

# Prenatal stress and the developing brain: Risks for neurodevelopmental disorders

BEA R. H. VAN DEN BERGH,<sup>a,b</sup> ROBERT DAHNKE,<sup>c</sup> AND MAARTEN MENNES<sup>d</sup>

<sup>a</sup>University of Leuven; <sup>b</sup>Belgian Department for Welfare, Public Health and Family; <sup>c</sup>University Hospital Jena; and <sup>d</sup>Radboud University

## Abstract

The prenatal period is increasingly considered as a crucial target for the primary prevention of neurodevelopmental and psychiatric disorders. Understanding their pathophysiological mechanisms remains a great challenge. Our review reveals new insights from prenatal brain development research, involving (epi)genetic research, neuroscience, recent imaging techniques, physical modeling, and computational simulation studies. Studies examining the effect of prenatal exposure to maternal distress on offspring brain development, using brain imaging techniques, reveal effects at birth and up into adulthood. Structural and functional changes are observed in several brain regions including the prefrontal, parietal, and temporal lobes, as well as the cerebellum, hippocampus, and amygdala. Furthermore, alterations are seen in functional connectivity of amygdala–thalamus networks and in intrinsic brain networks, including default mode and attentional networks. The observed changes underlie offspring behavioral, cognitive, emotional development, and susceptibility to neurodevelopmental and psychiatric disorders. It is concluded that used brain measures have not yet been validated with regard to sensitivity, specificity, accuracy, or robustness in predicting neurodevelopmental and psychiatric disorders. Therefore, more prospective long-term longitudinal follow-up studies starting early in pregnancy should be carried out, in order to examine brain developmental measures as mediators in mediating the link between prenatal stress and offspring behavioral, cognitive, and emotional problems and susceptibility for disorders.

The early environment, starting in utero, can represent major risk factors for a lifetime of physical, psychiatric, and neurological problems for the individual (Braun et al., 2017; Gluckman, Hanson, Cooper, & Thornburg 2008; Hanson & Gluckman, 2011; Howard, Molyneaux, et al., 2014; Seckl, 2007; Van den Bergh, 2011; Van den Bergh et al., 2017). While according to DSM-5 (American Psychiatric Association, 2013), neurodevelopmental disorders are a group of disorders whose onset and clinical expression occur in childhood (Thapar, Cooper, & Rutter, 2017), other psychiatric disorders are now being considered within the context of a neurodevelopmental disorder, even if their onset occurs in adolescence or adulthood (Deoni et al., 2014; Meredith, 2015; Thomason et al., 2015).

Since its inception, the field of developmental psychopathology has emphasized the dynamic and complex interactions between an individual and its developmental environment in shaping almost all forms of psychopathology (Cicchetti & Rogosch, 1996; Hyde, 2015; Sroufe & Rutter, 1984). For instance, an important tenet of the developmental psychopathology perspective is that early life adversity may

provide serious challenges to the species-typical organism–environment “coactions” that play important roles in the emergences and timing of normal developmental change (Cicchetti, Handley, & Rogosch, 2015, p. 553). The developmental origins of health and disease research field also adopts the developmental plasticity idea and extends it up into the prenatal life period. Initially a lowered birth weight was taken as a proxy measure of prenatal environmental exposure in most studies. A lowered birth weight was shown to be a risk factor for the development of cardiovascular and metabolic diseases such as arterial hypertension, coronary heart disease, obesity, and type 2 diabetes (Barker, 1990) as well as mental health problems such as depression (Thompson, Syddall, Rodin, Omond, & Barker, 2001) and schizophrenia (Rifkin, Lewis, Jones, Toone, & Murray, 1994). Although much less examined, accumulating evidence mostly gathered during the last decade is now revealing prenatal environmental exposure effects on early brain and behavior development in humans. The developmental origins of behavior, health, and disease has recently been proposed (Van den Bergh, 2011), and examples of its empirical testing have been described (Van den Bergh, 2016). It is generally accepted that behavioral problems, neurodevelopmental issues, and psychiatric disorders are driven by atypical brain functions, which in turn reflect alterations in underlying brain structure and circuitry (van Essen & Barch, 2015). It is now also being accepted that these alterations may have their origin in the earliest periods of brain development. It is an important aim of our paper to reveal important new insights

\*This article has been corrected since its initial publication. See doi:10.1017/S0954579418001402

This research was supported by funding from EU FP7/Health.2011.2.22-2 and GA2798219 (to B.v.d.B.).

Address correspondence and reprint requests to: Bea R. H. Van den Bergh, Health Psychology, KU Leuven, University of Leuven, Tiensestraat 102, B-3000 Leuven, Belgium; E-mail: [Bea.vandenbergh@kuleuven.be](mailto:Bea.vandenbergh@kuleuven.be).

gained from this growing field of research; we particularly focus on new findings from research in humans.

Many authors point explicitly to the role of maternal stress, anxiety, and depressive symptoms (Bowers & Yehuda, 2016; Kofink, Boks, Timmers, & Kas, 2013; Lewis, Galbally, Gannon, & Symeonides, 2014; O'Connor, Monk, & Fitelson, 2014; Van den Bergh et al., 2005) and maternal mental illness during pregnancy (Howard, Molyneaux, et al., 2014; Jones, Chandra, Dazzan, & Howard, 2014; Stein et al., 2014) as risk factors that may lead to functional and structural brain changes in the offspring (Charil, Laplante, Vaillancourt, & King, 2010; Franke et al., 2017; Scheinost, Sinha, et al., 2016; Van den Bergh et al., 2017). In these review papers, offspring brain alterations are seen as mediating the link between prenatal exposure to maternal distress, and offspring behavioral, cognitive, and emotional development and susceptibility to neurodevelopmental and psychiatric disorders. Functional and structural brain measures of their regional connectivity may be seen as putative footprints or biomarkers of prenatal stress, altering risk or resilience for neurodevelopmental and psychiatric disorders. Maternal psychological distress in the studies reviewed refers to (a) general or pregnancy-specific anxiety, and depressive symptoms; (b) major life events experienced by the mother (such as illnesses or deaths in the close family, financial and relationship problems, house moves, car accident, etc.); (c) a psychiatric diagnosis of a current or past anxiety or depression disorder; and (d) exposure to a disaster (maternal hardship due to a natural disaster) and the subjective distress and cognitive appraisal related to it. Maternal distress is measured with one of the following methods: (a) self-report distress and/or life events questionnaire, (b) a physician's diagnosis based on medical chart information or on a diagnostic psychiatric instrument, and/or (c) a physiological variable (such as cortisol).

Before reviewing studies on potential brain biomarkers of prenatal stress, we provide a short overview of brain imaging techniques and analysis methods. In the next section, we describe new insights into functional genomics and into a typical developmental trajectory of the human brain. We focus on recent neuroimaging studies and on studies starting from computational stimulation models that enable researchers to quantify the critical physical phenomena that are necessary to induce folding and predict gyral wavelength and gyrification indices. Next, we present human prenatal stress studies that have explicitly examined alteration in offspring brain development and share knowledge about the putative prenatal origins of neurodevelopmental and psychiatric disorders. Finally, we present ways in which future research may contribute to further mechanistic understanding of how prenatal stress exposure may lead to aberrant brain circuitry and describe clinical and societal responsibilities associated with prenatal stress.

### The Developing Brain: Imaging Techniques and Analysis Methods

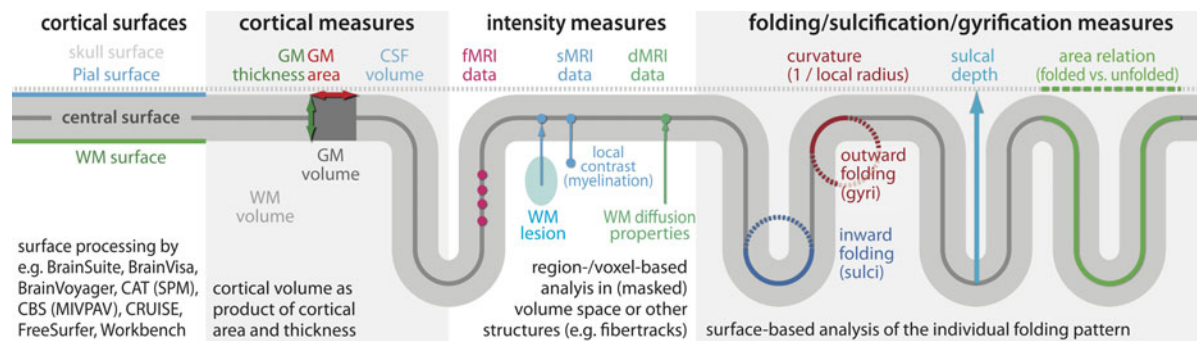
Magnetic resonance imaging (MRI) allows a variety of protocols with different contrasts to investigate structural (sMRI),

functional (fMRI), and diffusion (dMRI) specific *in vivo* properties of the brain by means of volume-, surface-, deformation-, or regions-wise analysis (Ashburner & Friston, 2000; Fischl, 2012; Gaser, Nenadic, Buchsbaum, Hazlett, & Buchsbaum, 2001). Given its noninvasive nature, MRI allows characterizing the human brain across the life span (see also the developing human connectome project at <http://www.developingconnectome.org>) ranging from *in utero* (e.g., Thomason et al., 2015) and longitudinal imaging, (e.g., Li, Nie, et al., 2014; Mills & Tamnes, 2014) to postmortem acquisitions (e.g., Kostović et al., 2014; Miller et al., 2011). Accordingly, MRI-based investigations have provided great contributions to our understanding of both typical and atypical brain structure and function.

In general, sMRI analyses focus on morphometric brain features such as thickness (e.g., Dahnke, Yotter, & Gaser, 2013; Fischl, 2012; Winkler et al., 2010), area (e.g., Fischl, 2012; Winkler et al., 2010), volume (e.g., Winkler et al., 2010), and gyrification (e.g., Li, Wang, et al., 2014; Schaer et al., 2008) of the brain's gray matter (GM), white matter (WM), and cerebrospinal fluid by means of voxel-based (e.g., Ashburner & Friston, 2000), surface-based (e.g., Fischl, 2012), deformation-based (e.g., Gaser et al., 2001), or region-based morphometry analyses (e.g., Fischl, 2012; Klein & Tourville, 2012) as illustrated in Figure 1. A reasonable resolution (about 1 mm or better), good tissue contrast, and artifact-free images (e.g., motion; Reuter et al., 2015) are required for accurate tissue classification (e.g., Ashburner & Friston, 2000), surface reconstruction (e.g., Dahnke et al., 2013; Fischl, 2012; Li, Wang, et al., 2014), and to normalize the individual anatomy to a common reference template or atlas (spatial registration; e.g., Ashburner & Friston, 2011; Avants et al., 2011; Klein & Tourville, 2012; Tardif et al., 2015). Besides tissue classification, quantitative imaging (e.g., Weiskopf et al., 2013) or specific contrast changes in the WM (e.g., the water myelin fraction; Billiet et al., 2015) or GM (e.g., Robinson et al., 2014) allows analysis of microstructural changes.

In general, dMRI aims to characterize WM fiber tracts by measuring diffusion patterns of water inside brain tissue. It allows to describe local fiber properties such as fractional anisotropy (FA; i.e., direction of diffusion), axial diffusivity (AD), neurite orientation dispersion and density imaging (NODDI) (e.g., Billiet et al., 2015; Jelescu et al., 2015; Kunz et al., 2014; Winston et al., 2014; Zhan et al., 2013), or fiber tracing (e.g., Billiet et al., 2015; Mori et al., 2002; Roalf et al., 2016; Yu et al., 2015).

In general, fMRI data are time series of volumes (4D data) that index changes in the oxygenation of the blood that allow analyzing brain activity patterns while the participant is performing specific tasks (e.g., visual perception; Kanwisher, McDermott, & Chun, 1997) or during a resting state (rs; e.g., Smith et al., 2009). Functional data can also be measured by other noninvasive methods such as electroencephalography (EEG; e.g., Hunter et al., 2012; Mulkey et al., 2015; Otte, Donkers, Braeken, & Van den Bergh, 2015; Stam & van Straaten, 2012; Tóth et al., 2017), magnetoencephalogra-



**Figure 1.** (Color online) Magnetic resonance imaging (MRI)-based measures. There are MRI-based measures to characterize structural and function changes of specific brain regions. Structural analysis uses the image contrast to classify the major tissue (GM, gray matter; WM, white matter; CSF, cerebrospinal fluid), map the individual brain to a common template (spatial normalization), and analyze volume, thickness, area, folding, or intensity changes. Surface-based analysis allows (a) separating cortical GM into area and thickness especially in case of thickness reduction and simultaneous area enlargement (Winkler et al., 2010), (b) the estimation of the myelination degree (Billiet et al., 2015; Robinson et al., 2014), and (c) the cortical folding that is expected to describe the properties of its early development in utero (Budday et al., 2015; Tallinen et al., 2016). Surface-based processing further allows improved normalization and smoothing that is typically required for statistical analysis (Ashburner & Friston, 2000). To measure WM changes diffusion MRI (dMRI) allows identifying of fiber properties. In addition, dMRI but also functional MRI (fMRI) can further be used to reconstruct structural dMRI and functional fMRI networks (connectomes) to understand the connectivity of different brain areas.

phy (e.g., Stam et al., 2014; Stam & van Straaten, 2012). Frontal alpha EEG asymmetry has been used as a rs-EEG measure (e.g., Müller, Kühn-Popp, Meinhardt, Sodian, & Paulus, 2015) while event-related potential studies (ERP), involving averaging the EEG activity time-locked to the presentation of a stimulus of some sort (visual, somatosensory, or auditory) are task-related EEG-measures (e.g., Hunter et al., 2012; Otte et al., 2015). EEGs record the electrical brain activity with superior temporal but lower spatial resolution compared to fMRI.

The functional units of the brain are networks of specialized, neural structures shown to communicate with each other (Poldrack, 2012). Conventional task-based analytical approaches use univariate models that calculate average responses of the brain to manipulations, either for a region of interest or at the whole-brain level (voxel-wise); these models enable localizing cognitive functions based on the blood oxygen level dependent response (Ogawa et al., 1992). However, a model that takes the connections between neural structures into account is likely to give a more valid description of neural responses than a model that assumes functional independence (Sporns, Chialvo, Kaiser, & Hilgetag, 2004). Accordingly, moving beyond characterizing individual regions, the brain is increasingly viewed as a connected organ, a connectome (Hagmann, 2005; Sporns, Tononi, & Kötter, 2005) where regions interact in larger networks in order to optimize behavior. This research field is often referred to as “connectomics” (Behrens & Sporns, 2012), a popular term to capture research aimed at describing structural and/or functional connections between nodes (i.e., specified regions) within the connected brain (Rubinov & Sporns, 2010; Watts & Strogatz, 1998). For a description of analytic connectivity methods, we refer to papers that describe important categories of analytic methods including seed-based correlations, independent component analysis, clustering, pattern classification, and lo-

cal method such as regional homogeneity and low-frequency fluctuations (e.g., Cole, Smith, & Beckmann, 2010; Margulies et al., 2010).

In neurodevelopmental studies (e.g., attention-deficit/hyperactivity disorder [ADHD]; Castellanos & Proal, 2012) two functional connectivity (FC) approaches are often used. Seed-based correlations examine correlations of time series between a region of interest (a “seed”) and remaining GM voxels. Yet, constraining seed selection remains a challenge as even minor variations matter. Independent component analysis is a second, popular alternative; four-dimensional imaging data are decomposed into three-dimensional spatial maps, each with associated time courses (Castellanos & Aoki, 2016). These maps of coherent spontaneous blood oxygen level dependent signals correspond to functional networks that are revealed by task-based fMRI (Smith et al., 2009), including the default mode network (DMN), which has been historically most often studied and includes the medial parietal (precuneus and posterior cingulate), bilateral inferior-lateral-parietal, and ventromedial frontal cortex (Smith et al., 2009). Other large-scale networks include the executive control, sensorimotor, and left and right attention networks. In human prenatal stress research this method is only beginning to be used.

In the context of connectivity analyses, graph theory is often used as one mathematical framework to quantitatively describe the topological organization of connectivity. Several complex network measures of brain connectivity identifying centrality, functional integration, and segregation have been described in Rubinov and Sporns (2010). Next to information about overall network infrastructure, specific features are also conveyed, such as which “nodes” (locations) within a system are central “hubs” of connectivity, linking numerous other units to one another. For example, graph analysis applied to fMRI data sets revealed that the human brain is organized

with “small-world” topology (van den Heuvel, Mandl, & Hulshoff Pol, 2008); this is a network “in which constituent nodes exhibit a large degree of clustering as well as relatively short distances between any two nodes of the system and is thought to reflect a balance between local processing and global integration of information” (Menon, 2013, p. 629). Another important finding is that the posterior cingulate and insular cortices are connectivity hubs (Fransson & Marrelec, 2008; Hagmann et al., 2008; Margulies et al., 2009; Menon, 2013). These topological brain connectivity measures have been used to characterize human brain development, starting in utero; their use in human prenatal stress research is being explored (see e.g., van den Heuvel & Thomason, 2016).

### Development of the Human Brain: Insights From Physical Modeling and Computational Simulation Studies and Advanced Neuroimaging Techniques

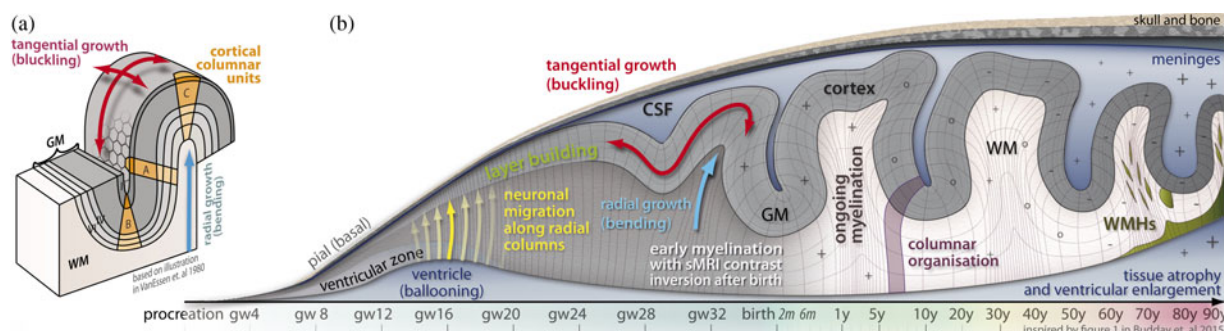
We highlight the importance of functional genomics for the brain developmental trajectory and then focus this section on new insights about brain development that are mainly gained by physical modeling and computational simulation studies and brain imaging studies. Brain development follows biomechanical rules and undergoes different major stages: ballooning and gyrification in utero (Bayly, Taber, & Kroenke, 2014; Budday, Steinmann, & Kuhl, 2015; Lewitus, Kelava, & Huttner, 2013; Striedter, Srinivasan, & Monuki, 2015; Tallinen & Biggins, 2015; Tallinen et al., 2016) and a subsequent scaling in childhood and adolescence (Franke, Luders, May, Wilke, & Gaser, 2012; Jiang & Nardelli, 2016), whereas further healthy changes are recognized as plasticity (short time; e.g., Gaser & Schlaug, 2003; Maguire et al., 2000; Reid, Sale, Cunnington, Mattingley, & Rose, 2017) and aging (long time; e.g., Franke, Ziegler, Klöppel, & Gaser, 2010; Ziegler, Ridgway, Dahnke, Gaser, & Alzheimer’s Disease Neuroimaging Initiative, 2014). These processes are illustrated in Figure 2. For a detailed overview of in utero development, we would like to refer to Budday et al. (2015) and give here just a short overview.

#### *Functional genomics of human brain development*

The building of the brain starts from a patch of cells; during embryonic and fetal life, complex developmental processes construct its initial architecture, which is dynamically changed during the whole life span (Collin & van den Heuvel, 2013; Silbereis, Pochareddy, Zhu, Li, & Sestan, 2016). Although it is generally known that from the first phase in development, that is, the zygote, the species-specific genome functions to guide development, recent large-scale analyses of the proteome/transcriptome and regulome have revealed that brain development entails very complex and sophisticated interactions between genetic, epigenetic, and environmental factors (for reviews, see Jiang & Nardelli, 2016; Kang et al., 2011; Pletikos et al., 2014; Silbereis et al., 2016; Ziats, Grosvenor, & Rennert, 2015). A most important

insight is that gene expression dynamics underlying early human brain development are spatially and temporally specific and involve complex regulatory processes at several levels, for example, coordinate progressive cell fate specification and tissue morphogenesis (Bernadskaya & Christiaen, 2016). The regulatory processes involve (a) transcription factors that operate in the context of complex gene regulatory networks, for example, transregulatory elements, which are genes that modify the expression of distant genes through intermolecular interaction and cis-regulatory modules, a stretch of DNA sequences where transcription factors bind and regulate expression of nearby genes; (b) epigenetic gene regulatory mechanisms (such as DNA methylation and histone modification, long noncoding RNAs, and short noncoding RNAs, including microRNAs; Geschwind & Flint, 2015). This means that major aspects of development such as proliferation, migration, and differentiation are not fully programmed genetically. According to Ben-Ari and Spitzer (2010, p. 486), they rely on “phenotypic checkpoints, i.e., times and places during development at which functional validation appropriate to the stage of the cells enables the process to go forward normally, take an alternative route, or become arrested.” According to Huang, Hu, Kauffman, Zhang, and Shmulevich (2009, p. 1), theoretical considerations as well as experimental evidence support the view “that cell fates (or commitment) are ‘high dimensional attractor states’ of the underlying molecular network.” There is emerging evidence from preclinical and from human postmortem, modeling and neuroimaging studies that, next to genetic risks (i.e., polygenic risks; common, rare, and de novo mutations), diverse extrinsic factors occurring in the prenatal and early postnatal life period presumed to enable changes in these complex regulatory processes, alter the spatiotemporal expression of gene patterns. The latter leads to alterations in patterning and regionalization of the GM, as well as in intracortical myelin and WM; all of these changes influence the developing structural and functional brain connectome in spatiotemporal specific ways (e.g., Bernadskaya & Christiaen, 2016). Negative environmental factors include maternal nutritional, medical and distress factors, and hypoxia/ischemia. Emerging evidence shows that such prenatally and early postnatally acquired “neuroanatomy” changes (Donovan & Basson, 2017) partially underlie behavioral and cognitive changes, including those observed in common neurodevelopmental and psychiatric disorders (Deoni et al., 2014; Ghiani & Faundez, 2017; Haroutunian et al., 2014; Ziats et al., 2015). Positive environmental factors, such as getting breastfed for a period of at least 3 months, is shown to have a developmental advantage (i.e., a positive effect on WM microstructure in late maturing frontal and association brain regions; Deoni et al., 2013). We refer to review papers (e.g., Bernadskaya & Christiaen, 2016; Geschwind & Flint, 2015; Parikshak, Gandal, & Geschwind, 2015) that specify how system biology and network approaches (e.g., by using graph theory) are applied to human genetics.





**Figure 2.** (Color online) Brain development and aging. (a) Cortical folding. (b) Illustration of brain development and aging process over lifetime, with focus on the prenatal life period. One of the essential times in development is the apical creation of neurons and their migration to the pial regions where they build the cortical gray matter layer. This immense tangential and radial growth causes folding of larger gyri and sulci between gestation weeks 24 and 34 (Budday et al., 2015). The myelination begins significantly after birth and ends after decades. The local folding (bending and buckling) compresses and stretches the cortical layers by keeping the volumes of each layer of the imaginary cortical columnar units A, B, and C (see part [a]) relatively similar (van Essen & Maunsell, 1980).

### Prenatal brain development phases: Ballooning and gyrification

The ballooning phase, from gestation week (gw) 3 to 15, is described by an intensive radial enlargement of the ventricle that compensates the simultaneous tangential growth of the intermediate zone and increases the brain surface without significant folding, where only the longitudinal and Sylvian fissures become prominent by radial growth (bending, i.e., forces below the developing cortex; see Figure 2). In gw 5 to 20, neurons are generated in the ventricular zone and migrate to the skull, where they create the cortical layer structure. At this time, the cortex shows a radial dMRI pattern, indicating low connectivity within the cortex (Budday et al., 2015; Jiang & Nardelli, 2016; Striedter et al., 2015), whereas the first large fiber tracts are becoming visible in the WM (e.g., in the corpus callosum; Wegiel et al., 2010).

After ballooning, the neurons start forming layer depending connections when neuronal migration is finished and the radial dMRI pattern gets lost (Budday et al., 2015, p. 6; Wegiel et al., 2010). This stage also involves the growth of neuronal dendrites and axons; the production and expansion of astrocytes, oligodendrocytes, and microglial cell; as well as the formation of synapses and the development of the vasculature system (Budday et al., 2015, p. 14). By these phenomena, the tangential growth of the outer cortex becomes prominent and causes tangential growth (buckling, i.e., forces within the developing cortex) and forms major structures such as the central sulcus around gw 24 (Budday et al., 2015; Tallinen et al., 2016). External forces due to limitations of the skull and meninges were found to have minor effects, and it is presumed that gyrification depends on tangential growth of the GM; this is referred to as the buckling theory (e.g., Bayly et al., 2014; Budday et al., 2015; Striedter et al., 2015; Tallinen et al., 2016). Recent experimental and computational growth models (Bayly et al., 2014; Budday et al., 2015; Tallinen et al., 2016; Toro, 2012) have shown promising results to explain the natural folding as an energy-minimizing process of radial and tangential sur-

face expansion that relies on the stiffness of the inner core (the WM), the cortical growing rate, and local cortical thickness. The bending of cortex is locally compensated by thickness changes of the cortical layer that finally guarantee an equal number of neurons independent of the local amount of folding (van Essen & Maunsell, 1980). Tallinen and Biggins (2015) demonstrate that soft and thinner structures and high growing rates lead to increases in “Z” folds (that are more typical for early development), whereas stiffer cores led to more complex “Y” folds (that are more typical for later development). Modification of these morphological properties may occur with alterations either in the gene network (e.g., by de novo mutations, in transcription factors) or in the epigenetic gene regulatory networks (e.g., DNA methylation, long noncoding RNAs, which may be caused by environmental factors); these changes may result in varying folding pattern that can be measured even in the adult brain (Bayly et al., 2014; Budday et al., 2015; Tallinen & Biggins, 2015; Tallinen et al., 2016). The gyrification occurs in synchrony with neuronal connectivity; that is, it starts after all neurons have reached their final position in the developing cortex between gw 17 and gw 47 (i.e., seventh week after birth in term-born babies; Budday et al., 2015). However, it is also the case that early neuronal migration “sets the stage” for the folding process as disturbed migration processes lead to disrupted neuronal connectivity and, hence, delayed growth and altered cortical formation of malformation (Budday et al., 2015, p. 14). The folding is nearly completed around birth in humans, and both tangential and radial growth is balanced again (Evans & Group, 2006; Li, Wang, et al., 2014; Tallinen & Biggins, 2015).

### The developing connectome: In utero and during infancy

With the use of rs-MRI, it has become recently possible to map FC of the human fetus in utero. Earlier studies in full-term (Fransson et al., 2007) and preterm infants (Doria et al., 2010; Smyser et al., 2010) had suggested the existence

of a “proto” or partial baseline DMN similar to the DMN observed in children and adults. Fetal rs-fMRI has confirmed the presence of primitive forms of functional networks by middle gestation, including the DMN. The connectivity of the posterior cingulate cortex may be seen as a precursor of the adult DMN (van den Heuvel & Thomason, 2016). Thomason et al. (2014) applied a graph theoretical approach to resting state FC of healthy fetuses, 19 to 39 gw. It was found that the fetal brain has a modular organization and that the modules overlap with functional systems observed postnatally. Compared to younger fetuses (<31 gw), in older fetuses (>31 gw) modularity decreases, and connectivity of the posterior cingulate to other brain networks becomes more negative. Modularity refers to the degree to which a network can be divided into nonoverlapping subsets of regions (i.e., “modules”) that are internally interactive, while sparsely connected with outside areas (Thomason et al., 2014, p. 2). The higher modularity in the younger fetuses is indicative of more segregated functional subnetworks. Fetal FC may develop according to a medial to lateral (Schöpf, Kasprian, Brugger, & Prayer, 2012; Thomason et al., 2013) and a posterior to anterior pattern (Jakab et al., 2014).

We refer to recent review papers for a detailed overview of the development of the brain connectome. Of the review papers that focus specifically on early life (i.e., prenatal and/or infancy and/or early childhood), some papers either focus on FC (e.g., Kipping, Tuan, Foisser, & Qiu, 2016; Menon, 2013; Power, Fair, Schlaggar, & Petersen, 2010; van den Heuvel & Thomason, 2016) or on structural connectivity (SC; e.g., Battalle et al., 2017). Other authors describe both structural and functional networks combined with the description of at least one of the following topics: (a) the development of the structure–function coupling (Hagmann, Grant, & Fair, 2012); (b) underlying neurobiological processes (Collin & van den Heuvel, 2013; Dubois et al., 2014; Vértes & Bullmore, 2015); (c) atypical connectome development in at-risk populations (Ball et al., 2014; Dennis & Thompson, 2013; Di Martino et al., 2014; Gupta, Gupta, & Shirasaka, 2016; Han, Chapman, & Krawczyk, 2016; Koyama et al., 2016); or (d) the environmental and genetic factors that influence the connectome (Atasoy, Donnelly, & Pearson, 2016; Richmond, Johnson, Seal, Allen, & Whittle, 2016). Studies that focus on the whole life span mostly compare brain development in infancy, childhood, adulthood and/or old age (e.g., Billiet et al., 2015; Lebel et al., 2012).

#### *Brain development across the life span: Short overview*

Over an individual’s lifetime, cortical thickness peaks between ages 3 and 9 (Walhovd, Fjell, Giedd, Dale, & Brown, 2016) and then shrinks slowly every year, whereas the WM continues to develop until the fourth to fifth decade (Billiet et al., 2015; Chang et al., 2015; Walhovd et al., 2016). The maturing and regressive biophysical changes in both GM and WM occur heterochronically in different brain regions (Haroutunian et al., 2014). Besides the global trend of tissue

atrophy with aging, brain plasticity allows an increase in local tissue volume by learning (Gaser & Schlaug, 2003; Maguire et al., 2000; Reid et al., 2017).

The WM can further degenerate as evidenced by MRI as WM hyperintensities with GM-like intensities in aging (Evans & Brain Development Cooperative Group, 2006; Habes et al., 2016), as well as in diseases such as multiple sclerosis (Schmidt et al., 2012) and Alzheimer disease (Ziegler et al., 2014). In neurodegenerative diseases such as Alzheimer disease, accelerated tissue atrophy was reported (Franke et al., 2010; Ziegler et al., 2014).

#### **Prenatal Stress Alters the Developmental Trajectory of the Brain: Brain Imaging Studies From Birth Until Adulthood**

There is an increasing number of prospective studies examining associations between prenatal exposure to maternal distress during pregnancy and offspring outcome measures such as motor development, cognition, neurocognitive functioning, learning problems, temperament, and mental health (Bock, Wainstock, Braun, & Segal, 2015; Bowers & Yehuda, 2016; Lewis et al., 2014; Van den Bergh et al., 2017). However, only a small number of the studies reviewed included brain measures. Studies explicitly examining offspring brain functional, structural, and brain connectome measures are promising in contributing crucial knowledge, (e.g., about alteration in specific brain regions or networks that may underlie the observed effects in the offspring). Moreover, especially the prospective longitudinal follow-up studies extending over a considerable postnatal time period may reveal changes in brain developmental trajectories. When available, information on mediating factors (such as genetic and epigenetic factors) and moderating factors (such as gender, postnatal maternal distress) will also be described.

#### *Results in newborns, infants, and preschoolers (Ages: 0–5 year)*

*Structural brain changes: sMRI and dMRI studies.* In the prospective longitudinal Growing up in Singapore Towards Healthy Outcomes (GUSTO), pregnant mothers were recruited at 13 weeks of pregnancy. Maternal depression and anxiety were measured at 26–28 weeks of pregnancy and when the child was 3 months and 1, 2, 3, and 4.5 years old. Effect of prenatal exposure to maternal distress was examined in infants ( $N =$  between 24 and 203) at 4–17 days after birth, at 6 months of age, and at 4.5 years with sMRI and/or dMRI of specific brain regions (Qiu, Anh, et al., 2015; Qiu et al., 2013; Rifkin-Graboi et al., 2013; Wen et al., 2017) or the whole brain (Chen et al., 2015; Qiu, Tuan, et al., 2015; Rifkin-Graboi et al., 2015).

Rifkin-Graboi et al. (2013) observed a significant association between maternal depression in pregnancy and microstructure of the amygdala, a region associated with stress reactivity and fear regulation (i.e., compared to neonates of

mothers with low-normal depression scores in pregnancy, FA in the left and right amygdala, and AD in right amygdala were lower in neonates of mothers with high depressive symptoms in pregnancy). However, no associations were found between maternal depression and volume of either the left or right amygdala. In contrast, the whole-brain analysis dMRI study of Rifkin-Graboi et al. (2015) revealed no effects of maternal anxiety during pregnancy on amygdala microstructure. However, maternal anxiety did predict alterations, that is, lower FA in WM fiber tracts in several regions: (a) dorsolateral prefrontal cortex and right insular cortex (regions important to cognitive–emotional responses to stress); (b) right middle occipital (important for sensory processing); (c) right angular gyrus, uncinate fasciculus, posterior cingulate, and parahippocampus (important for social cognition, social–emotional functioning); and (d) right cerebellum (important for sensorimotor learning and higher cognitive function). Moreover, high maternal anxiety was associated with lower AD in the left lateral orbitofrontal cortex and left inferior cerebellar peduncle and with higher AD in the genu of the corpus callosum. As the latter study did not reveal significant effects of depression on FA or AD values after corrections for multiple testing, the authors concluded that the effects found may be anxiety specific (Rifkin-Graboi et al., 2015). Tentative support was found for associations of five lateralized clusters (right insular, inferior frontal, middle occipital, middle temporal, and parahippocampal) with offspring internalizing problems (but not externalizing behavior) at 1 year of age (Rifkin-Graboi et al., 2015).

Qiu, Tuan, et al. (2015) observed an effect of maternal anxiety during pregnancy on neonatal cortical morphology that was moderated in several ways by functional variants of the catechol-O-methyltransferase (*COMT*) gene, which regulates catecholamine signaling in the prefrontal cortex and is implicated in anxiety, pain, and stress responsivity. The A-val-G (AGG) haplotype moderated the positive link between maternal anxiety and thickness of the right ventrolateral prefrontal, right parietal cortex, and precuneus. The G-met-A (GAA) haplotype modulated the negative link between maternal anxiety and thickness of the dorsolateral prefrontal cortex, and bilateral precentral gyrus.

In the study by Chen et al. (2015), for each mother and each infant in the GUSTO cohort they identified the participant's polymorphism of the brain-derived neurotrophic factor (*BDNF*) Val66 gene, that is, Met/Met, Met/Val, or Val/Val. *BDNF* is a neurotrophin known to underlie synaptic plasticity in the central nervous system (Chen et al., 2015). They also conducted a genome-wide DNA analysis on umbilical cord samples, using Human Methylation450 Bead Chip Array, which allows quantification of methylation status with single-base resolution across 482,421 cytosine–phosphate–guanine sites (CpGs) and 3,091 non-CpGs (Chen et al., 2015 p. 140). A first result showed that 148,890 CpGs had an absolute methylation difference of 15% between the three genotypic groups of the *BDNF* Val66Met gene. Second, the strength of the associations between maternal anxiety and

neonatal DNA methylation was found to be different for different polymorphisms of the infant (not maternal) Val66Met gene; that is, in the Met/Met polymorphism group, there was a greater impact of antenatal maternal anxiety on the DNA methylation than in both other groups. Third, it was found that 9 of the 18 brain-volume measures used had significantly different numbers of variable CpGs (i.e., right amygdala, left hippocampus, left thalamus, left caudate, right midbrain, right cerebellum, left total WM, left GM, and right GM). For instance, there were significantly more CpGs where methylation levels covaried with right amygdala volume among Met/Met compared with both Met/Val and Val/Val carriers; in contrast, more CpGs covaried with left hippocampus volume in Val/Val infants compared with infants of the Met/Val or Met/Met genotype.

While Qiu et al. (2013) found no effect of prenatal exposure to maternal anxiety on hippocampal volume at birth or at 6 months of age, maternal anxiety was negatively correlated with hippocampal growth between 0 and 6 months. Furthermore, a positive association was observed between postnatal maternal anxiety and offspring right hippocampal growth and a negative one between postnatal maternal anxiety and left hippocampal growth at 6 months of age (Qiu et al., 2013).

Finally, Wen et al. (2017) showed sex-specific effects of exposure to maternal depression in the 4.5-year-olds of the GUSTO cohort. A positive association was found between maternal depressive symptoms during pregnancy and larger right amygdala volume in girls, but not in boys. Furthermore, a positive association was found between postnatal maternal depressive symptoms and higher right amygdala FA in the whole sample and in girls, but not in boys.

In a second cohort, recruited at Columbia University Medical Center in the city of New York, maternal depression as measured between 34 and 37 weeks of pregnancy with a self-reported questionnaire was used to define a depression group versus a not depressed group. Infants of both groups were examined at age of 5.8 weeks (Posner et al., 2016). Results of dMRI tractography demonstrated decreased structural connectivity between the right amygdala and the right ventral prefrontal cortex in the infants prenatally exposed to maternal depression ( $n = 18$ ) compared to nonexposed infants ( $n = 39$ ; Posner et al., 2016).

A third cohort was followed up until 2.6 to 5.1 years of age, as part of the Alberta Pregnancy Outcome and Nutrition Study (Lebel et al., 2016). It was found that women's depression at 17 weeks of pregnancy was associated with preschoolers' cortical thinning in the right inferior frontal and middle temporal region, and with radial diffusivity and mean diffusivity in WM emanating from the inferior frontal area. However, the latter association was no longer significant after correction for postpartum depression. Postpartum depression was related with cortical thinning in preschoolers' right superior frontal cortical thickness and with diffusivity in WM originating from that region; the effect remained significant after correction for prenatal maternal depression. Maternal de-



pression at 11 or 32 weeks of pregnancy was not significantly related with altered GM or WM structure (Lebel et al., 2016).

In a fourth cohort, the Generation R study, brain imaging data of  $N = 654$  6- to 10-year-old children prenatally exposed to maternal depressive symptoms (El Marroun et al., 2016) were analyzed. Maternal and paternal depressive symptoms was measured at 20.6 weeks of gestation and when the children were 3 years old. It was found that prenatal exposure to maternal depressive symptoms was associated (a) with a thinner cortex in the frontal area of the left hemisphere, (b) with larger cortical surface in a caudal middle frontal area in proximity to the area showing decreased cortical thickness, and (c) with gyrification measures (however, after correction for several covariates, this association did not remain significant). No association was found between prenatal exposure to maternal depressive symptoms and volumetric measures in core structures of the limbic system. Furthermore, no association was found with brain morphology measures and either prenatal paternal depressive symptoms or maternal depressive symptoms at age 3 years.

*Functional brain development: fMRI and EEG studies.* Scheinost, Kwon, et al. (2016) used rs-fMRI and whole-brain seed connectivity to compare FC in extremely premature born infants (born <28 gw) who, furthermore, were exposed to maternal anxiety or stress during pregnancy, with FC in a control group of very premature neonates (born <32 gw) and a term control group. Prenatal stress exposure was coded as a binary variable, that is, whether or not a diagnosis of depression and/or anxiety was retrieved in the maternal medical card. Compared to term controls, preterm born infants without exposure to maternal distress show decreased FC from the left amygdala to other subcortical regions (thalamus, hypothalamus, brainstem, and insula). Compared to the latter group, preterm infants prenatally exposed to maternal distress showed lower FC of the left amygdala to the thalamus, hypothalamus, and peristate cortex. In the study by Posner et al. (2016), rs-fMRI measures demonstrated that at the mean age of 5.8 weeks, infants prenatally exposed to maternal depression demonstrated increased inverse FC between the amygdala and the dorsal prefrontal cortex, bilaterally. Further analyses showed that changes in amygdala–prefrontal cortex connectivity were associated with an increase in fetal heart rate reactivity to a mild maternal stressor that had been measured at 34–37 weeks of gestation (Posner et al., 2016). Qiu, Anh, et al. (2015) showed that 6-month-old infants of the GUSTO cohort, born to mothers with higher depressive symptoms during pregnancy, had greater FC of the amygdala with several other brain regions: the left temporal cortex, insula, bilateral anterior cingulate, medial orbitofrontal and ventromedial prefrontal cortices. These networks are known to underlie depression in children and adults, and the authors suggest that their rs-fMRI data may foreshadow future neurodevelopmental and psychiatric disorder (Qiu, Anh, et al., 2015).

Other studies examined asymmetry of left and right frontal alpha EEG and stability of EEG patterns in infants. There is a

large literature on the effects of early adversity, including effects of prenatal and/or postnatal depression on offspring frontal EEG asymmetry, but results are inconsistent (see, for reviews, e.g., Field & Diego, 2008; Field, Hernandez-Reif, & Diego, 2006; Peltola et al., 2014). For instance, while greater relative right frontal EEG activation has been associated with maternal depression in pregnancy in some studies (e.g., Lusby, Goodman, Bell, & Newport, 2014), other studies concluded that the number of postpartum months of depression was a stronger predictor of infant frontal EEG asymmetry at 14 months of age than prepartum months of maternal depression (e.g., Dawson, Frey, Panagiotides, Osterling, & Hessel, 1997). Field et al. (2010), studied the effect of comorbid depression and anxiety during pregnancy (measured at 20 gw), and it was found that neonates of the comorbid and depressed groups had greater relative right frontal EEG measures than neonates of the anxiety and nondepressed groups. In the study of Lusby et al. (2014), participants all met DSM-IV criteria for depression or another mood disorder and were enrolled before 16 gw. Offspring EEG was recorded during baseline, feeding, and play at age 3 and 6 months. They observed an interaction effect of maternal prenatal and postnatal depressive symptoms on asymmetry in EEG patterns in 3- and 6-month-old infants, showing that prenatal depressive symptoms and infant EEG asymmetry scores were significantly associated among women with high postpartum depressive symptoms but not in those with low postnatal depressive symptoms. They concluded that asymmetry scores were mostly consistent across contexts (but not from baseline to feeding and play at 6 months) and stable across ages (but not during feeding). In another study of the same cohort, the moderating effect of prenatal maternal anxiety on associations between infants' frontal EEG asymmetry and temperamental negative affectivity across infants' first year of life was studied. The findings showed that behavioral and psychophysiological outcomes co-occur over the course of infancy and are different for infants of high versus low levels of maternal anxiety. For instance, infant negative affectivity and frontal EEG asymmetry were negatively associated at 3 months of age and positively associated by 12 months of age in the high depression mothers, while there was no association between infant negative affectivity and EEG at any age in the low depressive groups (Lusby, Goodman, Yeung, Bell, & Stowe, 2016). In a GUSTO cohort study, Soe et al. (2016) studied the effects of prenatal and postnatal depression, using EEG measures. They concluded that neither prenatal nor postnatal maternal depressive symptoms independently predicted neither frontal EEG activity nor FC in 6- and 18-month-old infants. However, higher levels of depressive symptoms postnatally, compared to the prenatal levels, were associated with greater right frontal activity and relative right frontal asymmetry at 6 months and with lower right frontal FC in 18-month-olds. Lower bilateral frontal FC at 18 months predicted higher externalizing problems, while lower right frontal FC predicted higher internalizing problems.



Some studies focused on the effects of maternal anxiety during pregnancy on infant auditory attention and measured offspring ERPs in reaction to auditory stimuli (Harvison, Molfese, Woodruff-Borden, & Weigel, 2009; Hunter et al., 2012; Otte et al., 2015; van den Heuvel, Donkers, Winkler, Otte, & Van den Bergh, 2015). Auditory attention is a key aspect of early neurocognitive functioning and is vital for later acquisition of speech and language competences (Benasich et al., 2006; Kushnerenko, Van den Bergh, & Winkler, 2013; Molfese, 2000). In the study of Harvison et al. (2009) subject recruitment occurred in the maternity ward, and they completed a self-report anxiety questionnaire. Voice recordings of the neonate's mother and of a stranger were used as auditory stimuli. Results indicated alterations in auditory attention, with more attention allocated to a stranger's voice compared to the mother's voice in infants born to mothers with high anxiety and the opposite pattern in infants of mothers who were low anxious during pregnancy. Hunter et al. (2012) reported diminished P50 response to auditory clicking in 3-month-old infants born to mothers diagnosed with anxiety disorder in pregnancy; this diminished response may reflect lower response inhibition during sensory gating.

In a cohort born in the Netherlands between 2010 and 2013 (van den Heuvel et al., 2015), pregnant women completed self-report questionnaires in each pregnancy trimester (i.e., <15 gw, at 16–22 gw, and at 31–37 gw). It was shown that 9-month-old infants exposed to mothers with high versus low levels of maternal anxiety at 15–22 gw allocated more attentional resources (i.e., higher N250) to a frequently occurring standard sounds in an oddball paradigm; this may indicate a lack of habituation to these sounds or a state of enhanced vigilance in infants born to highly anxious mothers. The opposite pattern (i.e., lower N250) was found for prenatal exposure to maternal mindfulness. Otte et al. (2015) studied the same 9-month-old infants; infants were presented with emotional facial expressions (happy/fearful) followed by either a congruent or an incongruent vocalization (happy/fearful). Results indicated that infants prenatally exposed to higher levels of maternal anxiety (i.e., < gw 15) displayed larger P350 amplitudes in response to fearful vocalizations, regardless of the type of visual prime. This response may indicate increased attention (or enhanced vigilance) to fearful vocalizations. When the children of this cohort were 4 years old, they were again examined; ERPs were recorded when they passively watched neutral, pleasant, and unpleasant pictures (van den Heuvel, Henrichs, Donkers, & Van den Bergh, *in press*). It was found that compared to children exposed to lower levels of maternal anxiety during pregnancy, children exposed to higher level of maternal anxiety devoted more attentional resources to neutral pictures. Responding to neutral stimuli as if they are threatening may indicate a negativity bias and/or may indicate that these children show enhanced vigilance. A state of enhanced vigilance in a safe environment may be a predictive marker for later onset of an anxiety disorder (Thayer, Åhs, Fredrikson, Sollers, & Wager, 2012).

### *Results in school-age children (Ages: 6–10 years)*

In this age group only studies measuring an effect on structural brain changes were performed. Pregnant women completed self-report questionnaires at 19, 25, and 31 gw. The offspring, born between 1998 and 2002 in Southern California, were followed up until 6 to 9 years after birth. Significant associations between maternal anxiety, cortisol or depression during pregnancy and offspring structural brain changes were reported in three publications. Buss, Davis, Muftuler, Head, and Sandman (2010) reported an association between pregnancy-specific anxiety at 19 gw and decreased GM volume in areas extending from the cortical to the occipital regions (i.e., prefrontal cortex, premotor cortex, medial temporal lobe, lateral temporal cortex, postcentral gyrus, cerebellum extending, middle occipital gyrus, and fusiform gyrus). Maternal cortisol at 15 gw was associated with increased right amygdala volume in girls only; amygdala volume even mediated the association between maternal cortisol and girls' emotional problems (Buss et al., 2012). However, maternal cortisol in pregnancy was not associated with child hippocampus volume (Buss et al., 2012). Anxiety measured at 25 or 31 gw or cortisol measured at 19, 21, 31, and 37 gw were not associated with structural brain changes (Buss et al., 2010, 2012). Sandman, Buss, Head, and Davis (2015) observed that maternal reports of depressive symptoms were associated with cortical thinning, in prefrontal, medial postcentral, lateral ventral precentral, and postcentral regions of the right hemisphere; the association of depressive symptoms at 25 gw were the strongest. Moreover, cortical thinning in prefrontal areas of the right hemisphere mediated the association between maternal depression and child externalizing behavior. The observed pattern of cortical thinning seemed to be similar to patterns in children, adolescents, and depressed patients (Sandman et al., 2015). According to the authors, the observed cortical thinning in children born to mothers with higher depressive symptoms during pregnancy may reflect accelerated brain maturation (Sandman et al., 2015, p. 331).

Sarkar et al. (2014) performed dMRI in preschoolers of women who underwent amniocentesis in a London clinic. Retrospectively assessed self-report of stress during pregnancy (measured at 17 months after birth) was associated with increased FA in the right uncinate fasciculus and decreased radial diffusivity in the right uncinate fasciculus, in 6- to 9-year-old children. These changes in WM microstructure were suggested to reflect hypermyelination in the uncinate fasciculus, which links the limbic region with the prefrontal cortex. Self-reported stress was not associated with control tract properties.

In the study by Davis, Sandman, Buss, Wing, and Head (2013), the effect of prenatal exposure to betamethasone (a synthetic glucocorticoid for fetal lung maturation, prescribed to women at risk for preterm delivery) administered between 24 and 34 gw (two doses of 12 mg, intramuscularly, 24 h apart) was examined. Only children born >37 gw were in-

cluded. Fetal glucocorticoid exposure was associated with bilateral cortical thinning in several regions (left insula, left supramarginal gyrus, left transverse temporal cortex such as the cingulate cortex, frontal regions, and the superior parietal cortex) as well as with unilateral thinning in several regions. The rostral anterior cingulate cortex, which is important for regulation of stress and emotions, showed the largest group differences; that is, it was 30% thinner in the exposed group. Furthermore, in the control group children with more affective problems had a thinner left rostral anterior cingulate cortex. The authors conclude that prenatal GC exposure alters the trajectory of fetal brain development, which may be a risk factor for the development of mental health problems

#### *Results in adolescents and adults (Ages: 15–40 years)*

In a Belgium study that recruited pregnant women in 1986 and 1987 in a university hospital, and in which self-report questionnaires were completed at 12–22 gw, 23–31 gw, and 32–40 gw, male offspring was examined at 17 and 20 years of age. It was found that maternal anxiety at 12–22 gw was positively associated with less efficient endogenous decision making, as reflected in EEG measures, at age 17 (Mennes, Van den Bergh, Lagae, & Stiers, 2009). As fMRI may have an important role in understanding pathophysiologic processes by analyzing the brain areas activated while performing specific tasks, a task-related fMRI study was conducted in the 20-year-olds. Results indicated that in contrast to offspring exposed to low or medium levels of maternal anxiety during pregnancy, in the offspring exposed to high maternal anxiety in pregnancy, a number of right lateralized clusters, including the inferior frontal junction, were not modulated during endogenous cognitive control tasks. Moreover, in this cohort, prenatal exposure to maternal anxiety at 12–22 gw also was found to be associated with increased perception of dyspnea (i.e., shortness of breath or breathlessness) at age 28 (von Leupoldt et al., 2017), which may indicate increased sensitivity of the autonomous nervous system.

Favaro, Tenconi, Degortes, Manara, and Santonastaso (2015) examined effects of maternal life events stress during pregnancy in healthy volunteers, all woman, between the age of 15 and 44. Stress experienced during pregnancy was retrospectively measured with a semistructured interview administered to the mothers of the participants. Greater maternal life event stress was associated with decreased GM volume in left medial temporal lobe (MTL) and both amygdalae, but not total volume of amygdala nor with GM or total volume of the hippocampus. Furthermore, life event stress during pregnancy was positively correlated with FC of the left medial temporal lobe and with the pregenual anterior cingulate cortex, which is connected to the default mode network (Qin et al., 2016; Teipel et al., 2010). FC between the left MTL and part of the left medial orbitofrontal cortex partially explained variance in offspring depressive symptoms. All results remained the same when only data from subjects over the age of 18 were analyzed.

#### *Conclusion: Prenatal Stress Alters Brain Developmental Trajectories*

We reviewed prospective longitudinal studies examining whether prenatal exposure to maternal distress is associated with offspring brain development as measured with different brain imaging techniques, from birth until adulthood. Most studies used self-report distress questionnaires as predictors, while other studies used a physician's diagnosis based on medical chart information (Scheinost, Kwon, et al., 2016), a diagnostic psychiatric instrument (Field et al., 2010; Hunter et al., 2012; Lusby et al., 2014, 2016), or a physiological variable (Buss et al., 2012; Davis et al., 2013) as predictor. Among different MRI-based techniques, sMRI such as voxel-based morphometry was used to quantify GM and WM volumes, while dMRI detects alterations in WM structure and indirectly in the architecture of fiber pathways. Finally, fMRI investigates brain activations in specific brain regions during cognitive and sensory tasks, and in FC when at rest, while task-related ERPs and rs-EEG records the electrical brain activity with superior temporal but lower spatial resolution compared to fMRI.

Structural brain alterations in the aftermath of prenatal stress are observed in the neonate and until adulthood. In the neonate, maternal distress is associated with changes in the WM microstructure in the amygdala, volume changes in amygdalae and hippocampus, which were influenced by variations in offspring *BDNF* gene and its methylation patterns (Chen et al., 2015; Qiu et al., 2013; Rifkin-Graboi et al., 2013), in WM microstructural changes and cortical thickness changes in prefrontal and parietal regions and corticolimbic structures of which some were influenced by variations in offspring *COMT* and offspring epigenome (Qiu, Tuan, et al., 2015; Rifkin-Graboi et al., 2015). Next to inducing atypical amygdala–prefrontal connectivity (Posner et al., 2016), reduced hippocampal growth effects in infants (Qiu et al., 2013), and larger right amygdalar volume in female preschoolers (Wen et al., 2017), maternal distress was also related to microstructural WM changes, that is, lower diffusivity in frontal and temporal regions in preschoolers (Lebel et al., 2016), and in the limbic–prefrontal region in childhood (Sarkar et al., 2014). GM volume reduction was seen in several cortical areas and in the cerebellum in childhood (Buss et al., 2010; El Marroun et al., 2016; Sandman et al., 2015), and in the left MTL and both amygdalae in adulthood (Favaro et al., 2015). Larger cortical surface area in a frontal region and gyrification changes (becoming insignificant after corrections for covariates) were also observed (El Marroun et al., 2016). Furthermore, maternal cortisol in pregnancy was associated with larger amygdala volume in children (Buss et al., 2012) while betamethasone intake was related to bilateral cortical thinning (Davis et al., 2013).

With regard to the functional measures, rs-EEG studies point to association of maternal distress in pregnancy to greater relative right frontal EEG activation, which, however, is in some studies only seen in infants of mothers with high postnatal depressive symptoms or with greater postnatal

than prenatal symptoms (Field et al., 2010; Lusby et al., 2014, p. 584; Lusby et al., 2016; Soe et al., 2016). Rs-fMRI studies show that prenatal exposure to maternal distress amplifies the decreased amygdala–thalamic FC seen in preterm born neonates (Scheinost, Kwon, et al., 2016) and increased FC of amygdala with several cortical and subcortical regions (Chen et al., 2015) in 6-month-olds, while rs-fMRI studies conducted in adulthood reveal changes in FC in large-scale intrinsic networks such as DMN (Favaro et al., 2015). Task-related EEG (ERPs) in infants show that prenatal exposure to maternal distress has an effect on specific ERP components indicating altered auditory attention and processing (Harvison et al., 2009) diminished inhibition during sensory gating (Hunter et al., 2012), enhanced vigilance to fearful vocalization (Otte et al., 2015), to standard sounds (van den Heuvel et al., 2015), and to neutral pictures (van den Heuvel et al., in press); they reveal specific deficits, such as in endogenous cognitive control in male adolescents (Mennes et al., 2009) an effect that moreover is confirmed by task-related fMRI measures (Mennes, Van den Bergh, Sunaert, Lagae, & Stiers, 2016) in early adulthood.

The functional, (micro)structural, and connectivity changes described in this section are seen as potentially mediating the link between maternal distress in pregnancy and offspring cognitive, behavioral, and emotional problems, that is, as markers of prenatal stress, increasing the risk for mental health problems. Some of the studies reviewed also examined the association of altered brain development with offspring behavior, that is, with an increase in fetal heart rate to a mild in utero perturbation (Posner et al., 2016; retrospective observation), with internalizing and/or externalizing problems (Lusby et al., 2016; Rifkin-Graboi et al., 2015; Sandman et al., 2015; Soe et al., 2016), with affective problems (Buss et al., 2012; Davis et al., 2013), and with cognition (Harvison et al., 2009; Hunter et al., 2012; Mennes et al., 2009, 2016; Otte et al., 2015; van den Heuvel et al., 2015, in press). With regard to timing effects, no firm conclusions can be reached because not all studies examined maternal emotions in the first, second, and/or third trimester. As indicated above, Figure 2 depicts the possible timing effects of the studies reviewed. For instance, studies showing an effect of exposure to maternal distress during the “ballooning phase” (gw 3 to 15) most probably exert their effect by inducing alterations in the epigenetic gene regulatory network of genes coding for proteins and of noncoding RNAs, involved in proliferation and migration of neurons in a spatiotemporal specific way. If exposure to maternal distress after gw 15 may lead to changes in epigenetic gene regulatory networks, it may affect all ongoing processes involved in building of the GM and WM: growth of neuronal dendrites and axons; forming of synapses; production and expansion of astrocytes, oligodendrocytes, and microglial cells; and development of the vascular system and formation of the layers in the cortex, as well as in the gyrification processes. These alternations influence the building of structural and functional networks, starting to be formed around middle gestation. For instance, Tho-

mason et al. (2017) using in utero rs-fMRI have shown that system-level FC was reduced in fetuses that would subsequently be born preterm. Effects of maternal distress in the early postnatal period likely influences ongoing WM growth and pruning of synapses in the cortex (GM); it is not known whether an infant whose brain was altered during prenatal development is more susceptible to negative environmental exposure, but most probably this is the case (Lebel et al., 2016). Several of the studies reviewed in this section started from the hypothesis that the hypothalamic–pituitary–adrenal axis is a most important underlying mechanism mediating the effects of prenatal stress exposure (Lupien et al., 2011), and accordingly they examined the offspring limbic system (including the amygdala, hippocampus, parahippocampus, and parts of the prefrontal cortex). Most of these studies showed an effect on the limbic system (albeit the effect shown was different across the studies), but in studies that used whole-brain analyses, other brain regions were shown to be affected as well. The observation that prenatal stress leads to alterations is a most important one, as altered FC in brain networks has been increasingly recognized as a marker of neurodevelopmental and psychiatric disorders (e.g., Cao, Wang, & He, 2015; van Essen & Barch, 2015).

While some studies found effects only in females (Buss et al., 2012; Wen et al., 2017), because few studies explicitly examined sex differences, no firm conclusions can be reached at this point. In both sexes effects of prenatal stress on brain development are found, but the effects may be different. However, the origins of these differences are not clear. Scheinost, Sinha, et al. (2016) point to sex-dependent differences in placental function, epigenetic mechanisms, responses of the transcriptome, processes mediating neuron–glial interactions, and differential responses in specific regions of the brain.

### Putative Prenatal Origins of Developmental and Psychiatric Disorders

Neurodevelopmental disorders have a strong genetic bias, but in recent years causal environmental factors have been identified. Genetic, epigenetic, and prenatal environmental exposure factors may interact during brain development vulnerability windows and lead to disorders in ways that are not yet clearly understood (Davis et al., 2016; Ghiani & Faundez, 2017; Van den Bergh et al., 2017). While basic and translation research that focused on task-related brain activation has increased our understanding of underlying neural substrates of particular cognitive, emotional, and social behavior and how their alternations underlie psychopathology, the focus has been shifted to the study of neural circuits rather than on specific brain regions (Barch & Carter, 2016). While disrupted or disconnected functional and structural brain networks have been found to underlie many known neurodevelopmental disorders (van Essen & Barch, 2015), much knowledge still needs to be gathered on what their biological underpinnings are and how they are related to the brain developmental trajectory. In animal models, causal links were

shown between prenatal stress exposure and alteration in neurogenesis, myelination, synaptic branching, lamination, and gyrification and in alterations in the excitatory/inhibitory (E/I) balance (see, e.g., Ben-Ari, 2008; Bock, Rether, Gröger, Xie, & Braun, 2014; Bock et al., 2015; Braun et al., 2017; Lussier & Stevens, 2016). All these processes are vital in building optimal functional and structural networks. Preclinical studies have also shown that compensatory mechanisms, for insults to which animals were exposed prenatally, take place at several levels. Such mechanisms may explain part of the phenotypical heterogeneity seen in many developmental disorders. Footprints of the early insult and/or of the way the brain responded may be “visible” in the brain, and may foreshadow the later emergence of neurological or psychiatric problems, long before symptoms are observed (Ben-Ari, 2008). In humans, “We lack, however, detailed information as to the electrical and/or morphological properties of misplaced neuronal ensembles and their connections to comprehend how they affect brain development” (Ben-Ari, 2008, p. 632).

Where prenatal origins of neurodevelopmental and psychiatric disorders were previously derived from careful examinations of postmortem brain cortical and subcortical brain tissue, in recent research these analyses are combined with advanced brain imaging techniques that enable to study (micro)structural changes in WM (e.g., Deoni et al., 2014). We describe some hypotheses that are being tested, hereby acknowledging the many considerations that researchers take into account when making inferences about putative prenatal origins (see, e.g., Deoni et al., 2014; Kostović, Judaš, & Sedmak, 2011).

In ASD research, cortical abnormalities in the mini-columnar organization of the cortex, in connectivity patterns, in density of specific types of neurons in specific layers of the cortex, and in the E/I imbalance are observed (e.g., Benes, Vincent, & Todtenkopf, 2001; Donovan & Basson, 2017; Karst & Hutsler, 2016; Kostović et al., 2014; Kostović, Sedmak, Vukšić, & Judaš, 2015) as well as changes in noncoding RNAs modulating gene expression/misregulation of genes expressed during early brain development (e.g., nSR100/SRRM4; Irimia et al., 2014; Ziats et al., 2015). These neurobiological changes may underlie changes in FC (Khan et al., 2013) and SC (Conti et al., 2017) observed in ASD, that is, reduced long-distance connectivity and local overconnectivity in frontotemporal and basal ganglia. In the same vein in *schizophrenia research*, abnormalities most probably originating in the subplate (a transient fetal human brain structure) have been detected (Suárez-Solá et al., 2009). In another study, a higher polygenic risk score for schizophrenia was significantly associated with a lower local gyrification index in the bilateral inferior parietal lobes (Erk et al., 2017). Furthermore, research over the last 10 years that combines EEG/magnetoencephalography in clinical populations with preclinical research has led to the conceptualization of schizophrenia as a disorder of WM (Haroutunian et al., 2014) associated with aberrant neural dynamics and disturbances in E/I

balance (Czepielewski, Wang, Gama, & Barch, 2017; Davis et al., 2016; Rutkowski et al., 2017; Wen, 2017).

The putative fetal origins of ADHD described earlier (Van den Bergh & Marcoen, 2004) referred to disturbed processes of proliferation, migration, and differentiation of neurons, which changes the timetable of expression of neurotransmitters phenotype and of neuropeptides, leading to changes in the E/I balance. Recent studies show the relevance of the study of common genetic networks underlying several neurodevelopmental disorders, for example, symptoms in ADHD, externalizing behaviors, and substance-use disorder (Arcos-Burgos, Vélez, Solomon, & Muenke, 2012). Some of the genes involved have a role in axon guidance during brain development (e.g., LPHN3, coding for latrophilin, a cell adhesion molecule; Seiradake, Jones, & Klein, 2016). Furthermore, a significant interaction was found between four LPHN3 tag single nucleotide polymorphisms and maternal stress in pregnancy; it was associated with ADHD and behavioral and cognitive dimensions related to ADHD (Choudhry et al., 2012). Alterations in rs-FC are seen, that is, in DMN, attention (dorsal, ventral, and salience), frontoparietal networks, and reward-related and amygdala-related circuits (for a review, see, e.g., Castellanos & Aoki, 2016). Recent whole-brain connectivity studies observed a distributed pattern of disrupted WM microstructural integrity separately involving frontal, striatal, and cerebellar brain regions (Hong et al., 2014) and involving the prefrontal cortex, insula, occipital, and somatosensory areas (Francx et al., 2016); these changes were associated with measures of attentional performances (Hong et al., 2014) and symptom severity (Francx et al., 2016).

Finally, fetal origins of dyslexia have long been hypothesized (e.g., Behan & Geschwind, 1985). Recent surface-based imaging techniques (i.e., sMRI to assess whole-brain vertex-wise cortical and local gyrification index) showed that compared to controls, 6- to 15-year-old children with dyslexia show decreased cortical thickness in previously identified reading areas (including bilateral occipitotemporal and occipitoparietal regions) as well as increased gyrification within the left occipitotemporal and right superior frontal cortex (Williams, Juranek, Cirino, & Fletcher, 2017). These observations may lend support to the fetal origins of dyslexia hypothesis as primary gyral patterns are largely determined prior to birth (Budday et al., 2015). However, the authors discuss the question whether the observed anatomical differences are due to aberrant structural development or are secondary changes from an impoverished reading experience, and they conclude that their findings of atypical gyrification in ventral reading areas tend to support a neurodevelopmental vulnerability in developmental dyslexia, whereas a regionally thinner cortex in dyslexia may be associated with either a developmental etiology or an impoverished reading experience (Williams et al., 2017, p. 8). The review of Mascheretti et al. (2017) included studies examining intermediate phenotypes, as provided by imaging data, as a target for researching disease-associated genetic variants, of which several have specific roles during brain development.



## Conclusion

Our review highlights the vulnerability of the brain to insults, during at least the prenatal and early postnatal periods of its protracted development. Studies using diverse brain imaging techniques have clearly revealed that maternal distress during pregnancy is associated with alterations in brain structure and function, in FC, and in intrinsic brain networks. Prospective longitudinal follow-up studies reviewed indicate that influences are seen until at least age 29, while retrospective studies showed effects until age 40. Results revealed by the different techniques were not always consistent. Although this inconsistency is partly related to the fact that each method has different possibilities and constrains and that studies differ in the model and postprocessing techniques used, it remains a concern for future research. Our review made it also clear that the influence of prenatal exposure to maternal anxiety needs to be seen in the broader framework of the genetic–epigenetic–environmental processing governing or guiding brain development over a protracted period of life. On the one hand, maternal distress during pregnancy is but one of the factors having an influence on the developmental trajectory of the brain, but on the other hand, it plays an important factor as it is potentially malleable, which clearly bears important clinical and societal responsibility toward expectant parents and their children.

### *Prenatal stress and the developing brain: Emerging topics*

Highlighting the vulnerabilities of the brain to insults may provide information on mechanisms linking maternal distress in pregnancy with symptoms of disturbed cognitive, emotional, and social behavior, and enhanced susceptibility to neurodevelopmental and psychiatric disorders. The existing evidence is inconclusive; there are still many questions unanswered, for example, about potential differential effects of anxiety, depression, and subjective or objective stress and about how they may be moderated by prepregnancy factors, paternal factors and maternal distress factors in the postnatal period. Specific timing effects and sex-specific effects are understudied (Brummelte, 2017; Van den Bergh et al., 2017). In order to validate the measures used with regard to sensitivity, specificity, accuracy, or robustness in predicting neurodevelopmental and psychiatric disorders, more prospective long-term longitudinal follow-up studies starting early in pregnancy and examining brain developmental measures as potential mediators of the link between prenatal stress and offspring behavioral and cognitive measures should be carried out. Brain imaging measures could be used as an intermediate phenotype. Combining existing tools, that is, using structural and functional measures as done in the study by Soe et al. (2016) or integrating EEG, fMRI, and behavioral measures to analyze them jointly (Turner, Forstmann, Love, Palmeri, & van Maanen, 2017; Turner, Rodriguez, Norcia, McClure, & Steyvers, 2016), will not only enhance our insight in the relation between task- and rs-MRI connectivity

measures but also in bidirectional brain–behavior interactions and their evolution over time (Johnson, Jones, & Gliga, 2015). Studying the same cohort across different ages, using measures of functional brain changes and neurocognitive function (e.g., Mennes, Stiers, Lagae, & Van den Bergh, 2006; Mennes et al., 2009, 2016; Van den Bergh et al., 2005, 2006), may reveal whether the observed cognitive phenotype is consistent over time or not, and what the brain correlates are. The methods described—involving systems biology, systems neuroscience, and gene network analysis—are a big challenge (Irimia et al., 2014; Mascheretti et al., 2017; Parikshak et al., 2015). Introducing them in human prenatal stress research and combining them with neuroimaging techniques will, when combined with the promotion of common strategies of acquiring and sharing high-quality neuroimaging, accelerate progress in characterizing typical and atypical FC and SC (van Essen & Barch, 2015). The human prenatal stress field and the broader field of developmental origins of health and disease (DOHaD) research are interdisciplinary research fields that will benefit from collaborations between system biology and neuroscience, psychology, network science, mathematicians, and computer scientists in trying to understand how prenatal exposure influences the nervous system at different levels of interaction of structures and functions (Swanson & Lichtman, 2016). At the same time, “it is also vital to be realistic about the limits that are attainable using current technologies” (van Essen & Barch, 2015, p. 165). For instance, while fetal imaging has already been used for a long time to show effects of maternal distress on fetal motor behavior and fetal behavioral states (e.g., by using ultrasound to measure fetal general movements and eye movements, and electrocardiography to measure fetal heart variability in a randomized controlled trial study; Van den Bergh et al., 1989), the exciting possibility to use imaging techniques to directly study fetal brain development in distressed versus nondistressed mothers is now actively being explored (Koyama et al., 2016; Scheinost, Sinha, et al., 2016; van den Heuvel & Thomason, 2016). Imaging the fetal brain is still a challenging task, especially in the young fetus as young fetuses move more than older fetuses. It requires extremely fast acquisition techniques that deal with motion and other artifacts in sMRI (Habas et al., 2010; Tourbier et al., 2017; Wright et al., 2014, 2015) in fMRI (Jakab et al., 2015), and in dMRI (Marami et al., 2017). The imaged slices require interslice motion correction that rearranges each slice (Habas et al., 2010; Oubel, Koob, Studholme, Dietemann, & Rousseau, 2010) and allows the reconstruction of super resolution images if multiple slice directions were imaged (Alansary et al., 2017; Tourbier et al., 2017; Wright et al., 2014). Besides image reconstruction, data preprocessing requires adapted parameters and modified templates that handle the large structural changes that occur during gestation (Wright et al., 2014, 2015). Deleting fetal imaging data with too much motion artifacts may be necessary to reach an acceptable signal–noise ratio. Researchers may also opt to analyze only brain imaging data obtained during fetal quiescence (van den Heuvel & Thomason, 2016).

Furthermore, a critical step for future research is a better characterization of maternal distress. While most studies examine the effect of either depression or anxiety, several authors propose to approach them in an integrative way during both, the pregnancy and postpartum, that is, in the peripartum period (e.g., Babb, Deligiannidis, Murgatroyd, & Nephew, 2015; Lusby et al., 2016; Wen et al., 2017). Finally, because it has been shown that next to the brain, several other biological systems are involved in mediating the effect of prenatal stress on the offspring, future studies in the prenatal stress field should include other relevant potential biomarkers, that is, neuroendocrine, inflammation, telomere length, gut–brain interaction, and autonomous nervous systems biomarkers (for a review, see Rakers et al., 2017; Van den Bergh et al., 2017). Intervention studies are needed to identify predictive markers of risk, resilience, and prognosis of neurodevelopmental disorders.

### *Prenatal stress: Societal and clinical challenges*

Neurodevelopmental and psychiatric disorders constitute a major burden to society (Falk et al., 2016). The prenatal life period is increasingly considered as a crucial target for the primary prevention of neurodevelopmental and psychiatric disorders. Next to contributing to basic science, results of studies on prenatal origins of cognitive, emotional, and social dysfunction have crucial public health implications as they are dealing with prenatal environmental risk factors that potentially can be modified (Howard, Piot, & Stein, 2014). For instance, individual, family, or group-based therapy les-

sens their distress levels, which in some studies has been shown to be beneficial for the offspring in infancy (e.g., Howard, Molyneaux, et al., 2014; Milgrom et al., 2015). The potential social and economic returns on investment are substantial, as 15% of pregnant women experience a depression or anxiety disorder (Bauer, Parsonage, Knapp, Iemmi, & Adelaja, 2014) and up to 30% experience high stress during pregnancy (Loomans et al., 2013), which may be related, for instance, to job strain, marital problems, death of a child, or death of another relative. In a recent UK report, the estimated costs, related to three major perinatal mental health conditions (i.e., depression, anxiety, and psychosis), are equivalent to about £10,000 for every singly birth in the country; 72% of these costs were related to adverse impacts on the child rather than the mother. These costs indicate the potential benefits of intervention in the perinatal life period. The costs of extra provision to bring perinatal mental health care up to the level of standards recommended in national guidance was estimated at £400 per average birth (Bauer et al., 2014). The authors argue that “even a relatively modest improvement in outcome as a result of better services would be sufficient to justify the additional spending on value for money grounds” (Bauer et al., 2014, p. 5). Knowledge that is gathered by prenatal stress research provides input to make recommendations to public health policymakers and health professionals. Yet, to understand who to target and how to design effective interventions that tackle effects of maternal stress/anxiety/depression in pregnancy, much more knowledge is needed about timing effects, mechanisms, and moderating factors.

## References

- Alansary, A., Rajchl, M., McDonagh, S. G., Murgasova, M., Damodaram, M., Lloyd, D. F., . . . Kainz, B. (2017). PVR: Patch-to-volume reconstruction for large area motion correction of fetal MRI. *IEEE Transactions on Medical Imaging*, *36*. doi:10.1109/TMI.2017.2737081
- American Psychiatric Association. (2013). *Diagnostic and statistical manual of mental disorders* (5th ed.). Washington, DC: Author.
- Arcos-Burgos, M., Vélez, J. I., Solomon, B. D., & Muenke, M. (2012). A common genetic network underlies substance use disorders and disruptive or externalizing disorders. *Human Genetics*, *131*, 917–929. doi:10.1007/s00439-012-1164-4
- Ashburner, J., & Friston, K. J. (2000). Voxel-based morphometry—The methods. *NeuroImage*, *11*(6, Pt. 1), 805–821. doi:10.1006/nimg.2000.0582
- Ashburner, J., & Friston, K. J. (2011). Diffeomorphic registration using geodesic shooting and Gauss-Newton optimisation. *NeuroImage*, *55*, 954–967. doi:10.1016/j.neuroimage.2010.12.049
- Atasoy, S., Donnelly, I., & Pearson, J. (2016). Human brain networks function in connectome-specific harmonic waves. *Nature Communications*, *7*, 10340. doi:10.1038/ncomms10340
- Avants, B. B., Tustison, N. J., Song, G., Cook, P. A., Klein, A., & Gee, J. C. (2011). A reproducible evaluation of ANTs similarity metric performance in brain image registration. *NeuroImage*, *54*, 2033–2044. doi:10.1016/j.neuroimage.2010.09.025
- Babb, J. A., Deligiannidis, K. M., Murgatroyd, C. A., & Nephew, B. C. (2015). Peripartum depression and anxiety as an integrative cross domain target for psychiatric preventative measures. *Behavioural Brain Research*, *276*, 32–44. doi:10.1016/j.bbr.2014.03.039
- Ball, G., Aljabar, P., Zebari, S., Tumor, N., Arichi, T., Merchant, N., . . . Counsell, S. J. (2014). Rich-club organization of the newborn human brain. *Proceedings of the National Academy of Sciences*, *111*, 7456–7461. doi:10.1073/pnas.1324118111
- Barch, D. M., & Carter, C. S. (2016). Functional and structural brain connectivity in psychopathology. *Biological Psychiatry: Cognitive Neuroscience and Neuroimaging*, *1*, 196–198. doi:10.1016/j.bpsc.2016.03.006
- Barker, D. J. (1990). The fetal and infant origins of adult disease. *British Medical Journal*, *301*, 1111.
- Batalle, D., Hughes, E. J., Zhang, H., Tournier, J. D., Tumor, N., Aljabar, P., . . . Counsell, S. J. (2017). Early development of structural networks and the impact of prematurity on brain connectivity. *NeuroImage*, *149*, 379–392. doi:10.1016/j.neuroimage.2017.01.065
- Bauer, A., Parsonage, M., Knapp, M., Iemmi, V., & Adelaja, B. (2014). *Costs of perinatal mental health problems* London: London School of Economics.
- Bayly, P. V., Taber, L. A., & Kroenke, C. D. (2014). Mechanical forces in cerebral cortical folding: A review of measurements and models. *Journal of the Mechanical Behavior of Biomedical Materials*, *29*, 568–581. doi:10.1016/j.jmbbm.2013.02.018
- Behan, P., & Geschwind, N. (1985). Dyslexia, congenital anomalies, and immune disorders: The role of the fetal environment. *Annals of the New York Academy of Sciences*, *457*, 13–18. doi:10.1111/j.1749-6632.1985.tb20796.x
- Behrens, T. E. J., & Sporns, O. (2012). Human connectomics. *Current Opinion in Neurobiology*, *22*, 144–153. doi:10.1016/j.comb.2011.08.005
- Ben-Ari, Y. (2008). Neuro-archaeology: Pre-symptomatic architecture and signature of neurological disorders. *Trends in Neurosciences*, *31*, 626–636. doi:10.1016/j.tins.2008.09.002
- Ben-Ari, Y., & Spitzer, N. C. (2010). Phenotypic checkpoints regulate neuronal development. *Trends in Neurosciences*, *33*, 485–492. doi:10.1016/j.tins.2010.08.005
- Benasich, A. A., Choudhury, N., Friedman, J. T., Realpe-Bonilla, T., Chojnowska, C., & Gou, Z. (2006). The infant as a prelinguistic model for lan-

- guage learning impairments: Predicting from event-related potentials to behavior. *Neuropsychologia*, *44*, 396–411. doi:10.1016/j.neuropsychologia.2005.06.004
- Benes, F. M., Vincent, S. L., & Todtenkopf, M. (2001). The density of pyramidal and nonpyramidal neurons in anterior cingulate cortex of schizophrenic and bipolar subjects. *Biological Psychiatry*, *50*, 395–406. doi:10.1016/S0006-3223(01)01084-8
- Bernadskaya, Y., & Christiaen, L. (2016). Transcriptional control of developmental cell behaviors. *Annual Review of Cell and Developmental Biology*, *32*, 77–101. doi:10.1146/annurev-cellbio-111315-125218
- Billiet, T., Vandenbulcke, M., Mädler, B., Peeters, R., Dhollander, T., Zhang, H., . . . Emsell, L. (2015). Age-related microstructural differences quantified using myelin water imaging and advanced diffusion MRI. *Neurobiology of Aging*, *36*, 2107–2121. doi:10.1016/j.neurobiolaging.2015.02.029
- Bock, J., Rether, K., Gröger, N., Xie, L., & Braun, K. (2014). Perinatal programming of emotional brain circuits: An integrative view from systems to molecules. *Frontiers in Neuroscience*, *8*, 11. doi:10.3389/fnins.2014.00011
- Bock, J., Wainstock, T., Braun, K., & Segal, M. (2015). Stress in utero: Prenatal programming of brain plasticity and cognition. *Biological Psychiatry*, *78*, 315–326. doi:10.1016/j.biopsych.2015.02.036
- Bowers, M. E., & Yehuda, R. (2016). Intergenerational transmission of stress in humans. *Neuropsychopharmacology*, *41*, 232–244. doi:10.1038/npp.2015.247
- Braun, K., Bock, J., Wainstock, T., Matas, E., Gaisler-Salomon, I., Fegert, J., . . . Segal, M. (2017). Experience-induced transgenerational (re-)programming of neuronal structure and functions: Impact of stress prior and during pregnancy. *Neuroscience & Biobehavioral Reviews*. Advance online publication. doi:10.1016/j.neubiorev.2017.05.021
- Brummelte, S. (2017). Introduction: Early adversity and brain development. *Neuroscience*, *342*, 1–3. doi:10.1016/j.neuroscience.2016.09.041
- Budday, S., Steinmann, P., & Kuhl, E. (2015). Physical biology of human brain development. *Frontiers in Cellular Neuroscience*, *9*. doi:10.3389/fncel.2015.00257
- Buss, C., Davis, E. P., Muftuler, L. T., Head, K., & Sandman, C. A. (2010). High pregnancy anxiety during mid-gestation is associated with decreased gray matter density in 6–9-year-old children. *Psychoneuroendocrinology*, *35*, 141–153. doi:10.1016/j.psyneuen.2009.07.010
- Buss, C., Davis, E. P., Shahbaba, B., Pruessner, J. C., Head, K., & Sandman, C. A. (2012). Maternal cortisol over the course of pregnancy and subsequent child amygdala and hippocampus volumes and affective problems. *Proceedings of the National Academy of Sciences*, *109*, E1312–E1319. doi:10.1073/pnas.1201295109
- Cao, M., Wang, Z., & He, Y. (2015). Connectomics in psychiatric research: Advances and applications. *Neuropsychiatric Disease Treatment*, *11*, 2801–2810.
- Castellanos, F. X., & Aoki, Y. (2016). Intrinsic functional connectivity in attention-deficit/hyperactivity disorder: A science in development. *Biological Psychiatry: Cognitive Neuroscience and Neuroimaging*, *1*, 253–261. doi:10.1016/j.bpsc.2016.03.004
- Castellanos, F. X., & Proal, E. (2012). Large-scale brain systems in ADHD: Beyond the prefrontal–striatal model. *Trends in Cognitive Sciences*, *16*, 17–26. doi:10.1016/j.tics.2011.11.007
- Chang, Y. S., Owen, J. P., Pojman, N. J., Thieu, T. U., Bukshpun, P., Wakahiro, M. L. J., . . . Mukherjee, P. (2015). White matter changes of neurite density and fiber orientation dispersion during human brain maturation. *PLOS ONE*, *10*, 1–23. doi:10.1371/journal.pone.0123656
- Charil, A., Laplante, D. P., Vaillancourt, C., & King, S. (2010). Prenatal stress and brain development. *Brain Research Reviews*, *65*, 56–79. doi:10.1016/j.brainresrev.2010.06.002
- Chen, L., Pan, H., Tuan, T. A., Teh, A. L., MacIsaac, J. L., Mah, S. M., . . . Holbrook, J. D. (2015). Brain-derived neurotrophic factor (BDNF) Val66Met polymorphism influences the association of the methylome with maternal anxiety and neonatal brain volumes. *Development and Psychopathology*, *27*, 137–150. doi:10.1017/S0954579414001357
- Cicchetti, D., & Rogosch, F. A. (1996). Equifinality and multifinality in developmental psychopathology. *Development and Psychopathology*, *8*, 587–600.
- Choudhry, Z., Sengupta, S. M., Grizenko, N., Fortier, M.-E., Thakur, G. A., Bellingham, J., & Jooper, R. (2012). LPHN3 and attention-deficit/hyperactivity disorder: Interaction with maternal stress during pregnancy. *Journal of Child Psychology and Psychiatry*, *53*, 892–902. doi:10.1111/j.1469-7610.2012.02551.x
- Cicchetti, D., Handley, E. D., & Rogosch, F. A. (2015). Child maltreatment, inflammation, and internalizing symptoms: Investigating the roles of C-reactive protein, gene variation, and neuroendocrine regulation. *Development and Psychopathology*, *27*, 553–566. doi:10.1017/S0954579415000152
- Cole, D. M., Smith, S. M., & Beckmann, C. F. (2010). Advances and pitfalls in the analysis and interpretation of resting-state FMRI data. *Frontiers in Systems Neuroscience*, *4*, 8. doi:10.3389/fnsys.2010.00008
- Collin, G., & van den Heuvel, M. P. (2013). The ontogeny of the human connectome: Development and dynamic changes of brain connectivity across the life span. *Neuroscientist*, *19*, 616–628. doi:10.1177/1073858413503712
- Conti, E., Mitra, J., Calderoni, S., Pannek, K., Shen, K. K., Pagnozzi, A., . . . Guzzetta, A. (2017). Network over-connectivity differentiates autism spectrum disorder from other developmental disorders in toddlers: A diffusion MRI study. *Human Brain Mapping*, *38*, 2333–2344. doi:10.1002/hbm.23520
- Czepliewski, L. S., Wang, L., Gama, C. S., & Barch, D. M. (2017). The relationship of intellectual functioning and cognitive performance to brain structure in schizophrenia. *Schizophrenia Bulletin*, *43*, 355–364. doi:10.1093/schbul/sbw090
- Dahnke, R., Yotter, R. A., & Gaser, C. (2013). Cortical thickness and central surface estimation. *NeuroImage*, *65*, 336–348. doi:10.1016/j.neuroimage.2012.09.050
- Davis, E. P., Sandman, C. A., Buss, C., Wing, D. A., & Head, K. (2013). Fetal glucocorticoid exposure is associated with preadolescent brain development. *Biological Psychiatry*, *74*, 647–655. doi:10.1016/j.biopsych.2013.03.009
- Davis, J., Eyre, H., Jacka, F. N., Dodd, S., Dean, O., McEwen, S., . . . Berk, M. (2016). A review of vulnerability and risks for schizophrenia: Beyond the two hit hypothesis. *Neuroscience and Biobehavioral Reviews*, *65*, 185–194. doi:10.1016/j.neubiorev.2016.03.017
- Dawson, G., Frey, K., Panagiotides, H., Osterling, J., & Hessler, D. (1997). Infants of depressed mothers exhibit atypical frontal brain activity: A replication and extension of previous findings. *Journal of Child Psychology and Psychiatry*, *38*, 179–186. doi:10.1111/j.1469-7610.1997.tb01852.x
- Dennis, E. L., & Thompson, P. M. (2013). Typical and atypical brain development: A review of neuroimaging studies. *Dialogues in Clinical Neuroscience*, *15*, 359–382.
- Deoni, S. C. L., Dean, D. C., III, Piryatinsky, I., O’Muircheartaigh, J., Waskiewicz, N., Lehman, K., . . . Dirks, H. (2013). Breastfeeding and early white matter development: A cross-sectional study. *NeuroImage*, *82*, 77–86. doi:10.1016/j.neuroimage.2013.05.090
- Deoni, S. C. L., Zinkstok, J. R., Daly, E., Ecker, C., Williams, S. C. R., & Murphy, D. G. M. (2014). White-matter relaxation time and myelin water fraction differences in young adults with autism. *Psychological Medicine*, *45*, 795–805. doi:10.1017/S0033291714001858
- Di Martino, A., Fair, D. A., Kelly, C., Satterthwaite, T. D., Castellanos, F. X., Thomason, M. E., . . . Milham, M. P. (2014). Unraveling the miswired connectome: A developmental perspective. *Neuron*, *83*, 1335–1353. doi:10.1016/j.neuron.2014.08.050
- Donovan, A. P. A., & Basson, M. A. (2017). The neuroanatomy of autism—A developmental perspective. *Journal of Anatomy*, *230*, 4–15. doi:10.1111/joa.12542
- Doria, V., Beckmann, C. F., Arichi, T., Merchant, N., Groppo, M., Turkheimer, F. E., . . . Edwards, A. D. (2010). Emergence of resting state networks in the preterm human brain. *Proceedings of the National Academy of Sciences*, *107*, 20015–20020. doi:10.1073/pnas.1007921107
- Dubois, J., Dehaene-Lambertz, G., Kulikova, S., Poupon, C., Hüppi, P. S., & Hertz-Pannier, L. (2014). The early development of brain white matter: A review of imaging studies in fetuses, newborns and infants. *Neuroscience*, *276*, 48–71. doi:10.1016/j.neuroscience.2013.12.044
- El Marroun, H., Tiemeier, H., Muetzel, R. L., Thijssen, S., van der Knaap, N. J. F., Jaddoe, V. W. V., . . . White, T. J. H. (2016). Prenatal exposure to maternal and paternal depressive symptoms and brain morphology: A population-based prospective neuroimaging study in young children. *Depression and Anxiety*, *33*, 658–666. doi:10.1002/da.22524
- Erk, S., Mohnke, S., Ripke, S., Lett, T. A., Veer, I. M., Wackerhagen, C., . . . Walter, H. (2017). Functional neuroimaging effects of recently discovered genetic risk loci for schizophrenia and polygenic risk profile in five RDoC subdomains. *Translational Psychiatry*, *7*, e997. doi:10.1038/tp.2016.272
- Evans, A. C., & Brain Development Cooperative Group. (2006). The NIH MRI study of normal brain development. *NeuroImage*, *30*, 184–202. doi:10.1016/j.neuroimage.2005.09.068



- Falk, A., Heine, V. M., Harwood, A. J., Sullivan, P. F., Peitz, M., Brustle, O., . . . Djurovic, S. (2016). Modeling psychiatric disorders: From genomic findings to cellular phenotypes. *Molecular Psychiatry*, *21*, 1167–1179. doi:10.1038/mp.2016.89
- Favaro, A., Tenconi, E., Degortes, D., Manara, R., & Santonastaso, P. (2015). Neural correlates of prenatal stress in young women. *Psychological Medicine*, *45*, 2533–2543. doi:10.1017/S003329171500046X
- Field, T., & Diego, M. (2008). Maternal depression effects on infant frontal EEG asymmetry. *International Journal of Neuroscience*, *118*, 1081–1108. doi:10.1080/00207450701769067
- Field, T., Diego, M., Hernandez-Reif, M., Figueiredo, B., Deeds, O., Ascencio, A., . . . Kuhn, C. (2010). Comorbid depression and anxiety effects on pregnancy and neonatal outcome. *Infant Behavior and Development*, *33*, 23–29. doi:10.1016/j.infbeh.2009.10.004
- Field, T., Hernandez-Reif, M., & Diego, M. (2006). Intrusive and withdrawn depressed mothers and their infants. *Developmental Review*, *26*, 15–30. doi:10.1016/j.dr.2005.04.001
- Fischl, B. R. (2012). FreeSurfer. *NeuroImage*, *62*, 774–781. doi:10.1016/j.neuroimage.2012.01.021
- Franx, W., Llera, A., Mennes, M., Zwiers, M. P., Faraone, S. V., Oosterlaan, J., . . . Beckmann, C. F. (2016). Integrated analysis of gray and white matter alterations in attention-deficit/hyperactivity disorder. *NeuroImage: Clinical*, *11*, 357–367. doi:10.1016/j.nicl.2016.03.005
- Franke, K., Luders, E., May, A., Wilke, M., & Gaser, C. (2012). Brain maturation: Predicting individual BrainAGE in children and adolescents using structural MRI. *NeuroImage*, *63*, 1305–1312. doi:10.1016/j.neuroimage.2012.08.001
- Franke, K., Van den Bergh, B. R. H., de Rooij, S. R., Nathanielsz, P. W., Witte, O. W., Roseboom, T. J., & Schwab, M. (2017). Effects of prenatal stress on structural brain development and aging in humans. *bioRxiv*. doi:10.1101/148916
- Franke, K., Ziegler, G., Klöppel, S., & Gaser, C. (2010). Estimating the age of healthy subjects from T1-weighted MRI scans using kernel methods: Exploring the influence of various parameters. *NeuroImage*, *50*, 883–892. doi:10.1016/j.neuroimage.2010.01.005
- Fransson, P., & Marrelec, G. (2008). The precuneus/posterior cingulate cortex plays a pivotal role in the default mode network: Evidence from a partial correlation network analysis. *NeuroImage*, *42*, 1178–1184. doi:10.1016/j.neuroimage.2008.05.059
- Fransson, P., Skiold, B., Horsch, S., Nordell, A., Blennow, M., Lagercrantz, H., & Aden, U. (2007). Resting-state networks in the infant brain. *Proceedings of the National Academy of Sciences*, *104*, 15531–15536. doi:10.1073/pnas.0704380104
- Gaser, C., Nenadic, I., Buchsbaum, B. R., Hazlett, E. A., & Buchsbaum, M. S. (2001). Deformation-based morphometry and its relation to conventional volumetry of brain lateral ventricles in MRI. *NeuroImage*, *13*(6, Pt. 1), 1140–1145. doi:10.1006/nimg.2001.0771
- Gaser, C., & Schlaug, G. (2003). Brain structures differ between musicians and non-musicians. *Journal of Neuroscience*, *23*, 9240–9245.
- Geschwind, D. H., & Flint, J. (2015). Genetics and genomics of psychiatric disease. *Science*, *349*, 1489–1494. doi:10.1126/science.aaa8954
- Ghiani, C. A., & Faundez, V. (2017). Cellular and molecular mechanisms of neurodevelopmental disorders. *Journal of Neuroscience Research*, *95*, 1093–1096. doi:10.1002/jnr.24041
- Gluckman, P. D., Hanson, M. A., Cooper, C., & Thornburg, K. L. (2008). Effect of in utero and early-life conditions on adult health and disease. *New England Journal of Medicine*, *359*, 61–73. doi:10.1056/NEJMra0708473
- Gupta, K. K., Gupta, V. K., & Shirasaka, T. (2016). An update on fetal alcohol syndrome—Pathogenesis, risks, and treatment. *Alcoholism: Clinical and Experimental Research*, *40*, 1594–1602. doi:10.1111/acer.13135
- Habas, P. A., Kim, K., Rousseau, F., Glenn, O. A., Barkovich, A. J., & Studholme, C. (2010). Atlas-based segmentation of developing tissues in the human brain with quantitative validation in young fetuses. *Human Brain Mapping*, *31*, 1348–1358. doi:10.1002/hbm.20935
- Habes, M., Erus, G., Toledo, J. B., Zhang, T., Bryan, N., Launer, L. J., . . . Davatzikos, C. (2016). White matter hyperintensities and imaging patterns of brain ageing in the general population. *Brain*, *139*(Pt. 4), 1164–1179. doi:10.1093/brain/aww008
- Hagmann, P. (2005). *From diffusion MRI to brain connectomics* (PhD dissertation, Ecole Polytechnique Fédérale de Lausanne).
- Hagmann, P., Cammoun, L., Gigandet, X., Meuli, R., Honey, C. J., Wedeen, V. J., & Sporns, O. (2008). Mapping the structural core of human cerebral cortex. *PLOS Biology*, *6*, e159. doi:10.1371/journal.pbio.0060159
- Hagmann, P., Grant, P., & Fair, D. (2012). MR connectomics: A conceptual framework for studying the developing brain. *Frontiers in Systems Neuroscience*, *6*. doi:10.3389/fnsys.2012.00043
- Han, K., Chapman, S. B., & Krawczyk, D. C. (2016). Disrupted intrinsic connectivity among default, dorsal attention, and frontoparietal control networks in individuals with chronic traumatic brain injury. *Journal of the International Neuropsychological Society*, *22*, 263–279. doi:10.1017/S1355617715001393
- Hanson, M., & Gluckman, P. (2011). Developmental origins of noncommunicable disease: Population and public health implications. *American Journal of Clinical Nutrition*, *94*(Suppl.), 1754S–1758S. doi:10.3945/ajcn.110.001206
- Haroutunian, V., Katsel, P., Roussos, P., Davis, K. L., Altschuler, L. L., & Bartzokis, G. (2014). Myelination, oligodendrocytes, and serious mental illness. *Glia*, *62*, 1856–1877. doi:10.1002/glia.22716
- Harvison, K. W., Molfese, D. L., Woodruff-Borden, J., & Weigel, R. A. (2009). Neonatal auditory evoked responses are related to perinatal band anxiety. *Brain and Cognition*, *71*, 369–374. doi:10.1016/j.bandc.2009.06.004
- Hong, S.-B., Zalesky, A., Fornito, A., Park, S., Yang, Y.-H., Park, M.-H., . . . Kim, J.-W. (2014). Connectomic disturbances in attention-deficit/hyperactivity disorder: A whole-brain tractography analysis. *Biological Psychiatry*, *76*, 656–663. doi:10.1016/j.biopsych.2013.12.013
- Howard, L. M., Molyneux, E., Dennis, C.-L., Rochat, T., Stein, A., & Milgrom, J. (2014). Non-psychotic mental disorders in the perinatal period. *Lancet*, *384*, 1775–1788. doi:10.1016/S0140-6736(14)61276-9
- Howard, L. M., Piot, P., & Stein, A. (2014). No health without perinatal mental health. *Lancet*, *384*, 1723–1724. doi:10.1016/S0140-6736(14)62040-7
- Huang, A. C., Hu, L., Kauffman, S. A., Zhang, W., & Shmulevich, I. (2009). Using cell fate attractors to uncover transcriptional regulation of HL60 neutrophil differentiation. *BMC Systems Biology*, *3*, 20. doi:10.1186/1752-0509-3-20
- Hunter, S. K., Mendoza, J. H., D’Anna, K., Zerbe, G. O., McCarthy, L., Hoffman, C., . . . Ross, R. G. (2012). Antidepressants may mitigate the effects of prenatal maternal anxiety on infant auditory sensory gating. *American Journal of Psychiatry*, *169*, 616–624.
- Hyde, L. W. (2015). Developmental psychopathology in an era of molecular genetics and neuroimaging: A developmental neurogenetics approach. *Development and Psychopathology*, *27*, 587–613. doi:10.1017/S0954579415000188
- Irimia, M., Weatheritt, R. J., Ellis, J., Parikhshak, N. N., Gonatopoulos-Pournatzis, T., Babor, M., . . . Blencowe, B. J. (2014). A highly conserved program of neuronal microexons is misregulated in autistic brains. *Cell*, *159*, 1511–1523. doi:10.1016/j.cell.2014.11.035
- Jakab, A., Kasprian, G., Schwartz, E., Gruber, G. M., Prayer, D., . . . Langs, G. (2015). Disrupted developmental organization of the structural connectome in fetuses with corpus callosum agenesis. *NeuroImage*, *111*, 277–288. doi:10.1016/j.neuroimage.2015.02.038
- Jakab, A., Schwartz, E., Kasprian, G., Gruber, G. M., Prayer, D., Schöpf, V., & Langs, G. (2014). Fetal functional imaging portrays heterogeneous development of emerging human brain networks. *Frontiers in Human Neuroscience*, *8*. doi:10.3389/fnhum.2014.00852
- Jelescu, I. O., Veraart, J., Adisetiyo, V., Milla, S. S., Novikov, D. S., & Fieremans, E. (2015). One diffusion acquisition and different white matter models: How does microstructure change in human early development based on WMTI and NODDI? *NeuroImage*, *107*, 242–256. doi:10.1016/j.neuroimage.2014.12.009
- Jiang, X., & Nardelli, J. (2016). Cellular and molecular introduction to brain development. *Neurobiology of Disease*, *92* (Pt. A), 3–17. doi:10.1016/j.nbd.2015.07.007
- Johnson, M. H., Jones, E. J. H., & Gliga, T. (2015). Brain adaptation and alternative developmental trajectories. *Development and Psychopathology*, *27*, 425–442. doi:10.1017/S0954579415000073
- Jones, I., Chandra, P. S., Dazzan, P., & Howard, L. M. (2014). Bipolar disorder, affective psychosis, and schizophrenia in pregnancy and the post-partum period. *Lancet*, *384*, 1789–1799. doi:10.1016/S0140-6736(14)61278-2
- Kang, H. J., Kawasawa, Y. I., Cheng, F., Zhu, Y., Xu, X., Li, M., . . . Sestan, N. (2011). Spatio-temporal transcriptome of the human brain. *Nature*, *478*, 483–489.
- Kanwisher, N., McDermott, J., & Chun, M. M. (1997). The fusiform face area: A module in human extrastriate cortex specialized for face perception. *Journal of Neuroscience*, *17*, 4302–4311.



- Karst, A. T., & Hutsler, J. J. (2016). Two-dimensional analysis of the supragranular layers in autism spectrum disorder. *Research in Autism Spectrum Disorders*, 32, 96–105. doi:10.1016/j.rasd.2016.09.004
- Khan, S., Gramfort, A., Shetty, N. R., Kitzbichler, M. G., Ganesan, S., Moran, J. M., . . . Kenet, T. (2013). Local and long-range functional connectivity is reduced in concert in autism spectrum disorders. *Proceedings of the National Academy of Sciences*, 110, 3107–3112. doi:10.1073/pnas.1214533110
- Kipping, J. A., Tuan, T. A., Föiiter, M. V., & Qiu, A. (2016). Asynchronous development of cerebellar, cerebello-cortical, and cortico-cortical functional networks in infancy, childhood, and adulthood. *Cerebral Cortex*, 12, 1–15.
- Klein, A., & Tourville, J. (2012). 101 labeled brain images and a consistent human cortical labeling protocol. *Frontiers in Neuroscience*, 6. doi:10.3389/fnins.2012.00171
- Kofink, D., Boks, M. P. M., Timmers, H. T. M., & Kas, M. J. (2013). Epigenetic dynamics in psychiatric disorders: Environmental programming of neurodevelopmental processes. *Neuroscience and Biobehavioral Reviews*, 37, 831–845. doi:10.1016/j.neubiorev.2013.03.020
- Kostović, I., Jovanov-Milošević, N., Radoš, M., Sedmak, G., Benjak, V., Kostović-Srzić, M., . . . Judaš, M. (2014). Perinatal and early postnatal reorganization of the subplate and related cellular compartments in the human cerebral wall as revealed by histological and MRI approaches. *Brain Structure and Function*, 219, 231–253. doi:10.1007/s00429-012-0496-0
- Kostović, I., Judaš, M., & Sedmak, G. (2011). Developmental history of the subplate zone, subplate neurons and interstitial white matter neurons: Relevance for schizophrenia. *International Journal of Developmental Neuroscience*, 29, 193–205. doi:10.1016/j.ijdevneu.2010.09.005
- Kostović, I., Sedmak, G., Vukšić, M., & Judaš, M. (2015). The relevance of human fetal subplate zone for developmental neuropathology of neuronal migration disorders and cortical dysplasia. *CNS Neuroscience and Therapeutics*, 21, 74–82. doi:10.1111/cns.12333
- Koyama, M. S., Di Martino, A., Castellanos, F. X., Ho, E. J., Marcelle, E., Leventhal, B., & Milham, M. P. (2016). Imaging the “at-risk” brain: Future directions. *Journal of the International Neuropsychological Society*, 22, 164–179. doi:10.1017/S1355617715001356
- Kunz, N., Zhang, H., Vasung, L., O’Brien, K. R., Assaf, Y., Lazeyras, F., . . . Hüppi, P. S. (2014). Assessing white matter microstructure of the newborn with multi-shell diffusion MRI and biophysical compartment models. *NeuroImage*, 96, 288–299. doi:10.1016/j.neuroimage.2014.03.057
- Kushnerenko, E. V., Van den Bergh, B. R. H., & Winkler, I. (2013). Separating acoustic deviance from novelty during the first year of life: A review of event-related potential evidence. *Frontiers in Psychology*, 4, 595. doi:10.3389/fpsyg.2013.00595
- Lebel, C., Gee, M., Camicioli, R., Wieler, M., Martin, W., & Beaulieu, C. (2012). Diffusion tensor imaging of white matter tract evolution over the lifespan. *NeuroImage*, 60, 340–352. doi:10.1016/j.neuroimage.2011.11.094
- Lebel, C., Walton, M., Letourneau, N., Giesbrecht, G. F., Kaplan, B. J., & Dewey, D. (2016). Parturition and postpartum maternal depressive symptoms are related to children’s brain structure in preschool. *Biological Psychiatry*, 80, 859–868. doi:10.1016/j.biopsych.2015.12.004
- Lewis, A. J., Galbally, M., Gannon, T., & Symeonides, C. (2014). Early life programming as a target for prevention of child and adolescent mental disorders. *BMC Medicine*, 12. doi:10.1186/1741-7015-12-33
- Lewitus, E., Kelava, I., & Huttner, W. B. (2013). Conical expansion of the outer subventricular zone and the role of neocortical folding in evolution and development. *Frontiers in Human Neuroscience*, 7. doi:10.3389/fnhum.2013.00424
- Li, G., Nie, J., Wang, L., Shi, F., Lyall, A. E., Lin, W., . . . Shen, D. (2014). Mapping longitudinal hemispheric structural asymmetries of the human cerebral cortex from birth to 2 years of age. *Cerebral Cortex*, 24, 1289–1300. doi:10.1093/cercor/bhs413
- Li, G., Wang, L., Shi, F., Lyall, A. E., Lin, W., Gilmore, J. H., & Shen, D. (2014). Mapping longitudinal development of local cortical gyrification in infants from birth to 2 years of age. *Journal of Neuroscience*, 34, 4228–4238. doi:10.1523/JNEUROSCI.3976-13.2014
- Loomans, E. M., van Dijk, A. E., Vrijkkotte, T. G. M., van Eijsden, M., Stronks, K., Gemke, R. J. B. J., & Van den Bergh, B. R. H. (2013). Psychosocial stress during pregnancy is related to adverse birth outcomes: Results from a large multi-ethnic community-based birth cohort. *European Journal of Public Health*, 23, 485–491.
- Lupien, S. J., Parent, S., Evans, A. C., Tremblay, R. E., Zelazo, P. D., Corbo, V., . . . Séguin, J. R. (2011). Larger amygdala but no change in hippocampal volume in 10-year-old children exposed to maternal depressive symptomatology since birth. *Proceedings of the National Academy of Sciences*, 108, 14324–14329. doi:10.1073/pnas.1105371108
- Lusby, C. M., Goodman, S. H., Bell, M. A., & Newport, D. J. (2014). Electroencephalogram patterns in infants of depressed mothers. *Developmental Psychobiology*, 56, 459–473. doi:10.1002/dev.21112
- Lusby, C. M., Goodman, S. H., Yeung, E. W., Bell, M. A., & Stowe, Z. N. (2016). Infant EEG and temperament negative affectivity: Coherence of vulnerabilities to mothers’ perinatal depression. *Development and Psychopathology*, 28(4, Pt. 1), 895–911. doi:10.1017/S0954579416000614
- Lussier, S. J., & Stevens, H. E. (2016). Delays in GABAergic interneuron development and behavioral inhibition after prenatal stress. *Developmental Neurobiology*, 76, 1078–1091. doi:10.1002/dneu.22376
- Maguire, E. A., Gadian, D. G., Johnsrude, I. S., Good, C. D., Ashburner, J., Frackowiak, R. S. J., & Frith, C. D. (2000). Navigation-related structural change in the hippocampi of taxi drivers. *Proceedings of the National Academy of Sciences of the United States of America*, 97, 4398–4403. doi:10.1073/pnas.070039597
- Maramba, B., Mohseni Salehi, S. S., Afacan, O., Scherrer, B., Rollins, C. K., Yang, E., . . . Gholipour, A. (2017). Temporal slice registration and robust diffusion-tensor reconstruction for improved fetal brain structural connectivity analysis. *NeuroImage*, 156(Suppl. C), 475–488. doi:10.1016/j.neuroimage.2017.04.033
- Margulies, D. S., Böttger, J., Long, X., Lv, Y., Kelly, C., Schäfer, A., . . . Villringer, A. (2010). Resting developments: A review of fMRI post-processing methodologies for spontaneous brain activity. *Magnetic Resonance Materials in Physics, Biology and Medicine*, 23, 289–307. doi:10.1007/s10334-010-0228-5
- Margulies, D. S., Vincent, J. L., Kelly, C., Lohmann, G., Uddin, L. Q., Biswal, B. B., . . . Petrides, M. (2009). Precuneus shares intrinsic functional architecture in humans and monkeys. *Proceedings of the National Academy of Sciences of the United States of America*, 106, 20069–20074. doi:10.1073/pnas.0905314106
- Mascheretti, S., De Luca, A., Trezzi, V., Peruzzo, D., Nordio, A., Marino, C., & Arrigoni, F. (2017). Neurogenetics of developmental dyslexia: From genes to behavior through brain neuroimaging and cognitive and sensorial mechanisms. *Translational Psychiatry*, 7, e987. doi:10.1038/tp.2016.240
- Mennes, M., Stiers, P., Lagae, L., & Van den Bergh, B. R. H. (2006). Long-term cognitive sequelae of antenatal maternal anxiety: Involvement of the orbitofrontal cortex. *Neuroscience and Biobehavioral Reviews*, 30, 1078–1086. doi:10.1016/j.neubiorev.2006.04.003
- Mennes, M., Van den Bergh, B. R. H., Lagae, L., & Stiers, P. (2009). Developmental brain alterations in 17 year old boys are related to antenatal maternal anxiety. *Clinical Neurophysiology*, 120, 1116–1122. doi:10.1016/j.clinph.2009.04.003
- Mennes, M., Van den Bergh, B. R. H., Snaert, S. S., Lagae, L., & Stiers, P. (2016). Antenatal maternal anxiety modulates the BOLD response in 20-year old adolescents during an endogenous cognitive control task. *bioRxiv*. doi:10.1101/087817
- Menon, V. (2013). Developmental pathways to functional brain networks: Emerging principles. *Trends in Cognitive Sciences*, 17, 627–640. doi:10.1016/j.tics.2013.09.015
- Meredith, R. M. (2015). Sensitive and critical periods during neurotypical and aberrant neurodevelopment: A framework for neurodevelopmental disorders. *Neuroscience and Biobehavioral Reviews*, 50, 180–188. doi:10.1016/j.neubiorev.2014.12.001
- Milgrom, J., Holt, C., Holt, C. J., Ross, J., Erickson, J., & Gemmill, A. W. (2015). Feasibility study and pilot randomised trial of an antenatal depression treatment with infant follow-up. *Archives of Women’s Mental Health*, 18, 717–730. doi:10.1007/s00737-015-0512-5
- Miller, K. L., Stagg, C. J., Douaud, G., Jbabdi, S., Smith, S. M., Behrens, T. E. J., . . . McNab, J. A. (2011). Diffusion imaging of whole, post-mortem human brains on a clinical MRI scanner. *NeuroImage*, 57, 167–181. doi:10.1016/j.neuroimage.2011.03.070
- Mills, K. L., & Tamnes, C. K. (2014). Methods and considerations for longitudinal structural brain imaging analysis across development. *Developmental Cognitive Neuroscience*, 9, 172–190. doi:10.1016/j.dcn.2014.04.004
- Molfese, D. L. (2000). Predicting dyslexia at 8 years of age using neonatal brain responses. *Brain Language*, 72, 238–245.
- Mori, S., Kaufmann, W. E., Davatzikos, C., Stieltjes, B., Amodei, L., Fredericksen, K., . . . van Zijl, P. C. M. (2002). Imaging cortical association

- tracts in the human brain using diffusion-tensor-based axonal tracking. *Magnetic Resonance in Medicine*, 47, 215–223.
- Mulkey, S. B., Yap, V. L., Bai, S., Ramakrishnaiah, R. H., Glasier, C. M., Bornemeier, R. A., . . . Bhutta, A. T. (2015). Amplitude-integrated EEG in newborns with critical congenital heart disease predicts preoperative brain magnetic resonance imaging findings. *Pediatric Neurology*, 52, 599–605. doi:10.1016/j.pediatrneurol.2015.02.026
- Müller, B. C. N., Kühn-Popp, N., Meinhardt, J., Sodian, B., & Paulus, M. (2015). Long-term stability in children's frontal EEG alpha asymmetry between 14-months and 83-months. *International Journal of Developmental Neuroscience*, 41, 110–114. doi:10.1016/j.ijdevneu.2015.01.002
- O'Connor, T. G., Monk, C., & Fitelson, E. M. (2014). Practitioner Review: Maternal mood in pregnancy and child development—Implications for child psychology and psychiatry. *Journal of Child Psychology and Psychiatry*, 55, 99–111. doi:10.1111/jcpp.12153
- Ogawa, S., Tank, D. W., Menon, R., Ellermann, J. M., Kim, S. G., Merkle, H., & Ugurbil, K. (1992). Intrinsic signal changes accompanying sensory stimulation: Functional brain mapping with magnetic resonance imaging. *Proceedings of the National Academy of Sciences of the United States of America*, 89, 5951–5955.
- Otte, R. A., Donkers, F. C. L., Braeken, M. A. K. A., & Van den Bergh, B. R. H. (2015). Multimodal processing of emotional information in 9-month-old infants: II. Prenatal exposure to maternal anxiety. *Brain and Cognition*, 95, 107–117. doi:10.1016/j.bandc.2014.12.001
- Oubel, E., Koob, M., Studholme, C., Dietemann, J.-L., & Rousseau, F. (2010). Reconstruction of scattered data in fetal diffusion MRI. In T. Jiang, N. Navab, J. P. W. Pluim, & M. A. Viergever (Eds.), *Medical image computing and computer-assisted intervention—MICCAI 2010: 13th International Conference, Beijing, China, September 20–24, 2010, Proceedings, Part I* (pp. 574–581). Berlin: Springer.
- Parikshak, N. N., Gandal, M. J., & Geschwind, D. H. (2015). Systems biology and gene networks in neurodevelopmental and neurodegenerative disorders. *Nature Reviews Genetics*, 16, 441–458. doi:10.1038/nrg3934
- Peltola, M. J., Bakermans-Kranenburg, M. J., Alink, L. R. A., Huffmeijer, R., Biro, S., & van IJzendoorn, M. H. (2014). Resting frontal EEG asymmetry in children: Meta-analyses of the effects of psychosocial risk factors and associations with internalizing and externalizing behavior. *Developmental Psychobiology*, 56, 1377–1389. doi:10.1002/dev.21223
- Pletikos, M., Sousa, A. M. M., Sedmak, G., Meyer, K. A., Zhu, Y., Cheng, F., . . . Sestan, N. (2014). Temporal specification and bilaterality of human neocortical topographic gene expression. *Neuron*, 81, 321–332. doi:10.1016/j.neuron.2013.11.018
- Poldrack, R. A. (2012). The future of fMRI in cognitive neuroscience. *NeuroImage*, 62, 1216–1220. doi:10.1016/j.neuroimage.2011.08.007
- Posner, J., Cha, J., Roy, A. K., Peterson, B. S., Bansal, R., Gustafsson, H. C., . . . Monk, C. (2016). Alterations in amygdala–prefrontal circuits in infants exposed to prenatal maternal depression. *Translational Psychiatry*, 6, e935. doi:10.1038/tp.2016.146
- Power, J. D., Fair, D. A., Schlaggar, B. L., & Petersen, S. E. (2010). The development of human functional brain networks. *Neuron*, 67, 735–748. doi:10.1016/j.neuron.2010.08.017
- Qin, S., Duan, X., Supekar, K., Chen, H., Chen, T., & Menon, V. (2016). Large-scale intrinsic functional network organization along the long axis of the human medial temporal lobe. *Brain Structure and Function*, 221, 3237–3258. doi:10.1007/s00429-015-1098-4
- Qiu, A., Anh, T. T., Li, Y., Chen, H., Rifkin-Graboi, A., Broekman, B. F. P., . . . Meaney, M. J. (2015). Prenatal maternal depression alters amygdala functional connectivity in 6-month-old infants. *Translational Psychiatry*, 5, e508. doi:10.1038/tp.2015.3
- Qiu, A., Rifkin-Graboi, A., Chen, H., Chong, Y. S., Kwek, K., Gluckman, P. D., . . . Meaney, M. J. (2013). Maternal anxiety and infants' hippocampal development: Timing matters. *Translational Psychiatry*, 3, e306. doi:10.1038/tp.2013.79
- Qiu, A., Tuan, T. A., Ong, M. L., Li, Y., Chen, H., Rifkin-Graboi, A., . . . Gluckman, P. D. (2015). COMT haplotypes modulate associations of antenatal maternal anxiety and neonatal cortical morphology. *American Journal of Psychiatry*, 172, 163–172. doi:10.1176/appi.ajp.2014.14030313
- Rakers, F., Rupprecht, S., Dreiling, M., Bergmeier, C., Witte, O. W., & Schwab, M. (2017). Transfer of maternal psychosocial stress to the fetus. *Neuroscience and Biobehavioral Reviews*. Advance online publication. doi:10.1016/j.neubiorev.2017.02.019
- Reid, L. B., Sale, M. V., Cunnington, R., Mattingley, J. B., & Rose, S. E. (2017). Brain changes following four weeks of unimanual motor training: Evidence from fMRI-guided diffusion MRI tractography. *Human Brain Mapping*, 9, 4302–4312. doi:10.1002/hbm.23514
- Reuter, M., Tisdall, M. D., Qureshi, A., Buckner, R. L., van der Kouwe, A. J. W., & Fischl, B. R. (2015). Head motion during MRI acquisition reduces gray matter volume and thickness estimates. *NeuroImage*, 107, 107–115. doi:10.1016/j.neuroimage.2014.12.006
- Richmond, S., Johnson, K. A., Seal, M. L., Allen, N. B., & Whittle, S. (2016). Development of brain networks and relevance of environmental and genetic factors: A systematic review. *Neuroscience and Biobehavioral Reviews*, 71, 215–239. doi:10.1016/j.neubiorev.2016.08.024
- Rifkin, L., Lewis, S., Jones, P., Toone, B., & Murray, R. (1994). Low birth weight and schizophrenia. *British Journal of Psychiatry*, 165, 357–362. doi:10.1192/bjp.165.3.357
- Rifkin-Graboi, A., Bai, J., Chen, H., Hameed, W. B. R., Sim, L. W., Tint, M. T., . . . Qiu, A. (2013). Prenatal maternal depression associates with microstructure of right amygdala in neonates at birth. *Biological Psychiatry*, 74, 837–844. doi:10.1016/j.biopsych.2013.06.019
- Rifkin-Graboi, A., Meaney, M. J., Chen, H., Bai, J., Hameed, W. B. R., Tint, M. T., . . . Qiu, A. (2015). Antenatal maternal anxiety predicts variations in neural structures implicated in anxiety disorders in newborns. *Journal of the American Academy of Child & Adolescent Psychiatry*, 54, 313–321. doi:10.1016/j.jaac.2015.01.013
- Roalf, D. R., Quarnley, M., Elliott, M. A., Satterthwaite, T. D., Vandekar, S. N., Ruparel, K., . . . Gur, R. E. (2016). The impact of quality assurance assessment on diffusion tensor imaging outcomes in a large-scale population-based cohort. *NeuroImage*, 125, 903–919. doi:10.1016/j.neuroimage.2015.10.068
- Robinson, E. C., Jbabdi, S., Glasser, M. F., Andersson, J., Burgess, G. C., Harms, M. P., . . . Jenkinson, M. (2014). MSM: A new flexible framework for Multimodal Surface Matching. *NeuroImage*, 100, 414–426. doi:10.1016/j.neuroimage.2014.05.069
- Rubinov, M., & Sporns, O. (2010). Complex network measures of brain connectivity: Uses and interpretations. *NeuroImage*, 52, 1059–1069. doi:10.1016/j.neuroimage.2009.10.003
- Rutkowski, T. P., Schroeder, J. P., Gafford, G. M., Warren, S. T., Weinshenker, D., Caspary, T., & Mulle, J. G. (2017). Unraveling the genetic architecture of copy number variants associated with schizophrenia and other neuropsychiatric disorders. *Journal of Neuroscience Research*, 95, 1144–1160. doi:10.1002/jnr.23970
- Sandman, C. A., Buss, C., Head, K., & Davis, E. P. (2015). Fetal exposure to maternal depressive symptoms is associated with cortical thickness in late childhood. *Biological Psychiatry*, 77, 324–334. doi:10.1016/j.biopsych.2014.06.025
- Sarkar, S., Craig, M. C., Dell'Acqua, F., O'Connor, T. G., Catani, M., Deeley, Q., . . . Murphy, D. G. M. (2014). Prenatal stress and limbic-prefrontal white matter microstructure in children aged 6–9 years: A preliminary diffusion tensor imaging study. *World Journal of Biological Psychiatry*, 15, 346–352. doi:10.3109/15622975.2014.903336
- Schaer, M., Cuadra, M. B., Tamarit, L., Lazeyras, F., Eliez, S., & Thiran, J.-P. (2008). A surface-based approach to quantify local cortical gyification. *IEEE Transactions on Medical Imaging*, 27, 161–170. doi:10.1109/TMI.2007.903576
- Scheinost, D., Kwon, S. H., Lacadie, C., Sze, G., Sinha, R., Constable, R. T., & Ment, L. R. (2016). Prenatal stress alters amygdala functional connectivity in preterm neonates. *NeuroImage: Clinical*, 12, 381–388. doi:10.1016/j.nicl.2016.08.010
- Scheinost, D., Sinha, R., Cross, S. N., Kwon, S. H., Sze, G., Constable, R. T., & Ment, L. R. (2016). Does prenatal stress alter the developing connectome? *Pediatric Research. Advance online publication*. doi:10.1038/pr.2016.197
- Schmidt, P., Gaser, C., Arsic, M., Buck, D., Förschler, A., Berthele, A., . . . Mühlau, M. (2012). An automated tool for detection of FLAIR-hyperintense white-matter lesions in Multiple Sclerosis. *NeuroImage*, 59, 3774–3783. doi:10.1016/j.neuroimage.2011.11.032
- Schöpf, V., Kaspran, G., Brugger, P. C., & Prayer, D. (2012). Watching the fetal brain at “rest.” *International Journal of Developmental Neuroscience*, 30, 11–17. doi:10.1016/j.ijdevneu.2011.10.006
- Seckl, J. R. (2007). Glucocorticoids, developmental “programming” and the risk of affective dysfunction. In E. R. De Kloet, M. S. Oitzl, & E. Vermetten (Eds.), *Progress in brain research* (Vol. 167, pp. 17–34). New York: Elsevier.
- Seiradake, E., Jones, E. Y. J., & Klein, R. (2016). Structural perspectives on axon guidance. *Annual Review of Cell and Developmental Biology*, 32, 577–608. doi:10.1146/annurev-cellbio-111315-125008

- Silbereis, J. C., Pochareddy, S., Zhu, Y., Li, M., & Sestan, N. (2016). The cellular and molecular landscapes of the developing human central nervous system. *Neuron*, 89, 248–268. doi:10.1016/j.neuron.2015.12.008
- Smith, S. M., Fox, P. T., Miller, K. L., Glahn, D. C., Fox, P. M., Mackay, C. E., . . . Beckmann, C. F. (2009). Correspondence of the brain's functional architecture during activation and rest. *Proceedings of the National Academy of Sciences*, 106, 13040–13045. doi:10.1073/pnas.0905267106
- Smyser, C. D., Inder, T. E., Shimony, J. S., Hill, J. E., Degan, A. J., Snyder, A. Z., & Neil, J. J. (2010). Longitudinal analysis of neural network development in preterm infants. *Cerebral Cortex*, 20, 2852–2862. doi:10.1093/cercor/bhq035
- Soe, N. N., Wen, D. J., Poh, J. S., Li, Y., Broekman, B. F. P., Chen, H., . . . Qiu, A. (2016). Pre- and post-natal maternal depressive symptoms in relation with infant frontal function, connectivity, and behaviors. *PLOS ONE*, 11, e0152991. doi:10.1371/journal.pone.0152991
- Sporns, O., Chialvo, D. R., Kaiser, M., & Hilgetag, C. C. (2004). Organization, development and function of complex brain networks. *Trends in Cognitive Sciences*, 8, 418–425. doi:10.1016/j.tics.2004.07.008
- Sporns, O., Tononi, G., & Kötter, R. (2005). The human connectome: A structural description of the human brain. *PLOS Computational Biology*, 1, e42. doi:10.1371/journal.pcbi.0010042
- Sroufe, L. A., & Rutter, M. (1984). The domain of developmental psychopathology. *Child Development*, 55, 17–19.
- Stam, C. J., Tewarie, P., van Dellen, E., van Straaten, E. C. W., Hillebrand, A., & van Mieghem, P. (2014). The trees and the forest: Characterization of complex brain networks with minimum spanning trees. *International Journal of Psychophysiology*, 92, 129–138. doi:10.1016/j.ijpsycho.2014.04.001
- Stam, C. J., & van Straaten, E. C. W. (2012). The organization of physiological brain networks. *Clinical Neurophysiology*, 123, 1067–1087. doi:10.1016/j.clinph.2012.01.011
- Stein, A., Pearson, R. M., Goodman, S. H., Rapa, E., Rahman, A., McCallum, M., . . . Pariente, C. M. (2014). Effects of perinatal mental disorders on the fetus and child. *Lancet*, 384, 1800–1819. doi:10.1016/S0140-6736(14)61277-0
- Striedter, G. F., Srinivasan, S., & Monuki, E. S. (2015). Cortical folding: When, where, how, and why? *Annual Review of Neuroscience*, 38, 291–307. doi:10.1146/annurev-neuro-071714-034128
- Suárez-Solá, M. L., González-Delgado, F. J., Pueyo-Morlans, M., Medina-Bolívar, O. C., Hernández-Acosta, N. C., González-Gómez, M., & Meyer, G. (2009). Neurons in the white matter of the adult human neocortex. *Frontiers in Neuroanatomy*, 3, 7. doi:10.3389/neuro.05.007.2009
- Swanson, L. W., & Lichtman, J. W. (2016). From Cajal to connectome and beyond. *Annual Review of Neuroscience*, 39. doi:10.1146/annurev-neuro-071714-033954
- Tallinen, T., & Biggins, J. S. (2015). Mechanics of invagination and folding: Hybridized instabilities when one soft tissue grows on another. *Physical Review E*, 92, 022720. doi:10.1103/PhysRevE.92.022720
- Tallinen, T., Chung, J. Y., Rousseau, F., Girard, N., Lefèvre, J., & Mahadevan, L. (2016). On the growth and form of cortical convolutions. *Nature Physics*. Advance online publication. doi:10.1038/nphys3632
- Tardif, C. L., Schäfer, A., Wachert, M., Dinse, J., Turner, R., & Bazin, P.-L. (2015). Multi-contrast multi-scale surface registration for improved alignment of cortical areas. *NeuroImage*, 111, 107–122. doi:10.1016/j.neuroimage.2015.02.005
- Teipel, S. J., Bokde, A. L. W., Meindl, T., Amaro, E., Soldner, J., Reiser, M. F., . . . Hampel, H. (2010). White matter microstructure underlying default mode network connectivity in the human brain. *NeuroImage*, 49, 2021–2032. doi:10.1016/j.neuroimage.2009.10.067
- Thapar, A., Cooper, M., & Rutter, M. (2017). Neurodevelopmental disorders. *Lancet Psychiatry*, 4, 339–346. doi:10.1016/S2215-0366(16)30376-5
- Thayer, J. F., Åhs, F., Fredrikson, M., Sollers, J. J., & Wager, T. D. (2012). A meta-analysis of heart rate variability and neuroimaging studies: Implications for heart rate variability as a marker of stress and health. *Neuroscience and Biobehavioral Reviews*, 36, 747–756. doi:10.1016/j.neubiorev.2011.11.009
- Thomason, M. E., Brown, J. A., Dassanayake, M. T., Shastri, R., Marusak, H. A., Hernandez-Andrade, E., . . . Romero, R. (2014). Intrinsic functional brain architecture derived from graph theoretical analysis in the human fetus. *PLOS ONE*, 9, e94423. doi:10.1371/journal.pone.0094423
- Thomason, M. E., Dassanayake, M. T., Shen, S., Katkuri, Y., Alexis, M., Anderson, A. L., . . . Romero, R. (2013). Cross-hemispheric functional connectivity in the human fetal brain. *Science Translational Medicine*, 5, 173ra124–173ra124. doi:10.1126/scitranslmed.3004978
- Thomason, M. E., Grove, L. E., Lozon, T. A., Jr., Vila, A. M., Ye, Y., Nye, M. J., . . . Romero, R. (2015). Age-related increases in long-range connectivity in fetal functional neural connectivity networks in utero. *Developmental Cognitive Neuroscience*, 11, 96–104. doi:10.1016/j.dcn.2014.09.001
- Thomason, M. E., Scheinost, D., Manning, J. H., Grove, L. E., Hect, J., Marshall, N., . . . Romero, R. (2017). Weak functional connectivity in the human fetal brain prior to preterm birth. *Scientific Reports*, 7, 39286. doi:10.1038/srep39286
- Thompson, C., Syddall, H., Rodin, I., Omond, C., & Barker, D. J. P. (2001). Birth weight and the risk of depressive disorder in late life. *British Journal of Psychiatry*, 179, 450–455. doi:10.1192/bjp.179.5.450
- Toro, R. (2012). On the possible shapes of the brain. *Evolutionary Biology*, 39, 600–612. doi:10.1007/s11692-012-9201-8
- Tóth, B., Urbán, G., Hádán, G. P., Márk, M., Török, M., Stam, C. J., & Winkler, I. (2017). Large-scale network organization of EEG functional connectivity in newborn infants. *Human Brain Mapping*, 38, 4019–4033. doi:10.1002/hbm.23645
- Tourbier, S., Velasco-Annis, C., Taimouri, V., Hagmann, P., Meuli, R., Warfield, S. K., . . . Gholipour, A. (2017). Automated template-based brain localization and extraction for fetal brain MRI reconstruction. *NeuroImage*, 155(Suppl. C), 460–472. doi:10.1016/j.neuroimage.2017.04.004
- Turner, B. M., Forstmann, B. U., Love, B. C., Palmeri, T. J., & van Maanen, L. (2017). Approaches to analysis in model-based cognitive neuroscience. *Journal of Mathematical Psychology*, 76(Pt. B), 65–79. doi:10.1016/j.jmp.2016.01.001
- Turner, B. M., Rodriguez, C. A., Norcia, T. M., McClure, S. M., & Steyvers, M. (2016). Why more is better: Simultaneous modeling of EEG, fMRI, and behavioral data. *NeuroImage*, 128, 96–115. doi:10.1016/j.neuroimage.2015.12.030
- Van den Bergh, B. R. H. (2011). Developmental programming of early brain and behaviour development and mental health: A conceptual framework. *Developmental Medicine and Child Neurology*, 53, 19–23. doi:10.1111/j.1469-8749.2011.04057.x
- Van den Bergh, B. R. H. (2016). Maternal anxiety, mindfulness, and heart rate variability during pregnancy influence fetal and infant development. In N. Reissland & B. S. Kisilevsky (Eds.), *Fetal development: Research on brain and behavior, environmental influences, and emerging technologies* (pp. 267–292). Cham, Switzerland: Springer.
- Van den Bergh, B. R. H., & Marcoen, A. (2004). High antenatal maternal anxiety is related to ADHD symptoms, externalizing problems, and anxiety in 8- and 9-year-olds. *Child Development*, 75, 1085–1097. doi:10.1111/j.1467-8624.2004.00727.x
- Van den Bergh, B. R. H., Mennes, M., Oosterlaan, J., Stevens, V., Stiers, P., Marcoen, A., & Lagae, L. (2005). High antenatal maternal anxiety is related to impulsivity during performance on cognitive tasks in 14- and 15-year-olds. *Neuroscience and Biobehavioral Reviews*, 29, 259–269. doi:10.1016/j.neubiorev.2004.10.010
- Van den Bergh, B. R. H., Mennes, M., Stevens, V., van der Meere, J., Borger, N., Stiers, P., . . . Lagae, L. (2006). ADHD deficit as measured in adolescent boys with a continuous performance task is related to antenatal maternal anxiety. *Pediatric Research*, 59, 78–82.
- Van den Bergh, B. R. H., Mulder, E. J. H., Visser, G. H. A., Poelmann-Weesjes, G., Bekedam, D. J., & Precht, H. F. R. (1989). The effect of (induced) maternal emotions on fetal behaviour: A controlled study. *Early Human Development*, 19, 9–19. doi:10.1016/0378-3782(89)90100-X
- Van den Bergh, B. R. H., van den Heuvel, M. I., Lahti, M., Braeken, M., de Rooij, S. R., Entringer, S., . . . Schwab, M. (2017). Prenatal developmental origins of behavior and mental health: The influence of maternal stress in pregnancy. *Neuroscience and Biobehavioral Reviews*. Advance online publication. doi:10.1016/j.neubiorev.2017.07.003
- van den Heuvel, M. I., Donkers, F. C., Winkler, I., Otte, R. A., & Van den Bergh, B. R. H. (2015). Maternal mindfulness and anxiety during pregnancy affect infants' neural responses to sounds. *Social Cognitive Affective Neuroscience*, 10, 453–460. doi:10.1093/scan/nsu075
- van den Heuvel, M. I., Henrichs, J., Donkers, F. C., & Van den Bergh, B. R. H. (in press). Children prenatally exposed to maternal anxiety devote more attentional resources to neutral pictures. *Developmental Science*. doi:10.1111/desc.12612
- van den Heuvel, M. I., Mandl, R., & Hulshoff Pol, H. (2008). Normalized cut group clustering of resting-state fMRI data. *PLOS ONE*, 3, e2001. doi:10.1371/journal.pone.0002001



- van den Heuvel, M. I., & Thomason, M. E. (2016). Functional connectivity of the human brain in utero. *Trends in Cognitive Sciences*. Advance online publication. doi:10.1016/j.tics.2016.10.001
- van Essen, D. C., & Barch, D. M. (2015). The human connectome in health and psychopathology. *World Psychiatry, 14*, 154–157. doi:10.1002/wps.20228
- van Essen, D. C., & Maunsell, J. H. R. (1980). Two-dimensional maps of the cerebral cortex. *Journal of Comparative Neurology, 191*, 255–281. doi:10.1002/cne.901910208
- Vértes, P. E., & Bullmore, E. T. (2015). Annual Research Review: Growth connectomics—The organization and reorganization of brain networks during normal and abnormal development. *Journal of Child Psychology and Psychiatry, 56*, 299–320. doi:10.1111/jcpp.12365
- von Leupoldt, A., Mangelschots, E., Niederstrasser, N. G., Braeken, M., Billiet, T., & Van den Bergh, B. R. H. (2017). Prenatal stress exposure is associated with increased dyspnea perception in adulthood. *European Respiratory Journal*. Advance online publication.
- Walhovd, K. B., Fjell, A. M., Giedd, J., Dale, A. M., & Brown, T. T. (2016). Through thick and thin: A need to reconcile contradictory results on trajectories in human cortical development. *Cerebral Cortex*. Advance online publication. doi:10.1093/cercor/bhv301
- Watts, D. J., & Strogatz, S. H. (1998). Collective dynamics of “small-world” networks. *Nature, 393*, 440–442. doi:10.1038/30918
- Wegiel, J., Kuchna, I., Nowicki, K., Imaki, H., Wegiel, J., Marchi, E., . . . Wisniewski, T. (2010). The neuropathology of autism: Defects of neurogenesis and neuronal migration, and dysplastic changes. *Acta Neuropathologica, 119*, 755–770. doi:10.1007/s00401-010-0655-4
- Weiskopf, N., Suckling, J., Williams, G. B., Correia, M. M., Inkster, B., Tait, R., . . . Lutti, A. (2013). Quantitative multi-parameter mapping of R1, PD, MT, and R2 at 3T: A multi-center validation. *Frontiers in Neuroscience, 7*, 95. doi:10.3389/fnins.2013.00095
- Wen, D. J., Poh, J. S., Ni, S. N., Chong, Y. S., Chen, H., Kwek, K., . . . Qiu, A. (2017). Influences of prenatal and postnatal maternal depression on amygdala volume and microstructure in young children. *Translational Psychiatry, 7*, e1103. doi:10.1038/tp.2017.74
- Wen, Z. (2017). Modeling neurodevelopmental and psychiatric diseases with human iPSCs. *Journal of Neuroscience Research, 95*, 1097–1109. doi:10.1002/jnr.24031
- Williams, V. J., Juraneck, J., Cirino, P., & Fletcher, J. M. (2017). Cortical thickness and local gyrification in children with developmental dyslexia. *Cerebral Cortex*. Advance online publication.
- Winkler, A. M., Kochunov, P. V., Blangero, J., Almasy, L., Zilles, K., Fox, P. T., . . . Glahn, D. C. (2010). Cortical thickness or grey matter volume? The importance of selecting the phenotype for imaging genetics studies. *NeuroImage, 53*, 1135–1146. doi: 10.1016/j.neuroimage.2009.12.028
- Winston, G. P., Micallef, C., Symms, M. R., Alexander, D. C., Duncan, J. S., & Zhang, H. (2014). Advanced diffusion imaging sequences could aid assessing patients with focal cortical dysplasia and epilepsy. *Epilepsy Research, 108*, 336–339. doi:10.1016/j.eplepsyres.2013.11.004
- Wright, R., Kyriakopoulou, V., Ledig, C., Rutherford, M. A., Hajnal, J. V., Rueckert, D., & Aljabar, P. (2014). Automatic quantification of normal cortical folding patterns from fetal brain MRI. *NeuroImage, 91*(Suppl. C), 21–32. doi:10.1016/j.neuroimage.2014.01.034
- Wright, R., Makropoulos, A., Kyriakopoulou, V., Patkee, P. A., Koch, L. M., Rutherford, M. A., . . . Aljabar, P. (2015). Construction of a fetal spatio-temporal cortical surface atlas from in utero MRI: Application of spectral surface matching. *NeuroImage, 120*(Suppl. C), 467–480. doi:10.1016/j.neuroimage.2015.05.087
- Yu, Q., Ouyang, A., Chalak, L., Jeon, T., Chia, J., Mishra, V., . . . Huang, H. (2015). Structural development of human fetal and preterm brain cortical plate based on population-averaged templates. *Cerebral Cortex*. Advance online publication. doi:10.1093/cercor/bhv201
- Zhan, J., Dinov, I. D., Li, J., Zhang, Z., Hobel, S., Shi, Y., . . . Liu, S. (2013). Spatial-temporal atlas of human fetal brain development during the early second trimester. *NeuroImage, 82*, 115–126. doi:10.1016/j.neuroimage.2013.05.063
- Ziats, M. N., Grosvenor, L. P., & Rennert, O. M. (2015). Functional genomics of human brain development and implications for autism spectrum disorders. *Translational Psychiatry, 5*, e665. doi:10.1038/tp.2015.153
- Ziegler, G., Ridgway, G. R., Dahnke, R., Gaser, C., & Alzheimer's Disease Neuroimaging Initiative. (2014). Individualized Gaussian process-based prediction and detection of local and global gray matter abnormalities in elderly subjects. *NeuroImage, 97*(Suppl. C), 333–348. doi:10.1016/j.neuroimage.2014.04.01