# Movement and activity patterns of the nurse shark, *Ginglymostoma cirratum*, in an oceanic Marine Protected Area of the South-western Atlantic

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The movement and activity patterns of the nurse shark, Ginglymostoma cirratum, a vulnerable species off Brazil, were investigated using mark-recapture and acoustic telemetry at an oceanic insular Marine Protected Area, the Fernando de Noronha Archipelago, Brazil. A total of 93 sharks were captured and tagged, ranging from 82 to 265 cm of total length (TL). Nurse sharks were captured throughout the year, and all life-stages used the insular shelf. Fifteen sharks (16% of the total) were recaptured after periods at liberty ranging from 3.5 h to 705 days, and the distances between tag and recapture locations ranged from 0.07 to 3.5 km. Site fidelity and movements of 10 sharks ranging from 107 to 265 cm TL were investigated for 18 months with an array of automated telemetry receivers. The mean period of detection of the monitored sharks was 66 days, ranging from 13 to 119 days. One individual 158 cm TL was monitored with active tracking for 17 days, with distances between daily locations ranging from 0.84 to 3.32 km, exhibiting movements similar to those of sharks monitored by automated telemetry. Despite remaining motionless or exhibiting short range movements for several hours or days, nurse sharks can be relatively wide-ranging, and protected areas alone cannot be the only conservation measure used to protect this species, which requires a set of protective measures, including fisheries management.

Keywords: Elasmobranch, mark-and-recapture, acoustic telemetry, Ginglymostomatidae, endangered species, Brazil

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## INTRODUCTION

The nurse shark, *Ginglymostoma cirratum* (Bonnaterre, 1788) is a large (at least 300 cm total length (TL)), bottom-dwelling species that inhabits continental and insular shelves along both sides of the tropical and subtropical Atlantic Ocean, and also considered to occur in the eastern Pacific Ocean until 2015 (Compagno, 2001). However, a recent study described the Pacific Ocean population as a distinct species, *Ginglymostoma unami* (Moral-Flores *et al.*, 2015) while *G. cirratum* is currently regarded to be restricted to the Atlantic Ocean (Moral-Flores *et al.*, 2015).

The nurse shark has been reported as one of the most abundant shark species in the coastal shallow waters off Florida, the Caribbean (Castro, 2000; Heithaus *et al.*, 2007) and north-eastern Brazil (Hazin *et al.*, 2000; Castro & Rosa, 2005; Afonso *et al.*, 2014). The species is also popular for ecotouristic viewing by divers, particularly off Florida, the Bahamas and the Caribbean, and it is kept for public display in aquariums due to its robustness and ability to survive many years in captivity (Compagno, 2001). Although *G. cirratum* is commonly found and fished in inshore waters

Corresponding author: R.C. Garla Email: rgarla@hotmail.com throughout its range, fisheries data on the species is scarce, probably due to the low commercial value of its meat and fins (Castro *et al.*, 1999; Castro, 2000), perhaps a reason that the species is listed as 'Data Deficient' by the IUCN Red List (Rosa *et al.*, 2006). Nevertheless, the western Atlantic population of *G. cirratum* is regarded as 'Near Threatened' (Rosa *et al.*, 2006) and 'Vulnerable' in Brazil following IUCN criteria, where abundance decline and regional extirpations have been reported (Rosa & Gadig, 2009).

Despite its distribution in shallow coastal waters and its value to the eco-tourism industry, the current knowledge of *G. cirratum* ecology is mainly limited to the western North Atlantic Ocean, where most studies have focused on mating behaviour (Klimley, 1980; Carrier *et al.*, 1994; Carrier & Pratt, 1998; Pratt & Carrier, 2001), growth (Carrier, 1985) and site fidelity and movements (Carrier & Luer, 1990; Chapman *et al.*, 2005). A review on *G. cirratum* biology can be found in Castro (2000).

Data regarding *G. cirratum* from the South Atlantic Ocean remain scarce, but recent studies have improved the knowledge about population structure and habitat use off northeastern Brazil (Castro & Rosa, 2005; Santander-Neto *et al.*, 2011; Ferreira *et al.*, 2013; Afonso *et al.*, 2014), as well as about the connectivity among western Atlantic populations (Karl *et al.*, 2011). However, given the current Vulnerable status of *G. cirratum* off Brazil (Rosa & Gadig, 2009), further research on the life history of this species is necessary to ensure effective management and give support to future conservation policies. In this sense, we herein report the results from traditional mark-recapture and acoustic telemetry studies aiming to obtain data on *G. cirratum* in terms of population structure, seasonal patterns of abundance, and description of movement and residence patterns in relation to Marine Protected Areas (MPA) boundaries.

## MATERIALS AND METHODS

## Study site

The study was conducted in the Fernando de Noronha Archipelago (FEN), an isolated group of volcanic islands located in the western tropical Atlantic, 345 km off the northeastern coast of Brazil ( $03^{\circ}51'S 32^{\circ}25'W$ ) (Figure 1A). FEN is under the influence of the South Equatorial Current, with annual mean seawater temperature and salinity of  $26^{\circ}C$  and 36 respectively. About 70% of the main island and the coastal waters spanning from the coastline to the 50 m isobath constitute a MPA (National Marine Park) since 1988 (Maida & Ferreira, 1997), where fishing is prohibited, and boat traffic and diving are regulated. The remaining portions of the main island coastline to the 50 m isobath constitute an Environmental Protection Area (EPA) designated for sustainable use (Figure 1B).

## Sampling

Fishing operations were performed in four tagging campaigns, from March to October 2000, in January and July 2001, from February to October 2002, and February 2003 at various locations around FEN, focusing mainly on the leeward side because it provides suitable oceanographic conditions most of the year. Handlines were used with 10/0 or 11/0 barbless circle hooks baited with pieces of locally abundant fishes (e.g. *Cephalopholis fulva, Caranx* and *Carangoides* spp.). Between June 2010 and January 2012, additional fishing sets



**Fig. 1.** (A) Fernando de Noronha Archipelago and its sustainable use (EPA) and marine protected (MPA) areas. (B) Location of the automated acoustic telemetry array moored at the lee side of the archipelago from July 2010 to January 2012. Receivers #6 and #8 were deployed on February 2011. R: indicates site of capture of sharks Nos 15 and 19 implanted with transmitters close to Rata Island.

were conducted but were restricted to waters 5-30 m in depth from the south-western portion of FEN's leeward side, which is included in the MPA. In addition, from June 2011 to January 2012, bottom longlines with 10 circle hooks 18/0 spaced every 25 m were deployed at both sides of FEN in depths ranging from 50 to 65 m. Soak times averaged 2.5 h.

The catch per unit of effort for handlines and longlines (CPUE = sharks captured/(number of hooks × time elapsed)) was calculated following Musick *et al.* (2005). For longlines the soak time was used as the time of fishing.

Captured sharks were brought alongside the boat, measured for fork length (FL) and total length (TL) to the nearest centimetre and sexed. Juveniles were fitted with numbered nylon-barbed tags (Hallprint, South Australia). Sharks larger than 150 cm TL were tagged with stainless-steel darttags (Floytag, USA). All lengths herein reported refer to TL unless when stated otherwise. The approximate life-stage of each shark was defined on basis of external morphology and the previous published data on the size at maturity (Castro, 2000). Juveniles were individuals that measured between 60 and 179 cm. Females between 180 and 222 cm TL were regarded as pre-adults and those larger than 223 cm TL as mature adults. Males between 180 and 213 cm TL were regarded as pre-adults and those larger than 214 cm TL, with fully calcified claspers, as adults.

#### Transmitter attachment and acoustic array

Sharks fitted with acoustic transmitters were caught with handlines from June 2010 to February 2011 at random locations of the south-western portion of leeward side inside the MPA (Figure 1B). After conducting the procedures described in the previous section, all sharks measuring less than 180 cm TL were brought on board and positioned with their ventral side facing upwards inside a PVC trough with a hose placed inside their mouths in order to pump seawater directly to the gills. Two specimens, one measuring 170 cm TL and the other 267 cm TL, were inverted alongside the boat. After periods ranging from 10 to 120 s, the inversion resulted in the onset of tonic immobility (Henningsen, 1994) allowing for relatively simple surgery for transmitter implantation. Coded acoustic transmitters (model CHP-87-L, 16 mm diameter  $\times$  80 mm, frequency 32–83 kHz, burst interval 45-60 s, lifespan 18 months; Sonotronics, USA) were implanted into the abdominal cavity of the animals through a 3-4 cm incision made in the abdominal wall anterior to the origin of the left pelvic fin. The incision was closed with absorbable nylon sutures. The whole procedure usually took 5-10 min. After surgery, sharks were re-inverted alongside the boat until they were able to swim autonomously and then released, typically after 2-5 min. Only sharks cleanly hooked in the mouth with no signs of significant injury were selected for transmitter implantation.

The presence of acoustically tagged sharks was continuously monitored from June 2010 to January 2012 by an overlapping linear array of eight underwater acoustic receivers (SUR-2, Sonotronics) deployed in the area where the sharks were tagged (Figure 1B). Receivers were moored directly to the substrate in depths between 10 to 31 m with shackles and chains and kept vertically stretched by a buoyed line. In all sites receivers were kept from 5 to 10 m above the bottom to inspect as much as possible of the bottom and water column. Preliminary field tests conducted both at night and daylight hours indicated that detection ranges varied from 400 to 500 m, so the receivers were moored  $\sim$ 400 from each other along a 5-km section of the leeward coast included in the MPA. The receivers were retrieved at 6-month intervals for data download and battery refurbishment.

The telemetry study was conducted over a 540 days period during which three temporal gaps occurred at site 3: the first and third gaps encompassed 101 days (between 14 October 2010 and 25 January 2011) and 64 days (11 November 2011 to 15 January 2012), respectively, and were caused by a great number of detections that took up the whole memory capacity of the receivers; the second gap encompassed 154 days (10 February to 15 July 2011) and was caused by malfunction of the receiver. At site 2 there was a gap of 153 days (9 February to 6 July 2011) also due to receiver malfunction. Receivers of both sites were replaced.

## Active tracking

One male juvenile nurse shark measuring 158 cm TL was captured with a handline and tracked with active acoustic telemetry to assess short-term, fine-scale movements. Upon capture, the shark was restrained, measured and sexed. A coded ultrasonic continuous transmitter (model CT-82-1-E, 16 mm diameter × 38 mm, lifespan 60 days, Sonotronics, USA) attached to a 3 cm plastic sheep tag (Allflex, USA) with a corrodible wire was fitted to the first dorsal fin of the shark following Cartamil et al. (2003). The handling time for measurement and tag attachment did not exceed 5 min. The shark was tracked for 8 h immediately after release from a 3.5 m skiff equipped with a portable directional hydrophone and receiver (models DH-4 and USR-96, Sonotronics, respectively). The geographic location of the shark was determined by positioning the vessel above the shark, which was verified when the strongest transmitter signal was oriented from underneath the vessel, and recording the position of the vessel every 15 min with a hand-held GPS. The shark was tracked daily for periods ranging from 4 to 11 h during 16 days. All recorded positions were plotted on a map, and the linear distances between successive positions and the activity space used during the tracking period were measured using Google Earth Pro 7.1.



Fig. 2. Length frequency distribution of *Ginglymostoma cirratum* caught at Fernando de Noronha Archipelago from March 2000 to February 2003 and from July 2010 to January 2012 (N = 93). Solid bars = males; open bars = females.

## Data analysis

Detections from all receivers were sorted by transmitter, date and receiver to generate a complete monitoring record for each individual. The total number of days over which each shark was detected in the array, the maximum number of consecutive days it was detected, the mean number of daily detections, and detection rates during diurnal and nocturnal periods were also assessed. Detection records were plotted against month to further investigate seasonal patterns of habitat use. The minimum dispersal range of each shark along the array was estimated by measuring the distance between the peripheries of the detection ranges of the two farthest receivers where the shark was detected. The activity space used by each shark within the array was also calculated. Measurements were made using Google Earth Pro 7.1.

Statistical analyses were performed with R software version 3.2.1, adopting a significance level of 5%. Data were log (x) transformed for shark total length and log transformed (x + 1) for CPUE to normalize and homogenize variances. Data normality was verified through Shapiro–Wilk test and data homoscedasticity with Bartlett test. One-way ANOVA were used to verify if shark length differed between seasons, sexes and sampling periods. Kruskal–Wallis tests were used to test for differences of CPUE between seasons, periods and fishing gear.

#### RESULTS

A total of 93 nurse sharks were captured and tagged during the two studied periods, including 81 juveniles, six pre-adults and six adults. Females ranged from 82 to 240 cm TL (N = 45, mean = 145.58, SD = 36.47) and males from 93 to 265 cm TL (N = 48, mean = 143.96, SD = 33.73) (Figure 2). There were no significant differences in mean TL between sampling periods (F = 0.07, df = 1, P = 0.932), sexes (F = 0.03, df = 1, P = 0.530).

Handline fishing at 232 sites over 5–30 m depth resulted in the capture of 87 *G. cirratum* at 44 of the sites (19%), with the CPUE per site ranging from 0 to 0.86 (mean = 0.04, SD = 0.11). There were no significant differences in the mean handline CPUE between sampling periods (Kruskal–Wallis = 1.093, P = 0.295), sexes (Kruskal–Wallis = 0.282, P =0.594) or seasons (Kruskal–Wallis = 0.202, P = 0.652). Longline fishing effort at 29 sites over 50 m depth resulted in the capture of six nurse sharks at five sites (17.2%), with the CPUE ranging from 0 to 0.1 (mean = 0.01, SD = 0.02).

Fifteen tagged *G. cirratum* (16% of 93 tagged animals) were recaptured after periods at liberty ranging from 3.5 h to 705 days (mean = 140.9 days, SD = 206.3 days; Figure 3). The distance between tag and recapture locations ranged from 0.07 to 1 km after 3.5 h and 705 days at liberty. One pre-adult female 201 cm TL was resigned during dives in a location distant 3.5 km from the tagging site on six different occasions after 53 to 362 days. Growth rates could be estimated for only four recaptured juveniles (Table 1), indicating a preliminary rate of 10.99  $\pm$  3.56 cm year<sup>-1</sup>.

Transmitters were implanted into four female and six male nurse sharks ranging from 107 to 265 cm TL (mean = 154.4 cm TL, SD = 46) (Table 2). Seven sharks were juveniles, and one was a mature male 265 cm TL with calcified claspers with sperm remnants. This adult male (No. 38) and two



Fig. 3. Recapture locations of *Ginglymostoma cirratum* at the Fernando de Noronha Archipelago (N = 20). 2000–2003. Stars ( $\star$ ) depict sharks that were recaptured at the same location of tagging. For all cases the associated numbers indicate the time, in days, at liberty since tagging and the estimated distance travelled (km). The solid circle ( $\bullet$ ) indicates multiple underwater sightings of a 201 cm TL female, and the arrow indicates the estimated distance travelled (km) between the tagging and sighting location. 2010–2012: Left panel: star ( $\star$ ) indicates the recapture of a shark at the release site 3.5 h after tagging; solid circle ( $\bullet$ ) indicate distance travelled (time at liberty and estimated distance travelled; solid triangles ( $\blacktriangle$ ) indicate two recaptures of the same individual; solid squares ( $\blacksquare$ ) indicate two recaptures of the same individual; solid squares ( $\blacksquare$ ) indicate the recapture of a shark tagged 10 days before and the arrow indicates the estimated distance travelled between the tagging and recapture location (km).

females (No. 15 and 19) were detected only once by receivers located 4 and 9 km, respectively, from their capture sites and were not included in further analyses.

Detection records of multiple individuals were concentrated at sites 2, 3, 5 and 6. In contrast, there were no detections from any shark on the receivers placed at the deepest site 4 (31 m) and at the north-eastern extremity of the array (site 8). Shark No. 1 was mainly detected by the receiver located at the site of its capture, while the remaining six sharks (Nos 11, 12, 16, 18 and 39) were detected at up to six neighbouring areas (Figure 4). Minimum linear distances travelled to neighbouring monitoring sites were conservatively estimated at 2.1 to 3.7 km and the maximum number of receivers visited per day was three. Detection records at the sites usually occurred throughout the diel cycle (Table 2).

There was evidence of emigration of *G. cirratum* away from FEN's south-western coastline. Sharks were continuously detected within the receiver array during 3 to 6 months, and

 Table 1. Growth rates of nurse sharks recaptured at Fernando de Noronha Archipelago.

TL (cm) at capture	TL (cm) at recapture	N days at large	Increment in size (cm)	Growth rate (cm year <sup>-1</sup> )
118	129	270	11	14.67
132	141	280	9	11.57
165	177	705	12	6.13
143	153	363	10	9.92

then returned after periods ranging from 3 to 8 months later. Figure 5 indicates a bimodal pattern of attendance at this part of the archipelago. The mean period of detection was 66 days and ranged from 13 to 119 days. The mean number of consecutive days a shark was detected by the array was 19, ranging from 2 to 51 days (Table 2). The mean activity spaces used within the array was 1.84 km<sup>2</sup>, ranging from 1.26 to 2.22 km<sup>2</sup> (Table 2).

The juvenile male 158 cm TL actively tracked was followed for periods of 4 to 11 h between 18 July and 4 August 2011, in total of 93 h (Figure 5). This shark was relocated daily until tracking sessions had to be prematurely terminated due to the failure of the outboard engine of the skiff. It remained most of the time in waters less than 10 m depth. In most of the tracking sessions the shark exhibited no discernible movement interspersed with slow displacement and occasional longer-range movements of short temporal duration. Those occurred during daytime or night-time. Periods of inactivity lasted from 15 min to 11 h. The activity space used by this shark during the 16-days tracking was  $0.84 \text{ km}^2$ , and the linear distance travelled was 15.81 km. Distances between daily locations ranged from 0.1 to 3.32 km (mean = 1.05 km, SD = 0.99 km).

#### DISCUSSION

Nurse sharks were captured throughout both sampling periods and seasons, suggesting that the species has a 

 Table 2.
 Nurse sharks monitored at Fernando de Noronha Archipelago between July 2010 and January 2012. ID indicates the transmitter number and TL is the total length.

ID	TL (cm)	Sex	Release date and site <sup>a</sup>	Total detections	No. days detected	Max. consecutive days detected	Mean daily detections	Daytime detections (%)	Nighttime detections (%)	Min. linear distance (km)	Max. receivers visited per day	Mean activity space (km²) <sup>b</sup>
1	137	М	07/17/10 (3)	1328	49	11	20.7 (1-234)	63.1	47.7	3.7	3	2.17
11	170	Μ	07/25/10 (5)	3339	98	9	30.8 (1-170)	53.1	46.9	3.7	3	2.22
12	183	Μ	02/16/11 (1)	104	13	2	7.9 (1-36)	5.8	94.2	2.1	1	1.26
16	138	М	07/22/10 (3)	3163	119	29	26.7 (1-210)	84.8	15.2	3.7	2	2.10
18	107	F	07/25/10 (5)	1870	110	21	16.3 (1-289)	63.6	36.4	3.7	2	1.97
27	115	F	02/08/11 (1)	1070	18	13	59.4 (1–196)	23.5	76.5	2.1	1	1.09
38	265	М	02/17/11 (1)	1	1	0		100	0	4	1	_
39	128	М	07/22/10 (3)	4921	55	51	89.5 (1-323)	46.6	53.4	3.7	1	2.10

<sup>a</sup>The number in parentheses indicates the site of capture (see Fig. 1 for location).

<sup>b</sup>Mean activity space in the array.

year-round occurrence at the insular shelf of FEN. Results indicate that all life-stages of *G. cirratum* use the insular shelf of the archipelago. A wide range of sizes was examined, from juveniles to adults. Although not captured, previous investigations have reported the occurrence of neonates in waters up to 15 m deep in distinct years (Garla *et al.*, 2009, 2015). Most of the individuals examined were juveniles from 100 to 179 cm TL, captured in depths of o-30 m. Individuals larger than 200 cm TL can be occasionally observed at this depth range, but were more commonly captured in depths over 25 m, regardless of the fishing gear used.

No significant differences were found between the length frequencies of the two sampling periods, demonstrating a relatively stable population structure during the study. The mean TL of nurse sharks in FEN (144.7 cm) was lower than those reported for Belize (Pikitch *et al.*, 2005) and other sites of north-eastern Brazil (Castro & Rosa, 2005; Santander-Neto *et al.*, 2011; Ferreira *et al.*, 2013). This variation in mean TL may be related to distinct growth parameters among populations, but may also reflect use of fishing gear. For example, Pikitch *et al.* (2005) and Ferreira *et al.* (2013) used larger 16/0 and 17/0 circle hooks as standard gear, in contrast to the 11/0 used in most of the sampling sessions in FEN.

No seasonal differences of sex ratio and mean TL were found, indicating that all size-classes can be found year-round in FEN. This lack of seasonal occurrence is similar to the condition observed in Ceará (Santander-Neto *et al.*, 2011), but distinct from Recife and Atol das Rocas, where seasonal differences have been reported (Castro & Rosa, 2005; Ferreira *et al.*, 2013).

The observed sex ratios of *G. cirratum* indicated that nearly equal numbers of males and females are present at FEN. The presence of mature individuals of both sexes and observations of reproductive aggregations of up to 15 nurse sharks >2 m TL in a shallow lagoon performing courtship behaviours during July–August 2015 demonstrate that FEN is used as a mating ground (RC Garla, personal communication). In the nearby Atol das Rocas mating activity was observed in May (Castro & Rosa, 2005). Thus, available data suggest that both the parturition and mating seasons of nurse sharks in oceanic insular areas of the western south Atlantic occur during the austral winter (Castro & Rosa, 2005; Garla *et al.*, 2009). This condition is distinct from that observed for sympatric carcharhinid sharks in those areas, whose neonates and adults with fresh mating scars were captured or observed during the austral summer (Freitas *et al.*, 2006; Garla *et al.*, 2009; RCG, personal communication). This pattern of distinct reproduction times of sympatric shark species using the same area has been suggested as a means to reduce interspecific competition and predation mortality during the first few months among species with great discrepancies in their size at birth (Motta *et al.*, 2005). Although this hypothesis needs further investigation, the sizes at birth of the sympatric *Negaprion brevirostris* and *Carcharhinus perezi* in FEN are nearly twice that of *G. cirratum* and the former are born in a later period of the year (Garla, 2004), suggesting that this reproductive tactic may occur.

The growth rate determined from recapture data was lower than that previously reported for nurse sharks in the Florida Keys  $(13.1 \pm 9 \text{ cm year}^{-1})$  (Carrier & Luer, 1990) and Recife  $(15.77 \pm 2.53 \text{ cm year}^{-1})$  (Ferreira *et al.*, 2013). However, the small sample size precludes further insights on the growth of *G. cirratum* in FEN and waits additional sampling to provide more accurate estimates.

Most of the tagged sharks were recaptured or resighted in close proximity to the tagging location, suggesting that G. cirratum individuals have restricted movements, as demonstrated by previous studies employing mark-and-recapture techniques (Pratt & Carrier, 2001; Pikitch et al., 2005; Ferreira et al., 2013). However, when analysing the recaptures together with the automated and active telemetry data, results indicate that nurse sharks exhibited site fidelity but not longterm residency to the array. Seven out of eight sharks fitted with acoustic transmitters repeatedly returned to the monitored area, but remained there for variable periods of time, a condition also observed in Belize and Recife (Chapman et al., 2005; Pikitch et al., 2005). Underwater observations and results of the actively tracked individual demonstrate that nurse sharks spend much of their time immobile close to structured bottoms or resting in holes, crevices and caves. This behaviour might result in part of the signals being blocked by solid structures between the transmitter and receiver. However, this would not happen for the entire periods of weeks or months without detections observed in the present study, and likely reflect emigration of the sharks out of the monitored area. All monitored sharks emigrated away from the array at some point, but unlike nurse sharks off Recife (Ferreira et al., 2013) there was no evidence of



Fig. 4. Distribution, in percentage, of acoustic detections of seven *Ginglymostoma cirratum* along an array of acoustic receivers deployed at the Fernando de Noronha Archipelago from July 2010 to January 2012. Inset bar graphs indicate the percentage of detections in each month at the receiver array and grey arrows indicate the month of transmitter implantation.



Fig. 5. Active track of one *Ginglymostoma cirratum* monitored during 16 consecutive days at the lee side of Fernando de Noronha Archipelago in July 2011. The left box indicates the linear distance travelled between successive days and the duration of each. For clarity purposes, only the first localization of each day is provided.

synchronized or seasonal migration. Each individual exhibited a distinct behaviour of visitation and emigration away from the array.

Previous studies have shown that *G. cirratum* can show relatively wide-ranging movements. The longest recorded distance travelled by the species was 541 km in Florida (Kohler & Turner, 2001), and 29.3 km in Belize, where a juvenile individual almost circumnavigated Glover's Reef Atoll in a 5-month period (Chapman *et al.*, 2005). This is further supported by the active tracking of a juvenile nurse shark in FEN, which moved 15.81 km along a stretch of the coastline inspected by four receivers of the telemetry array in a 16-day period. Thus, results indicate *G. cirratum* likely move along wide sections of the insular shelf and use areas outside the protected zones in FEN where they can be exposed to fisheries. In this sense, future research should assess residency and movements over periods longer than 2 years to completely evaluate the conservation benefits of MPAs areas to populations of *G. cirratum*.

Nonetheless, findings are relevant for a better understanding and have management implications for the species in FEN and at other sites of its distribution. In this sense, MPAs are important to the early life-stages of *G. cirratum*, such as neonates and small juveniles, but they cannot be the only conservation measure used to protect this species. Despite remaining motionless or exhibiting short range movements for several hours or days, nurse sharks can be relatively wide-ranging (Chapman *et al.*, 2005) and require different types of protection throughout their life cycle. These should include the use of MPAs, collaborative fisheries monitoring, and fisheries management practices such as the live release of incidentally caught individuals, along with law enforcement.

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#### REFERENCES

- Afonso A.S., Andrade H.A. and Hazin F.H.V. (2014) Structure and dynamics of the shark assemblage off Recife, Northeastern Brazil. *PLoS ONE* 9, e102369.
- Carrier J.C. (1985) Nurse sharks of Big Pine Key: comparative success of three types of external tags. *Florida Scientist* 48, 146–154.
- Carrier J.C. and Luer C.A. (1990) Growth rates in the nurse shark, Ginglymostoma cirratum. Copeia 3, 686–692.
- Carrier J.C. and Pratt H.L. (1998) Habitat management and closure of a nurse shark breeding and nursery ground. *Fisheries Research* 39, 209–213.
- **Carrier J.C., Pratt H.L. and Martin L.K.** (1994) Group reproductive behavior in free-living nurse shark, *Ginglymostoma cirratum. Copeia* 3, 646–656.

- **Castro A.L.F. and Rosa R.S.** (2005) Use of natural marks on population estimates of nurse shark, *Ginglymostoma cirratum*, at Atol das Rocas Biological Reserve, Brazil. *Environmental Biology of Fishes* 72, 213–221.
- **Castro J.I.** (2000) The biology of the nurse shark, *Ginglymostoma cirratum*, off the Florida east coast and the Bahamas Islands. *Environmental Biology of Fishes* 58, 1–22.
- **Castro J.I., Woodley C.M. and Brudek R.L.** (1999) A preliminary evaluation of the status of shark species. FAO Fisheries Technical Paper. No. 380. Rome: FAO. 72 pp.
- Cartamil D.P., Vaudo J.J., Lowe C.G., Wetherbee B.M. and Holland K.N. (2003) Diel movement patterns of the Hawaiian stingray, *Dasyatis lata*: implications for ecological interactions between sympatric elasmobranch species. *Marine Biology* 142, 841–847.
- Chapman D.D., Pikitch E.K., Babcock E. and Shivji M.S. (2005) Marine reserve design and evaluation using automated acoustic telemetry: a case-study involving coral reef-associated sharks in the Mesoamerican Caribbean. *Marine Technology Society Journal* 39, 42-55.
- Compagno L.J.V. (2001) Sharks of the world. An annotated and illustrated catalogue of shark species know to date. No. 1, Volume 2. Bullhead, mackerel and carpet sharks (Heterodontiformes, Lamniformes and Orectolobiformes). Rome: FAO Species Catalogue for Fishery Purposes.
- Ferreira L.C., Afonso A.S., Castilho P.C. and Hazin F.H.V. (2013) Habitat use of the nurse shark, *Ginglymostoma cirratum*, off Recife, Northeast Brazil: a combined survey with longline and acoustic telemetry. *Environmental Biology of Fishes* 96, 735–745.
- Freitas R.H.A., Rosa R.S., Gruber S.H. and Wetherbee B.M. (2006) Early growth and juvenile population structure of lemon sharks *Negaprion brevirostris* in the Atol das Rocas Biological Reserve, off north-east Brazil. *Journal of Fish Biology* 68, 1319–1332.
- Garla R.C. (2004) Ecologia e conservação dos tubarões do Arquipélago de Fernando de Noronha, com ênfase no tubarão-cabeça-de-cesto, Carcharhinus perezi. PhD Thesis. UNESP, Rio Claro, SP, Brasil.
- Garla R.C., Garcia J. Jr., Lopes N.P. and Veras L.B. (2009) Fernando de Noronha as an insular nursery area for lemon sharks, *Negaprion brevirostris*, and nurse sharks, *Ginglymostoma cirratum*, in the equatorial western Atlantic Ocean. *Marine Biodiversity Records* 2, 1-4.
- Garla R.C., Garrone-Neto D. and Gadig O.B.F. (2015) Defensive strategies of neonate nurse sharks, *Ginglymostoma cirratum*, in an oceanic archipelago of the Western Central Atlantic. *Acta Ethologica* 18, 167–171.
- Hazin F.H.V., Wanderley J.A.M. Jr. and Mattos S.M.C. (2000) Distribuição e abundância relativa de tubarões no litoral do Estado de Pernambuco, Brasil. *Arquivos de Ciências do Mar* 33, 33-42.
- Heithaus M.R., Burkholder D., Hueter R.E., Heithaus L.I., Pratt H.W. Jr. and Carrier J.C. (2007) Spatial and temporal variation in shark communities of the lower Florida Keys and evidence for historical population declines. *Canadian Journal of Fisheries and Aquatic Sciences* 64, 1302–1313.
- Henningsen A.D. (1994) Tonic immobility in 12 elasmobranchs: use as an aid in captive husbandry. *Zoo Biology* 13, 325–332.

- Karl S.A., Castro A.L.F. and Garla R.C. (2011) Population genetics of the nurse shark (*Ginglymostoma cirratum*) in the western Atlantic. *Marine Biology* 159, 489–498.
- Kohler N.E. and Turner P.A. (2001) Shark tagging: a review of conventional methods and studies. *Environmental Biology of Fishes* 60, 191– 224.
- Klimley A.P. (1980) Observation of courtship and population in the nurse shark, *Ginglymostoma cirratum*. Copeia 4, 878–882.
- Maida M. and Ferreira B.P. (1997) Coral reefs of Brazil: an overview. Proceedings of the 8th International Coral Reef Symposium 1, 263–274.
- Moral-Flores L.F.D., Ramírez-Antonio E., Angulo A. and de León G.P.P. (2015) *Ginglymostoma unami* sp. nov. (Chondrichthyes: Orectolobiformes: Ginglymostomatidae): una especie nueva de tiburón gata del Pacífico oriental tropical. *Revista Mexicana Biodiversidad* 86, 48–58.
- Motta F.S., Gadig O.B.F., Namora R.C. and Braga F.M.S. (2005) Size and sex compositions, length-weight relationship, and occurrence of the Brazilian sharpnose shark, *Rhizoprionodon lalandii*, caught by artisanal fishery from southeastern Brazil. *Fisheries Research* 74, 116–126.
- Musick J.A. (2005) Management of sharks and their relatives (Elasmobranchii). In Musick J. and Bonfil R. (eds) *Management techniques for Elasmobranch fisheries*. FAO Fisheries Technical Paper No. 474. Rome: FAO, pp. 1–8.
- Pikitch E.K., Chapman D.D., Babcock E.A. and Shivji M.S. (2005) Habitat use and demographic population structure of elasmobranchs at a Caribbean atoll (Glover's Reef, Belize). *Marine Ecology Progress Series* 302, 187–197.
- **Pratt H.L. and Carrier J.C.** (2001) A review of elasmobranch reproductive behavior with a case study on the nurse shark, *Ginglymostoma cirratum. Environmental Biology of Fishes* 60, 157–188.
- Rosa R.S., Castro A.L.F., Furtado M., Monzini J. and Grubbs R.D. (2006) *Ginglymostoma cirratum*. In *IUCN 2016. IUCN Red List of Threatened Species*. Available at http://www.iucnredlist.org (accessed 24 February 2016).
- Rosa R.S. and Gadig O.B.F. (2009) Ginglymostoma cirratum (Bonnaterre, 1788). In Machado A.B.M., Drumond G.M. and Paglia A.P. (eds) Livro Vermelho da Fauna Brasileira Ameaçada de Extinção., Brasília, Brasil: Ministério do Meio Ambiente, pp. 28–29.

#### and

Santander-Neto J., Shinozaki-Mendes R., Silveira L.M., Juca-Queiroz B., Furtunato-Neto M.A.A. and Faria V.V. (2011) Population structure of nurse sharks, *Ginglymostoma cirratum* (Orectolobiformes), caught off Ceará State, Brazil, south-western Equatorial Atlantic. *Journal of the Marine Biological Association of the United Kingdom* 91, 1193-1196.

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