Elsevier
- Ma JF, Takahashi E (2002) Soil, Fertilizer, and Plant Silicon Research in Japan. Amsterdam: Elsevier. 294 p
- Ma JF, Tamai K, Yamaji N, Mitani N, Konishi S, Katsuhara M, Ishiguro M, Murata Y, Yano M (2006) A silicon transporter in rice. *Nature* 440:688–691
- Ma JF, Yamaji N (2006) Silicon uptake and accumulation in higher plants. *Trends Plant Sci* 11:392–397
- Ma JF, Yamaji N, Mitani N, Tamai K, Konishi S, Fujiwara T, Katsuhara M, Yano M (2007) An efflux transporter of silicon in rice. *Nature* 448:209–212
- MacKown CT, Jones TA, Johnson DA, Monaco TA, Redinbaugh MG (2009) Nitrogen uptake by perennial and invasive annual grass seedlings: nitrogen form effects. *Soil Sci Soc Am J* 73:1864–1870
- Madronich S, McKenzie RL, Björn LO, Caldwell MM (1998) Changes in biologically active ultraviolet radiation reaching the Earth's surface. *J Photochem Photobiol B* 46:5–19
- Mangla S, Sheley RL, James JJ (2011) Field growth comparisons of invasive alien annual and native perennial grasses in monocultures. *J Arid Environ* 75:206–210
- Martínez-Ruiz C, Fernández-Santos B, Gómez-Gutiérrez JM (2001) Effects of substrate coarseness and exposure on plant succession in uranium-mining wastes. *Plant Ecol* 155:79–89
- Massey FP, Ennos AR, Hartley SE (2006) Silica in grasses as a defence against insect herbivores: contrasting effects on folivores and a phloem feeder. *J Anim Ecol* 75:595–603
- Massey FP, Hartley SE (2006) Experimental demonstration of the antiherbivore effects of silica in grasses: impacts on foliage digestibility and vole growth rates. *Proc R Soc Lond B Biol Sci* 273:2299–2304
- Massey FP, Hartley SE (2009) Physical defences wear you down: progressive and irreversible impacts of silica on insect herbivores. *J Anim Ecol* 78:281–291
- Massey FP, Massey K, Roland Ennos A, Hartley SE (2009) Impacts of silica-based defences in grasses on the feeding preferences of sheep. *Basic Appl Ecol* 10:622–630
- Mayland HF, Shewmaker GE (2001) Animal health problems caused by silicon and other mineral imbalances. *J Range Manage* 54:441–446
- McKell CM, Robison JP, Major J (1962) Ecotypic variation in medusahead, an introduced annual grass. *Ecology* 43:686–698
- McNaughton SJ, Tarrants JL, McNaughton MM, Davis RD (1985) Silica as a defense against herbivory and a growth promoter in African grasses. *Ecology* 66:528–535
- Meyer MD, Schiffman PM (1999) Fire season and mulch reduction in a California grassland: a comparison of restoration strategies. *Madrono* 46:25–37

- Miller HC (1996) Demography of medusahead on two soil types: potential for invasion into intact native communities. M.S. thesis. Corvallis, OR: Oregon State University. 47 p
- Miller HC, Clausnitzer D, Borman MM (1999) Medusahead. Pages 271–281 in RL Sheley, JK Petroff, eds. *Biology and Management of Noxious Rangeland Weeds*. 1st ed. Corvallis: Oregon State University Press
- Mitani N, Ma JF (2005) Uptake system of silicon in different plant species. *J Exp Bot* 56:1255–1261
- Mitani N, Yamaji N, Ma JF (2008) Identification of maize silicon influx transporters. *Plant Cell Physiol* 50:5–12
- Monaco TA, Osmond TM, Dewey SA (2005) Medusahead control with fall- and spring-applied herbicides on northern Utah foothills. *Weed Technol* 19:653–658
- Montes-Sánchez JJ, Van Miegroet H, Villalba JJ (2017) Effects energy supplementation and time on use of medusahead by grazing ewes and their lambs. *Rangeland Ecol Manag* 70:380–387
- Montes-Sánchez JJ, Villalba JJ (2017) Understanding medusahead low intake and palatability through in vitro digestibility and fermentation kinetics. *Animal* 11:1930–1938
- Mueggler WF (1972) Influence of competition on the response of bluebunch wheatgrass to clipping. *J Range Manage* 25:88–92
- Mugford ST, Barclay E, Drurey C, Findlay KC, Hogenhout SA (2016) An immuno-suppressive aphid saliva protein is delivered into the cytosol of plant mesophyll cells during feeding. *Mol Plant-Microbe Interact* 29:854–861
- Nafus AM, Davies KW (2014) Medusahead ecology and management: California annual grasslands to the Intermountain West. *Invasive Plant Sci Manag* 7:210–221
- Nagajyoti PC, Lee KD, Sreekanth TVM (2010) Heavy metals, occurrence and toxicity for plants: a review. *Environ Chem Lett* 8:199–216
- Namaganda M, Krekling T, Lye KA (2009) Leaf anatomical characteristics of Ugandan species of *Festuca* L. (Poaceae). *S Afr J Bot* 75:52–59
- National Research Council (2000) *Nutrient Requirements of Beef Cattle*. 7th rev. ed. Update 2000. Washington, DC: National Academies Press. 248 p
- National Research Council (2007a) *Nutrient Requirements of Horses*. 6th rev. ed. Washington, DC: National Academies Press. 360 p
- National Research Council (2007b) *Nutrient Requirements of Small Ruminants: Sheep, Goats, Cervids, and New World Camelids*. Washington, DC: National Academies Press. 384 p
- Neary DG, Klopatek CC, DeBano LF, Ffolliott PF (1999) Fire effects on below-ground sustainability: a review and synthesis. *For Ecol Manag* 122:51–71
- Nelson JR, Wilson AM (1969) Influence of age and awn removal on dormancy of medusahead seeds. *J Range Manage* 22:289–290
- Neumann D (2003) Silicon in plants. Pages 149–160 in WEG Müller, ed. *Silicon Biomineralization*. Berlin: Springer
- Nuernberger T, Lipka V (2005) Non-host resistance in plants: new insights into an old phenomenon. *Molec Plant Pathol* 6:335–345
- O'Neill C, Clarke G, Hodges G, Jordan P, Newman R, Pan Q-Q, Liu F-S, Ge M, Chang Y-M, Toulson E (1982) Silica fragments from millet bran in mucosa surrounding oesophageal tumours in patients in northern China. *Lancet* 319:1202–1206
- O'Neill CH, Hodges GM, Riddle PN, Jordan PW, Newman RH, Flood RJ, Toulson EC (1980) A fine fibrous silica contaminant of flour in the high oesophageal cancer area of north-east Iran. *Int J Cancer* 26:617–628
- Palm CA (1988) *Mulch Quality and Nitrogen Dynamics in an Alley Cropping System in the Peruvian Amazon*. Ph.D dissertation. Raleigh, NC: North Carolina State University
- Peters EF, Bunting SC (1994) Fire conditions pre- and postoccurrence of annual grasses on the Snake River Plain. Pages 31–36 in SB Monsen, SG Kitchen, eds. *Proceedings—Ecology and Management of Annual Rangelands*. Gen. Tech. Rep INT-GTR-313. Ogden, UT: U.S. Department of Agriculture, Forest Service
- Pierson FB, Blackburn WH, Van Vactor SS (2007) Hydrologic impacts of mechanical seeding treatments on sagebrush rangelands. *Rangeland Ecol Manag* 60:666–674
- Provenza FD, Villalba JJ, Bryant JP (2002) Making the match: from biochemical diversity to landscape diversity. Pages 387–421 in *Landscape Ecology and Resource Management: Making the Match*. New York: Island Press
- Rains DW, Epstein E, Zasoski RJ, Aslam M (2006) Active silicon uptake by wheat. *Plant Soil* 280:223–228
- Raven JA (1983) The transport and function of silicon in plants. *Biol Rev* 58:179–207
- Reglero MM, Monsalve-González L, Taggart MA, Mateo R (2008) Transfer of metals to plants and red deer in an old lead mining area in Spain. *Sci Total Environ* 406:287–297
- Rémus-Borel W, Menzies JG, Bélanger RR (2009) Aconitate and methyl aconitate are modulated by silicon in powdery mildew-infected wheat plants. *J Plant Physiol* 166:1413–1422
- Rinella MJ, Bellows SE, Roth AD (2014) Aminopyralid constrains seed production of the invasive annual grasses medusahead and ventenata. *Rangeland Ecol Manag* 67:406–411
- Rinella MJ, Davy JS, Kyser GB, Mashiri FE, Bellows SE, James JJ, Peterson VF (2018) Timing aminopyralid to prevent seed production controls medusahead (*Taeniatherum caput-medusae*) and increases forage grasses. *Invasive Plant Sci Manag* 11:61–68
- Rodrigues FÁ, Benhamou N, Datnoff LE, Jones JB, Bélanger RR (2003) Ultrastructural and cytochemical aspects of silicon-mediated rice blast resistance. *Phytopathology* 93:535–546
- Rodrigues FÁ, McNally DJ, Datnoff LE, Jones JB, Labbé C, Benhamou N, Menzies JG, Bélanger RR (2004) Silicon enhances the accumulation of diterpenoid phytoalexins in rice: a potential mechanism for blast resistance. *Phytopathology* 94:177–183
- Savage DE, Young JA, Evans RA (1969) Utilization of medusahead and downy brome caryopses by chukar partridges. *J Wildl Manag* 33:975–978
- Savant NK, Snyder GH, Datnoff LE (1996) Silicon management and sustainable rice production. Pages 151–199 in DL Sparks, ed. *Advances in Agronomy*. San Diego, CA: Academic Press
- Schantz M, Sheley R, Hardegree S (2019) Restoring perennial grasses in medusahead habitat: role of tilling, fire, herbicides, and seeding rate. *Rangeland Ecol Manag* 72:249–259
- Sebastian DJ, Nissen SJ, De Souza Rodrigues J (2016a) Pre-emergence control of six invasive winter annual grasses with imazapic and indaziflam. *Invasive Plant Sci Manag* 9:308–316
- Sebastian DJ, Nissen SJ, Sebastian JR, Beck KG (2017a) Seed bank depletion: the key to long-term downy brome (*Bromus tectorum* L.) management. *Rangeland Ecol Manag* 70:477–483
- Sebastian DJ, Nissen SJ, Sebastian JR, Meiman PJ, Beck KG (2017b) Preemergence control of nine invasive weeds with aminocyclopyrachlor, aminopyralid, and indaziflam. *Invasive Plant Sci Manag* 10:99–109
- Sebastian DJ, Sebastian JR, Nissen SJ, Beck KG (2016b) A potential new herbicide for invasive annual grass control on rangeland. *Rangeland Ecol Manag* 69:195–198
- Sheley RL, Bingham BS, Svejcar TJ (2008) Crested wheatgrass defoliation intensity and season on medusahead invasion. *Rangeland Ecol Manag* 61:211–217
- Sheley RL, Carpinelli MF, Morghan KJR (2007) Effects of imazapic on target and nontarget vegetation during revegetation. *Weed Technol* 21:1071–1081
- Shen X, Zhou Y, Duan L, Li Z, Eneji AE, Li J (2010) Silicon effects on photosynthesis and antioxidant parameters of soybean seedlings under drought and ultraviolet-B radiation. *J Plant Physiol* 167:1248–1252
- Shewmaker GE, Mayland HF, Rosenau RC, Asay KH (1989) Silicon in C-3 grasses: effects on forage quality and sheep preference. *J Range Manage* 42:122–127
- Smith GS, Nelson AB, Boggino EJ (1971) Digestibility of forages in vitro as affected by content of “silica.” *J Anim Sci* 33:466–471
- Stoncipher CA, Panter KE, Villalba JJ (2016) Effect of protein supplementation on forage utilization by cattle in annual grass-dominated rangelands in the Channeled Scablands of eastern Washington. *J Anim Sci* 94:2572–2582
- Stott DE, Kassim G, Jarrell WM, Martin JP, Haider K (1983) Stabilization and incorporation into biomass of specific plant carbons during biodegradation in soil. *Plant Soil* 70:15–26
- Suding KN, Stanley Harpole W, Fukami T, Kulmatiski A, MacDougall AS, Stein C, van der Putten WH (2013) Consequences of plant-soil feedbacks in invasion. *J Ecol* 101:298–308
- Sweet SB, Kyser GB, DiTomaso JM (2008) Susceptibility of exotic annual grass seeds to fire. *Invasive Plant Sci Manag* 1:158–167

- Swenson CF, Le Tourneau D, Erickson LC (1964) Silica in medusahead. *Weeds* 12:16–18
- Takahashi E, Ma JF, Miyake Y (1990) The possibility of silicon as an essential element for higher plants. *Comments on Agricultural and Food Chemistry* 2:99–102
- Tamai K, Ma JF (2008) Reexamination of silicon effects on rice growth and production under field conditions using a low silicon mutant. *Plant Soil* 307:21–27
- Torell PJ, Erickson LC, Haas RH (1961) The medusahead problem in Idaho. *Weeds* 9:124–131
- Tubaña BS, Heckman JR (2015) Silicon in soils and plants. Pages 7–51 in Rodrigues FA, Datnoff LE, eds. *Silicon and Plant Diseases*. Cham, Switzerland: Springer
- Vallentine JF, Stevens AR (1994) Use of livestock to control cheatgrass—a review. Pages 202–206 in Monsen SB, Kitchen SG, eds. *Proceedings—Ecology and Management of Annual Rangelands*. Gen. Tech. Rep INT-GTR-313. Ogden, UT: U.S. Department of Agriculture, Forest Service
- Van Soest PJ (1994) *Nutritional Ecology of the Ruminant*. Ithaca, NY: Cornell University Press. 496 p
- Van Soest PJ (2006) Rice straw, the role of silica and treatments to improve quality. *Anim Feed Sci Technol* 130:137–171
- Van Soest PJ, Jones LHP (1968) Effect of silica in forages upon digestibility. *J Dairy Sci* 51:1644–1648
- Vasquez E, Sheley R, Svejcar T (2008) Nitrogen enhances the competitive ability of cheatgrass (*Bromus tectorum*) relative to native grasses. *Invasive Plant Sci Manag* 1:287–295
- Villalba JJ, Burritt EA (2015) Intake of medusahead by sheep: influence of supplements, silica and individual animal variation. *Invasive Plant Sci Manag* 8:151–159
- Vlamis J, Williams DE (1967) Manganese and silicon interaction in the graminiae. *Plant Soil* 27:131–140
- Wagner F (1940) The importance of silicic acid for the growth of some cultivated plants, their metabolism, and their susceptibility to true mildews. *Phytopathologische Zeitschrift* 12:427–479
- Wang Y, Stass A, Horst WJ (2004) Apoplastic binding of aluminum is involved in silicon-induced amelioration of aluminum toxicity in maize. *Plant Physiol* 136:3762–3770
- Whisenant SG (1990) Changing fire frequencies on Idaho's Snake River Plains: ecological and management implications. Pages 4–10 in McArthur ED, Romney EM, Smith SD, Tueller PT, eds. *Proceedings - Symposium on Cheatgrass Invasion, Shrub Die-Off, and Other Aspects of Shrub Biology and Management*. General Technical Report INT-GTR-276. Ogden, UT: USDA Forest Service
- Wu J-W, Shi Y, Zhu Y-X, Wang Y-C, Gong H-J (2013) Mechanisms of enhanced heavy metal tolerance in plants by silicon: a review. *Pedosphere* 23:815–825
- Yalçın N, Sevinç V (2001) Studies on silica obtained from rice husk. *Ceramics International* 27:219–224
- Ye J, Yan C, Liu J, Lu H, Liu T, Song Z (2012) Effects of silicon on the distribution of cadmium compartmentation in root tips of *Kandelia obovata* (S., L.) Yong. *Environ Pollut* 162:369–373
- Yoshida S, Ohnishi Y, Kitagishi K (1962) Histochemistry of silicon in rice plant. *Soil Sci Plant Nutr* 8:1–5
- Young JA (1992) Ecology and management of medusahead (*Taeniatherum caput-medusae* ssp. *asperum* [SIMK.] Melderis). *Great Basin Nat* 52:245–252
- Young JA, Clements CD, Nader G (1999) Medusahead and clay: the rarity of perennial seedling establishment. *Rangelands* 21:19–23
- Young JA, Evans RA, Eckert RE (1969) Emergence of medusahead and other grasses from four seeding depths. *Weed Sci* 17:376–379
- Young K, Mangold J (2008) Medusahead (*Taeniatherum caput-medusae*) outperforms squirreltail (*Elymus elymoides*) through interference and growth rate. *Invasive Plant Sci Manag* 1:73–81