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Review

Cite this article: Li C *et al.* (2020) Functions and mechanism of noncoding RNA in the somatic cells of the testis. *Zygote* **28**: 87–92. doi: 10.1017/S0967199419000650

Received: 23 July 2019 Revised: 20 August 2019 Accepted: 12 September 2019 First published online: 2 December 2019

Keywords:

Leydig cells; miRNAs; ncRNAs; Sertoli cells

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Functions and mechanism of noncoding RNA in the somatic cells of the testis

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Summary

ncRNAs are involved in numerous biological processes by regulating gene expression and cell stability. Studies have shown that ncRNAs also contribute to spermatogenesis. Leydig cells (LCs) and Sertoli cells (SCs) are somatic cells of the testis that support spermatogenesis and are vital to male fertility. In this review, we summarized the findings from studies on ncRNAs in SCs and LCs. In SCs, ncRNAs play key roles in phagocytosis, immunoprotection and development of SCs. In LCs, ncRNAs are involved in steroidogenesis, in particular production of testosterone as well as development of LCs. Here, we discuss the possible target genes and functions of ncRNAs in both types of cells. These ncRNAs regulate the expression of target genes or mRNA coding sequence regions, resulting in a chain reaction that influences cell function. In addition, microRNAs, lncRNAs, piRNA-like RNAs (pilRNAs) and natural antisense transcripts (NATs) are discussed in this review. In summary, we suggest that these ncRNAs might act in coordination to control spermatogenesis and maintain the environmental homeostasis of the testis.

Introduction

Because of the work of the Encyclopedia of DNA Elements (ENCODE) Project Consortium, we have a comprehensive understanding of DNA sequences in the human genome (The ENCODE Project Consortium, 2004, 2012; Birney et al., 2007). Eighty per cent of the genome is transcribed into RNA, while only a small proportion encodes proteins (The ENCODE Project Consortium, 2012). Noncoding RNAs, formerly called 'junk', have been proven to conduct 'pervasive transcription' and contain a great deal of functional regulatory elements (Ohno, 1972; Kapranov et al., 2007; Neph et al., 2012). According to the length of noncoding RNA, they are divided into long noncoding RNAs (lncRNAs; >200 nt) and small noncoding RNAs. Small noncoding RNAs include microRNAs (miRNAs), endogenous small interfering RNAs (endosiRNAs) and PIWI-interacting RNAs (piRNAs) (Lucas and Raikhel, 2013). Noncoding RNAs (ncRNAs) play important roles in gene expression and stability in processes from embryonic development to adult homeostasis (Geisler and Coller, 2013; Patil et al., 2014). According to their functional features, ncRNAs are divided into housekeeping ncRNAs and regulatory ncRNAs. The regulatory ncRNAs comprise miRNAs, siRNAs, lncRNAs, piRNAs and intermediate ncRNAs such as small nucleolar RNAs. They are expressed in specific cells or a specific stage during cell development and differentiation or in response to environmental stimuli (Brosnan and Voinnet, 2009; Guan et al., 2013). Increasing evidence shows that ncRNAs are involved in spermatogenesis and maintenance of male fertility in germ cells (de Mateo and Sassone-Corsi, 2014; Salviano-Silva et al., 2018). Here we summarize the findings of studies on the noncoding RNAs involved in male fertility in somatic cells.

Sertoli cells

Niche cells, which are devoted to subtle coordination of the testicular microenvironment, are mostly composed of SCs. SCs, which are proximal to spermatogonial stem cells (SSCs), play auxiliary roles in spermatogenesis as 'mother' or 'nurse' cells for SSCs by supplying structural, immunological and nutritional support (Oatley and Brinster, 2012; Hai *et al.*, 2014). As supportive cells, SCs also produce a great number of growth factors and define the fate of SSCs, as stem cell factors (SCFs), bone morphogenetic proteins (BMPs) or glial cell line-derived neurotrophic factors (GDNFs) (Jan *et al.*, 2012; Hai *et al.*, 2014). Moreover, SCs are important constituents of the blood-testis barrier, which is an essential ultrastructure for male fertility (Setchell, 2008; McCabe *et al.*, 2016). The biological functions of SCs have been summarized as follows: they expand SSCs as feeder cells and activate SSC differentiation, phagocytosis and immunoprotection (Zhang *et al.*, 2007; Hai *et al.*, 2014). In 2013, a study using computer-assisted annotation of the small RNA transcriptome in murine SCs was completed (Ortogero *et al.*, 2013). Large numbers of studies have also shown that the disruption in the

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expression of noncoding RNAs in SCs might effect male fertility and these studies also made efforts to determine the functions of SCs in spermatogenesis and male fertility.

microRNAs in Sertoli cells

A microRNA is a small RNA of approximately 22 nucleotides in length (Kim, 2005). Growing evidence has shown that microRNAs regulate many biological processes by influencing post-transcriptional gene expression, such as mRNA degradation, translational repression, DNA methylation and chromatin modification (Krol *et al.*, 2010). MicroRNAs target specific mRNAs and stimulate the degeneration or inhibition of translated mRNAs (Tay *et al.*, 2008). Recently, some studies have shown that miRNAs are also involved in modulating many functions in SCs.

Apoptosis and phagocytosis

Phagocytosis is indispensable for the maintenance of tissue homeostasis. Sufficient evidence has shown that phagocytosis by SCs plays an essential role in the development and differentiation of germ cells. More than one-half of spermatogenic stem cells was cleared and degraded by SCs (Wang et al., 2006). Several miRNAs have been reported to be involved in conventional phagocytosis or LC3associated phagocytosis (LAP) or cell apoptosis (Jovanovic and Hengartner, 2006; Niu et al., 2011). miR-471-5p regulates the level of Dock180, which interacts with autophagy-related proteins and make up LC3-dependent phagocytic complexes. It has been proven that SCs recruit autophagy-related proteins via LAP and that these proteins play crucial roles in the clearance of apoptotic germ cells. Overexpression of miR-471-5p in SCs from transgenic mice increased the number of apoptotic germ cells and damaged male fertility as miR-471-5p targets Dock180, LC3, Atg12, Rab5, Rubicon and Becn1 and represses their expression. This evidence suggested that SC phagocytosis and the clearance of apoptotic germ cells are regulated by miR-471-5p and its target proteins (Panneerdoss et al., 2017).

miR-758 and miR-98-5p were predicted to be participants in germ cell apoptosis by binding to the 3'UTR of *mitogen-activated protein kinase 11 (MAPK11, p38 \beta isoform*) gene. The expression of MAPK11 in SCs could induce the expression of tumour necrosis factor α (TNF- α), which interacts with TNF receptor 1 (TNFR1) and leads to germ cell apoptosis (Chen *et al.*, 2016). In the SCs of underfed sheep, the lack of let-7/miR-98 increased the expression of Fas mRNA and Fas protein and the presence of let-7/ miR-98 reduced cell sensitivity to Fas-induced apoptosis (Wang *et al.*, 2011). Overexpressed miR-202-3p increased the number of apoptotic SCs and inhibited the proliferation and synthesis function of SCs by targeting LRP6 and cyclin D1 (Yang C *et al.*, 2019). In addition, miR-125a-3p, miR-872 and miR-24 possibly induced cell apoptosis by targeting SOD-1, a Cu/Zn superoxide dismutase in SCs (Papaioannou *et al.*, 2011).

Proliferation and development:

Dicer, an RNaseIII endonuclease, has been deemed crucial for miRNA production (Bernstein *et al.*, 2001). Studies have shown that it is highly correlated with maturation and survival of SCs, which suggests that the development, survival and function of SCs might be regulated by miRNAs related to Dicer (Papaioannou *et al.*, 2009, 2011). Dicer is critical to SCs as well as to spermatogenesis. The absence of Dicer leads to infertility. Several miRNAs (miR-299, miR-381, miR-409-5p, miR-376a and miR-674) in SCs were suppressed in Dicer ablated mice,

indicating that these RNAs may take part in the development and function of SCs (Papaioannou *et al.*, 2009).

miR-34c has been proved to be involved in the development of male germ cells (Lian et al., 2012). In fact, the development of seminiferous tubules also requires the participation of miR-34c. Platelet-derived growth factor receptor alpha (PDGFRA), which is mainly expressed in support cells such as SCs, has been identified as one target gene of miR-34c in swine. In addition, the expression level of PDGFRA in 2 days was much higher than it was in 5 months, indicating that PDGFRA may be involved in the early stages of SC development (Zhang X et al., 2015). miR-762 also played an important role in pig testis. miR-762 promoted immature SC proliferation and controlled apoptosis by targeting the 3'UTR of ring finger protein 4 (RNF4), thereby decreasing AR protein expression and the transcriptional regulatory AR activity in SCs (Ma C et al., 2016). In addition, miR-762 could also promote DNA damage repair in SCs (Ma C et al., 2016). As mentioned above, miR-202-3p also inhibited the proliferation and synthesis functions of SCs (Yang C et al., 2019).

Compared with those of patients with obstructive azoospermia (OA), the SCs of patients with Sertoli-cell-only syndrome (SCOS) expressed higher levels of miRNA-133b. Studies have shown that miR-133b promotes SC proliferation in humans by targeting GLI3 and activating cyclin B1 and cyclin D1 (Yao *et al.*, 2016). In addition, the level of miR-375 expression is negatively correlated with the mRNA levels of rearranged L-*myc* fusion (RLF) and hypoxia-induced gene domain protein 1A (HIGD1A) in pig SCs. MTS analysis showed that miR-375 might inhibit SC proliferation (Guo *et al.*, 2018).

Junction of the blood-testis barrier

The BTB is one of the most impermeable blood-tissue barriers in the living body, It is which was composed of tight junctions (TJs), gap junctions (GJs) and desmosome-like junctions and adherens junctions (AJs). The AJs include the basal tubulobulbar complex (basal TBC) and the basal ectoplasmic specialization (basal ES) (Wong and Cheng, 2005; Cheng and Mruk, 2012). The BTB separates the seminiferous epithelium into basal and apical parts. Spermatogenesis involves five orderly processes and the BTB provides a physical barrier that compartmentalizes three of these processes: the cell cycle that enables the spermatocyte transition from the zygotene to the diplotene stage, the procession of round spermatids to spermatozoa and spermiation. In addition, it enables these processes to progress in an immune-privileged environment, indicating that the BTB plays an important role in spermatogenesis and male fertility (Cheng and Mruk, 2012). In mice, considerable evidence has proven that the miR-17-92 cluster plays an important role in spermatogenesis (Bjork et al., 2010; Tong et al., 2012). miR-20a, a member of the miR-17-92 cluster, might be related to genistein (GEN)-induced abnormal spermatogenesis (Gu et al., 2017). As the target gene of miR-20a, Limk1 (Gu et al., 2017), is involved in the RhoB/ROCK/LIMK1 pathway and regulates the adherens junction dynamics of Sertoli germ cells (Lui et al., 2003).

Other findings about microRNAs in Sertoli cells

Circular RNAs (circRNAs) have loop structures and perform important functions in many biological processes (Rybak-Wolf *et al.*, 2015; Ebbesen *et al.*, 2016), including those of inflammatory reactions (Ng *et al.*, 2016). Evidence has shown that circRNA-9119 acts as a miRNA sponge and inhibits miR-136 and miR-26a expression. miR-136 and miR-26a inhibited the expression of retinoic acid inducible gene-I (RIG-I) and Toll-like receptor 3 (TLR3)

by binding to the 3'UTR of RIG-I and TLR3 respectively in Sertoli cells and Leydig cells (LCs). By targeting TLR3 and RIG-I during orchitis in SCs and LCs, miR-136 and miR-26a modulate circRNA-9119-mediated inflammatory reactions, indicating that miRNAs and circRNAs are crucial to the immune microenvironment (Qin *et al.*, 2019).

miR-202-5p, regulated by the testis-determining factor SOX9, is a let-7 family member (Wainwright *et al.*, 2013). miR-202-5p is selectively expressed in SCs. The expression of miR-202-5p is related to testicular development and maturation (Dabaja *et al.*, 2015). However, no miR-202-5p expression was detected in SCs from SCOS patients. The distinct expression of levels of miR-202-5p in fertile and infertile men suggests that it might play crucial roles in normal male fertility (Dabaja *et al.*, 2015).

A study on the expression of miRNAs and their target genes in SCs after exposure to nonylphenol (NP) has been completed. The expression of 186 miRNAs is significantly distinct from that of the control group. In addition, it has been proven that miR-135a* can mediate the generation of reactive oxygen species (ROS) by regulating the Wnt/beta-catenin signalling pathway (Choi *et al.*, 2011). In 2018, a study showing the regulation of microRNA signalling by doxorubicin in LCs and SCs was also reported (Akinjo *et al.*, 2018).

Long noncoding RNAs

LncRNAs are longer than 200 nucleotides, with tissue- or cell-type specificity and without protein-coding capacity (Derrien et al., 2012). Large numbers of lncRNAs have been identified, however few of these have been deeply explored (Ma L et al., 2015; Reon et al., 2016; Liu et al., 2019). The main functions of lncRNAs were summarized as mediators of nuclear trafficking, altering splicing, changing mRNA stability and translation at both the transcriptional and post-transcriptional levels (Ayupe et al., 2015). For post-transcriptional regulation, lncRNAs also organized protein complexes to influence cell signalling and regulate allosteric proteins (Geisler and Coller, 2013). The Catsper1 gene, expressed in male germ cells, is essential for sperm motility and fertilization. Evidence suggests that the promoter of the Catsper1 gene can regulate a new gene, named Catsperlau (Catsperl antisense upstream transcript). Analysis of the whole genome sequence revealed that Catsper1au has 1402 bp and is a polyadenylated lncRNA with no intron. It is found in the nucleus of SCs and germ cells of adult male mouse testis as well as in LCs, suggesting that it might have an effect on spermatogenesis and male fertility (Jimenez-Badillo et al., 2017). Nevertheless, the detailed mechanism is still unclear and remains to be explored. Notably, many lncRNAs have been found in SCs, but their specific targets are also unknown (Yang et al., 2018).

PiRNA-like RNAs

PIWI-interacting RNAs (piRNAs) are small noncoding RNAs that are exclusively expressed in the germ cells of mammalian gonads. They have been regarded for a long time as germ-cell-specific small RNAs. The piRNAs are categorized into repetitive sequencederived piRNAs and non-repetitive sequence-derived piRNAs (Unhavaithaya *et al.*, 2009). The former silences transposons by DNA methylation and the latter is intergenic or intragenic and some have other non-repetitive regions with relatively unclear functions (Aravin *et al.*, 2007; Carmell *et al.*, 2007). However, in 2014, a novel class of somatic small RNAs, which were similar to piRNAs, were detected in somatic cells and named piRNA-like RNAs (pilRNAs). The pilRNAs have a distinct ping–pong signature and might target mRNAs 3'UTRs in a unique and complementary way (Ortogero *et al.*, 2014). These data provided one possibility that pilRNAs, whose functions are similar to those of germ cell piRNAs in SCs, might play a role in male fertility.

Leydig cells

Androgen greatly influences male health. Lack of testosterone can influence general health in males, such as by downregulating bone density, impairing muscle mass, injuring cognitive function and damaging immunity (Huhtaniemi, 2014). As males age, serum testosterone levels decrease gradually following an increased in serum follicle stimulating hormone (FSH) levels that either enhance or do not affect LH levels, indicating that the loss of testosterone results from changes in the ability of LCs to respond to LH. The loss is at the gonadal level rather than the hypothalamic-pituitary level (Wang et al., 2017). Many chronic and age-related clinical symptoms are associated with a low level of androgen, including cardiovascular diseases, obesity and metabolic syndrome (Kupelian et al., 2006; Saad and Gooren, 2009; Kloner et al., 2016). Testosterone is mainly produced by LCs. It plays a critical role in maintaining secondary sexual characteristics and spermatogenesis regulation in adults (Matzkin et al., 2013).

The LCs are divided into fetal LCs and postnatal LCs. The LH and hypothalamic–pituitary–gonadal axis regulates testosterone synthesis in adult interstitial LCs (Huhtaniemi I, 2015). However, fetal LCs produce androstenedione instead of testosterone directly without the presence of 17β -hydroxysteroid dehydrogenase-type 3 and androstenedione is converted into testosterone by fetal SCs (Shima *et al.*, 2013). Recent studies have shown that large numbers of ncRNAs are also expressed in LCs, Here, we summarized the findings of ncRNA function in LCs.

MicroRNAs

MicroRNAs related to LC steroidogenesis

Basic fibroblast growth factors (bFGF), including acidic and basic fibroblast growth factors, play diverse and specific roles in specific stages of LC steroidogenesis (Laslett et al., 1997). It was reported that five miRNAs (miR-29a, miR-29c, miR-142-3p, miR-451 and miR-335) are regulated by both bFGF and LH and are involved in the regulation of androgen production in immature LCs (Liu et al., 2014). miR-142-3p plays a vital role in cAMP production and PKA biological function to influence the cAMP/PKA signalling cascade, a secondary messenger pathway for steroid synthesis (Huang et al., 2009; Manna et al., 2014). Scavenger receptor class B type I (SR-BI), a HDL (high-density lipoprotein) receptor, is essential for the selective uptake of HDL CEs (cholesteryl esters) in steroidogenic cells (Shen et al., 2018). The expression of SR-BI and the selective uptake of HDL CEs were inhibited after the transfection of pre-miRNA-125a and pre-miRNA-455 in LCs, implying that miRNA-125a and miRNA-455 also play roles in steroidogenesis. Evidence has shown that miRNA-125a and miRNA-455 can bind to the 3'UTR of the SR-BI gene and negatively regulate SR-BI functions in rat steroidogenic cells. The two miRNAs were sensitive to changes in trophic hormones (ACTH or gonadotropin) and cAMP (Hu et al., 2012). In addition, treatment with Bt2cAMP increased the levels of miRNA-96, miRNA-132, miRNA-182 miRNA-183 and miRNA-212 and decreased the expression levels of miRNA-19a and miRNA-138 in MLTC-1 cells. All of these miRNAs can be found in the adrenal glands and are sensitive hormones similar to ACTH. In addition, miRNA-132 and miRNA-214 could inhibit

the expression of SREBP-1c and LDLR by combining with the 3'UTR of SREBP-1c and LDLR respectively (Hu *et al.*, 2013).

MicroRNAs associated with Leydig cell development

miR-140-3p was the most highly expressed miRNA expressed in a sexually dimorphic pattern, while the expression level of miR-140-5p in the testis was low. Evidence showed that the absence of miR-140-5p/miR-140-3p increased the number of mouse LCs, indicating that miR-140-3p and miR-140-5p might be related to the development of gonad as well as testis differentiation in mice (Rakoczy *et al.*, 2013). As explained above, miR-136 and miR-26a played roles in the immune microenvironment, which is crucial to LC development (Qin *et al.*, 2019).

Other findings of microRNAs in Leydig cells

Currently, in the zearalenone (ZEN)-exposed TM3 LC line, the analysis of miRNAs expression has been completed. Approximately 197 miRNAs were found to be significantly distinct from those of the control group. The predicted target genes participate in many signalling pathways, suggesting that ZEN, an important environmental pollutant, is regulated by miRNAs in LCs.(Wang M *et al.*, 2019)

Long noncoding RNAs

In 2018, in total, 33,883 lncRNAs were identified from sheep testes. The sheep LCs with knocked out lncRNA TCONS_00863147 expressed lower levels of *PRKCD* (*protein kinase C,delta*), indicating that the *PRKCD* could interact with lncRNA TCONS_00863147 in a trans-activation mechanism and then played a role in spermatogenesis (Yang *et al.*, 2018). This study also revealed a large number of lncRNAs in the LCs, however their specific roles in male fertility remain unknown.

Tesra, a novel testis-specific lncRNA in mice, has been proved to be present in germ cells and the cytoplasm of LCs, as shown by *in situ* hybridization. *Tesra* activated *Prss42/Tessp-2* gene expression by binding to the *Prss42/Tessp-2* promoter and then enhancing promoter activity. *Prss42/Tessp-2* played important roles in the progression of meiosis as well as in germ cell survival. It was found that *Tesra*, similar to other lncRNAs, might recruit histone modification enzymes or transcription factors such as *GClnc1* to the *Prss42/Tessp-2* promoter region. However the specific mechanism of *Tesra* in LCs is still unknown (Satoh *et al.*, 2019).

Natural antisense transcripts

Natural antisense transcripts (NAT) are RNA sequences that complement a sense transcript and either encode a protein or do not encode a protein (Balbin *et al.*, 2015; Latge *et al.*, 2018). In fact, many NATs were mistakenly regarded as lncRNAs (Latge *et al.*, 2018). Similar to lncRNAs, NAT expression was regulated by promoters and enhancers. Notably, their sense genes or the neighbouring genes are closely connected to their expression levels (Lin *et al.*, 2015). Growing evidence implicates NATs as participants with a unique mechanism of action in gene expression (Pelechano and Steinmetz, 2013; Nishizawa *et al.*, 2015; Latge *et al.*, 2018).

Translocator protein (Tspo), with rate-limiting step activity in steroidogenesis in LC steroidogenesis, can transport cholesterol into mitochondria (Chung *et al.*, 2013). Evidence showed that the expression of the *Tspo gene* and its function in steroidogenesis were regulated by a NAT that was specific for *Tspo* (*Tspo*-NAT) in LCs (Fan and Papadopoulos, 2012). The extension of the SINE (short interspersed repetitive element) B2 element-mediated transcript formed *Tspo*-NAT in mouse tumour LCs. It has been proven

that endogenous *Tspo*-NAT was more likely to suppress endogenous Tspo levels. In addition, the evidence also revealed that the expression of *Tspo*-NAT was regulated by cAMP and in this way maintained Tspo at a proper level for optimal LCs steroid production (Fan and Papadopoulos, 2012).

The steroidogenic acute regulatory (StAR) protein is a key protein that transports cholesterol located in mitochondria from outer membrane to the inner membrane (Manna *et al.*, 2013). *StAR* NAT was complementary to the spliced StAR sense 3.5-kb transcript and was highly expressed in LC and steroidogenic tissues. Evidence has shown that the *StAR* RNAs sense strands and the *StAR* RNAs antisense strands might be regulated in coordination as they were both expressed in the same cells. It has been proven that *StAR* NAT could downregulate the expression of StAR protein, as well as progesterone, by regulating cAMP (Castillo *et al.*, 2011). In this way, *StAR* RNAs play a role in regulating steroid biosynthesis.

Conclusion

With the development of large-scale genomic technologies and bioinformatics analyses, an increasing number of ncRNAs have been identified in SCs and LCs. Noncoding RNAs especially miRNAs including Dicer-dependent and Dicer-independent miRNAs in SCs play key roles in phagocytosis, immunoprotection and SCs development. These miRNAs are essential for the junction of BTB which maintains the testicular microenvironment for spermatogenesis LC ncRNAs are involved in steroidogenesis and the production of testosterone as well as development of LCs. Many miRNAs directly target genes involved in steroidogenesis and many of these are regulated by cAMP. Recently several novel lncRNAs such as Tesra have been identified. In addition, the discovery of NATs has provided another prospect for the regulation of gene expression. Compared with the vast number of ncRNAs in somatic cells, the numbers of current functional studies are exploring only the tip of the iceberg. In addition, the discovery of pilRNAs, which are similar to piRNAs and are present in somatic cells, open new horizons for researchers.

Acknowledgements. The author thank members of the Basic Medical College of Nanchang University, China for their help.

Author contribution. Chunjie Li designed and wrote the review. Baiqi Chen assisted in writing the manuscript. Jing Wang carefully revised the article.

Financial support. This study was supported by National Natural Science Foundation of China (no. 81660332), Natural Science Foundation of Jiangxi province (no. 20151BAB205057) and Health and Family Planning Project of Jiangxi province (no. 20155634).

Conflicts of interest. The authors have no conflicts of interest.

Ethical standards. Not applicable.

References

- Akinjo OO, Gant TW and Marczylo EL (2018) Perturbation of microRNA signalling by doxoRubiconin in spermatogonial, Leydig and Sertoli cell lines *in vitro. Toxicol Res* 7, 760–70.
- Aravin AA, Sachidanandam R, Girard A, Fejes-Toth K and Hannon GJ (2007) Developmentally regulated piRNA clusters implicate MILI in transposon control. *Science* 316(5825), 744–7.
- Ayupe AC, Tahira AC, Camargo L, Beckedorff FC, Verjovski-Almeida S and Reis EM (2015) Global analysis of biogenesis, stability and sub-cellular localization of lncRNAs mapping to intragenic regions of the human genome. *RNA Biol* **12**, 877–92.

- Balbin OA, Malik R, Dhanasekaran SM, Prensner JR, Cao X, Wu YM, Robinson D, Wang R, Chen G, Beer DG et al. (2015) The landscape of antisense gene expression in human cancers. *Genome Res* 25, 1068–79.
- Bernstein E, Caudy AA, Hammond SM and Hannon GJ (2001) Role for a bidentate ribonuclease in the initiation step of RNA interference. *Nature* 409(6818), 363–6.
- Birney E, Stamatoyannopoulos JA, Dutta A, Guigo R, Gingeras TR, Margulies EH, Weng Z, Snyder M, Dermitzakis ET, Thurman RE *et al.* (2007) Identification and analysis of functional elements in 1% of the human genome by the ENCODE pilot project. *Nature* 447(7146), 799–816.
- Bjork JK, Sandqvist A, Elsing AN, Kotaja N and Sistonen L (2010) miR-18, a member of Oncomir-1, targets heat shock transcription factor 2 in spermatogenesis. *Development* 137, 3177–84.
- Brosnan CA and Voinnet O (2009) The long and the short of noncoding RNAs. *Curr Opin Cell Biol* **21**, 416–25.
- Carmell MA, Girard A, van de Kant HJ, Bourc'his D, Bestor TH, de Rooij DG and Hannon GJ (2007) MIWI2 is essential for spermatogenesis and repression of transposons in the mouse male germline. Dev Cell 12, 503–14.
- Castillo AF, Fan J, Papadopoulos V and Podesta EJ (2011) Hormone-dependent expression of a steroidogenic acute regulatory protein natural antisense transcript in MA-10 mouse tumor Leydig cells. *PLoS One* **6**, e22822.
- Chen Y, Zhou Y, Wang J, Wang L, Xiang Z, Li D and Han X (2016) Microcystin-leucine arginine causes cytotoxic effects in Sertoli cells resulting in reproductive dysfunction in male mice. *Sci Rep* **6**, 39238.
- Cheng CY and Mruk DD (2012) The blood-testis barrier and its implications for male contraception. *Pharmacol Rev* 64, 16–64.
- Choi JS, Oh JH, Park HJ, Choi MS, Park SM, Kang SJ, Oh MJ, Kim SJ, Hwang SY and Yoon S (2011) miRNA regulation of cytotoxic effects in mouse Sertoli cells exposed to nonylphenol. *Reprod Biol Endocrinol* 9, 126.
- Chung JY, Chen H, Midzak A, Burnett AL, Papadopoulos V and Zirkin BR (2013) Drug ligand-induced activation of translocator protein (TSPO) stimulates steroid production by aged brown Norway rat Leydig cells. *Endocrinology* 154, 2156–65.
- Dabaja AA, Mielnik A, Robinson BD, Wosnitzer MS, Schlegel PN and Paduch DA (2015) Possible germ cell-Sertoli cell interactions are critical for establishing appropriate expression levels for the Sertoli cell-specific microRNA, miR-202-5p, in human testis. *Basic Clin Androl* 25, 2.
- de Mateo S and Sassone-Corsi P (2014) Regulation of spermatogenesis by small noncoding RNAs: role of the germ granule. Semin Cell Dev Biol 29, 84–92.
- Derrien T, Johnson R, Bussotti G, Tanzer A, Djebali S, Tilgner H, Guernec G, Martin D, Merkel A, Knowles DG et al. (2012) The GENCODE v7 catalog of human long noncoding RNAs: analysis of their gene structure, evolution and expression. *Genome Res* 22, 1775–89.
- Ebbesen KK, Kjems J and Hansen TB (2016) Circular RNAs: identification, biogenesis and function. *Biochim Biophys Acta* 1859, 163–8.
- The ENCODE Project Consortium (2004) The ENCODE (ENCyclopedia Of DNA Elements) Project. *Science* **306**(5696), 636–40.
- The ENCODE Project Consortium (2012) An integrated encyclopedia of DNA elements in the human genome (2012) *Nature* **489**(7414), 57–74.
- Fan J and Papadopoulos V (2012) Transcriptional regulation of translocator protein (Tspo) via a SINE B2-mediated natural antisense transcript in MA-10 Leydig cells. *Biol Reprod* 86, 147.
- Geisler S and Coller J (2013) RNA in unexpected places: long noncoding RNA functions in diverse cellular contexts. Nat Rev Mol Cell Biol 14, 699–712.
- Gu H, Wu W, Yuan B, Tang Q, Guo D, Chen Y, Xia Y, Hu L, Chen D, Sha J et al. (2017) Genistein up-regulates miR-20a to disrupt spermatogenesis via targeting Limk1. Oncotarget 8, 58728–37.
- Guan D, Zhang W, Zhang W, Liu GH and Belmonte JC (2013) Switching cell fate, ncRNAs coming to play. *Cell Death Dis* **4**, e464.
- Guo J, Liu X, Yang Y, Liang M, Bai C, Zhao Z and Sun B (2018) miR-375 down-regulation of the rearranged L-myc fusion and hypoxia-induced gene domain protein 1A genes and effects on Sertoli cell proliferation. *Asian Australas J Anim Sci* **31**, 1103–9.
- Hai Y, Hou J, Liu Y, Liu Y, Yang H, Li Z and He Z (2014) The roles and regulation of Sertoli cells in fate determinations of spermatogonial stem cells and spermatogenesis. *Semin Cell Dev Biol* 29, 66–75.

- Hu Z, Shen WJ, Kraemer FB and Azhar S (2012) MicroRNAs 125a and 455 repress lipoprotein-supported steroidogenesis by targeting scavenger receptor class B type I in steroidogenic cells. *Mol Cell Biol* **32**, 5035–45.
- Hu Z, Shen WJ, Cortez Y, Tang X, Liu LF, Kraemer FB and Azhar S (2013) Hormonal regulation of microRNA expression in steroid producing cells of the ovary, testis and adrenal gland. *PLoS One* 8, e78040.
- Huang B, Zhao J, Lei Z, Shen S, Li D, Shen GX, Zhang GM and Feng ZH (2009) miR-142-3p restricts cAMP production in CD4⁺CD25⁻ T cells and CD4⁺CD25⁺ Treg cells by targeting AC9 mRNA. *EMBO Rep* **10**, 180–5.
- Huhtaniemi IT (2014) Andropause—lessons from the European male ageing study. Ann Endocrinol 75, 128–31.
- Huhtaniemi I (2015) A short evolutionary history of FSH-stimulated spermatogenesis. *Hormones* 14, 468–78.
- Jan SZ, Hamer G, Repping S, de Rooij DG, van Pelt AM and Vormer TL (2012) Molecular control of rodent spermatogenesis. *Biochim Biophys* Acta 1822, 1838–50.
- Jimenez-Badillo SE, Oviedo N, Hernandez-Guzman C, Gonzalez-Mariscal L and Hernandez-Sanchez J (2017) Catsper1 promoter is bidirectional and regulates the expression of a novel lncRNA. *Sci Rep* 7, 13351.
- Jovanovic M and Hengartner MO (2006) miRNAs and apoptosis: RNAs to die for. Oncogene 25, 6176–87.
- Kapranov P, Cheng J, Dike S, Nix DA, Duttagupta R, Willingham AT, Stadler PF, Hertel J, Hackermuller J, Hofacker IL *et al.* (2007) RNA maps reveal new RNA classes and a possible function for pervasive transcription. *Science* **316**(5830), 1484–8.
- Kim VN (2005) MicroRNA biogenesis: coordinated cropping and dicing. Nat Rev Mol Cell Biol 6, 376–85.
- Kloner RA, Carson C, 3rd, Dobs A, Kopecky S and Mohler ER, 3rd (2016) Testosterone and cardiovascular disease J Am Coll Cardiol 67, 545–57.
- Krol J, Loedige I and Filipowicz W (2010) The widespread regulation of microRNA biogenesis, function and decay. Nat Rev Genet 11, 597–610.
- Kupelian V, Page ST, Araújo AB, Travison TG, Bremner WJ and McKinlay JB (2006) Low sex hormone-binding globulin, total testosterone and symptomatic androgen deficiency are associated with development of the metabolic syndrome in nonobese men. J Clin Endocrinol Metab 91, 843–50.
- Laslett AL, McFarlane JR and Risbridger GP (1997) Developmental response by Leydig cells to acidic and basic fibroblast growth factor. J Steroid Biochem Mol Biol 60(3–4), 171–9.
- Latge G, Poulet C, Bours V, Josse C and Jerusalem G (2018) Natural antisense transcripts: molecular mechanisms and implications in breast cancers. *Int J Mol Sci* 19, pii: E123.
- Lian C, Sun B, Niu S, Yang R, Liu B, Lu C, Meng J, Qiu Z, Zhang L and Zhao Z (2012) A comparative profile of the microRNA transcriptome in immature and mature porcine testes using Solexa deep sequencing. *FEBS J* **279**, 964–75.
- Lin S, Zhang L, Luo W and Zhang X (2015) Characteristics of antisense transcript promoters and the regulation of their activity. *Int J Mol Sci* **17**, 9.
- Liu H, Yang Y, Zhang L, Liang R, Ge RS, Zhang Y, Zhang Q, Xiang Q, Huang Y and Su Z (2014) Basic fibroblast growth factor promotes stem Leydig cell development and inhibits LH-stimulated androgen production by regulating microRNA expression. J Steroid Biochem Mol Biol 144(Pt B), 483–91.
- Liu H, Wang R, Mao B, Zhao B and Wang J (2019) Identification of lncRNAs involved in rice ovule development and female gametophyte abortion by genome-wide screening and functional analysis. *BMC Genome* 20, 90.
- Lucas K and Raikhel AS (2013) Insect microRNAs: biogenesis, expression profiling and biological functions. *Insect Biochem Mol Biol* 43, 24–38.
- Lui WY, Lee WM and Cheng CY (2003) Sertoli-germ cell adherens junction dynamics in the testis are regulated by RhoB GTPase via the ROCK/ LIMK signalling pathway. *Biol Reprod* 68, 2189–206.
- Ma C, Song H, Yu L, Guan K, Hu P, Li Y, Xia X, Li J, Jiang S and Li F (2016) miR-762 promotes porcine immature Sertoli cell growth via the ring finger protein 4 (RNF4) gene. *Sci Rep* 6, 32783.
- Ma L, Li A, Zou D, Xu X, Xia L, Yu J, Bajic VB and Zhang Z (2015) LncRNAWiki: harnessing community knowledge in collaborative curation of human long noncoding RNAs. *Nucl Acids Res* **43**, D187–92.
- Manna PR, Cohen-Tannoudji J, Counis R, Garner CW, Huhtaniemi I, Kraemer FB and Stocco DM (2013) Mechanisms of action of hormonesensitive lipase in mouse Leydig cells: its role in the regulation of the steroidogenic acute regulatory protein. J Biol Chem 288, 8505–18.

- Manna PR, Slominski AT, King SR, Stetson CL and Stocco DM (2014) Synergistic activation of steroidogenic acute regulatory protein expression and steroid biosynthesis by retinoids: involvement of cAMP/PKA signalling. *Endocrinology* **155**, 576–91.
- Matzkin ME, Yamashita S and Ascoli M (2013) The ERK1/2 pathway regulates testosterone synthesis by coordinately regulating the expression of steroidogenic genes in Leydig cells. *Mol Cell Endocrinol* **370**(1–2), 130–7.
- McCabe MJ, Tarulli GA, Laven-Law G, Matthiesson KL, Meachem SJ, McLachlan RI, Dinger ME and Stanton PG (2016) Gonadotropin suppression in men leads to a reduction in claudin-11 at the Sertoli cell tight junction. *Hum Reprod* **31**, 875–86.
- Neph S, Vierstra J, Stergachis AB, Reynolds AP, Haugen E, Vernot B, Thurman RE, John S, Sandstrom R, Johnson AK *et al.* (2012) An expansive human regulatory lexicon encoded in transcription factor footprints. *Nature* 489(7414), 83–90.
- Ng WL, Marinov GK, Liau ES, Lam YL, Lim YY and Ea CK. (2016) Inducible RasGEF1B circular RNA is a positive regulator of ICAM-1 in the TLR4/LPS pathway. *RNA Biol* **13**, 861–71.
- Nishizawa M, Ikeya Y, Okumura T and Kimura T (2015) Post-transcriptional inducible gene regulation by natural antisense RNA. *Front Biosci* **20**, 1–36.
- Niu Z, Goodyear SM, Rao S, Wu X, Tobias JW, Avarbock MR and Brinster RL (2011) MicroRNA-21 regulates the self-renewal of mouse spermatogonial stem cells. *Proc Natl Acad Sci USA* 108, 12740–5.
- Oatley JM and Brinster RL (2012) The germline stem cell niche unit in mammalian testes. *Phys Rev* **92**, 577–95.
- Ohno S (1972) So much 'junk' DNA in our genome. *Brookhaven Symp Biol* 23, 366–70.
- Ortogero N, Hennig GW, Langille C, Ro S, McCarrey JR and Yan W (2013) Computer-assisted annotation of murine Sertoli cell small RNA transcriptome. *Biol Reprod* 88, 3.
- Ortogero N, Schuster AS, Oliver DK, Riordan CR, Hong AS, Hennig GW, Luong D, Báo J, Bhetwal BP, Ro S et al. (2014) A novel class of somatic small RNAs similar to germ cell pachytene PIWI-interacting small RNAs. J Biol Chem 289, 32824–34.
- Panneerdoss S, Viswanadhapalli S, Abdelfattah N, Onyeagucha BC, Timilsina S, Mohammad TA, Chen Y, Drake M, Vuori K, Kumar TR et al. (2017) Crosstalk between miR-471-5p and autophagy component proteins regulates LC3associated phagocytosis (LAP) of apoptotic germ cells. Nat Commun 8, 598.
- Papaioannou MD, Pitetti JL, Ro S, Park C, Aubry F, Schaad O, Vejnar CE, Kuhne F, Descombes P, Zdobnov EM et al. (2009) Sertoli cell Dicer is essential for spermatogenesis in mice. *Dev Biol* 326, 250–9.
- Papaioannou MD, Lagarrigue M, Vejnar CE, Rolland AD, Kuhne F, Aubry F, Schaad O, Fort A, Descombes P, Neerman-Arbez M et al. (2011) Loss of Dicer in Sertoli cells has a major impact on the testicular proteome of mice. *Mol Cell Proteom* 10, M900587mcp900200.
- Patil VS, Zhou R and Rana TM (2014) Gene regulation by noncoding RNAs. Crit Rev Biochem Mol Biol 49, 16–32.
- Pelechano V and Steinmetz LM (2013) Gene regulation by antisense transcription. Nat Rev Genet 14, 880–93.
- Qin L, Lin J and Xie X (2019) CircRNA-9119 suppresses poly I:C induced inflammation in Leydig and Sertoli cells via TLR3 and RIG-I signal pathways. *Mol Med* 25, 28.
- Rakoczy J, Fernandez-Valverde SL, Glazov EA, Wainwright EN, Sato T, Takada S, Combes AN, Korbie DJ, Miller D, Grimmond SM et al. (2013) MicroRNAs-140-5p/140-3p modulate Leydig cell numbers in the developing mouse testis. *Biol Reprod* 88, 143.
- Reon BJ, Anaya J, Zhang Y, Mandell J, Purow B, Abounader R and Dutta A (2016) Expression of lncRNAs in low-grade gliomas and glioblastoma multiforme: an *in silico* analysis. *PLoS Med* **13**, e1002192.
- Rybak-Wolf A, Stottmeister C, Glazar P, Jens M, Pino N, Giusti S, Hanan M, Behm M, Bartok O, Ashwal-Fluss R et al. (2015) Circular RNAs in the mammalian brain are highly abundant, conserved and dynamically expressed. Mol Cell 58, 870–85.

- Saad F and Gooren L (2009) The role of testosterone in the metabolic syndrome: a review. J Steroid Biochem Mol Biol 114(1–2), 40–3.
- Salviano-Silva A, Lobo-Alves SC, Almeida RC, Malheiros D and Petzl-Erler ML (2018) Besides pathology: long noncoding RNA in cell and tissue homeostasis. *Noncoding RNA* 4(1), E3.
- Satoh Y, Takei N, Kawamura S, Takahashi N, Kotani T and Kimura AP (2019) A novel testis-specific long noncoding RNA, Tesra, activates the Prss42/Tessp-2 gene during mouse spermatogenesis dagger. *Biol Reprod* 100, 833–48.
- Setchell BP (2008) Blood-testis barrier, junctional and transport proteins and spermatogenesis. Adv Exp Med Biol 636, 212–33.
- Shen WJ, Azhar S and Kraemer FB (2018) SR-B1: a unique multifunctional receptor for cholesterol influx and efflux. Ann Rev Physiol 80, 95–116.
- Shima Y, Miyabayashi K, Haraguchi S, Arakawa T, Otake H, Baba T, Matsuzaki S, Shishido Y, Akiyama H, Tachibana T et al. (2013) Contribution of Leydig and Sertoli cells to testosterone production in mouse fetal testes. *Mol Endocrinol* 27, 63–73.
- Tay Y, Zhang J, Thomson AM, Lim B and Rigoutsos I (2008) MicroRNAs to Nanog, Oct4 and Sox2 coding regions modulate embryonic stem cell differentiation. *Nature* 455(7216), 1124–8.
- Tong MH, Mitchell DA, McGowan SD, Evanoff R and Griswold MD (2012) Two miRNA clusters, miR-17-92 (miRc1) and miR-106b-25 (miRc3), are involved in the regulation of spermatogonial differentiation in mice. *Biol Reprod* **86**, 72.
- Unhavaithaya Y, Hao Y, Beyret E, Yin H, Kuramochi-Miyagawa S, Nakano T and Lin H (2009) MILI, a PIWI-interacting RNA-binding protein, is required for germ line stem cell self-renewal and appears to positively regulate translation. *J Biol Chem* **284**, 6507–19.
- Wainwright EN, Jorgensen JS, Kim Y, Truong V, Bagheri-Fam S, Davidson T, Svingen T, Fernandez-Valverde SL, McClelland KS, Taft RJ et al. (2013) SOX9 regulates microRNA miR-202-5p/3p expression during mouse testis differentiation. *Biol Reprod* 89, 34.
- Wang H, Wang H, Xiong W, Chen Y, Ma Q, Ma J, Ge Y and Han D (2006) Evaluation on the phagocytosis of apoptotic spermatogenic cells by Sertoli cells *in vitro* through detecting lipid droplet formation by Oil Red O staining. *Reproduction* **132**, 485–92.
- Wang M, Wu W, Li L, He J, Huang S, Chen S, Chen J, Long M, Yang S and Li P (2019) Analysis of the miRNA expression profiles in the zearalenoneexposed TM3 Leydig cell line. *Int J Mol Sci* 20, 635.
- Wang S, Tang Y, Cui H, Zhao X, Luo X, Pan W, Huang X and Shen N (2011) Let-7/miR-98 regulate Fas and Fas-mediated apoptosis. *Genes Immun* 12, 149–54.
- Wang Y, Chen F, Ye L, Zirkin B and Chen H (2017) Steroidogenesis in Leydig cells: effects of aging and environmental factors. *Reproduction* 154, R111–22.
- Wong CH, Cheng CY (2005) The blood-testis barrier: its biology, regulation and physiological role in spermatogenesis. *Curr Topics Dev Biol* 71, 263–96.
- Yang C, Yao C, Tian R, Zhu Z, Zhao L, Li P, Chen H, Huang Y, Zhi E, Gong Y et al. (2019) miR-202–3p regulates Sertoli cell proliferation, synthesis function and apoptosis by targeting LRP6 and cyclin D1 of Wnt/beta-catenin signaling. Mol Ther Nucl Acids 14, 1–19.
- Yang H, Wang F, Li F, Ren C, Pang J, Wan Y, Wang Z, Feng X and Zhang Y (2018) Comprehensive analysis of long noncoding RNA and mRNA expression patterns in sheep testicular maturation. *Biol Reprod* **99**, 650–61.
- Yao C, Sun M, Yuan Q, Niu M, Chen Z, Hou J, Wang H, Wen L, Liu Y, Li Z et al. (2016) miRNA-133b promotes the proliferation of human Sertoli cells through targeting GLI3. Oncotarget 7, 2201–19.
- Zhang X, Zhao W, Li C, Yu H, Qiao Y, Li A, Lu C, Zhao Z and Sun B (2015) Differential expression of miR-34c and its predicted target genes in testicular tissue at different development stages of swine. *Asian Australas J Anim Sci* 28, 1532–6.
- Zhang Z, Shao S and Meistrich ML (2007) The radiation-induced block in spermatogonial differentiation is due to damage to the somatic environment, not the germ cells. J Cell Physiol 211, 149–58.