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Pyrrolidine-Acridine hybrid in Artemisinin-based combination: a pharmacodynamic study

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Aiming to develop new artemisinin-based combination therapy (ACT) for malaria, antimalarial effect of a new series of pyrrolidine-acridine hybrid in combination with artemisinin derivatives was investigated. Synthesis, antimalarial and cytotoxic evaluation of a series of hybrid of 2-(3-(substitutedbenzyl)pyrrolidin-1-yl)alkanamines and acridine were performed and mode of action of the lead compound was investigated. *In vivo* pharmacodynamic properties (parasite clearance time, parasite reduction ratio, dose and regimen determination) against multidrug resistant (MDR) rodent malaria parasite and toxicological parameters (median lethal dose, liver function test, kidney function test) were also investigated. 6-Chloro-N-(4-(3-(3,4-dimethoxybenzyl)pyrrolidin-1-yl)butyl)-2-methoxyacridin-9-amine (15c) has shown a dose dependent haem bio-mineralization inhibition and was found to be the most effective and safe compound against MDRmalaria parasite in Swiss mice model. It displayed best antimalarial potential with artemether (AM)*in vitro*as well as*in vivo*. The combination also showed favourable pharmacodynamic properties and therapeutic response in mice withestablished MDR malaria infection and all mice were cured at the determined doses. The combination did not show toxicity at the doses administered to the Swiss mice. Taken together, our findings suggest that compound 15c is a potentialpartner with AM for the ACT and could be explored for further development.

Key words: Malaria, multidrug resistant, pyrrolidine-acridine hybrid, ACT, combination therapy, pharmacodynamics.

INTRODUCTION

Malaria continues to be the most prevalent parasitological diseases across the world, representing a global public health problem with approximately 300 million cases and 0.6 million deaths annually (World Health Organization, 2014). Although efforts to develop vaccine against the disease are yet to be translated to clinics, malaria can be effectively cured by chemotherapy. Unfortunately there was extensive resistance of malaria parasite to most of the available antimalarials including chloroquine, mefloquine, amodiaquine, pyrimethamine, sulfadoxine, atovaquone or their combinations thereof in specific areas. As a result, new antimalarials representing artemisinin or its derivatives, which kill the parasite rapidly became the drug of choice. However, artemisinin monotherapy has been banned to prevent the origin of resistance and only

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combination therapy is recommended, which also reduce transmission (White, 2008a, b). However, there are now evidences that parasite has developed resistance against artemisinin in Cambodia, the Lao People's Democratic Republic, Myanmar, Thailand and Viet Nam (Noedl et al. 2008; Okombo et al. 2014; Wells et al. 2015; WHO, 2016). Continued use of antimalarials with high level of resistance as the partner drug in artemisinin-based combination therapy (ACT) may enhance the chances of further drug resistance thereby increasing the malaria mortality rate (Bloland, 2001; White, 2008a, b). Therefore, development of new antimalarial agents directed towards potential and safe targets and their effectiveness in combination therapy could be a successful way forward in curing the malaria infection (Fong et al. 2015).

The acridine core is a privileged unit in medicinal chemistry as it is represented in several analogues endowed with diverse biological activities including antileishmanial, antitrypanosomal (Gamage *et al.* 1994), antitumour, (Baguley *et al.* 1981), antiprion (May *et al.* 2006) and anti-Alzheimer (Fang *et al.* 2008). In context of their antimalarial activity, different modes of action of acridine derivatives have been proposed viz. DNA intercalation (Chen et al. 1978), haem-binding (Dorn et al. 1998) and inhibition of topoisomerase II (Gamage et al. 1994). An acridine-based antimalarial pyronaridine, was demonstrated to be effective against chloroquine resistant strain of Plasmodium falciparum (P. falciparum) (Chang et al. 1992). Later, we and other independent groups also reported the antimalarial effect of pyronaridine against multidrug resistant (MDR) malaria parasite (Looareesuwan et al. 1996; Ringwald et al. 1996; Dutta et al. 2000; Tripathi et al. 2000). Recently pyronaridine-artesunate combination had been shown to be effective and safe for uncomplicated malaria in different clinical trials (Duparc et al. 2013) and this has been commercialized as Pyramax[®]. In a separate study it was reported that pyrrolidinoaminoalkane class of compounds show effectiveness as CQ-resistance reversers (De et al. 1993; Walter et al. 1993; Batra and Bhaduri, 1997). We anticipated that covalently linking the pyrrolidinoaminoalkane to acridine would offer a new series of hybrid compounds, which may display antimalarial effect. It may be noted that literature include reports of antimalarial effect of hybrid compounds wherein the 9-aminoacridine is covalently linked to polyarylmethyl group III & IV (Gemma et al. 2009), imidazole group V (Fattorusso et al. 2008), artemisinin VI & VII (Jones, 2009) or quinoline VIII (Kumar et al. 2010) (Fig. 1A). In this context we prepared a series of hybrid compounds belonging to prototype IX in which the pyrrolidinoaminoalkane subunit is tethered to 6-chloro-2-methoxyacridine at 9-position and investigated their antimalarial activity (Fig. 1B). The most active compound of the series was further examined for its potential as partner to artemisinin derivatives for the ACT. The drug interaction, pharmacodynamics and toxicological parameters of the hybrid compound and artemether (AM) were studied to assess the dose and regimen.

MATERIALS AND METHODS

Chemicals

AlbumaxII, Sybr Green-I, were purchased from Invitrogen. RPMI-1640, hypoxanthine and saponin were bought from Sigma Aldrich. Tween-20 and Giemsa's stain were procured from Merck, India. The chemicals used in the synthesis were either procured from Sigma-Aldrich or Spectrochem, India whereas the solvents were procured from Spectrochem, India, Central Drug House, India or Merck, India.

Animals

Laboratory Animal Centre, CSIR-Central Drug Research Institute, Lucknow, India. Animals were housed in polypropylene cages (~100 cm² areas per mouse) on paddy husk saw dust with proper ventilation, 12 h light/dark cycle and food water *ad libitum*. Housing temperature was maintained at 22–24 °C and humidity was controlled to 45–60%. All studies were conducted under approved protocol by the Institutional Animal Ethics Committee of CSIR-Central Drug Research Institute, Lucknow, India.

Chemistry

General Procedure for the synthesis of N-(2-(3-benzylpyrrolidin-1-yl)alkyl)-6-chloro-2-methoxyacridin-9-amines 13a-e-15a-e as exemplified for 6-chloro-N-(4-(3-(3,4-dimethoxybenzyl)pyrrolidin-1-yl)butyl)-2-methoxyacridin-9-amine 15c. To a solution of 12c (1.0 g, 3.42 mmol) in acetonitrile (20 mL) were added Et₃N (1.75 mL, 13.7 mmol) and 6,9-dichloro-2-methoxyacridine (0.95 g, 3.42 mmol). The reaction mixture was heated at reflux and after completion as monitored by thin layer chromatography (TLC), the solvent was removed under vacuum. The residue was extracted with CHCl₃ (3 $mL \times 50 mL$) and water (50 mL) and the organic phases were combined, dried over Na2SO4 and concentrated in vacuum to obtain the crude product. Purification of the crude product by column chromatography over basic alumina using MeOH/ CHCl₃ (1:99, v/v) as eluent afforded 1.26 g (69%) of 15c as yellow oil.

6-Chloro-N-(4-(3-(3,4-dimethoxybenzyl)pyrrolidin-1-yl) butyl)-2-methoxyacridin-9-amine (15c). $R_f = 0.53$ (MeOH/CHCl₃, 10:90, v/v); IR (Neat) $v_{\text{max}} = 3321$ (NH) cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ = 1.65-1.72 (m, 2H, CH₂), 1.80-1.84 (m, 5H, 2× CH₂ and CHH), 2·13-2·18 (m, 1H, CHH), 2·45-2.56 (m, 4H, $2 \times CH_2$), 2.59-2.76 (m, 3H, CH_2 and CH), 3.72-3.77 (m, 2H, CH₂), 3.84 (s, 3H, OCH₃), 3.85 (s, 3H, OCH₃), 3.95 (s, 3H, OCH₃), 6.67-6.78 (m, 3H, ArH), 7.26-7.28 (m, 2H, ArH), 7.41 (dd, 1H, $\mathcal{J}_1 = 2.6$ Hz, $\mathcal{J}_2 = 9.4$ Hz, ArH), 7.98– 8.05 (m, 3H, ArH); ¹³C NMR (75 MHz, CDCl₃) $\delta = 26.6, 29.7, 30.5, 39.2, 41.2, 50.8, 53.5, 54.2,$ 55.7, 56.0, 56.1, 60.2, 100.2, 111.3, 112.1, 115.7, 117.8, 120.5, 124.3, 128.1, 131.3, 133.8, 134.9, 146.8, 147.4, 148.4, 148.9, 150.3, 155.8; ESI-MS m/z = 534.3 $[M + H]^+;$ ESI-HR-MS m/z =534·2518, calcd. for $C_{31}H_{36}ClN_3O_3$ $[MH]^+$ 534·2523.

In vitro *antimalarial evaluation*. The compounds were evaluated for antimalarial activity against 3D7 (CQ-sensitive) as well as K1 (CQ-resistant) strains of *P. falciparum* using Malaria SYBR Green I nucleic acid staining dye based fluorescence (MSF) assay as mentioned by Singh *et al.* (2011).

Outbred Swiss mice of 6-8 weeks (20-22 g and either sex) were procured from National



Pyrrolidino aminoalkane derivarive of Acridine

Fig. 1. (A) Structure of acridine-based hybrid compounds, (B) proposed structure of acridine tethered pyrrolidino aminoalkane.

Experiments were performed in triplicates and repeated at least three times.

Cytotoxicity of the com-Cytotoxicity assay. pounds was carried out using Vero cell line (C1008; Monkey kidney fibroblast) following the method as mentioned in Sharma et al. (2014). The cells were incubated with compound-dilutions for 72 h and MTT ((3-(4,5-dimethylthiazol-2-yl)-2,5diphenyltetrazolium bromide) was used as reagent for detection of cytotoxicity. Fifty per cent cytotoxic concentration (CC₅₀) was determined using nonlinear regression analysis of dose response curves pre-programmed Excel spreadsheet. using Selectivity index (SI) was calculated as

$$\mathrm{SI} = \mathrm{CC}_{50}/\mathrm{IC}_{50}$$

In vitro *combinations assay.* In vitro interaction of newly synthesised compound (15c) and antimalarials

arteether (AE), AM and artesunate (AS) were determined according to Pandey *et al.* (2013). Experiments were performed in triplicates and repeated at least three times. Fractional inhibitory concentration (FIC) was interpreted by the following formula and subsequent isobologram were plotted.

FIC = Conc. of drug in combination to produce $IC_{50}/Conc.$ of drug alone require to produce IC_{50} .

The sum FIC (Σ FIC) value for each of the preparations determined by the following formula was used to classify the drug-drug interaction.

$$\Sigma FIC = \frac{IC_{50} \text{ of drug A in combination}}{IC_{50} \text{ of drug A alone}} + \frac{IC_{50} \text{ of drug B in combination}}{IC_{50} \text{ of drug B alone}}$$

 Σ FIC < 1 represents synergism, Σ FIC = 1 represents additive interaction while >1 was considered as antagonistic (Bell, 2005; Kelly *et al.* 2009). Mean Σ FIC for all the tested ratios were used to classify the overall trend of the interaction.

In vivo -day 4 parasitaemia suppression test. For in vivo evaluation, drugs were administered orally as Tween 80-water formulation. Different groups of Swiss mice (5 mice/group) were inoculated with 5×10^5 Plasmodium yoelii nigeriensis MDR infected erythrocytes obtained in anticoagulant through cardiac puncture from infected mice. After 2–4 h of infection, these groups were treated with daily oral doses for 4 days (0, 1, 2, and 3) at 100 mg kg⁻¹ day⁻¹. Mice in the control group were injected with parasites only and treated with vehicle. Parasitaemia was monitored on day 4 by Giemsa stained thin blood smears and suppression was determined relative to the control group.

Determination of antimalarial response of the lead combinations against P. yoelii nigeriensis MDR.

Different groups of Swiss mice (10 mice per group) were inoculated with 5×10^5 *P. yoelli nigeriensis* MDR -infected erythrocytes. For simultaneous drug response assessment, these groups were treated with daily oral doses of alone partners (AM or 15c) and their combinations at 2–4 h after infection whereas treatment was done after observation of parasitaemia in the blood of infected mice (5 mice per group) for the evaluation of therapeutic antimalarial response. Mice in the control group were injected with parasites only and treated with vehicle.

Parasitaemia was monitored by Giemsa stained thin blood smears on pre-determined days and survival of the mice was duly recorded (Tripathi *et al.* 2011).

Determination of pharmacodynamic properties.

The pharmacodynamic properties (parasite reduction ratio (PRR), parasite clearance time (PCT), recrudescence studies etc.) were explored according to the protocol defined by White (1997) with some modifications. Briefly, a calculated amount of parasites was administered intraperitonially to a group of 5 Swiss mice and parasitaemia was determined on the successive days. Upon appearance of parasitaemia in the blood, single dose of the different combinations as well as individual drugs were given. Giemsa's stained thin blood smears were examined under 100X objective (oil immersion). The PRR was calculated as per the formula

$$au = P_0/P_2$$

 P_0 is parasite burden per micro litre (μ L) at the time of drug treatment.

 P_2 is parasite burden μ L after 24 h of drug treatment.

The parasite burden was calculated as follows:

Parasite burden (P) per $\mu L = 4 \times 10^6 \times \text{parasit-aemia } 100^{-1}$ (The Swiss mice have 4×10^6 RBCs per μL of blood).

The PCT (*T*) was determined with the following assumption: if the blood volume of an average mouse of 20–22 g is assumed to be 2.0 mL and *P* is the parasite count per μ L, then the total parasite burden (*B*) is given by:

$$B = 2 \times 10^3 \times P = 2P \times 10^3$$

Therefore,

$$\log_e B = \log_e P + 7 \cdot 6$$

If $\tau = P0/P_2$, then the time (*T*) in days for which parasites are present in the body (with an asexual cycle of ~1 day) is given by:

$$T = (\log_e P + 7 \cdot 6) / \log_e \tau$$

T is the time for which therapeutic concentration of an antimalarial drug must be present in the host plasma, P is the parasite burden per μ L and τ is parasite reduction ratio.

Determination of median lethal dose. Median lethal dose (LD_{50}) was determined according to the method of Srivastava *et al.* (2005) with some modifications. Briefly, higher oral doses of test compound was given to four groups of 5 Swiss mice (20–22 g) and housed according to ethical guidelines. Mice were observed for 2 days and mortality was duly recorded. On the basis of mortality in the different groups, LD_{50} was calculated with the help of Prism software.

Biochemical evaluation of liver, kidney and blood parameters. Four groups of Swiss mice (5 mice per group) were treated with alone partners (200 mg kg⁻¹ day⁻¹ 15c or 60 mg kg⁻¹ day⁻¹ AM) or their combinations for 4 days. Blood was withdrawn by cardiac puncture and allowed to stand undisturbed for 30 min. Serum was separated and levels of urea, blood urea nitrogen, total bilirubin, creatinine, alanine transaminase, aspartate aminotransferase and alkaline phosphatase were estimated using fully automated biochemical analyser (Merck-selectra junior).

Assessment of haem bio-mineralization inhibition.

Haem biomineralization inhibition in the extract of *P. yoelii*-infected erythrocytes was assayed by a method described by Agrawal *et al.* (2002) and the quantification of hemozoin was done using an extinction coefficient of 91 mm⁻¹ cm⁻¹ at 400 nm.

Statistical analysis

Logit regression analysis of dose response curves using pre-programmed Excel spreadsheet (a kind Table 1. *In vitro* antimalarial activity of pyrrolidinoaminoalkane derivatives of acridine against CQsensitive (3D7) strain and CQ resistant (K1) strain of *P. falciparum*. Experiments were performed in triplicates and repeated 3 times.



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Compound no.	R^1	R^2	R^3	n	IC ₅₀ against Pf3D7 (nм)	SI for <i>Pf</i> 3D7	IC ₅₀ against <i>Pf</i> K1 (nM)	SI for <i>Pf</i> K1	LogP	CLogP
13a	Н	Н	Н	0	258	43	351	32	5.79	7.78
13b	Н	OMe	Н	0	311	27	342	25	5.66	7.57
13c	Н	OMe	OMe	0	190	41	109	37	5.53	7.44
13d	OMe	OMe	OMe	0	39	377	3492	1.6	5.41	7.08
13e	OCH_2O		Н	0	54.0	246	3267	11	5.57	7.75
14a	Н	Н	Н	1	279	532	1117	3.6	5.89	8.05
14b	Н	OMe	Н	1	360	30	1361	13	5.77	7.82
14c	Н	OMe	OMe	1	28	120	78.4	23	5.64	7.71
14d	OMe	OMe	OMe	1	3.7	2829	9.5	609	5.51	7.35
14e	OCH_2O		Н	1	432	1.5	1558	10	5.67	8.01
15a	Н	Н	Н	2	80	2246	2998	18	6.35	8.10
15b	Н	OMe	Н	2	17	628	10.9	495	6.22	7.85
15c	Н	OMe	OMe	2	30.3	140	9.6	236	6.09	7.75
15d	OMe	OMe	OMe	2	9.3	19011	5.2	3400	5.97	7.40
15e	OCH_2O		Н	2	1.7	1767	109.	14	6.12	8.06
CQ			-	-	6.3	23 305	253	495	3.73	5.06

gift from Drugs for Neglected Diseases Initiative-Geneva) was done for IC_{50} assessment. Mean, s.D. and s.E.M. were also calculated using Excel spread-sheet. Two tailed student *t*-test was performed for significance.

RESULTS

Chemistry

The synthesis of the title compounds is outlined in the Supplementary Material (Scheme 1). The dimesylates 6a-e were prepared from substituted benzaldehyde and γ -butyrolactone via the reported procedure (Batra *et al.* 2000). Reaction of 6a-e with several mono-Boc protected diamines afforded the Boc-protected pyrrolidinoaminoalkanes 7a-e-9a-e. Deprotecting the Boc-group in 7–9 with 4 N aq. HCl gave the free amines 10a-e-12a-e. Treating these free amines with 6,9-dichloro acridine in the presence of Et₃N in MeCN as medium produced the final products 13a-e-15a-e in moderate to good yields. All prepared compounds were investigated for their *in vitro* and *in vivo* antimalarial efficacy.

In vitro antimalarial activity and cytotoxicity

The compounds 13a-e-15a-e were subjected to in vitro antimalarial assay against CQ-sensitive

(3D7) as well as CQ resistant (K1) strains of *P. falciparum*. The antimalarial activity (IC₅₀, nM) was quantified as 50% inhibition of parasite growth and the results are reported in Table 1. The two points of variations were the phenyl ring and the alkane spacer. The five substitutions on the phenyl ring in compounds of 13a-e-15a-e, included hydrogen, 4-methoxy, 3,4-dimethoxy, 3,4,5-trimethoxy and 3,4-methylenedioxy, whereas the three variations in the alkane spacer included n = 0, 1, 2. As illustrated in Table 1, out of 15 compounds from the series, 8 compounds displayed antimalarial efficacy between 1.7 and 54 nm against 3D7 strain of P. falciparum. In general compounds (15) having butyl spacer displayed relatively potent activity as compared with the compounds bearing propyl (14) or ethyl spacer (13). On the other hand compounds bearing substitutions on the phenyl ring displayed better antimalarial effect as compared to the compounds with unsubstituted phenyl ring (13a, 14a, 15a). All compounds from 13a-e-15a-e besides having potent antimalarial efficacy displayed good SI. Compound 14d, which displayed the activity with $IC_{50} = 3.7$ nM showed SI of 2829, which was considered to be safe. Similarly 15d was also found to be safe having also good SI value of 19011.

Next the antimalarial effect of these compounds against CQ-resistant (K1) strain of *P. falciparum* was investigated. The IC₅₀ of CQ for this strain was



Fig. 2. Antimalarial potential of selected 3-(substituted benzyl)-pyrrolidino-aminoalkanes (A) Parasitaemia suppression on day4 at 100 mg kg⁻¹ × 4 doses against *P.yoelii nigeriensis* multidrug resistant (MDR) infected Swiss mice. (B) Mice survival graph.



Fig. 3. Dose dependent inhibitory activity of 15c on haem bio-mineralization. Error bars represent mean \pm standard deviation.

observed to be 253 nM. Compounds 14c-d and 15b-e displayed IC₅₀, which was several folds better as compared with CQ. Interestingly compound 15e, which had better efficacy than CQ against CQS strain was found to be relatively less potent as compared with other analogues. Nevertheless compounds 14d and 15d bearing trimethoxy substitution on the phenyl ring in the pyrrolidine moiety elicited potent antimalarial effect with IC₅₀ of 9·53 and 5·2 nM, respectively. Compound 15d also had a good SI of 3400 as compared with 495 of CQ. However, several of these compounds (13d-e, 14a-b, 14b, 15a and 15e) were less efficacious in K1 when compared with their activity in 3D7. Interestingly, 15c was found to have more potent antimalarial activity in CQ-resistant K1 strain.

Day 4 Parasitaemia suppression

Based on the results of the *in vitro* assays of these compounds, a few of them were examined for their *in vivo* antimalarial activity against MDR strain *P. yoelii nigeriensis* in outbred Swiss mice at a dose of 100 mg kg⁻¹ day⁻¹ × 4 days *via* oral route and the results are presented in Fig. 2. It was observed that all compounds at a dose of 100 mg kg⁻¹ × 4 days dose showed >90% suppression of parasitaemia on day 4. However, compound *15c* having dimethoxy substitution on the phenyl ring and 4-carbon spacer could make survival of 3 out of 5 mice beyond 28 days of observation period.

Mode of action and in vitro antimalarial interaction of 15c with artemisinin derivatives

In an attempt to investigate the possible mode of action of 15c, the haem bio-mineralization inhibition assay was performed wherein 15c showed a dose dependent haem bio-mineralisation inhibitory activity against *P. yoelii nigeriensis* (Fig. 3). Antimalarial activity of 15c in combination with artemisinin derivatives AM, AE and AS was investigated



Fig. 4. Isobologram representing antimalarial interaction of 15c with artemisinin derivatives with (A) artemether, (B) arteether, (C) artesunate. Diagonal lines presents the boundaries of synergy (<1), additivity (1) and antagonism (>1).

against 3D7 strain of *P. falciparum*. Interaction of 15c with AM and AE were found to be slightly synergistic (mean Σ FIC 0.68 ± 0.16 and 0.80 ± 0.15 , respectively) whereas it was slightly antagonistic with AS (mean Σ FIC 1.43 ± 0.31). From the results it was concluded that the differences between the interactions of 15c with the 3 artemisinins were minor. These interactions were supported by the isobolograms representing the interactions between the partner drugs (Fig. 4).

Table 2. An	timalarial pote	ntial (prophylacti	c) of 15c with	artemether a	igainst P. yoei	lii nigeriensis m	ultidrug resist.	ant in Swiss n	nice.		
Dena/	Does (Ma ba^{-1}	Parasitaemia (Mea	n±s.D.) at diff	erent days (nur	nber of mice w	ithout parasitaeı	mia/total survive	ed mice)		Mean survival	70
combination	× 4 days)	Day4	Day7	Day10	Day14	Day18	Day21	Day24	Day28	time (Days)	o, cure
15c + AM	100 + 25	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \; (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	>28	100
	50 + 25	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	>28	100
;	100 + 12.5	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \; (10/10)$	$0 \pm 0 \ (10/10)$	>28	100
	50 + 12.5	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	>28	100
	100 + 6.25	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	>28	100
	50 + 6.25	$0 \pm 0 \; (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \; (10/10)$	$0 \pm 0 \ (10/10)$	>28	100
:	-+25	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	$0 \pm 0 \ (10/10)$	>28	100
	- + 12.5	$0 \pm 0 \ (10/10)$	$0.002 \pm .006$	9.4 ± 26.89	0 ± 0 (8/8)	7.06 ± 7.95	0 ± 0 (4/4)	0.58 ± 0.63	5 ± 7.07 (2/3)	21.6	20
			(9/10)	(8/10)		(4/8)		(2/4)			
	-+6.25	$0 \pm 0 \; (10/10)$	0.42 ± 1.04	6.5 ± 12.31	5.71 ± 13.99	3.67 ± 5.67	$21 \cdot 25 \pm 38 \cdot 6$	0.8 ± 1.13	0 ± 0 (2/2)	19.6	20
			(6/10)	(6/8)	(6/7)	(3/6)	(3/4)	(2/3)			
;	100 + -	$0.05 \pm 0.12 \ (7/10)$	0.9 ± 1.58	10.25 ± 23.5	10.33 ± 15.76	9.9 ± 12.97	$0.3 \pm 0.4 \; (1/4)$	$0.01 \pm .01$	$0.002 \pm .004$	20.2	10
			(6/10)	(4/10)	(2/6)	(1/5)		(2/4)	(3/4)		
	50 +-	0.2 ± 0.25 (3/10)	26.4 ± 27.2	$41 \cdot 2 \pm 37 \cdot 08$	15.9 ± 14.1	$0.8 \pm 0.0 \ (0/1)$	$0 \pm 0 \; (1/1)$	$0.02 \pm 0 \; (0/1)$	$0 \pm 0 \ (1/1)$	13.4	0
			(0/10)	(0/0)	(0/2)						
Control		$39.1 \pm 18.6 \ (0/10)$								5.7	0

In vivo antimalarial potential of 15c with AM against MDR rodent malaria parasite

Antimalarial potential of oral 15c with AM, was evaluated against MDR P. yoelii nigeriensis in random-bred Swiss mice. Compound 15c individually and its different combinations with AM were administered orally to different groups of mice. 15c at $100 \text{ mg kg}^{-1} \text{ day}^{-1}$ and AM at 12.5 mg kg^{-1} day⁻¹ separately for 4 days could produce only 10, and 20% cure, respectively, while the combination of lower doses of 15c (50 mg kg⁻¹ day⁻¹) and AM $(6.25 \text{ mg kg}^{-1} \text{ day}^{-1})$ for 4 days, produced 100% cure. Additionally, the mean survival time for alone drugs at higher concentrations (100 mg kg^{-1}) 15c or 12.5 mg kg^{-1} AM) were also short (~20 days) in comparison with the combination at lower doses (50 mg kg⁻¹ 15c with 6.25 mg kg⁻¹ AM; >28 days) (Table 2).

In vivo Pharmacodynamics of 15c with AM

Pharmacodynamic study showed a gradual decrease in parasitaemia with the single dose of 30 and 60 $mg kg^{-1} AM$ alone and its combination with 200 mg kg⁻¹ 15c while a gradual increase was observed in 15c group after 12 h of dosing, After 24 h, there was negligible parasitaemia in the AM alone and combination group whereas in 15c alone group, all the mice possess significant parasitaemia in their blood (Table 3). Divided doses of $15c \ 100 \ \mathrm{mg \ kg^{-1}}$ at every 12 h could not produce any significant improvements in the results (data not shown).

On the basis of above observations, pharmacodynamic properties, i.e. PRR and PCT were calculated (Table 3). The PCT was much lower and PRR was higher in the combination groups (200 + 60) compared with alone groups. PRR for 30 and 60 mg kg^{-1} AM alone were 12 and 17.3, respectively, while 15c could not reduce the parasite load. Combination of 30 mg kg^{-1} of AM with 200 mg kg^{-1} of 15c was not found as much pharmacodynamically favourable over alone treatment with a PRR value of 14. On the other hand, combination of 200 $mg kg^{-1}$ 15c with 60 mg kg^{-1} AM had reduced the parasite load significantly having the PRR value 880. The calculated PCT in the most potent combination 60 mg kg⁻¹ AM + 200 mg kg⁻¹ 15c is found to be much lower than AM alone (2.8 vs 6.5 days).

Therapeutic antimalarial potential of 15c with AM against MDR rodent malaria parasite

15c and AM combinations were also evaluated and optimized against established infection of P. yoelii nigeriensis in Swiss mice. The combination (200 $mg kg^{-1} 15c + 60 mg kg^{-1}$ AM) was having the lowest PCT (2.8 days) and highest PRR (880) was found as most appropriate combination.

Drug/combination $(M_{1}, 1, -1)$	Mean % para	isitaemia at diff	ferent time inte	ervals \pm s.d.			
$(Mg kg \times I day)$ 15c + AM	0 h	6 th hour	12 th hour	18 th hour	24 th hour	PRR	PCT (Days)
200 + 30	1.26 ± 0.46	1.38 ± 0.38	1.02 ± 0.52	0.42 ± 0.25	0.09 ± 0.1	14	7.05
200 + 60	1.76 ± 0.38	1.52 ± 0.71	0.82 ± 0.41	0.16 ± 0.16	0.002 ± 0.004	880	2.8
-+ 30	0.86 ± 0.44	1.26 ± 0.69	0.72 ± 0.56	0.31 ± 0.21	0.07 ± 0.12	12	7.2
-+60	1.04 ± 0.49	1.24 ± 0.55	0.48 ± 0.39	0.10 ± 0.12	0.06 ± 0.005	17.3	6.5
200 + -	0.98 ± 0.61	2.07 ± 1.2	2.78 ± 1.7	3.92 ± 2.2	12.2 ± 6.34	NA^{a}	NA^{a}
Control	0.72 ± 0.43	3.16 ± 2.3	3.9 ± 2.8	6.42 ± 4.09	$25{\cdot}6\pm18{\cdot}6$	_	-

Table 3. In vivo pharmacodynamics of 15c with artemether.

PCT, parasite clearance time; PRR, parasite reduction ratio.

^a Parasitaemia not reduced.

When 200 mg kg⁻¹ 15c + 60 mg kg⁻¹ AM was given to infected Swiss mice for 4 days, no parasitaemia was observed throughout the study and all the mice in the group survived beyond the observation period of 28 days. While the combination having higher PCT of 7.05 (200 mg kg⁻¹ 15c + 30mg kg⁻¹ AM) when given for 4 days, recrudescence occurred on day 24 and cure rate was also reduced to 80%. In alone groups maximum 20% cure was observed and mean survival time was also lower than the combination groups (>28 vs <20 days), which further indicated the strong antimalarial potential of 15c with AM (Table 4).

In vivo toxicity of 15c in combination with AM

On the basis of mortality in the different groups of treated mice, LD_{50} of newly synthesized compound 15c was calculated and it was 10 times higher than its curative dose in combination (data not shown).

The potent antimalarial combination (200 mg kg⁻¹ 15c + 60 mg kg⁻¹ AM) had no significant effect on key parameters of liver and kidney. All the liver and kidney parameters were found to be normal for both the combinations (data not shown).

DISCUSSION

The increase of resistance by the parasite to common antimalarial chemotherapy was the major reason for WHO advocating the use of ACT as the first-line of drugs in endemic areas. Unfortunately, there are now reports that the malaria parasite has developed resistance to artemisinin derivatives too (Noedl et al. 2008; Wells et al. 2015). Furthermore, the continued application of ineffective antimalarial as a partner drug in ACT may increase the chances of drug resistance thereby leading to rise in malaria mortality. Therefore, ACT with a new partner drug directed towards safer target might be effective and well tolerated (Bloland, 2001; White, 2008a, b). The present study concerns with exploration of pyrrolidinoaminoalkane-acridine hybrids as a partner in ACT and its pharmacodynamic studies.

The antimalarial potential of mepacrine (quinacrine), an acridine derivative was reported 80 years ago. It had been demonstrated to not only inhibit the asexual stages of malaria parasite but also display inhibitory effect on gametocytes. Notably, it is found to be effective against chloroquine and pyrimethamine resistant malaria parasite (Chavalitshewinkoon-Petmitr et al. 2001; Schlitzer, 2007). More recently, pyronaridine another acridine derivative incorporating the pyrrolidine unit in combination with AS is being used in clinics as Pyramax[®] and is considered to be a promising ACT for uncomplicated falciparum malaria (Croft et al. 2012). Acridine derivatives show wide spectrum effect on malaria parasite metabolic pathways such as inhibition of hemozoin (β -hematin) formation, mitochondrial bc1 complex and DNA topoisomerase II and interaction with DNA (Ramharter et al. 2008; Valdés, 2011; Padmanaban et al. 2012). Therefore we envisaged to prepare a new series of acridine hybrid, which carried the pyrrolidine unit, which in itself had shown the potential as CQresistance reverser. We found that all the compounds displayed potent antimalarial activity in vitro. Among the series, compound 15c displayed the best in vivo antimalarial potential. As a consequence compound 15c was selected to examine the *in vitro* as well as in vivo antimalarial efficacy in combination with the three most active and widely used artemisinine derivatives viz. AM, AE and AS. It is known that if the combination of two drugs has Σ FIC of <1, then the combination is termed as synergistic whereas if it is 1 it is considered to be additive. During the assessment of the combinations it was found that mean Σ FIC value for the 15c + AM was 0.68 whereas it was 0.80 for 15c + AE thereby indicating the overall slight synergistic antimalarial potential of 15c in combination with AM or AE in vitro. The Σ FIC value for 15c + AS combination was found to be >1 suggesting a marginal trend towards antagonism. This might be due to higher in vitro antimalarial activity of AS than AM or AE (~2 times; our unpublished data) and 15c failed to enhance the activity of AS above that level.

Table 4. T	herapeutic ant	ımalarıal poi	tential of 150	c with artemeti	her against <i>P</i> .	yoeln ngerens	as multidrug r	esistant in S	owiss mice.			
D	D_{000}/M_{\odot}	Parasitaemi	a (Mean‰ \pm s.	D.) at different o	days (number of	f mice without ₁	parasitaemia/tota	al survived m	nice)		Moon and M	
Drug/ combination	$\log^{-1} \times 4 \text{ days}$	0 h	Day4	Day7	Day10	Day14	Day18	Day21	Day24	Day28	time (Days)	% cure
15c + AM	200 + 60	0.5 ± 0.32	$0 \pm 0 \ (5/5)$	$0 \pm 0 \ (5/5)$	$0 \pm 0 \ (5/5)$	$0 \pm 0 \ (5/5)$	$0 \pm 0 \ (5/5)$	$0 \pm 0 \ (5/5)$	$0 \pm 0 \ (5/5)$	$0 \pm 0 (5/5)$	>28	100
ŝ	200 + 30	1.02 ± 0.32	$0 \pm 0 \ (5/5)$	$0 \pm 0 \ (5/5)$	$0 \pm 0 \; (5/5)$	$0 \pm 0 \ (5/5)$	$0 \pm 0 \ (5/5)$	$0 \pm 0 \ (5/5)$	0.56 ± 0.0	$0 \pm 0 (5/5)$	>28	80
	-+ 60	0.81 ± 0.38	$0 \pm 0 \ (5/5)$	0.54 ± 0.75	14.28 ± 17.33	25.25 ± 16.75	$0.1 \pm 0.0 \ (0/1)$	$0 \pm 0 (1/1)$	(1/1) (1/1) (1/1)	$0\pm0(1/1)$	16.6	20
	-+30	0.76 ± 0.34	0 ± 0 (5/5)	(3/3) 13.14 ± 25.53	19.16 ± 18.36	Died					12.6	0
;	200+-	0.38 ± 0.28	0.04 ± 0.08	5.18 ± 3.52	(0/5) 32.3 ± 17.68	$38 \cdot 3 \pm 29 \cdot 52$	32.33 ± 32.04	$0 \pm 0 (1/1)$	$0 \pm 0 (1/1)$	$0 \pm 0 (1/1)$	19.2	20
Control	I	(0,0) 0.4 ± 0.38 (0,5)	$90 \pm 0 (0/1)$	(c/u) Died	(c n)	(+/0)	(7/1)				5.2	0
		(clar										

The synergistic *in vitro* interaction of 15c with AM invoked us to combine 15c with AM for its antimalarial response *in vivo* against MDR malaria parasite. It was encouraging to discover that the combination of 15c with AM enhanced the antimalarial response of both the partners. Compound 15ccould cure only 10% of the Swiss mice infected with *P. yoelii nigeriensis* MDR at 100 mg kg⁻¹ × 4 days and AM at its 12.5 mg kg⁻¹ × 4 days dose produced only 20% cure. Interestingly, a combination with lower doses of same partners (50 mg kg⁻¹ × 4 days 15c with 6.25 mg kg⁻¹ × 4 days AM) potentiate the curative effect up to 100% and also enhanced the mean survival time.

The inappropriate treatment strategies with antimalarials induce the problems of recrudescence, toxicity of a particular drug and its improper therapeutic response. Therefore the pharmacokineticpharmacodynamic (PK-PD) properties (curative doses, maximum effect produced by the drug, parasite multiplication rate, parasite reduction ratio, PCT and recrudescence study) of the drug should be explored appropriately before its application in the patients. As a consequence, to determine a proper dose and regimen, the pharmacodynamic studies of the combination of 15c + AM were carried out and the pharmacodynamic parameters were found to be favourable in the group treated with 200 mg kg⁻¹ 15c in combination with 60 mg kg^{-1} AM in comparison with alone treatment. The PCT was lower in the combination group (2.8 days in combination vs 6.5 days in alone group) while PRR was higher (880 in combination vs 17.3 in alone group). However, combination of 200 mg kg⁻¹ 15c with 30 mg kg⁻¹ AM could not show any improvement in pharmacodynamics properties over alone treatment. Low PCT and high PRR values indicate the combination to be pharmacological as well as pharmacodynemic worth pursuing (White, 1997).

Our *in vivo* screening data for established infection also validated the pharmacodynamic studies. When 200 mg kg⁻¹ 15c + 60 mg kg⁻¹ AM was given for 4 consecutive days all the infected mice were cured and no parasitaemia was observed till the end of the experiment, while 200 mg kg⁻¹ 15c + 30 mg kg⁻¹ AM (required PCT 7.05 days) when given for 4 days there was only 80% cure. However, both the combinations were better than their individual effects. Moreover, low PCT and high PRR value may be considered helpful in reducing the chances of drug resistance (White, 1997).

Most of the acridine derivatives are associated with toxicity. For example, Loiseau and Nguyen (1996) reported that a new series of acridine derivatives, which possess potent antimalarial activity against murine malaria model showed a high level of toxicity. Moreover, single oral dose toxicology studies with Pyramax in the rat displayed toxicity characteristically reflected by decreased body weight gain, diarrhoea and soft stools at $\geq 1000 \text{ mg kg}^{-1}$ and chromaturia at 2000 mg kg⁻¹. These observations were in line with the inherent acute toxicity expressed by pyronaridine alone (Croft *et al.* 2012). Interestingly, compound *15c* has not shown hepatotoxicity or nephrotoxicity and displayed acceptable safety index, both *in vitro* and *in vivo* (LD₅₀ ~ 2000 mg kg⁻¹ alone and safety index of ~10 in combination).

In summary, we have synthesized and investigated the antimalarial efficacy of a new series of acridine-based hybrids. The compounds displayed oral efficacy against MDR *P. yoelii migeriensis* in mice. Further it was shown that one of the derivatives (15c) serves as good partner in ACT in particular with AM. However, detailed absorptiondistribution-metabolism-excretion and clinical safety studies are required to utilize this combination for further development.

SUPPLEMENTARY MATERIAL

The supplementary material for this article can be found at http://dx.doi.org/10.1017/S0031182016000937.

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