Effects of extreme cold and aridity on soils and habitability: McMurdo Dry Valleys as an analogue for the Mars Phoenix landing site

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Abstract: The McMurdo Dry Valleys are among the driest, coldest environments on Earth and are excellent analogues for the Martian northern plains. In preparation for the 2008 Phoenix Mars mission, we conducted an interdisciplinary investigation comparing the biological, mineralogical, chemical, and physical properties of wetter lower Taylor Valley (TV) soils to colder, drier University Valley (UV) soils. Our analyses were performed for each horizon from the surface to the ice table. In TV, clay-sized particle distribution and less abundant soluble salts both suggested vertical and possible horizontal transport by water, and microbial biomass was higher. Alteration of mica to short-order phyllosilicates suggested aqueous weathering. In UV, salts, clay-sized materials, and biomass were more abundant near the surface, suggesting minimal downward translocation by water. The presence of microorganisms in each horizon was established for the first time in an ultraxerous zone. Higher biomass numbers were seen near the surface and ice table, perhaps representing locally more clement environments. Currently, water activity is too low to support metabolism at the Phoenix site, but obliquity changes may produce higher temperatures and sufficient water activity to permit microbial growth, if the populations could survive long dormancy periods ($\sim 10^6$ years).

Received 3 November 2010, accepted 5 August 2011, first published online 4 January 2012

Key words: dry permafrost, habitability, Taylor Valley, University Valley

Introduction

The McMurdo Dry Valleys (MDV) comprise the largest ice-free region of Antarctica (4800 km²), are aptly characterized as a cold desert, and have been considered as an astrobiology analogue for Mars. They are located in southern Victoria Land between the Ross Sea/McMurdo Sound and the Polar Plateau/East Antarctic Ice Sheet (Fig. 1) and consist of a series of generally east–west trending, glacially formed valleys that have been previously categorized into three soil moisture zones, determined largely by elevation and climate influences: subxerous, xerous and ultraxerous (Campbell & Claridge 1982).

The subxerous zone is the warmest and wettest, and occurs primarily near the coast. Ice-cemented soil is found within 1 m of the surface, and the soil thaws down to the ice table each summer. The ultraxerous zone occurs at high elevation (generally > 1000 m) near the polar plateau. Ground temperatures exceed 0°C only very near the surface, although loose ice-free soil that is < 0°C, termed "dry permafrost", extends to greater depths (e.g. McKay *et al.* 1998).

Commonly, dry permafrost transitions abruptly to icecemented ground at depths down to about 1 m. Dry permafrost may be unique to the MDV on Earth, but it is ubiquitous on Mars due to much lower Martian temperatures. The Phoenix (PHX) spacecraft investigated dry permafrost in Mars' polar regions in 2008 (Smith *et al.* 2008), increasing the importance of understanding the unique MDV environment as a Mars analogue.

Moving from subxerous low coastal valleys to ultraxerous high inland valleys, the soils in the MDV represent a long-term natural experiment revealing the geochemical effects and biological consequences of progressively less abundant liquid water. Our primary goal was to understand how soil chemistry, mineralogy, structure, and habitability change with the influence of water between these two end member locations, and to use that information for comparison to an even colder, drier location - the Martian northern plains. In particular, we focused on four factors: 1) soluble salts, 2) minerals, 3) clay-sized particle distribution, and 4) biomass, each sensitive in distinct ways to traces of mobile water.



Fig. 1. McMurdo Dry Valleys. Stars indicate sites of the field locations described in this paper: Taylor Valley pit 2 at $77^{\circ}36'06.1$ "S, $163^{\circ}08'19.6$ "E; elevation 21 ± 5.5 m and University Valley pit 7 at $77^{\circ}51'43.6$ "S, $160^{\circ}42'23.8$ "E; elevation: 1683 ± 5.9 m.

To address our objectives, we conducted a systematic, interdisciplinary investigation of soil pits dug in Lower Taylor Valley (subxerous; TV) and in University Valley (ultraxerous; UV; Fig. 1). We chose to conduct and present our full analyses, including biomass estimates, on two end member pits, although six total pits in TV and UV were excavated and analysed. While UV and TV do exhibit heterogeneity, our purpose was to examine the influence of varying water amount in each location and its effect on the quantity and type of soluble salts, mineralogy, clay-sized particle distribution, and biomass, for which we believe end



Fig. 2. Taylor Valley, with the Commonwealth Glacier in the foreground, and Lake Fryxell on the left. The approximate position of the soil pit is indicated by the yellow star.

member pits are sufficient and generally representative of the climate zone of interest, based on literature and comparison to our other pits in these valleys. In all our pits, we identified and sampled each soil horizon down to, and including, the ice-cemented layer, which constituted the bottom of our pits. Analyses included both *in situ* (i.e. in the field) and standard laboratory techniques. Here we describe the detailed analyses of these two end member soil profiles, identify the chemical, compositional, and physical properties that are most sensitive to trace water, and use relative abundance of microbial biomass to identify soil horizons that are more hospitable to life than others within the same pit. We then compare these results to similar analyses of the Martian dry permafrost.

Site characteristics

Taylor Valley (TV) is a low-elevation, coastal valley, 60 km long in total length (Fig. 2), receiving 1–75 mm water equivalent (weq) snow accumulation (precipitation plus windblown snow deposition) annually (Fountain *et al.* 2010, Table I). The summer air temperature is frequently $> 0^{\circ}$ C, and liquid water is present annually, as soil pore water and surface streams.

The active layer in TV, (soil zone in which temperature annually exceeds 0°C), extends to 25–50 cm depth (Marchant & Head 2007), and in most locations, liquid water is stable throughout the soil column for part of each year. Water infiltration from snowmelt, which is concentrated in the lees and small-scale topographic lows that trap blowing snow, is spatially heterogeneous, as are the resultant chemical and physical properties of the soil. Liquid water can drain the entire soil column, and flows laterally across the surface of the ice table during summer (Wilson 1979). This flushing is the primary means by which mass is removed from the TV soil column.



Fig. 3. University Valley, looking toward the headwall and the permanent snowfield. The approximate position of the soil pit is indicated by the yellow star.

Climatic zone	Annual av. $T_s^{a, b}$	Summer av. T_s^{a}	Active layer thickness	Annual av. T_a	Summer av. T_a^{a}	Snow accumulation (mm weq yr ⁻¹)	Annual R_H^{a}
Lower Taylor Valley Subxerous	-18°C	+2°C	$\sim 25 \mathrm{cm}^{\mathrm{a}}$	-20.2°C ^d	-2°C	20–100 ^c	63%
University Valley Ultraxerous	-25°C	-5°C	$\sim 12 \text{ cm}^{\text{b}}$	-25°C ^b	-15°C ^b	Unknown	43%

Table I. Summary of characteristics of Taylor and University valleys.

^a Marchant & Head (2007); T_s = surface temperature.

^b Marinova *et al.* (2011); T_s = surface temperature.

^c Fountain *et al.* (2010), Table I; weg = water equivalent.

^d Doran *et al.* (2002); T_a = air temperature.

University Valley (\sim 1700 m elevation; Fig. 3) is a small lateral hanging valley (\sim 1 x 3 km) that feeds into Beacon Valley (BV), located about 75 km from the coast. There is a permanent snowfield near the valley headwall that is probably maintained by accumulation of snow blown off the polar plateau (McKay 2009). University Valley is colder and dryer than TV with average annual air and soil temperatures of \sim -25°C and average summer surface temperatures of only \sim -5°C (Marinova *et al.* 2011).

There is a substantial uncertainty in the average annual snow accumulation at our UV soil pit. Annual snow accumulation has not been measured in either UV or nearby BV, but McKay (2009) argued that snow accumulates at the site of our soil pit, 500 m down-valley from the permanent snowfield, only once every 2–3 years. The snow accumulation gradient moving inland from the coast (Fountain *et al.* 2010) implies between 0 (unmeasurable) and \sim 53 mm weq yr⁻¹ accumulation, but this gradient has not been verified outside TV. Forecast modelling of annual precipitation by Powers *et al.* (2003) predicts that UV gets more snow than TV, perhaps up to 100 mm weq yr⁻¹.

The key climatic difference between subxerous (lower TV) and ultraxerous (UV) regimes is the maximum air temperature. Measurements in BV (500 m lower in elevation that UV) suggest that the air temperature in UV never reaches 0°C (Doran *et al.* 2002), precluding large-scale melting. A significant but unknown fraction of the snow sublimates (Fountain *et al.* 2010). Ground temperatures in UV do however, reach 0°C (e.g. Marinova *et al.* 2011), and whatever trace snow remains in contact with bare ground can melt and seep into the subsurface. In fact, this process provides enough liquid water to sustain the

cryptoendolithic communities in the Beacon sandstone walls of UV (Friedmann 1982).

Meltwater production in UV is spatially restricted, by the same small-scale topographic trapping that operates in TV, and probably occurs no more than a few afternoons each year. In UV, the active layer is ≤ 10 cm deep (McKay *et al.* 1998). Typically, the active layer in ultraxerous valleys does not reach ground ice, the soil column is not flushed, and even the most soluble salts accumulate.

Permanently "dry" soil is expected in ultraxerous valleys, in that part of the soil column between the active layer and the ice table. At the nanometer to micron scale however, all grain surfaces are covered with thin films of adsorbed water molecules. These films form at solid surfaces due to the attraction between the polar water molecule and the grain surfaces (van der Waals forces), and are ubiquitous on any surface exposed to water vapour. Adsorbate films exhibit an equilibrium thickness that depends on temperature and the relative humidity of the pore gas (Anderson & Tice 1972). An entirely distinct process, known as premelting, likewise results in a mobile, unfrozen layer of water molecules covering the surface of all ice grains, particularly near their melting point (Rempel et al. 2004). Adsorbed and premelted water films are universal under appropriate conditions, and the environments of both TV and UV are such that unfrozen films are probably ubiquitous. The capacity of these films to facilitate geologic and biologic processes, as compared to the periodic bulk liquid water that occurs in the subxerous valleys, is a key focus of this study.

Although the ultraxerous MDV are among the coldest and most arid sites on Earth, they are considerably more clement than the PHX site on Mars (68°N). The soil surface

Table II. Location, geomorphic surface, parent material and age of surface for Taylor and University valleys.

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	Lat (S)	Long (E)	Elevation (m)	Slope	Landform/parent materials	Till/parent material composition	Age
Lower Taylor Valley pit 2	77°36'6.1"	163°08'19.6"	21 ± 5	4–5%	foot slope/glacial till-glaciolacustrine	granite, gneiss, schist, sandstone, dolerite	11–15 ky ^a
University Valley pit 7	77°51'43.6"	160°42'23.8"	1683 ± 6	<1%	valley floor/colluvial materials from valley walls	sandstone, dolerite	^b Unknown

^a Approximate age range based upon proximity to dated sites by Hall & Denton (2000); Ross Sea Drift material.

^b Materials have not been dated; inflationary surface from infilling by colluvial processes from valley walls.





Fig. 4. Locations of the soil pits (yellow stars) in Taylor Valley. Pit 2 at $77^{\circ}36'06.1"$ S, $163^{\circ}08'19.6"$ E, elevation $21 \pm 5.5 \text{ m}$ (top) and University Valley pit 7 at $77^{\circ}51'43.6"$ S, $160^{\circ}42'23.8"$ E, elevation: $1683 \pm 5.9 \text{ m}$ (bottom). The location of the Beacon Valley (BV) meteorology station is marked with a green circle (bottom; $77^{\circ}51'43.6"$ S, $160^{\circ}42'23.8"$ E).

at the PHX site reached only $\sim -23^{\circ}$ C near summer solstice, and the maximum temperature of the ground ice ($\sim 5 \text{ cm}$ depth) probably does not exceed -53°C (Mellon *et al.* 2009, Zent *et al.* 2009). Models have suggested the possibility that at high obliquity, surface temperatures at the PHX latitude may be similar to present-day UV. There is considerable uncertainty in these models, although regolith temperatures in the -13 to $+7^{\circ}$ C range are commonly predicted (e.g. Zent 2008). Because there is water in the Martian system (polar caps, subsurface, and atmospheric water vapour), unfrozen



Fig. 5. a. Taylor Valley surface prior to digging pit 2 (top), Taylor Valley pit 2 after excavation (bottom). b. University Valley surface prior to digging pit 7 (top), University Valley pit 7 after excavation (bottom).

water should be present as well, and, could be as abundant as that in present day UV when temperatures warm with obliquity changes.

Methods

Site and excavation description

The field sites described in this paper are shown on Figs 1–4 (yellow stars). Several pits were dug in each valley during summer 2007–08. We will discuss in detail the results of pit 2 in lower TV, near Lake Fryxell (Figs 2 & 5a, Table II), and pit 7 in UV (Figs 3 & 5b, Table II). Both pits were in polygon centres, experience lower stress and less cryoturbation than locations near the polygon edges (Sletten *et al.* 2003), and are representative of the climate zone in which they were dug.

In each pit, excavation was performed down to and including the ice-cemented layer, of which an area about 0.5 m² was exposed. Soils were characterized using standard methods (*Soil Survey Laboratory Methods Manual*, USDA 2004; http://soils.usda.gov/technical/lmm/, accessed 2009).

Soil master horizons were designated as D, B, or C. The D horizon designation was used to represent a desert pavement (Bockheim 1997). Lowercase letters were used as suffixes to designate specific kinds of master horizons and layers. For example, the *ff* suffix was used to represent dry permafrost. Definitions of master horizons and suffixes are found in *Keys to Soil Taxonomy* (Chapter 18, http://soils.usda.gov/technical/classification/tax_keys/index.html, accessed 2009).

At each location, soil horizons were sampled using sterile tools, and stored in sterile sample containers. The sterilization protocols were those used by the Johnson Space Center curation facilities (Astromaterials Research Office 2006) and total organic carbon was monitored during the cleaning process to ensure that organic materials were removed. Samples intended for biological analyses were placed in Whirl-Pak[®] sterile bags. The layers were sampled from the bottom up in order to avoid cross-contamination from falling material. Approximately 2 kg of material was excavated from each horizon and shipped frozen back to the United States. The ice-cemented ground was also sampled using a sterile hammer and chisel (2–5 cm of ice-cemented material was removed). The soil just above the ice table in TV was damp at the time of excavation, either due to melting of the ice table, or lateral migration of liquid water along the ice table, or both (Wilson 1979). All samples were stored, shipped, and maintained in a frozen state. A total of five horizons were identified and sampled in UV pit 7 and a total of 8 horizons in TV pit 2.

Some measurements were conducted in the field, primarily using Phoenix flight spare instruments, but the majority of our analyses were performed at our home institutions or at commercial labs.

In situ measurements

Soil thermal properties were taken from *in situ* (i.e. in the field) measurements made with the PHX Thermal and Electrical Conductivity Probe (TECP; Zent *et al.* 2009, 2010). Wet chemistry analyses for soluble ions, except ClO_4^- , were also performed in the field using the PHX Wet Chemistry Laboratory (WCL; Kounaves *et al.* 2009). Use of flight spare instruments allowed us to both test the instruments in a Mars analogue environment, and simultaneously deliver on-site analyses.

Soil thermal properties, volumetric heat capacity (ρC) and thermal conductivity (K), were determined via the transient heated needle technique (de Vries 1952). Soil temperatures and thermophysical properties are first order controls on the propagation of energy through the soil, and as such play a key role in determining the active layer, as well as the thickness of unfrozen water films. These data were used to constrain thermal models of the soil pits in support of our sample analyses. Due to power-supply issues, extended TECP measurements were made only in BV. The ρC and K measured in BV allowed the model to reproduce independent measurements of subsurface temperature in UV (Marinova et al. 2011), so this proxy was acceptable. TECP measurements were collected over a 21-hour period on 2 and 3 January 2008. Raw data for each channel were converted to physical units via the functions established for the TECP unit during flight calibration (Zent et al. 2009).

A series of WCL analyses for Na⁺, K⁺, Mg²⁺, Ca²⁺, Cl⁻, pH, and electrical conductivity (EC) were performed on samples from pits in lower TV to guide the sampling approach. Soluble ion chemistry and abundance were found to be different in each soil horizon, supporting the need to analyze each horizon. Subsequent comparison of the WCL *in situ* data to laboratory Ion Chromatographic (IC) analyses of the same samples (see below) found comparable results.

Mineralogy

Soil samples were fractioned into five sand-sized (2–1, 1–0.5, 0.5–0.25, 0.25–0.09, 0.09–0.053 mm), three silt-sized (53–20, 20–5, 5–2 μ m) and two clay-sized (2–0.2, < 0.2 μ m) fractions following the method of Jackson (1985) with the exception that a high-pH dispersion agent was omitted

to minimize dissolution of amorphous aluminosilicates. Mineralogy of all soil fractions was determined by X-ray diffraction (XRD) analysis on a Scintag XDS2000 X-ray Diffractometer using CuK α radiation from 2–70°2 θ . Sand and silt samples were prepared as random-oriented mounts. Bulk soils (< 2 mm size fraction) and sand-sized fractions were ground by hand to $< 53 \,\mu m$ prior to XRD analysis. The clay-sized fractions were saturated with Mg^{2+} and K^{+} and pipetted onto glass slides for preferred-orientation mounts. Mg-saturated clav-sized fractions were X-raved at ambient temperature followed by glyceration, which causes 2:1 expandable phyllosilicates to expand to a characteristic d-spacing. K-saturated clay-sized fractions were X-rayed at ambient temperature, followed by heating to 110°C, 300°C, and 550°C (Jackson 1985). This procedure was performed on each soil horizon for both pits. Sub-samples of the $< 125 \,\mu m$ fraction of untreated bulk soils were ground by hand to $< 53 \,\mu\text{m}$ and analysed by XRD to determine mineralogy, including salts.

Quantitative estimates of the total crystalline and amorphous phases were determined by the Rietveld method (Rietveld 1969) using a Panalytical X'Pert Pro MPD Diffractometer using Cu $K\alpha$ radiation operated at 45 kV and 40 mA current. Representative portions of each bulk soil (<2 mm size fraction) were ground by hand with an agate mortar and pestle in ethanol to a fine powder (<53 µm) and a 10% by weight spike of alumina (Al₂O₃) was added as an internal calibration standard.

Particle sizes and specific surface area

Soils from each layer were also analysed to determine their particle-size distribution and specific surface area. The particle-size analysis was conducted using a Beckman Coulter LS 13 320 Laser Diffraction Particle Size Analyzer that measures particles from 0.4 μ m to 2 mm over 116 size bins. For particle size analysis, the samples were prepared by soaking them for 24 hours in DI water on a rotating mixer and then adding sodium hexametaphosphate as a dispersing agent and mixing overnight (XRD analysis on size separates were done on samples that were segregated without using a dispersant to preserve any amorphous material present). Each sample was run four times and averaged. Specific surface area measurements were carried out on unmodified soil samples using the standard BET method, measuring nitrogen adsorption to determine the surface area of the sample in m² g⁻¹.

Ion chemistry

Soluble ions, except ClO_4^- , were determined by measurements made on aqueous extracts of soil samples, which were leached at room temperature for 1 hr at a 1:1 weight ratio of soil to water and were extracted through a 0.2 µm filter.

Analysis of the soluble anion species was performed using a Dionex ICS-2000 Reagent FreeTM IC equipped

Horizon	Depth (cm)	Amorphous (%)	Gypsum	Nitratine	Ca-Carbonate	
Lower Taylor Valley pit 2						
D	0-1	4				
Bw	1–9	4	Present			
2Db	9-12	5	Present			
2Bbw1	12–17	7	Present	Present		
2Bwb2	17-20	10				
2Bwb3	20–24	5				
2Bwb4	24–28	10		Present		
2Bwbfm	28 +	15			Present	
University Valley pit 7						
Bwz	1–9	5	Present	Present		
CBffij1	9–20	5	Present	Present		
CBffjj2	20-34	4	Present	Present		
Cfm	34+	5	Present	Present		

Table III. Secondary weathering phases in ADV soils. Semi-quantitative estimates of amorphous materials from Rietveld analysis of XRD patterns. Crystalline salts were identified by XRD but quantities are at trace levels.

with a Dionex Ionpac AS18 analytical column (250 x 4 mm inside diameter, I.D.), an AG18 guard column (50 x 4 mm I.D.), and an Anion Self-Generating Suppressor 300 (4 mm) with a suppression current of 105 mA. The eluent was 20 mM potassium hydroxide with a flow-rate of 1.0 ml min⁻¹. Calibration curves using three standards were generated for the anionic species: F^- , Cl^- , Br^- , NO_3^- , NO_2^- , SO_4^{2-} , and PO_4^{3-} . Initially, all solutions were run undiluted, however, due to the presence of high concentrations of certain anions, it was necessary in most cases to re-analyse the solutions following a 100-fold dilution with 18 M Ω cm⁻¹ water. The ClO₄ was determined by IC as described in Kounaves *et al.* (2010a).

Cation species were analysed using a Perkin Elmer Analyst 800 Atomic Absorbance Spectrometer outfitted with cation specific Hollow Cathode lamps. Calibration curves using five standards were generated for: Na^+ , K^+ , Mg^{2+} , and Ca^{2+} . Initially, all solutions were diluted 1:2 with a 0.2% HNO₃ solution. All standards and necessary further dilutions were made with 0.1% HNO₃.

Scanning Electron Microscopy (SEM) and Transmission Electron Microscopy (TEM)

Silt-sized fractions of the soil samples were mounted on sticky tape attached to a stub and then carbon-coated for morphological analysis using a JSM 5910LV scanning electron microscope equipped with an IXRF energy dispersive X-ray spectroscopy (EDS) system. Clay-sized materials from selected soil horizons were dispersed in water and pipetted onto a carbon film on a copper grid. They were examined without coating under a JEOL

Table IV. Soluble ions, pH, electrical conductivity. Concentrations of all ions, except ClO_4^- , are for 1 g soil leached with 1 ml water. For ClO_4^- 1 g soil was leached with 5 ml water.

Horizon	Depth	pН	EC (mhos/cm) (mM)	Mg (mM)	Ca (mM)	K (mM)	Na (mM)	Cl (mM)	SO ₄ (mM)	NO ₃ (μM)	ClO ₄ ^a (µM)
	(cm)										
Taylor Valley											
D	0-1	7.6	2.8	0.658	2.25	0.92	29.2	27.8	6.9	0.4	nd
Bw	1-9	7.4	8.2	8.27	14.2	2.48	75.3	84.6	29.3	2.9	nd
2Db	9-12	7.9	4.4	3.62	1.25	1.18	50.5	54.3	6.2	2.6	nd
2Bbwb1	12-17	7.5	6.8	6.91	1.22	2.35	68.2	94.7	1.1	6.5	2.2
2Bwb2	17-20	8.0	4.2	2.55	0.349	1.64	45.5	54.8	1.0	2.4	nd
2Bwb3	20-24	8.3	1.0	0.078	0.035	0.43	11.3	9.5	0.4	0.5	nd
2Bwb4	24-28	8.6	0.5	0.053	0.035	0.36	6.9	1.3	0.6	0.1	nd
2Bwbfm	28 +	8.7	0.8	0.946	0.474	1.93	8.4	3.5	0.4	0.5	0.57
University Val	ley										
D	0-1										0.33
Bwz	1-9	6.9	8.7	34.2	21.3	3.17	55.4	26.2	39.0	81.1	1.3
CBffjj1	9–20	6.9	8.5	36.0	19.9	3.20	42.2	30.7	40.8	68.2	0.50
CBffjj2	20-34	6.9	3.0	6.42	4.09	0.49	17.1	8.74	12.0	11.5	0.26
Cfm	34+	7.3	3.4	7.04	6.29	0.69	18.0	8.35	17.0	10.0	0.20

nd = not detected at the IC method LOD of $0.05 \,\mu\text{M}$ in the solution.

^a Kounaves et al. 2010a.



Fig. 6. a. Taylor Valley. (Top) Soluble ion concentrations from the aqueous extracts (extracted 1:1). Horizontal grey/white bars indicate the soil horizons. Dark grey indicates the ice table. (Bottom) Clay-sized fraction (percent*100), and perchlorate concentration (ppb) in aqueous extract. Perchlorate was only detected in horizons 2*Bbw* and 2*Bbfm*, the ice table. b. University Valley. (Top) Soluble ion concentrations from the aqueous extracts (extracted 1:1). Horizontal grey/white bars indicate the soil horizons. Dark grey indicates the ice table. No analysis of the D horizon was done. (Bottom) Clay-sized fraction (percent*100), and perchlorate concentration (ppb*10) in aqueous extract.

2000FX scanning transmission electron microscope operated at 200 kV at 10nA beam current.

Biology

Phospolipid Fatty Acids (PLFA) were extracted and purified at Microbial Insights, Inc. Soils were lyophilized, and lipids extracted in methanol:chloroform. The extract was filtered and the polar lipid fraction was recovered, purified by thin layer chromatography and analysed by gas chromatography (GC) with the inclusion of internal quantitation standards (White *et al.* 1979). Following White *et al.* (1979), the PLFA in the samples were separated by GC into particular metabolic groups, and the resulting PLFA "profile" reflects the proportions (as percentage) of the groups of organisms present in the sample.

An estimate of biomass was made from PLFA data by applying a conversion factor $(2 \times 10^4 \text{ cells/pmole})$ based on cells grown in laboratory media (White *et al.* 1979), where it was shown to be independent of microbial group; we assume that the microbes from the MDV behave similarly. For this reason, and because we do not know the specific

extraction efficiency for our Antarctic soils (especially dry permafrost and clay mineral rich soils) the PFLA biomass determinations are considered estimates only.

As a comparison point, DNA-based biomass estimates were made using quantitative PCR. Extraction of DNA from soil and quantitation assays was also performed at Microbial Insights, Inc. A calibration curve was obtained by using serial dilutions of DNA of known concentration and the values obtained for each sample were compared with the standard curve to determine the initial DNA concentration. Cell numbers were estimated assuming an average molecular weight of 660 for a base pair in double-stranded DNA, and an average of 3.5 16S rRNA gene operons per bacterial cell (Balkwill *et al.* 1988), although we do not know how well these assumptions apply to the organisms present at our field sites.

Support modelling

In order to assess the subsurface environment at UV, a soil thermal model was developed, using the TECP-measured thermal conductivity (*K*) (~ 0.4 W m⁻¹ K⁻¹) and volumetric



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heat capacity (ρC) (~ 1.7 x 10⁶ J m⁻³ K⁻¹). The simple model assumes a planar, semi-infinite half space, and solves the surface energy balance and thermal conduction equations:

$$I(1-A) - \varepsilon \sigma T^4 = \left. K \frac{dT}{dz} \right|_{z=0} \tag{1}$$

$$\frac{d^2T}{dz^2} = \frac{K}{\rho C} \frac{dT}{dz}$$
(2)

A simple shadow mask accounts for shadowing by the walls of UV. The albedo and emissivity are taken from Campbell et al. (1997), and were tuned slightly to reproduce the soil temperatures reported by Marinova et al. (2011, $\alpha = 0.26$, $\varepsilon = 1.0$). The lower boundary condition is fixed at -14°C. based on measured ice temperatures. The model does not attempt to couple heat and mass transfer that would occur during drainage of snowmelt, because critical parameters such as the hydraulic conductivity, soil matric potential and infiltration rate are not known for the undisturbed soil column.

Results

Lower Taylor Valley

At our sampling site in lower TV (pit 2), with a depth to ground ice of 28 cm, eight soil horizons were identified and sampled: 0-1 cm D, 1-9 cm Bw, 9-12 cm 2Db, 12-17 cm 2Bwb1, 17-20 cm 2Bwb2, 20-24 cm 2Bwb3, 24-28 cm 2Bwb4, and 28+ cm 2Bwbfm (Fig. 5a, Tables III & IV).

Soil particle sizes generally decreased with depth in the soil column in pit 2 (and in the other pits examined in lower TV), but the size frequency distribution in the 20-24 cm horizon was distinctly different from all other horizons. The sand-sized fraction decreased with depth throughout the profile, except for the 20-24 cm layer, in which the relative abundance of sand increased at the expense of the silt- and clay-sized fraction. The silt fraction increased with depth, except at the 20-24 cm layer, in which it declined. The claysized fraction increased with depth going from 4% (by volume) at the surface to 45% of the $<2 \,\text{mm}$ material in the ice-cemented soil (Fig. 6a), and showed a substantial increase in the bottom two layers (> 24 cm). The USDA textural class of the $< 2 \,\mathrm{mm}$ fraction moved from sand at the surface to clay at depth with sandy loam and sandy clay loam soils at intermediate depths. The specific surface area of the soils also increased with depth, with a surface value of $1.8 \text{ m}^2 \text{ g}^{-1}$ that increased to $93 \text{ m}^2 \text{g}^{-1}$ in the ice-cemented soil.

via PLFA and DNA for TV pit 2.

PLFA and DNA for UV pit 7.

b. Biomass trends as determined via

The mineralogy of the sand- and silt-sized fractions was dominated by feldspars, mica, quartz, pyroxene, and amphibole, and there were trace amounts of kaolinite and vermiculite. Pyrite framboids (microscopic aggregates of pyrite grains in spheroidal clusters resembling a raspberry) were identified throughout the profile by SEM, and became more prominent in the 17-20 cm layer and below. Fe-oxide staining was also seen at the 17-20 cm depth on surfaces of pebbles and rocks. XRD and TEM analyses indicated that the clay-sized fraction was dominated by X-ray amorphous or short-order, 2:1 phyllosilicates (Table III), along with mica, feldspar, amphibole, kaolinite, and vermiculite. The X-ray

amorphous or short-order, 2:1 phyllosilicates were also seen in our other TV pits. TEM indicated that mica is weathering first to vermiculite and then to short order 2:1 phyllosilicates in the 0–1 cm horizon. The vermiculite was seen to be forming topotaxially (i.e. in mutual orientation) on the mica and the 2:1 short order phyllosilicates are associated with the vermiculite near the edges of the altered mica particles. TEM/EDS analyses indicate the loss of K in the vermiculitic layers and the formation of short-order 2:1 phyllosilicates, suggesting the weathering of K out of the mica interlayers accompanied by a reduction in layer charge.

The crystalline salts identified by XRD analysis of TV soils were gypsum in the 1–9, 9–12 and 12–17 cm horizons and nitratine in the 12–17 and 24–28 cm horizons. In the lowest horizon ~ 5 wt% aragonite was detected by XRD and Ca-rich particles were confirmed by SEM/EDS analyses. This is consistent with the more alkaline pH in this horizon (see Table IV). XRD did not detect any sulphate-bearing species in the lowest horizon or any chloride phases in any horizon.

Soluble ions in the TV soil column were dominated by Na⁺ and Cl⁻ (Fig. 6a), particularly in the 1–9 cm and 12–17 cm horizons. The 1–9 cm horizon is by far the most saline - $SO_4^{2^-}$ and all cation species were most abundant in this layer, with Cl⁻ showing a large secondary peak in this layer as well. The maximum Cl⁻ and NO₃⁻ concentrations were measured in the 12–17 cm horizon. Perchlorate (ClO₄⁻) was detected in this horizon and at the ice table interface, but was not detected elsewhere in the soil column (Table IV). Very small enrichments of Cl⁻, K⁺, Ca²⁺ and Mg²⁺ ions were found at the ice interface.

In the TV soils, the PLFA profile showed a peak in the 1-9 cm horizon (Fig. 7a), nearly an order of magnitude decrease in the lowest two soil layers (20–24 cm and 24–28 cm), and a sharp return to higher values at the ice table interface (28+ cm). The DNA trend shows two maxima, one at 9–12 cm and the other at 20–24 cm, and a sharp decrease in the 24–28 cm layer with only a modest return in the ice table interface layer.

University Valley

In UV, the depth to ground ice was 34 cm. The soil horizons analysed were: 0-1 cm D, 1-9 cm Bwz, 9-20 cm CBjjff1, 20-34 cm CBjjff2, and 34 + cm Cfm (Fig. 5b, Tables III & IV). The 1-9 cm horizon showed evidence of oxidation in the form of a strong red hue caused by iron oxides. Sand- and silt-sized fractions were dominated by quartz with lesser amounts of feldspar, pyroxene, and mica. Silt fractions also contained trace amounts of laumontite, pyrophyllolite, chlorite, and kaolinite.

The sand-sized fraction of the <2 mm material was between 50–75% throughout the UV profile. There was less sand in the 10–19 cm horizon. The silt-sized fraction showed an increase in percentage with depth in the column. However, not including the surface pavement, the claysized fraction was 2–3 times higher in the top two layers (1–19 cm; Fig. 6b) compared to the lowest two layers. The soils have a USDA (<2 mm fraction) class of either a sandy loam or a sandy clay loam. The specific surface area of UV soils showed little variation, with values ranging from a minimum of $3.8 \text{ m}^2 \text{ g}^{-1}$ in the soil layer immediately above the ice layer and a maximum of $4.2 \text{ m}^2 \text{ g}^{-1}$ in the layer of maximum clay concentration.

XRD results of clay-sized fractions were dominated by mica, with lesser amounts of amorphous materials (Table III), quartz, pyrophyllite, kaolinite, chlorite, and hematite. Gypsum and nitratine were identified by XRD in all horizons of the UV soil.

Soluble ions in UV peaked in the near subsurface (1–9 cm horizon), consistent with previous observations of similar high-elevation locations. Soluble anions were dominated by NO_3^- , which peaked in the 1–9 cm horizon and was probably paired with Na⁺ that also showed a strong concentration in this layer. Maximum SO_4^{2-} and Cl⁻ occurred in the 9–20 cm horizon, as did Mg²⁺ and K⁺. Ca²⁺ was equally abundant in the uppermost two horizons, but all soluble ions decreased precipitously in the 20–34 cm horizon. Although no indurated saltpans were found in the soil profile, we did observe some salt encrustations on the bottoms of rocks in the upper soil horizons. All horizons contained soluble ClO₄⁻ (Table IV), which also peaked in the 1–9 cm horizon.

 Ca^{2+} and SO_4^{2-} showed secondary accumulations at the ice table interface (34+ cm), and SEM found two remarkably pristine-looking gypsum crystals in this horizon. We discuss possible explanations for apparently autochthonous gypsum at depths where liquid water is not expected in the Discussion. XRD did not detect any carbonates, consistent with the less alkaline pH observed in UV.

The highest biomass measurements in UV were obtained from the 'surface' (0–1 cm) soil layer. The PLFA data show that below that layer there were two secondary peaks: the 1–9 cm layer and the ice table interface (34+ cm). These two horizons were $\sim 5x$ higher than the intervening horizons. Comparison of estimates based on total DNA with those based on PLFA (Fig. 7b) showed that the DNA method consistently gave higher values for all horizons in UV, but that both had higher values at the surface and near the ice table (20–34+ cm for DNA and 34+ for PLFA), with lower values in the intermediate layers. In the lowest horizons, the biomass rose 2–200+ times higher (based on PLFA or DNA, respectively) than that measured in the middle of the soil profile.

Discussion

Mineralogy, alteration, and particle properties

In TV, our mineralogical results are consistent with a warmer, wetter, and more habitable environment for microbes than UV.



Fig. 8. Graphical depiction of PLFA-based microbial community composition for each layer in TV pit 2 and UV pit 7. The data indicate a greater number of different microbial groups present in the TV soil pit than in the UV pit. The yellow, orange, and grey markers represent microbial groups that are exclusively or predominantly anaerobic in metabolic capability. The yellow marker present in the soil surface sample from UV represents the aerobic group Actinomycetes rather than the anaerobic sulphate reducing bacteria (SRB). In contrast, in the TV soils, yellow represents SRB, consistent with the pyrite framboids present within the soils (also absent in UV) that are indicative of active SRB metabolism and thereby of anoxic conditions.

The presence of abundant X-ray amorphous phases and/or short-order 2:1 phyllosilicates suggests weathering in a moist environment. Additionally, the increase with depth of mineral specific surface area is due to higher clay-sized particle abundance and an increase in the abundance of X-ray amorphous phases.

The trend of greater clay-sized content with depth (Fig. 6a) may indicate either translocation of clays down the profile by water or layering of fines from glaciolacustrine sedimentation. The sharp increase in abundance of clay-sized particles below the 20–24 cm horizon may also reflect transport of clay-sized particles via water flow along the ice table.

There is evidence that the oxidation state of the TV soils is highly variable, presumably on an annual timescale. We observed ferric iron staining on pebble and rock surfaces in the 17–20 cm horizon, which is clear evidence of oxidation processes. We also observed pyrite framboids in all layers, but they became more prevalent in the 17-20 cm horizon and below. These can form both biotically and abiotically, but in either case their formation requires anoxic conditions, suggesting periodic saturation. Although pyrite framboids can form abiotically (e.g. in hydrothermal vents or volcanogenic deposits), our interpretation is that these are probably biogenic in origin because, at low temperature, pyrite framboids are most stable when associated with organic material (Sawlowicz 2000). Further, the commonly occurring growth patterns of sulphate reducing bacteria (SRB), present in TV (Figs 8 & 9), as micro-colonies enclosed in extracellular polymeric substances leads to the distinctive framboidal appearance (MacLean et al. 2008).

In UV, there is less evidence of aqueous alteration of minerals. The UV sand- and silt-sized phases, with the possible exception of kaolinite, represent local parent rock fragments or phases that formed by hydrothermal alteration during the intrusion of the Ferrar Dolerite into the Beacon Sandstone. The clay-sized fraction abundance peaks at the 1–9 cm depth in UV (Fig. 6b) and then drops, as opposed to increasing in depth as in the TV profile, indicating that bulk liquid water does not routinely percolate deeper than this layer. Unlike TV soils, mica in UV soil has not undergone weathering to short-order 2:1 phyllosilicates suggesting minimal liquid water interaction (also seen in



<sup>Fig. 9. PLFA-based groupings of microorganisms at the ice table interface from two wet permafrost environments (Arctic and Taylor Valley) and a dry permafrost environment (University Valley), expressed in percent of total.
PLFA-based biomass measurements for these samples were: 4.53 x 10⁷ (Arctic), 2.97 x 10⁷ (TV), 1.16 x 10⁵ (UV). Note that biomass at the ice table in UV was two orders of magnitude lower than at the wet permafrost sites.</sup>

our other UV pit). Instead, the basaltic glass from the physical weathering of the Ferrar Dolerite is the probable candidate for the amorphous phase detected by XRD. However, the presence of hematite and the salts throughout the UV profile and the concentration of salts near the surface indicate that liquid water has played a role in the formation of these secondary phases.

Chemistry

Soluble ion species in the McMurdo Dry Valleys are principally comprised of Ca^{2+} , Mg^{2+} , Na^+ , SO_4^{2-} , NO_3^- , and Cl-, and the IC analyses that we performed were consistent with that. In TV (all four pits), soluble anions are dominated by Cl⁻, which correlates with Na⁺, and probably represents a marine aerosol source (Fig. 6a). Sulphate is a factor of four less abundant in the aqueous extraction, in all TV pits, which probably reflects the low solubility of sulphates in general. Marine aerosols are the probable source of SO_4^{2-} ; strongly deliquescent species such as Na_2SO_4 and NaHSO₄ commonly act as condensation nucleii for aerosols and precipitate as an isotopically-identifiable source of salts (Bao & Marchant 2006). Nitrates are the least abundant of the three principal anions in the TV soil (< 10% of the total Cl⁻; true for all TV pits). Antarctic nitrate is primarily atmospheric in origin, the result of NOx oxidation and precipitation on the Antarctic ice cap as HNO₃ (Michalski et al. 2005). Nitrates are delivered from the polar plateau to the MDV by katabatic winds. Most nitrates are highly soluble, and are rapidly flushed through the soil column in TV by free-flowing liquid water.

The salt-enriched 12–17 cm horizon in the TV soils is characteristic of an enrichment observed throughout much of the subxerous zone in lower Taylor and Wright valleys, and was seen in all four TV pits examined. It is probably due to water infiltration (Bockheim 1997). The small Cl⁻ increase (seen in all four TV pits) and the biomass peak at the ice table probably reflect the occurrence of liquid water, due either to melting or lateral flow.

In UV, NO₃⁻ is the most abundant soluble anion at the top of the soil column. SO_4^{2-} is most abundant below 20 cm, and Cl⁻ is least abundant (~ 30% of the total NO₃⁻; Fig. 6b). The dominance of NO₃⁻ and SO₄²⁻ is typical of ultraxerous soils (Bockheim 1997), and our other UV pit was consistent with this. This reflects, in part, the diminishing importance of local marine aerosols with distance from the coast, and a proportionately greater contribution from stratospheric aerosols, as well as the absence of drainage throughout the soil column.

In general, the concentration of Mg^{2+} , NO_3^- , and SO_4^- are higher in the UV soils (by factors of 2–20) than in TV (Fig. 6a & b), whereas K⁺, Na⁺, and Cl⁻ are lower. Claridge & Campbell (1977) found that soils derived from dolerite had higher concentrations of Ca²⁺, Mg²⁺, and Na⁺ than other soils, with Ca²⁺ and Mg²⁺ attributed to the

weathering of ferromagnesian minerals in the Ferrar Dolerite, a process we would expect in UV. Na⁺ is more abundant than Mg^{2+} or Ca²⁺ in the UV soil (true for both UV pits), and all three are much higher than K⁺.

The correlation of Ca^{2+} with SO_4^{2-} throughout the UV soil (in both UV pits) suggests precipitation of gypsum $(K_{sp} = 2.4 \times 10^5)$. Indeed XRD confirmed its presence throughout the pit profile and, surprizingly, two pristine gypsum crystals were seen with SEM near the ice table, suggesting the presence of liquid water in that horizon at some time. Further discussion and modelling of water in the UV soil column is presented below.

One of the interesting and unexpected results of the chemical analyses of the UV soils was the high level of perchlorate (ClO_4^-) measured in at least some horizons in both valleys. The search for natural ClO_4^- in the MDV was motivated by the unexpected discovery of ClO_4^- on Mars by PHX (Hecht *et al.* 2009, Kounaves *et al.* 2010b), and our findings are comparable to those at the PHX site. ClO_4^- was distributed throughout the UV soil column (Fig. 6b; true for both UV pits). However, in TV, it was discontinuously distributed; it was only detected in the 12–17 cm horizon and at the ice table interface layer (28+ cm). Similarly, it was discontinuously distributed in all the TV pits. Since perchlorate is highly soluble, and is retained in UV's ultraxerous soils, its presence throughout the profile is an indication of the relative lack of liquid water influence.

Across all UV samples (in both pits) there is a correlation between NO₃⁻ and ClO₄⁻ abundances ($r^2 = 0.85$), supporting the hypothesis that ClO₄⁻ is globally formed in the atmosphere, but survives in the soil only in hyperarid environments. The discovery of this natural ClO₄⁻ in the MDV and its potential presence over the entire ice cap has significant implications for its origin and interaction with aqueous environments and microbial communities (Kounaves *et al.* 2010a).

Biology

Our biological data were meant to provide 1) an indication of whether microbial biomass was or was not present and its relative (not absolute) abundance among layers in each pit, and 2) a broad indication of the major types of metabolic and structural groups present as measurable by the PLFA method. This was not meant to be a characterization of microbial community composition or diversity but, rather, since the same methods were used for each and every soil sample, to offer an inter-comparative way of delineating heterogeneity. Such information can guide future work in which the microbial communities and diversity could be characterized using DNA and RNA based phylogenetic approaches. These results, while not definitive, establish the presence of microorganisms in all sampled horizons down to and including the ice table in both UV and TV - a fact that has not previously been reported for ultraxerous zones.

Our PLFA analysis showed that microbes are more numerous and also represent a greater number of metabolic and structural groups in the subxerous, lower TV soils than the ultraxerous UV soils (Fig. 8): TV had both aerobic and anaerobic microbial groups. In contrast, anaerobic microbial groups were undetectable in the dry, oxidized soils of UV. In TV, the greater amount of liquid water allows drainage and pooling leading to saturated zones in the soil, which subsequently become anoxic, resulting in a heterogeneous environment that supports a greater number of microbial groups. In the subxerous MDV soils, recent studies demonstrated that the microbial content approached that of temperate desert soils, $\sim 10^8$ cells g⁻¹, (Cowan *et al.* 2002) and phylogenetic analyses (Aislabie et al. 2008) showed that these communities are remarkably diverse, a fact that relates to the stability and abundance of liquid water.

The microbial population in TV is, in fact, on a broad level more similar to the Arctic population, a known wet and cold environment, than to population in UV. Figure 9 shows a comparison of the ice-table laver PFLA results for TV. UV. and an Arctic permafrost location. The Arctic samples were collected in midsummer from the bottom of the active layer (the very top of the ice-cemented ground) at a site with patterned ground (79.40451°N, 90.78466°W, 116 m elevation) on a flank about one hundred metres above Expedition Fjord, down slope from the McGill High Arctic Research Station (for a description of the area around the Station see Pollard et al. 1999, Heldmann et al. 2005). They were analysed by the same company and in the same way as our MDV samples, and are consistent with the findings of other researchers (e.g. Steven et al. 2006). Both the Arctic and TV samples, which come from a "wet" permafrost environment where periodic water saturation and therefore anoxic conditions occur near the ice table, had both aerobic and anaerobic microbial groups. In contrast, anaerobic microbial groups were undetectable in the dry, oxidized soils of UV.

We hypothesize that the biomass differences between TV and UV are strongly influenced by the amount of available water for microbes to use. However, other factors may also play a role, such as differences in soil geochemistry and mineralogy, especially the availability of necessary ions and soluble metal species that may be more readily accessible and exploitable by microorganisms in the wetter environment of the TV soils.

The generally higher biomass estimates given by the DNA-based method as compared to PLFA estimates (Fig. 7) may be due to several reasons. Previous studies have shown that, even in Antarctica, where molecules may be expected to resist degradation longer than in warmer regions, PLFA do not persist very long in a measurable form after the death of the organisms which formed them (Cowan *et al.* 2002). Therefore, our PLFA results may represent live biomass only while the DNA estimate includes both live and dead biomass. Secondly, the PLFA may show lower extraction efficiency or the models used to relate PLFA amount to microbial cell



Fig. 10. SEM image showing a pristine, apparently autochthonous gypsum crystal from the +34 cm horizon in UV.

numbers are not appropriate for the soils we studied. However, the key findings are that biomass is present in every layer sampled in both valleys, and numbers are higher near the surface and near the ice-cemented ground than in the intervening layers in UV.

University Valley soil history and habitability

While there is a body of knowledge about the high-elevation Dry Valleys and their dry permafrost environment, our UV



Fig. 11. Soil temperatures from University Valley, January 2010. Data are from Marinova *et al.* (2011). The 0°C isotherm does not reach beyond ~ 10 cm. At the ice table (36 cm), temperatures do not exceed -12°C.



Fig. 12. a. Modelled subsurface temperatures at the University Valley pit for midsummer (~3 January). **b.** Modelled water activity, a_w (z-axis), for two conditions. The upper surface represents snowmelt and infiltration, while the lower surface, calculated for the same thermal history, applies to dry, post-infiltration conditions. Both activity profiles are anchored at $\sim a_w = 0.87$ by the ice table at 34 cm. The contour for $a_w = 0.6$ is marked (pink).

findings show some differences from other reports of ultraxerous soils, indicating that the soil history and habitability of this valley may be unique. First, none of the commonly seen salt accumulations were found. Second, and even more peculiar, evidence was seen of autochthonous gypsum in soil horizons that are far too cold and dry to permit subaqueous precipitation (Fig. 10; also seen in the other UV pit). Third, our horizon-by-horizon sampling showed the greatest microbial population in the surface layer with secondary peaks in microbial population in the 1–9 cm layer and in the two bottom layers, including the ice table; biomass depth profiles in the ultraxerous soils have not been previously reported, only soil surface biomass estimates exist.

In an attempt to explain the autochthonous gypsum, we consider the possibility that it indicates occasional

penetration of liquid water all the way to the ice table. The maximum depth reached by liquid water on infiltrating a frozen soil depends upon its temperature and the rate of infiltration. Meltwater entering the soil profile will drain under gravity, filling pores and dissolving salts until either, a) gravity is equalled by matric suction, or b) the water freezes. We use our sub-surface thermal model to investigate the sub-surface temperatures and water activity during midsummer in UV. Figure 11 shows soil temperatures in UV recorded for the month of January 2010 two years after our samples were collected (Marinova et al. 2011). These were used to ground-truth our model. The soil pit in which these data were recorded is at 77°51'35.04"S, 160°43'1.14"E; elevation: 1684 m. The depth to ground ice was 36 cm (cf. 34 cm at pit 7). The distance between our pit (pit 7) and the pit used by Marinova et al. (2011) is 358.8 m, and the bearing is 222°31'. At no time during January 2010 did the 0°C isotherm reach as deep as 10 cm. The temperature at the ice table was $\sim -14^{\circ}$ C throughout the month.

The sub-surface thermal model predicts the sub-surface temperature field for a full day, near the warmest part of the year (3 January), and reproduces the main features observed in the data, specifically that the surface temperature exceeds 0°C during the warmest part of the day and the maximum penetration of the 0°C isotherm is < 10 cm (Fig. 12a). In addition, the temperature of the ice table, at 34 cm, is fixed by the measurements shown in Fig. 11.

This modelling suggests, in spite of the uncertainty in total snow accumulation at UV and compounded by the uncertainty in the ratio of sublimated vs melted snow, that substantial snowmelt and infiltration events that allow water to reach the ice table are rare. If liquid water is to percolate to the ice table through the frozen overburden, latent heat of freezing must release enough heat to raise the soil temperature to 0°C. Integrating the temperature profile between 8 and 34 cm, and using the measured volumetric heat capacity of the soil $(1.7 \times 10^6 \text{ Jm}^{-3} \text{ K}^{-1})$, the required heat is equivalent to freezing a 5 cm column of liquid water, assuming the water enters the soil column at 0°C. This seems an unlikely daily infiltration rate in the current climate. The instability of liquid water at depth is consistent with the maximum abundance of salts that occur within the 1-9 cm horizon and also with the observation that visible oxidation was confined to the uppermost 10 cm of the profile.

If it is problematic to get liquid water at the depth of the gypsum in the current environment that may not always have been the case. UV soils are derived from the sandstone and dolerite found in the valley walls, consistent with Linkletter *et al.* (1973) and we interpret the valley floor to be the result of an inflationary process since the friable Beacon Sandstone weathers relatively rapidly. Therefore, this horizon may have formed early in the history of these soils and been subsequently buried and preserved by addition of sandstone and dolerite. The scarcity of rocks and pebbles in the soil

column, the absence of an indurated saltpan, and the preponderance of sand-sized quartz, argue that inflation has played a significant role in establishing the soil column. For an inflationary surface, older material is progressively buried, and the autochthonous $CaSO_4 \cdot 2H_2O$ may have been precipitated at a time when it was within the active layer, and liquid water a common occurrence.

In an attempt to explain the UV microbial population peaks in the uppermost subsurface and near the ice table, we hypothesize that these peaks may be the result of adaptation to two water reservoirs: occasional surface snow with rare meltwater, and the essentially invariant ground ice, which maintains high relative humidity at the bottom of the soil column.

Assuming the microbial biomass peaks represent more clement environments, the thermal model can again be used to estimate bounding values of the water activity (defined as the partial pressure of water, relative to the saturation vapour pressure over liquid water, even at temperatures $< 0^{\circ}$ C). The activity of water, a_w , is considered a key environmental parameter for microbial survival and a bulk value of $a_w = 0.6$ represents the lowest limit for which growth has been seen ($a_w = 0.61$; Grant 2004).

In summer, one may imagine that some snow cover might melt. However, snow cover is limited in UV (McKay 2009), and for much of the summer, relatively dry conditions would be expected. We capture these two conditions in Fig. 12b. The upper surface indicates a condition where surface snow is melting and infiltrating whereas the lower surface represents drier summertime conditions. For the case in which surface snow melts (upper surface), where $T > 0^{\circ}$ C, it is assumed liquid water is present and $a_w = 1$ (surface down to ~ 8 cm for a portion of the day). The lower boundary condition is essentially invariant, and set by the temperature of the ice table at -14°C, which fixes $a_w \approx 0.87$. We assume that sufficient water is present throughout the soil to saturate the pore spaces. Thus, when snow is melting, a_w is greater than the biological bulk threshold value of 0.6 for the entire soil column for most of the day. These conditions however, are unlikely to be long-lived and they evolve to much harsher conditions once the snowmelt has ended.

Drier summertime conditions are represented in the lower surface of Fig. 12b. The relative humidity of the atmosphere as measured by the TECP is used as the upper boundary, and the same sub-surface ice layer sets the lower boundary; the a_w equals 0.6 from the surface to about 28 cm depth or 6 cm above the ice layer. Between 28 cm and the ice table, $a_w > 0.6$, and summer temperatures are -14 to -10°C. Thus, near the ground ice table there exists a unique and viable habitat, and microbial growth is possible continuously throughout much of the summer. We interpret the increase in biomass near the ice table to be a consequence of this clement environment. However, it should be noted that microorganisms deposited at the

surface could be progressively buried and reach the ice table after some 5000 to 50000 years (Mahaney *et al.* 2001). In the cold dry environment of the UV soils, they could remain viable but dormant for this entire time.

Intermediate soils (13 cm < z < 28 cm predicted in the model) experience neither infiltration of melt, nor the consistently high relative humidity of the lowest layer, and our data show substantially smaller biomass in the 9–20 cm horizon. The detection of PLFA and DNA molecules throughout the soil column may mean that there are microenvironments in the "dry" permafrost in which sufficient unfrozen water is occasionally present to allow metabolic processes, or that the remains of dead cells from the surface can be transported down through the soil column.

University Valley and the Phoenix landing site

Qualitatively, both UV and the PHX landing site in the Martian Arctic can be characterized as dry permafrost overlying ice-cemented ground, and this was one of the initial motivations to test Phoenix hardware in UV.

In the Martian Arctic today, water is primarily in vapour and ice form, with perhaps discontinuous and immobile unfrozen water, due to the very cold temperatures. In fact, these low temperatures restrict the activity of water to an annual range of $a_w \sim 0.6$ –0.1. However, at UV, calculations based on vapour pressure over ice show that soil pores at the ice table range annually between $a_w \sim 0.9-0.74$. Obliquity changes may increase the Martian Arctic temperatures to become similar to those in UV today, but whether the a_w increases with this temperature increase is extremely uncertain, due to our poor understanding of global and boundary layer dynamics at high obliquity, as well as the details and kinetics of Martian global water redistribution. It is however, at least plausible that the PHX site could support metabolic activity of microbes during periods of high obliquity. One limitation on the habitability of the PHX site might be the requirement that, due to the obliquitychange cycle, microbial communities might need to survive dormancy periods on the order of 10^6 years (Zent 2008). Diurnal temperature changes across most of Mars are extreme in the current epoch, but temperature variations are less extreme at high latitudes, even less beneath a few centimetres of highly insulating regolith, and less again at high obliquity. Consequently, diurnal temperature variations are not thought to be a limiting factor for habitability.

If, in a given orbital cycle period, Mars had sufficient a_w to support microbial activity, we consider the very different soil chemistry at UV and PHX, and metabolic pathways that might be available. In UV, the soil is continually inoculated with organic compounds and cells from the Beacon Sandstone cryptoendoliths (Nienow & Friedmann 1993). It is therefore probable that the microbial population at UV primarily metabolizes through heterotrophic oxidation of those organics. The presence of organics on

the Martian surface, however, is far less clear. Unaltered carbon delivered to the Martian surface via meteoritic infall is estimated to be 2.4×10^5 kg yr⁻¹ (Flynn 1996), but organics have not been confirmed by robotic exploration. Further, recent results by Navarro-Gonzalez et al. (2010) indicate that organics may have been contained in the Viking samples, but would not have been detected if some perchlorate were present in those samples as well. Thus, since there is still no concrete evidence for organic compounds in the Martian regolith, a putative Martian biota might require an inorganic carbon source (e.g. atmospheric CO₂) as well as a plausible metabolic pathway. We suggest that in the absence of preformed organic carbon in the Martian soil the perchlorate discovered by PHX could provide an electron acceptor. The PHX soils have < 1% perchlorate (Hecht *et al.* 2009) and there is no toxicity to microbes even at much higher concentrations. Experiments with a microbial culture obtained from a contaminated deep vadose zone showed that the organisms were able to reduce perchlorate and had no detrimental effects at levels up to 2% perchlorate (Gal et al. 2008). Therefore, the levels of perchlorate found at the PHX site do not necessarily preclude habitability.

Terrestrial perchlorate-reducing bacteria can reduce perchlorate with a number of different electron donors: hydrogen, a variety of organic species, Fe^{2+} , and H_2S (Coates & Achenbach 2004). Autotrophic (i.e. those which use inorganic carbon sources) perchlorate-reducing bacteria have been found to be widespread in natural, uncontaminated environments (Miller & Logan 2000) so that autolithotrophic metabolism using H₂, CO₂, and perchlorate is plausible. Mars is a basaltic planet, so one might hypothesize that soluble Fe²⁺ from unweathered primary minerals would be available in soil solution for microbial metabolism. Although Kounaves et al. (2010b) reported $Fe^{2+} \le 10^{-4} M$ (the detection limits) in the PHX WCL data, this amount was measured in bulk soil and is higher than that exploitable by microorganisms in soils on Earth. Micro-zones such as those at the ice/soil particle interface might have exploitable ferrous iron unmeasurable by the WCL. In summary, it is plausible that the appropriate a_w can exist in certain epochs, and that there exists metabolic pathways for microbes, even in the absence of organics.

Conclusions

We present the results of a comprehensive, horizon-byhorizon analysis of the soil column from 1 of 4 subxerous lower TV pits and 1 of 2 ultraxerous UV pits which investigated the effects of ever-decreasing unfrozen water in cold deserts. These results serve as comparison points for the investigation of dry permafrost on Mars.

Where the air and soil column warm to $>0^{\circ}$ C annually, liquid water is abundant. Soluble salts were less abundant, and the vertical distribution of clay-sized fractions suggested translocation via liquid water. Mineral assemblages of pyrite and Fe-oxide were evidence of recent (probably seasonal) variations in the redox state of the soil. Soil biomass in subxerous soils was nearly equivalent to those found in temperate soils and similar to arctic permafrost ice table results. The number of metabolic and structural groups was also higher, which argues that a variety of soil microenvironments were occupied, and that the extreme low temperatures in the MDV, *per se*, are no significant barrier to habitability.

In UV, slightly cooler soil and air temperatures severely limit the occurrence of water in the soil. As a result, soluble salts, clay-sized particles, and biomass are all concentrated near the surface, where the limited annual liquid water occurs. Salts are more abundant than in subxerous soils, soluble salts particularly so. Soil biomass is far less abundant and there are fewer metabolic and structural groups than in TV, but nowhere did we find a sterile soil horizon, or one lacking claysized particles and soluble ions. Further, the lack of altered mica particles indicates minimal aqueous alteration, unlike the mica in the lower TV soils. Under such challenging conditions, microbial populations may be measurably enhanced in microenvironments that are marginally less stressful in terms of a_w , such as near the ice table.

Although the PHX landing site is currently too cold to support a_w values necessary for metabolism, orbital variations may periodically bring the soil to within the T- a_w domain of terrestrial psychrophiles. With the presence of organics uncertain, but possible, autotrophic perchlorate reducers could be the possible source of primary production. Survival of long, but indeterminate dormancy periods between high obliquities would be essential.

Acknowledgements

The authors would like to thank two anonymous reviewers for very thorough reviews of this paper that helped substantially improve it. Part of this work was conducted at the Jet Propulsion Laboratory/Caltech under a grant funded through the International Polar Year research opportunity with NASA. We thank the NSF Office of Polar Programs and the men and women of the US Antarctic Program for outstanding support of field operations.

References

- AISLABIE, J.M., JORDAN, S. & BARKER, G.M. 2008. Relation between soil classification and bacterial diversity in soils of the Ross Sea region, Antarctica. *Geoderma*, 144, 9–20.
- ANDERSON, D.M. & TICE, A.R. 1972. Predicting unfrozen water contents in frozen soils from surface area measurements. *Highway Research Record*, 393, 12–18.
- ASTROMATERIALS RESEARCH OFFICE. 2006. Astromaterials curation facility cleaning procedures for contamination control. Doc. # JSC-03243, Rev. D. Houston, TX: NASA Johnson Space Center.
- BALKWILL, D.L., LEACH, F.R., WILSON, J.T., MCNABB, J.F. & WHITE, D.C. 1988. Equivalence of microbial biomass measures based on membrane lipid and cell wall components, adenosine triphosphate, and direct counts in subsurface aquifer sediments. *Microbial Ecology*, 16, 73–84.

- BAO, H. & MARCHANT, D.R. 2006. Quantifying sulfate components and their variations in soils of the McMurdo Dry Valleys, Antarctica. *Journal of Geophysical Research*, 111, 10.1029/2005JD006669.
- BOCKHEIM, J.G. 1997. Properties and classifications of cold desert soils from Antarctica. Soil Science Society of America Journal, 61, 224–231.
- CAMPBELL, I.B. & CLARIDGE, G.G.C. 1982. The influence of moisture on the development of soils of the cold deserts of Antarctica. *Geoderma*, 28, 221–238.
- CAMPBELL, D.I., MACCULLOCH, R.J.L. & CAMPBELL, I.B. 1997. Thermal regimes of some soils in the McMurdo Sound region. In LYONS, W.B., HOWARD-WILLIAMS, C. & HAWES, I., eds. Antarctica, ecosystem processes in Antarctic ice-free landscapes. Rotterdam: Balkema, 45–55.
- CLARIDGE, G.G.C. & CAMPBELL, I.B. 1977. The salts in Antarctic soils, their distribution and relationship to soil processes. *Soil Science*, **123**, 377–384.
- COATES, J.D. & ACHENBACH, L.A. 2004. Microbial perchlorate reduction: rocket-fuelled metabolism. *Nature Reviews Microbiology*, 2, 569–580.
- COWAN, D.A., RUSSELL, N.J., MAMAIS, A. & SHEPPARD, D.M. 2002. Antarctic Dry Valley mineral soils contain unexpectedly high levels of microbial biomass. *Extremophiles*, 6, 431–436.
- DE VRIES, D.A. 1952. A nonstationary method for determining thermal conductivity of soil *in situ. Soil Science*, **73**, 83–89.
- DORAN, P.T., PRISCU, J.C., LYONS, W.B., WALSH, J.E., FOUNTAIN, A.G., MCKNIGHT, D.M., MOORHEAD, D.L., VIRGINIA, R.A., WALL, D.H., CLOW, G.D., FRITSEN, C.H., MCKAY, C.P. & PARSONS, A.N. 2002. Antarctic climate cooling and terrestrial ecosystem response. *Nature*, **415**, 517–520.
- FLYNN, G.J. 1996. The delivery of organic matter from asteroids and comets to the early surface of Mars. *Earth, Moon, and Planets*, 72, 469–474.
- FOUNTAIN, A.G., NYLEN, T.H., MONAGHAN, A., BASAGIC, H.J. & BROMWICH, D. 2010. Snow in the McMurdo Dry Valleys, Antarctica. *International Journal of Climatology*, **30**, 633–642.
- FRIEDMANN, E.I. 1982. Endolithic microorganisms in the Antarctic cold desert. Science, 215, 1045–1053.
- GAL, H., RONEN, Z., WEISBROD, N., DAHAN, O. & NATIV, R. 2008. Perchlorate degradation in contaminated soils and the deep unsaturated zone. *Soil Biology & Biochemistry*, **40**, 1751–1757.
- GRANT, W.D. 2004. Life at low water activity. *Philosophical Transactions* of the Royal Society London, B359, 1249–1267.
- HALL, B.L. & DENTON, G.H. 2000. Radiocarbon chronology of Ross Sea Drift, eastern Taylor Valley, Antarctica: evidence for a grounded ice sheet in the Ross Sea at the last glacial maximum. *Geografiska Annaler*, A82, 305–336.
- HECHT, M.H., KOUNAVES, S.P., QUINN, R.C., WEST, S.J., YOUNG, S.M.M., MING, D.W., CATLING, D.C., CLARK, B.C., BOYNTON, W.V., HOFFMAN, J., DEFLORES, L.P., GOSPODINOVA, K., KAPIT, J. & SMITH, P.H. 2009. Detection of perchlorate and the soluble chemistry of Martian soil at the Phoenix lander site. *Science*, **325**, 65–69.
- HELDMANN, J.L., MCKAY, C.P., POLLARD, W.H., ANDERSEN, D.T. & TOON, O.B. 2005. Annual development cycle of an icing deposit and associated perennial spring activity on Axel Heiberg Island, Canadian High Arctic. *Arctic, Antarctic, and Alpine Research*, 37, 127–135.
- JACKSON, M.L. 1985. Soil chemical analysis advanced course, 2nd ed. Madison, WI: University of Wisconsin, 930 pp.
- KOUNAVES, S.P., HECHT, M.H., WEST, S.J., MOROOKIAN, J., YOUNG, S., QUINN, R., GRUNTHANER, P., WEN, X., WEILERT, M., CABLE, C.A., FISHER, A., GOSPODINOVA, K., KAPIT, J., STROBLE, S., HSU, P., CLARK, B.C., MING, D.W. & SMITH, P.H. 2009. The MECA wet chemistry laboratory on the 2007 Phoenix Mars Scout lander. *Journal of Geophysical Research*, 114, E00A19.
- KOUNAVES, S.P., STROBLE, S.T., ANDERSON, R.M., MOORE, Q., CATLING, D.C., DOUGLAS, S., MCKAY, C.P., MING, D.W., SMITH, P.H., TAMPPARI, L.K. & ZENT, A.P. 2010a. Discovery of natural perchlorate in the Antarctic Dry Valleys and its global implications. *Environmental Science & Technology*, 44, 2360–2364.

- KOUNAVES, S.P., HECHT, M.H., KAPIT, J., GOSPODINOVA, K., DEFLORES, L., QUINN, R., BOYNTON, W.V., CLARK, B.C., CATLING, D.C., HREDZAK, P., MING, D.W., MOORE, Q., SHUSTERMAN, J., STROBLE, S., WEST, S.J. & YOUNG, S.M.M. 2010b. The wet chemistry experiments on the 2007 Phoenix Mars Scout lander mission: data analysis and results. *Journal of Geophysical Research*, **115**, 10.1029/2009JE003424.
- LINKLETTER, G., BOCKHEIM, J. & UGOLINI, F.C. 1973. Soils and glacial deposits in the Beacon Valley, southern Victoria Land, Antarctica, New Zealand. *Journal of Geology and Geophysics*, 16, 90–108.
- MacLean, L.C.W., TYLISZCZAK, T., GILBERT, P.U.P.A., ZHOU, D., PRAY, T.J., ONSTOTT, T.C. & SOUTHAM, G. 2008. A high-resolution chemical and structural study of framboidal pyrite formed within a low-temperature bacterial biofilm. *Geobiology*, 6, 471–480.
- MAHANEY, W.C., DOHM, J.M., BAKER, V.R., NEWSOM, H.E., MALLOCH, D.V., HANCOCK, R.G.V., CAMPBELL, I., SHEPPARD, D. & MILNER, M.W. 2001. Morphogenesis of Antarctic paleosols: Martian analogue. *Icarus*, 154, 113–130.
- MARCHANT, D.R. & HEAD III, J.W. 2007. Antarctic Dry Valleys: microclimate zonation, variable geomorphic processes, and implications for assessing climate change on Mars. *Icarus*, **192**, 187–222.
- MARINOVA, M.M., MCKAY, C.P., HELDMANN, J.L., DAVILA, A.F., ANDERSEN, D.T., JACKSON, W.A., LACELE, D., PAULSON, G., POLLARD, W.H. & ZACNY, K. 2011. Sublimation-dominated active layers in the highlands of the Antarctic Dry Valleys and implications for other sites. *Lunar and Planetary Science Conference*, Abstract # 2644, 2 pp. (available at http://www.lpi.usra.edu/meetings/lpsc2011/pdf/2644.pdf)
- McKAY, C.P. 2009. Snow recurrence sets the depth of dry permafrost at high elevations in the McMurdo Dry Valleys of Antarctica. *Antarctic Science*, 21, 89–94.
- MCKAY, C.P., MELLON, M.T. & FRIEDMANN, E.I. 1998. Soil temperatures and stability of ice-cemented ground in the McMurdo Dry Valleys, Antarctica. *Antarctic Science*, 10, 31–38.
- MELLON, M.T., ARVIDSON, R.E., SIZEMORE, H.G., SEARLS, M.L., BLANEY, D.L., CULL, S., HECHT, M.H., HEET, T.L., KELLER, H.U., LEMMON, M.T., MARKIEWICZ, W.J., MING, D.W., MORRIS, R.V., PIKE, W.T. & ZENT, A.P. 2009. Ground ice at the Phoenix landing site: stability state and origin. *Journal of Geophysical Research*, **114**, 10.1029/2009JE00341.
- MICHALSKI, G., BOCKHEIM, J.G., KENDALL, C. & THIEMENS, M. 2005. Isotopic composition of Antarctic Dry Valley nitrate: implications for NOx sources and cycling in Antarctica. *Geophysical Research Letters*, **32**, L13817.
- MILLER, J.P. & LOGAN, B.E. 2000. Sustained perchlorate degradation in an autotrophic, gas-phase, packed-bed reactor. *Environmental Science & Technology*, 34, 3018–3022.
- NAVARRO-GONZÁLEZ, R., VARGAS, E., DE LA ROSA, J., RAGA, A.C. & MCKAY, C.P. 2010. Reanalysis of the Viking results suggests perchlorate and organics at mid-latitudes on Mars. *Journal of Geophysical Research*, **115**, 10.1029/2010JE003599.
- NIENOW, J.A. & FRIEDMANN, E.I. 1993. Terrestrial lithophytic (rock) communities. In FRIEDMANN, E.I., ed. Antarctic microbiology. New York: Wiley-Liss, 343–412.
- POLLARD, W.H., OMELON, C., ANDERSEN, D.T. & MCKAY, C.P. 1999. Perennial spring occurrence in the Expedition Fiord area of western Axel Heiberg Island, Canadian High Arctic. *Canadian Journal of Earth Sciences*, **36**, 1–16.
- POWERS, J.G., MONAGHAN, A.J., CAYETTE, A.M., BROMWICH, D.H., KUO, Y. & MANNING, K.W. 2003. Real time mesoscle modeling over Antarctica: the Antarctic mesoscale prediction system. *Bulletin of the American Meteorological Society*, 84, 1533–1545.
- REMPEL, A.W., WETTLAUFER, J.S. & WORSTER, M.G. 2004. Premelting dynamics in a continuum model of frost heave. *Journal of Fluid Mechanics*, 498, 227–244.
- RIETVELD, H.M. 1969. A profile refinement method for nuclear and magnetic structures. *Journal of Applied Crystology*, **2**, 65–71.
- SAWLOWICZ, Z. 2000. Framboids: from their origin to application. *Prace Mineralogzne*, **88**, 1–80.

- SLETTEN, R.S., HALLET, B. & FLETCHER, R.C. 2003. Resurfacing time of terrestrial surfaces by the formation and maturation of polygonal patterned ground. *Journal of Geophysical Research*, **108**, 10.1029/2002JE001914.
- SMITH, P.H., TAMPPARI, L., ARVIDSON, R.E., BASS, D., BLANEY, D., BOYNTON, W., CARSWELL, A., CATLING, D., CLARK, B., DUCK, T., DEJONG, E., FISHER, D., GOETZ, W., GUNNLAUGSSON, P., HECHT, M., HIPKIN, V., HOFFMAN, J., HVIID, S., KELLER, H., KOUNAVES, S., LANGE, C.F., LEMMON, M., MADSEN, M., MALIN, M., MARKIEWICZ, W., MARSHALL, J., MCKAY, C., MELLON, M., MICHELANGELI, D., MING, D., MORRIS, R., RENNO, N., PIKE, W.T., STAUFER, U., STOKER, C., TAYLOR, P., WHITEWAY, J., YOUNG, S. & ZENT, A. 2008. Introduction to special section on the Phoenix Mission: landing site characterization experiments, mission overviews, and expected science. *Journal of Geophysical Research*, **113**, 10.1029/2008JE003083.
- STEVEN, B., LÉVEILLÉ, R., POLLARD, W.H. & WHYTE, L.G. 2006. Microbial ecology and biodiversity in permafrost. *Extermophiles*, **10**, 259–267.

- WHITE, D.C., DAVIS, W.M., NICKELS, J.S., KING, J.D. & BOBBIE, R.J. 1979. Determination of the sedimentary microbial biomass by extractable lipid phosphate. *Oecologia*, 40, 51–62.
- WILSON, A.T. 1979. Geochemical problems of the Antarctic dry areas. *Nature*, 280, 205–208.
- ZENT, A.P. 2008. An historical search for thin H₂O films at the Phoenix landing site. *Icarus*, **196**, 385–408.
- ZENT, A.P., HECHT, M.H., COBOS, D.R., WOOD, S.E., HUDSON, T.L., MILKOVICH, S.M., DEFLORES, L.P. & MELLON, M.T. 2010. Initial results from the thermal and electrical conductivity probe (TECP) on Phoenix. *Journal of Geophysical Research*, **115**, 10.1029/2009JE003420.
- ZENT, A.P., HECHT, M.H., COBOS, D.R., CAMPBELL, G.S., CAMPBELL, C.S., CARDELL, G., FOOTE, M.C., WOOD, S.E. & MEHTA, M. 2009. Thermal and Electrical Conductivity Probe (TECP) for Phoenix. *Journal of Geophysical Research*, **114**, 10.1029/2007JE00305.