

New insights in the taxonomy of Mediterranean *Diodora* (Mollusca, Gastropoda, Fissurellidae)

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The taxonomy of Mediterranean populations of Diodora is assessed based on new molecular (COI and 28S) data. The recently described Diodora demartiniorum Buzzurro & Russo, 2005, is found to be a valid species restricted to the Gulf of Gabès (Tunisia) but possibly occurring also on the coast of Libya. However, specimens from the Aegean Sea previously identified as D. demartiniorum are molecularly (and morphologically pseudocryptically) distinct and represent a previously unrecognized species here described as D. giannispadai n. sp. It is hypothesized that the current distribution of these two species corresponds to glacial refuges during Pleistocene climate changes.

Keywords: Fissurellidae, *Diodora*, endemism, Gulf of Gabès, Aegean Sea, new species

Submitted 14 January 2016; accepted 6 June 2016; first published online 4 July 2016

INTRODUCTION

The family Fissurellidae includes 55 genera and over 500 valid species (WoRMS, 2016). Although fissurellids have been routinely included in phylogenetic analyses of (veti)gastropods (e.g. Harasewych *et al.*, 1997; Colgan *et al.*, 2000, 2003; McArthur & Harasewych, 2003; Geiger & Thacker, 2005; Aktipis & Giribet, 2010) taxon sampling has been very loose; these studies included just one or a few fissurellid species. The genus *Diodora* occurs in tropical and temperate shallow waters all over the world, and three species have been classically recognized in the Mediterranean (Bucquoy *et al.*, 1882–1886 (as *Fissurella*); Ghisotti & Melone, 1969; Sabelli *et al.*, 1990, 1992; Poppe & Goto, 1991): *Diodora graeca* (Linnaeus, 1758), *D. italica* (Defrance, 1820) and *D. gibberula* (Lamarck, 1822). Two further taxa *D. dorsata* (Monterosato, 1878) and *D. producta* (Monterosato, 1880) listed as 'doubtful species' by Sabelli *et al.* (1992) were treated as valid by Giannuzzi-Savelli *et al.* (1994) and Buzzurro & Russo (2005). In addition, a couple of Indo-Pacific migrants, *Diodora rupellii* (G. B. Sowerby I, 1834) and *D. funiculata* (Reeve, 1850), are also recorded from the Mediterranean, but – as is frequent with Lessepsian migrants (Zenetos *et al.*, 2010) – are restricted to the coasts of the Levant (Barash & Danin, 1992; Mienis, 2002). Whereas no new fissurellid had been discovered in the Mediterranean since the 19th century, a sixth native species, *Diodora demartiniorum* Buzzurro & Russo, 2005,

was quite unexpectedly recently described from the Gulf of Gabès and the Aegean Sea. Its status as a valid species was accepted by Delongueville & Scaillet (2006) and Cecalupo *et al.* (2008). The subtlety of shell characters used to delimit species of fissurellids, in general, and of *Diodora* in particular, explains why the status of *D. producta* or *D. dorsata* has remained uncertain for over a century (listed as 'doubtful species' by Sabelli *et al.* (1992) but treated as valid by Giannuzzi-Savelli *et al.* (1994) and Buzzurro & Russo (2005)), as it requires considerable personal experience to discriminate Mediterranean *Diodora* species beyond the 'classical three' (*D. graeca*, *D. italica*, *D. gibberula*). The state of the art of Mediterranean *Diodora* taxonomy is thus to recognize 4–6 native and two introduced species, all exclusively discriminated based on shell characters. The comprehensive molecular fissurellid phylogeny by Aktipis *et al.* (2011) included 30 terminal taxa, of which two were Mediterranean species of *Diodora* (*D. graeca*, *D. gibberula*), but the purpose of these authors was not to discuss species-level systematics.

Diodora demartiniorum is one of 33 mollusc species currently regarded as valid and endemic to the Gulf of Gabès (Cecalupo *et al.* 2008). All were defined exclusively based on shell characters, and the only molecular approach to the systematics of putative Gulf of Gabès endemics is by Barco *et al.* (2013). Targeting the small muricids of the genus *Ocinebrina*, they recovered some species as valid and others as synonyms. In this context, the purpose of the present work is to revisit the status and distribution of the Mediterranean species of *Diodora* using molecular markers: Is *D. demartiniorum* a valid species? What is the status of the populations of *D. demartiniorum* from the Aegean Sea that have been attributed to it?

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MATERIALS AND METHODS

Samples and primary morphological identifications

Live animals were collected specifically for this work from the Gulf of Gabès (Djerba, Tunisia; 9 specimens, personal collects, 2012 and 2013); the Alboran Sea (Cabo de Palos and El Mohon, Prov. of Murcia, Spain, 2 specimens; Islas Chafarinas, off Morocco, 1 specimen; J. Esteban leg. 2014); and the Aegean Sea (Gulf of Strimonikos and Limnos I., Greece; 3 specimens; K. Papavasileiou leg. 2014) (Figure 1). The soft parts were detached from the shell and preserved for molecular analysis in 95–98% ethanol. The shell was photographed, morphologically identified by Gianni Spada, Paolo Russo and/or Marco Oliverio, and catalogued as a voucher in Muséum National d'Histoire Naturelle (MNHN, Paris) (Tunisian and Greek material) or Museo Nacional de Ciencias Naturales (MNCN, Madrid) (Spanish material).

The locality data, voucher catalogue numbers and GenBank accession numbers for each of the 15 specimens are presented in Table 1.

In addition, to increase the size of the dataset analysed, the sequences of seven Mediterranean specimens of *Diodora* produced by Giribet & Wheeler (2002) and Aktipis *et al.* (2011) were downloaded from GenBank. We found problems with the specimen corresponding to the sequence HM771613.2 identified by Aktipis *et al.* (2011) as *D. graeca* fell with specimens identified by them as *D. cayenensis* (a species from the tropical western Atlantic). After correspondence with the authors, we concluded that contamination and/or misidentification was responsible for this result, and we decided to discard this sequences from our analyses. Further, the two specimens corresponding to the sequences AF120632 and AY923915 identified as *D. graeca* respectively by Giribet & Wheeler (2002) and by Geiger & Thacker (2005), fell with specimens identified by Aktipis & Giribet (2010) (and also by us) as *D. gibberula*, and we have also decided to discard these

sequences in our analyses. This leaves four specimens from GenBank with reliable and usable sequences.

Sequencing and amplification

DNA was extracted from a piece of muscle tissue using the DNeasy 96 Tissue kit (Qiagen). A fragment of Cytochrome Oxidase I (COI) mitochondrial gene and a fragment of 28S rDNA nuclear gene were amplified using respectively universal primers LCO1490 (5'-GGTCAACAAATCATAAAGATATTGG-3') and HCO2198 (5'-TAAACTTCAGGGTGACC AAAAATCA-3') (Folmer *et al.*, 1994), and C1' (5'-ACCCGC TGAATTTAAGCAT-3') (Jovelin & Justine, 2001) and D2 (5'-TCCGTGTTTCAAGACGG-3') (Dayrat *et al.*, 2001). All PCR reactions were performed in 25 µl, containing 3 ng of DNA, 1× reaction buffer, 2.5 mM MgCl₂, 0.26 mM dNTP, 0.3 µM of each primer, 5% DMSO and 1.5 units of Q-Bio Taq, QBiogene. Thermocycles consisted of an initial denaturation step at 94°C for 4 min, followed by 40 cycles of denaturation at 94°C for 30 s, annealing at 48°C for COI gene and 60°C for 28S rDNA gene for 40 s and extension at 72°C for 1 min. The final extension was at 72°C for 5 min. PCR products were purified and sequenced by the Eurofins sequencing facility. Both directions were sequenced to confirm accuracy of each sequence.

Phylogenetic analyses

COI and 28S genes were edited using Codon Code Aligner version 4.0.4, and automatically aligned using Clustal W implemented in BioEdit version 7.0.5.3 (Larkin, 2007). The accuracy of automatic alignments was confirmed by eye. To delimit species an online version available at <http://wwwabi.snv.jussieu.fr/public/abgd/> of the Automatic Barcode Gap Discovery method (ABGD; Puillandre *et al.*, 2012) was used, with default parameters. It automatically detects, when present, a gap in the pairwise distribution of the genetic distances calculated between each pair of specimens. This gap, referred to as the barcode gap, is supposed to correspond to a



Fig. 1. Location of the localities of Mediterranean fissurellids discussed in this paper.

Table 1. List of species, collection locality, specimen voucher numbers and GenBank accession numbers.

Species	Locality	Voucher number	COI numbers	28S numbers
<i>Diodora gibberula</i> (Lamarck, 1822)	Girona, Spain	DNA102554	HM771614	HM771493
<i>Diodora gibberula</i> (Lamarck, 1822)	Tossa de Mar, Spain	DNA101154	HM771615	HM771494
<i>Diodora gibberula</i> (Lamarck, 1822)	Girona, Spain	DNA101961	GQ160752	GQ160637
<i>Diodora gibberula</i> (Lamarck, 1822)	Girona, Spain	DNA103317	---	HM771492
<i>Diodora graeca</i> (Linnaeus, 1758)	Cabo de Palos, Murcia, Spain	MNCN 85530	*****	*****
<i>Diodora graeca</i> (Linnaeus, 1758)	Elmohon, Murcia, Spain	MNCN 85531	*****	---
<i>Diodora gibberula</i> (Lamarck, 1822)	Islas Chafarinas, Spain	MNCN 85532	*****	---
<i>Diodora italica</i> (Defrance, 1820)	Sidi Jmour, Djerba, Gulf of Gabès, Tunisia	IM-2013-31730	*****	---
<i>Diodora italica</i> (Defrance, 1820)	Sidi Jmour, Djerba, Gulf of Gabès, Tunisia	IM-2013-31824	*****	---
<i>Diodora italica</i> (Defrance, 1820)	Sidi Jmour, Djerba, Gulf of Gabès, Tunisia	IM-2013-31826	*****	---
<i>Diodora italica</i> (Defrance, 1820)	Sidi Jmour, Djerba, Gulf of Gabès, Tunisia	IM-2013-31827	*****	---
<i>Diodora italica</i> (Defrance, 1820)	Sidi Jmour, Djerba, Gulf of Gabès, Tunisia	IM-2013-31967	*****	*****
<i>Diodora italica</i> (Defrance, 1820)	Houmet Souk, Djerba, Gulf of Gabès, Tunisia	IM-2013-31973	*****	*****
<i>Diodora demartiniorium</i> Buzzurro & Russo, 2005	Sidi Jmour, Djerba, Gulf of Gabès, Tunisia	IM-2013-31829	*****	---
<i>Diodora demartiniorium</i> Buzzurro & Russo, 2005	Sidi Jmour, Djerba, Gulf of Gabès, Tunisia	IM-2013-31972	*****	*****
<i>Diodora demartiniorium</i> Buzzurro & Russo, 2005	Sidi Jmour, Djerba, Gulf of Gabès, Tunisia	IM-2013-31823	*****	*****
<i>Diodora giannispadai</i> n. sp. (present study)	Limnos Island, North Aegean Sea, Greece	IM-2013-32768	---	*****
<i>Diodora giannispadai</i> n. sp. (present study)	Limnos Island, North Aegean Sea, Greece	IM-2013-32769	---	*****
<i>Diodora dorsata</i> (Monterosato, 1878)	Strimonikos, North Aegean Sea, Greece	IM-2013-32767	*****	*****
Outgroups				
<i>Emarginula octaviana</i> Coen, 1939	Girona, Spain	DNA103540	HM771604	HM771476
<i>Fissurella nubecula</i> (Linnaeus, 1758)	Houmt Souk, Djerba, Gulf of Gabès, Tunisia	IM-2009-30650	*****	*****
<i>Fissurella nubecula</i> (Linnaeus, 1758)	Vilanova, Spain	DNA102544	HM771625.1	---

threshold between intra- and interspecific distances and is used to propose species hypotheses. For each gene, a phylogenetic tree was built, using Bayesian Analysis (BA), consisting of six Markov chains (5,000,000 generations each with a sampling frequency of one tree each hundred generations) run in two parallel analyses using MrBayes version 3.1.2 (Ronquist & Huelsenbeck, 2003). The chain temperature was set at 0.2. When the log-likelihood scores were found to stabilize, a consensus tree was calculated after omitting the first 25% trees as burn-in.

RESULTS

Molecular analysis

We obtained 16 COI sequences (13 newly produced, three from GenBank), and 28S sequences for 12 (eight newly produced, four from GenBank); we have not been able to obtain sequences for both loci in some specimens. The COI sequences resulted in a 658 bp alignment. The pairwise distance values are presented in Table 2. The Bayesian tree (Figure 2A) shows five different lineages, (a) to (e), each recognized as a distinct species hypothesis by ABGD and each including sequences separated by genetic distances lower than 3% (and often 1% or less). Sequences from different groups are separated by genetic distances greater than 5%. When including several specimens, each lineage corresponds to a well-supported clade (Posterior Probabilities $PP > 0.98$). The first clade ($PP = 1$) corresponds to a group (a) which includes two specimens from Spain that diverge from all the other species hypotheses. The second clade ($PP = 0.99$) includes four genetic groups (b to e), of which (b) from Djerba ($PP = 0.99$) is sister to the other three together and separated from them by COI distances $> 10\%$. The remaining three genetic

groups (c, $PP = 0.99$; d, $PP = 1$; e, $PP = 0.98$) are separated by genetic distances around 7%; two closely related clades contains specimens collected respectively in Spain and Strimonikos (Northern Aegean Sea) and the last group contains only one specimen.

Using the *a priori* identifications, the COI groups are morphologically identifiable as, respectively, (a) *D. graeca*, (b) *D. demartiniorium*, (c) *D. italica*, (d) *D. gibberula* and (e) *D. dorsata*. There is no material identifiable as *D. producta*.

The 12 remaining 28S sequences resulted in a 700 bp alignment. Although not all specimens were successfully sequenced for both COI and 28S, the same highly supported ($PP > 95$) clades (a, b, c, d and e) are found (Figure 2B), now with the addition of a sixth group (f) corresponding to specimens morphologically identifiable as *D. demartiniorium* from Limnos Island (no COI sequences could be obtained for the latter).

Altogether, the molecular data unambiguously indicate the following:

- (1) They confirm the distinctiveness of the three 'classical' species (*D. graeca*, *D. italica*, *D. gibberula*).
- (2) They confirm the validity of *D. demartiniorium* based on specimens from the type locality, and suggest a further cryptic species corresponding to clade (f) from Limnos Island and morphologically identified as '*D. demartiniorium*'.
- (3) They suggest the existence of further species of *Diodora* in the Mediterranean (one specimen in Figure 2 tentatively identified as *D. dorsata*, see below).

SYSTEMATICS

Superfamily Fissurelloidea Fleming, 1822

Family Fissurellidae Fleming, 1822

Genus *Diodora* Gray, 1821

Diodora demartiniorium Buzzurro & Russo, 2005
(Figures 3E–H)

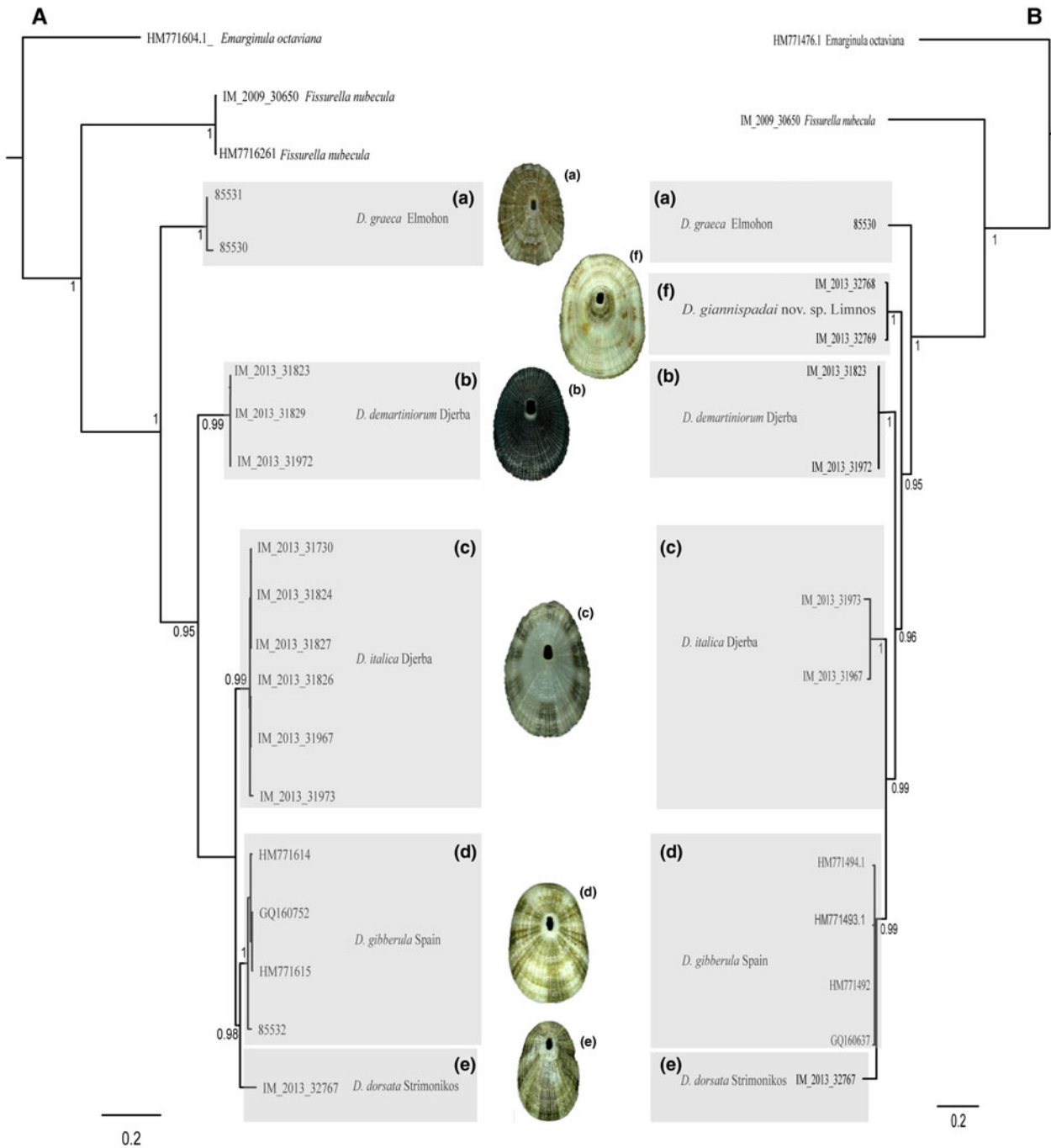


Fig. 2. Bayesian trees of COI (A) and 28S (B) genes. Probabilities (superior to 0.5) and bootstrap values (superior to 50) are given for each node. Genetics groups are lettered from (a) to (f).

DISTRIBUTION

Throughout the Mediterranean. Distribution in the eastern basin and Aegean Sea not known in detail.

DESCRIPTION (BASED ON SEQUENCED ADULT SPECIMENS FROM DJERBA)

Shell limpet-shaped, markedly narrower towards the anterior part, solid, elongated, sculptured with 28 to 36 widely spaced primary radial ribs, irregularly shaped, with secondary riblets in their interspaces. Anal pore up to 4–6 mm long, distance to anterior and posterior shell edge respectively 15–16 mm, and 26–27 mm. Peristome concave, non-adherent to the plane.

Inner background colour uniformly beige; outer shell surface cream with visibly darker bands radiating from the apex. Dimensions of the shell: length 38–40 mm, width 21–22 mm, height 7.5–8 mm.

Diodora giannispadai sp. nov.
(Figures 2A–D)

TYPE MATERIAL

Holotype, a sequenced specimen, MNHN IM-2013-32769; paratype MNHN IM-2013-32768.

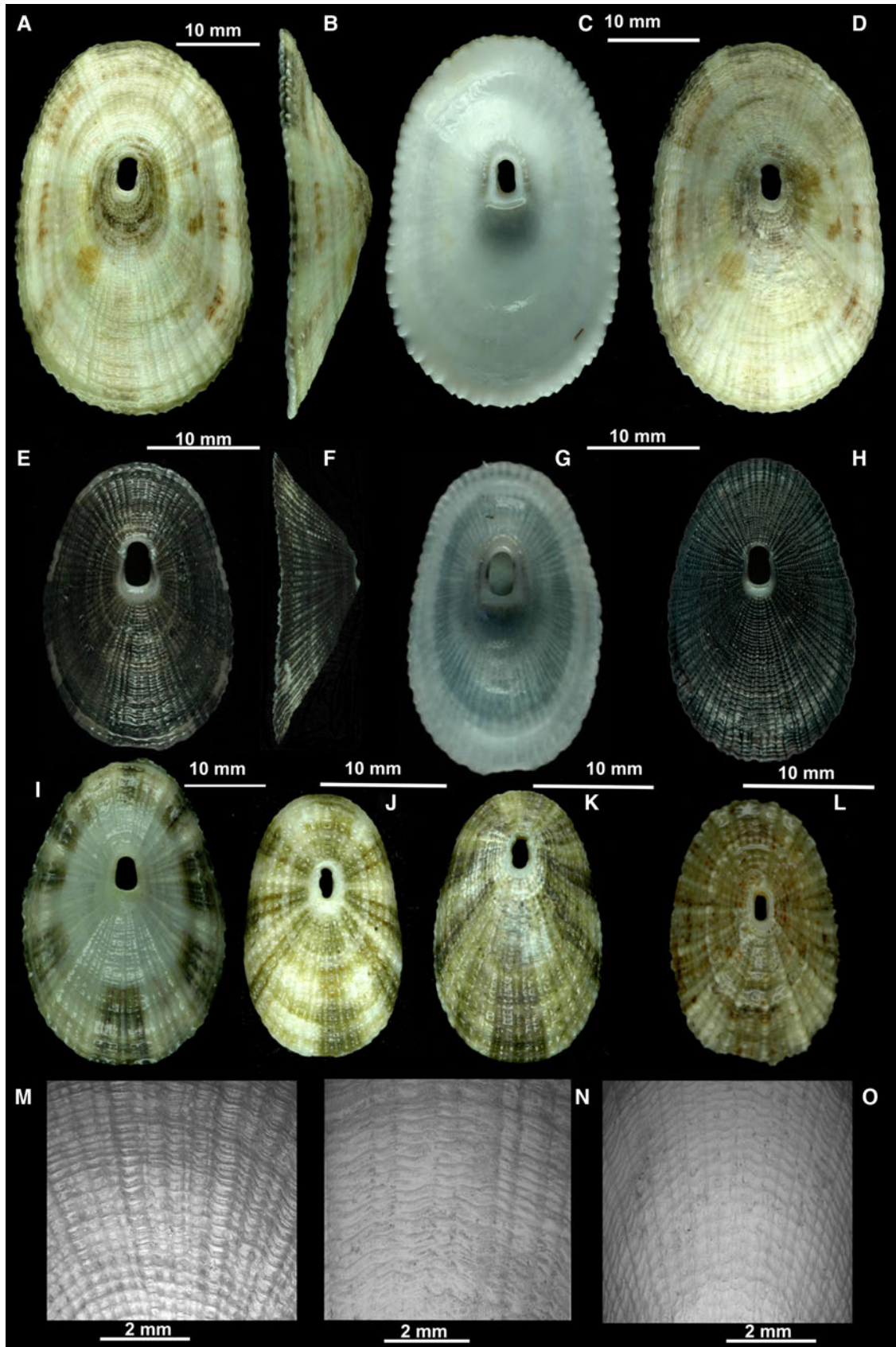


Fig. 3. Shells of Mediterranean fissurellids sequenced. A–C: *Diodora giannispadai* n. sp. (holotype, MNHN 2013-32769). Limnos Island. D: *Diodora giannispadai* n. sp. (paratype, MNHN 2013-32768). Limnos Island. E–H: *Diodora demartiniyorum*. Djerba (MNHN IM-2013-31972, IM-2013-31829). I: *Diodora italica*. Djerba. (MNHN IM-2013-31967). J: *D. gibberula*. Islas Chafarinas. Spain (MNCN 85532). K: *D. dorsata*. Strimonikos. Greece (MNHN IM-2013-32767). L: *D. greaca*. Cabo de Palos. Murcia. Spain (MNCN 85530). M: ribs of *D. italica* (MNHN IM-2013-31967). N: ribs of *Diodora giannispadai* n. sp. (holotype, MNHN 2013-32769). O: ribs of *D. demartiniyorum*. Djerba (MNHN IM-2013-31972).

TYPE LOCALITY

Limnos I., Greece (39°55'N, 25°15'E); shallow subtidal.

DIAGNOSIS

Shell sculptured with widely spaced primary radial ribs and secondary riblets in the interspaces. Peristome totally adherent to the plane. Anal pore elongated, broad.

DESCRIPTION (OF HOLOTYPE)

Shell limpet-shaped, very solid, elongated, sculptured with 70 unequal, widely and evenly spaced primary radial ribs and secondary riblets in their interspaces. Anal pore elongated, broad, length 7 mm long, distance to anterior and posterior shell edge respectively 15 and 30 mm. Peristome slightly concave, especially in anterior part. Inner background colour evenly light beige; outer shell surface cream with darker, greenish to brownish, bands radiating from the apex. Dimensions of the holotype: length 45.0 mm, width 28.0 mm, height 13.6 mm; paratype: length 48.0 mm, width 30.0 mm, height 13.6 mm.

REMARKS

Diodora giannispadai differs from *D. demartiniorium* and *D. italica* by its broadly spaced ribbed with secondary riblets in between; the concentric (commarginal) cords may also be fewer and more widely spaced, although there are too few specimens to judge intraspecific variation (Table 3). Specimens of *D. italica* occasionally reach 52 mm at least in the northern Adriatic (Buzzurro & Russo 2005), and are comparable in size to *D. giannispadai*, but *D. demartiniorium* from the Gulf of Gabès rarely exceeds 35 mm and appears to be smaller on average.

DISTRIBUTION

Beside the sequenced holotype and paratype from Limnos, specimens from Strimonikos (coll. P. Mel, cited by Buzzurro & Russo 2005; not seen), Samothrace (coll. F. Rubio; not seen) and Simi (Giannuzzi-Savelli *et al.*, 1994, as *D. italica*) are very probably referable to the new species: the first two localities are in the North Aegean near the type locality, while Simi (also spelled Symi) is in the south-eastern part of the Aegean Sea. It is not known which other species of *Diodora* *D. giannispadai* is syntopic or sympatric with. Manousis (2012) illustrates *D. demartiniorium*, *D. dorsata*, *D. gibberula*, *D. graeca*, *D. italica* and *D. producta* from Greece, but gives no details on their distribution within Greek waters. The material from Iskendrun (Turkey) referred to *D. demartiniorium* by Delongueville & Scaillet (2006) may or may not be conspecific; its identity should be verified with molecular markers.

ETYMOLOGY

The new species is named after Gianni Spada, a longtime promoter of Mediterranean malacology in the heydays of Società Malacologica Italiana, and a participant of fieldwork in the Gulf of Gabès in 1982 with the senior author, and again in 2013 with the first author. Gianni generously helped the present project with identifications, advice and opinions.

DISCUSSION

The same divergent monophyletic groups are discriminated by the COI and 28S genes and they are congruent with the shell characters classically used to identify Mediterranean keyhole limpets, even though three of these groups are morphologically similar to each other: clade b (*Diodora demartiniorium*) from the Gulf of Gabès, clade c (*D. italica* from the Gulf of Gabès, Figure 3I), clade f (*D. giannispadai* from the Aegean Sea). As there is congruence between the mitochondrial dataset, nuclear dataset and shell morphology, we treat these three groups as three distinct species.

Diodora italica and *D. demartiniorium* were collected syntopically in Djerba and they are separated by COI distances greater than 10%, well over the threshold commonly found (i.e. > 3%) between different gastropod species (Hebert *et al.*, 2003; Puillandre *et al.*, 2009; Crocetta *et al.*, 2012; Barco *et al.*, 2013), and well over the threshold found between the other *Diodora* species in our dataset. As they are also recognizable by subtle, but constant morphological differences, there is no doubt that they are two separate species. The 28S data also indicate that the Aegean Sea specimens morphologically identified as *D. demartiniorium* are not conspecific with *D. demartiniorium* from the type locality, the Gulf of Gabès. As no name is available to designate this North Aegean species, it has been described as new.

The status of the other Mediterranean *Diodora* is more problematic. Our data are sufficient to show that there are more species of *Diodora* in the Mediterranean, but how many and what names should be applied to them can of course not be determined from the single specimen in our molecular dataset. Its identification as *D. dorsata* (Figure 3K) is tentative, and the whole issue should be revisited with more specimens from more localities.

As emphasized already by Sabelli & Taviani (1981), the Gulf of Gabès is geologically very young. During the last glacial maximum and resulting low sea levels, the shallow Gulf was completely exposed, and it can be reasonably assumed that the Gabès ecosystem, as we know it today, is not older than 6000–8000 years when sea level rose sufficiently to flood the exposed platform (Morhange & Pirazzoli, 2005; Barco *et al.*, 2013). Quoting Barco *et al.* (2013) who tested the validity and endemism of the small muricid *Ocenebrina hispidula*, there are several possible hypotheses on the putative Gulf of Gabès endemics: (1) it is a distinct species that has originated elsewhere and is now restricted to the Gulf; (2) it is a distinct species that has originated elsewhere, has colonized the Gulf and is also present elsewhere in the Mediterranean; (3) it is a distinct species that has originated in the Gulf very recently; or (4) it is not a distinct species, and the Gulf of Gabès populations are merely ecophenotypes resulting from the extreme environmental conditions in the Gulf. Our results confirm the validity of *D. demartiniorium*, and therefore exclude hypothesis (4) above. However, just like Barco *et al.* (2013), our results do not positively favour one or the other of hypotheses (1)–(3). The possible occurrence of *D. demartiniorium* in Tripoli (Libya) (Giannuzzi-Savelli *et al.*, 1994, see above) suggests that the range of *D. demartiniorium* includes the Gulf of Syrte. The Gulf of Syrte has a narrow continental shelf that drops quickly to deep water, and thus has always remained marine during Glacial low sea levels. We hypothesize that *D. demartiniorium* in fact originated in the Gulf of Syrte during Pleistocene low sea levels and subsequently

Table 3. Comparative morphological data of the shells of Mediterranean *Diodora*.

	<i>D. demartinorum</i>	<i>D. italica</i>	<i>D. giannispadai</i>	<i>D. gibberula</i>	<i>D. dorsata</i>	<i>D. graeca</i>
Average adult dimensions	length 35 mm. width 21 mm. height 7.5 mm	length 39 mm. width 21.5 mm. height 8 mm	length 45.5 mm. width 21.5 mm. height 13.5 mm	length 20 mm. width 12 mm. height 6.2 mm	length 13 mm. width 5 mm. height 3.2 mm	length 21 mm. width 10.5 mm. height 5 mm
Sculpture	75 to 78 equal. closely and evenly spaced primary radial ribs of even size. with 2 secondary riblets per interspace. Commarginal cords fine and evenly spaced	28 to 36 widely spaced primary radial ribs. of uneven size. with 1 to 3 secondary riblets per interspace. Commarginal cords coarse and unevenly spaced	70 unequal. widely and evenly spaced primary radial ribs. with 3 secondary riblets per interspace. Commarginal cords coarse and unevenly spaced	46 to 52 coarse and uneven ribs. with 0 to 3 uneven secondary riblets. Commarginal cords broadly spaced. forming pustules at intersection with ribs	overall sculpture as in <i>D. gibberula</i> . 42 radial ribs	18–20 widely spaced radial ribs with 3 secondary riblets per interspace. Commarginal cords forming a regular reticulation with intersected ribs
Anal pore	rounded-elongated; distance to anterior shell edge 40–52% of distance to posterior shell edge	elongated; distance to anterior shell edge 55–61% of distance to posterior shell edge	elongated; distance to anterior shell edge c. 50% of distance to posterior shell edge	shape variable; distance to anterior shell edge c. 46% of distance to posterior shell edge	shape variable; distance to anterior shell edge c. 26% of distance to posterior shell edge	elongated. rounded anteriorly and truncated posteriorly; distance to anterior shell edge c. 70% of distance to posterior shell edge
Colour	inner background colour beige. darker apically; outer shell surface dark grey with greenish bands radiating from the apex	inner background colour evenly beige; outer shell surface cream with darker bands radiating from the apex	inner background colour evenly light beige; outer shell surface cream with darker. greenish to brownish. bands radiating from the apex	inner background colour beige; outer shell surface white-yellow with darker bands radiating from the apex	inner background colour evenly beige; outer shell surface light grey with greenish bands radiating from the apex	inner background colour white with outer bands faintly visible through; outer shell surface grey with darker to greenish bands radiating from the apex
Profile	low; peristome adherent to the plane	elevated; convex (peristome not adherent to the plane)	low; peristome adherent to the plane	elevated; convex (peristome not adherent to the plane)	elevated; convex (peristome not adherent to the plane)	elevated; convex (peristome not adherent to the plane)

invaded the Gulf of Gabès. Its apparent endemism there results from a sustained interest by malacologists in the Gulf of Gabès ever since Monterosato (1879) whereas, by contrast, the coasts of Libya remain essentially unexplored and its biota undocumented. The occurrence of another large *Diodora* in the Aegean Sea (and possibly the Gulf of Iskendrun) raises the possibility that populations of *Diodora* remained trapped in various Mediterranean refuges during glacial times, underwent genetic drift and speciated during this/these phase(s) of isolation, as found by other gastropods (e.g. *Dendropoma*, Calvo *et al.*, 2015). In this respect, it is noteworthy that the North Adriatic has at least a couple of endemic molluscs (Sabelli & Taviani, 2013), and it would be most interesting to test genetically the identity of *Diodora italica* from the northern Adriatic which, like the Gulf of Gabès, dried out during glacial times and was subsequently reinvaded.

The systematics of the Mediterranean *Diodoras* thus ends up considerably more complex than the long-accepted three-species group. Beside the recognition of *D. demartiniorum* and *D. giannispadai* as distinct species, the present study also points to the existence of at least another species (here tentatively identified as *D. producta*). Clearly, molecular data for more morphs from more localities are critically needed.

ACKNOWLEDGEMENTS

The material for molecular studies in this paper derived from fieldwork in the Gulf of Gabès conducted specifically for this research, and we are grateful to everyone in the field party, Philippe Maestrati, Emmanuel Vassard, Gianni Spada, Marco Oliverio and Jean Pierre Miquel for their help in sampling. Special thanks go to Kyriakos Papavasileiou who collected specimens in the northern Aegean, and Junes Esteban who collected specimens in Murcia and Chafarinas. Paolo Russo sent photos and offered expert opinion. Virginie Héros, Barbara Buge and Philippe Maestrati helped with the curation of vouchers and access to MNHN historical material. The molecular part of the work was carried out at Service de Systematique Moléculaire (UMS 2700 CNRS, MNHN).

FINANCIAL SUPPORT

This project was partly funded by the MNHN ATM 'Barcode' (PIs: S. Samadi and J.N. Labat).

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