Phylogenetic relationships between the genera Aphidius and Lysaphidus (Hymenoptera: Braconidae: Aphidiinae) with description of Aphidius iranicus sp. nov.

Željko Tomanović¹

Institute of Zoology, Faculty of Biology, University of Belgrade, Studentski trg 16, 11000 Belgrade, Serbia

Ehsan Rakhshani

Department of Plant Protection, College of Agriculture, Zabol University, Zabol, P.O. Box 998615-538, I.R., Iran

Petr Starý

Institute of Entomology, Academy of Sciences of the Czech Republic, Branišovská 31, 37005 České Budějovicé, Czech Republic

Nickolas G. Kavallieratos

Laboratory of Agricultural Entomology, Department of Entomology and Agricultural Zoology, Benaki Phytopathological Institute, 8 Stefanou Delta str., 14561 Kifissia, Attica, Greece

Ljubiša Ž. Stanisavljević, Vladimir Žikić

Institute of Zoology, Faculty of Biology, University of Belgrade, Studentski trg 16, 11000 Belgrade, Serbia

Christos G. Athanassiou

Laboratory of Agricultural Zoology and Entomology, Agricultural University of Athens, 75 Iera Odos, 11855 Athens, Attica, Greece

Abstract—We analyzed the phylogenetic relationships between eight *Aphidius* Nees and six *Lysaphidus* Smith species on the basis of 12 morphological characters by parsimony analysis. The consensus tree does not support the generic status of *Lysaphidus*. *Aphidius iranicus*, **sp. nov.**, associated with *Titanosiphon bellicosum* Nevsky on *Artemisia absinthium* L. from Iran, is described. The new parasitoid species is described and illustrated by line drawings, and its diagnostic characters are discussed. The taxonomic position of the subgenus *Tremblayia* Tizado and Núñez-Pérez is also considered. *Tremblayia* and *Lysaphidus* are newly classified as synonyms of *Aphidius*. The following new or revised combinations are proposed: *Aphidius adelocarinus* Smith, **comb. rev.**, *A. ramythirus* Smith, **comb. rev.**, *A. rosaphidis* Smith, **comb. rev.**, *A. viaticus* (Sedlag), **comb. nov.**, *A. arvensis* (Starý), **comb. nov.**, and *A. erysimi* (Starý), **comb. nov.**

Résumé—Nous avons analysé les relations phylogénétiques entre huit espèces d'Aphidius Nees et six espèces de Lysaphidus Smith se basant sur 12 caractères morphologiques par analyse de parsimonie. L'arbre de consensus ne supporte pas le statut générique de Lysaphidus. Aphidius iranicus sp. nov., provenant d'Iran et associé avec Titanosiphon bellicosum Nevsky sur Artemisia absinthium L., est décrit. Cette nouvelle espèce de parasitoïde est décrite et illustrée en dessin linéaire et ses caractères diagnostiques sont discutés. La position taxonomique du sous-genre Tremblayia Tizado et Núñez-Pérez est aussi considérée. Lysaphidus et Tremblayia sont récemment classifiés en tant que synonymes d'Aphidius. Les combinaisons suivantes nouvelles ou de révision sont donc proposées : Aphidius adelocarinus Smith, comb. rev., A. ramythirus Smith, comb. rev., A. rosaphidis Smith, comb. rev., A. viaticus (Sedlag), comb. nov., A. arvensis (Starý), comb. nov. et A. erysimi (Starý), comb. nov.

Received 24 January 2006. Accepted 17 January 2007.

¹Corresponding author (e-mail: ztoman@bf.bio.bg.ac.yu).

Can. Entomol. 139: 297-307 (2007)

Introduction

The genus Aphidius Nees is one of the most diverse taxa within the subfamily Aphidiinae (Hymenoptera: Braconidae). Aphidius species attack a wide range of aphid hosts (Starý 1981; Tomanović et al. 2003; Kavallieratos et al. 2004). To date, over 100 species have been described worldwide. The evolution of small body size in Aphidiinae resulted in the loss of many useful morphological characters compared with their larger braconid ancestors (Müller et al. 1999). Within the Aphidiinae, Aphidius represents the most problematic group taxonomically, with a very limited number of reliable characters relevant for species separation (Starý 1973; Pungerl 1986; Kambhampati Mackauer 1988; Unruh et al. 1989; Tomanović et al. 2003). The taxonomic and phylogenetic relationships between Aphidius and the closely related genera Lysaphidus Smith, Euaphidius Mackauer, and Diaeretiella Starý are also still unresolved (Kambhampati et al. 2000; Sanchis et al. 2000).

Lysaphidus was originally described as a subgenus of Aphidius (Smith 1944). Starý (1960a) gave Lysaphidus generic status on the basis of wing venation and some head characters. Later, several new species were described from the Palaearctic region (Starý 1960b; Takada 1966). Recently, Smith et al. (1999), Kambhampati et al. (2000), Sanchis et al. (2000), and Shi and Chen (2005) studied several genes and found nothing to support the generic status of Lysaphidus.

Lysaphidus species (like many other aphidiines, including some Aphidius species) have been found in association with host aphids (Hemiptera: Aphididae) on species of Artemisia L. and related plants of the family Asteraceae in the Nearctic (Pike et al. 1997) and Palaearctic regions (Klausnitzer 1968; Takada 1968; Starý 1973, 2006; Tizado and Núñez-Pérez 1994; Shi and Chen 2005). An Aphidius species was reported from Titanosiphon neoartemisiae Takahashi from Taiwan (Gahan 1926). Tizado and Núñez-Pérez (1994) described Aphidius (Tremblayia) artemisicola from Titanosiphon artemisiae (Koch) in Spain. The other records of this species are from Serbia, where it was reared from Macrosiphoniella sp. on Artemisia vulgaris L. (Tomanović et al. 2003: Kavallieratos et al. 2004). Only a relatively limited number of

Aphidiinae have so far been detected in the Middle and Far East (Starý 1975, 1979; Takada 1979; Raychaudhuri *et al.* 1982; Starý *et al.* 1998, 2000).

In the present paper we discuss the taxonomic position of and phylogenetic relationships between *Lysaphidus* and *Aphidius* species associated with host aphids on *Artemisia* spp. and related plants and the relationship between *Aphidius* and *Tremblayia* Tizado and Núñez-Pérez on the basis of morphological characters. We describe a new *Aphidius* species reared from the *Artemisia absinthium* L. – *Titanosiphon bellicosum* Nevsky association in Iran.

Materials and methods

Collection and deposition of specimens

Samples of plants supplying aphid colonies containing both live and mummified aphids were collected during 1960–2004. Live aphids were killed and preserved in a mixture of 90% ethanol and 75% lactic acid in a ratio of 2:1 (Eastop and van Emden 1972) for later identification. The remaining aphid colonies were maintained in the laboratory until parasitoid emergence. Dissected parasitoid specimens were mounted on slides in Canada balsam for later identification. The external structure of emerged parasitoids was studied using an Olympus BH2 phase-contrast microscope and illustrated with a drawing tube (Trdan 2002; Kavallieratos et al. 2005).

The material examined in this study is deposited in the collection of the Institute of Zoology, Faculty of Biology, University of Belgrade (Serbia and Montenegro); the collection of P. Starý (České Budějovicé, Czech Republic); and the collection of the Laboratory of Agricultural Zoology and Entomology, Agricultural University of Athens (Greece). In the course of our study we examined the paratype and topotypes of *Aphidius (Tremblayia) artemisicola* (deposited in the collection of the Department of Animal Biology, University of León, Spain).

Phylogenetic analysis

A cladistic parsimony analysis of *Aphidius* and *Lysaphidus* species parasitizing host aphids on *Artemisia* spp. and related plants was undertaken. Four additional *Aphidius* species (*A. aquilus* Mackauer, *A. hortensis* Marshall, *A. ervi* Haliday, and *A. matricariae* Haliday)

were also included in the analysis. In total, we analyzed 14 species (eight species of Aphidius and six of Lysaphidus) (Table 1) on the basis of 12 morphological characters. Characters with two states were coded as polymorphic. Ephedrus persicae Froggatt and Toxares deltiger (Haliday) were designated as the outgroups because, based on morphology and life-history traits, both species can be considered basal among extant Aphidiinae (Mackauer 1961; Starý 1981; Gärdenfors 1986). The phylogenetic analyses proceeded from a maximum parsimony search using PAUP* 4.0b10 (Swofford 2003), with all positions equally weighted. We calculated the consistency index, retention index, and rescaled consistency index as indicators of the extent of homoplasy in the data as well as their consistency. We performed a bootstrap analysis with 1000 replicates. The tree was visualized and printed using TreeView (Page 1996).

Character description

The morphological terminology used in the phylogenetic analysis and the parasitoid species description is based on Huber and Sharkey (1993) and Kavallieratos *et al.* (2001). Twelve characters were coded using original coding strategies or coded characters compiled from the literature (Quicke and van Achterberg 1990; Quicke and Belshaw 1999) (Table 2). Plesiomorphic character states are coded as "0".

- 1. Number of segments of maxillary palp: 0 = 4; 1 = 3 (Fig. 1).
- 2. Number of segments of labial palp: 0 = 3; 1 = 2; 2 = 1 (Fig. 1).
- 3. Flagellomere 1 (\bar{F}_1) : 0 = less than 3.0 times as long as wide (ratio between length of F_1 and its width basally); 1 = more than 3.0 times as long as wide (Figs. 2, 3). More elongated flagellomeres in general represent an adaptation for host searching.
- 4. Number of longitudinal placodes on flagel-lomere 1 (F₁): 0 = 1 or more (Fig. 4); 1 = 0 (Figs. 2, 3).
- 5. Number of longitudinal placodes on flagellomere 2 (F₂): 0 = 3 or more (Fig. 4); 1 = 0-2 (Figs. 2, 3).
- 6. Forewing stigma: 0 = less than 3.0 times as long as wide (ratio between length of forewing stigma and width at Radial sector (Rs)) (Fig. 5); 1 = elongated, more than 3.0 times as long as wide (Fig. 6).

7. Forewing vein R1: 0 = subequal or equal to length of stigma (Fig. 5); 1 = less than half length of stigma (Fig. 6).

Reduction of wing venation represents an apomorphic character state.

- 8. Forewing M+m-cu vein: 0 = developed throughout (Fig. 5); 1 = not developed or partly developed under r-m vein (Figs. 6, 7).
- 9. Propodeum: 0 = areolate (Fig. 8); 1 = carinate (Figs. 9, 10); 2 = smooth or with small divergent carina at the base (Fig. 11).
- 10. Petiole: 0 = subquadrate, less than 3.0 times as long as wide at spiracles (Fig. 12); 1 = more elongated, more than 3.0 times as long as wide at spiracles.
- 11. Ovipositor sheath: 0 = dorsal margin straight; 1 = dorsal margin clearly concave (Figs. 13, 14). We assumed that a curved and longer ovipositor sheath is an apomorphic character state that allows easy access to the host aphid during oviposition (Starý 1976).
- 12. Ovipositor sheath: 0 = short (Fig. 13); 1 = elongated (Fig. 14).

Results

Aphidius iranicus Rakhshani and Starý, sp. nov.

Type material

Holotype (female): Iran, Rostam-Abad (Guilan Province), 3.v.2004, reared from *Titanosiphon bellicosum* Nevsky on *Artemisia absinthium* L., coll. E. Rahkshani (Collection of the Institute of Zoology, University of Belgrade). Paratype (one female): same data as holotype, slide-mounted (Collection of the Institute of Zoology, University of Belgrade).

Etymology

The new species is named after the country of origin, Iran.

Diagnosis

The new species is similar to *Aphidius artemisicola* Tizado and Núñez-Pérez in having the propodeum with an incomplete central areola (Fig. 9) and two divergent distal carinae but differs clearly from it in the range of the tentorial index (0.35-0.40 in A. iranicus vs. 0.43-0.50 in A. artemisicola), the ratio between lengths of F_1 and F_2 $(F_1 1.1-1.2 \text{ times as long as } F_2 \text{ in } 1.1-1.2 \text{ times as long as } 1.1 \text{ and } 1.1 \text{ are the proposed of the proposed$

Table 1. Aphidiinae species used in phylogenetic analysis.

Species	Aphid host	Country
Aphidius absinthii Marshall Aphidius artemisicola Tizado and Núñez-Pérez	Macrosiphoniella spp. Titanosiphon artemisiae (Koch), Macrosiphoniella sp.	Serbia, Czech Republic, Russia Spain, Serbia
Aphidius aquilus Mackauer Aphidius ervi Haliday Aphidius iranicus sp. nov.	Callaphis flava Mordvilko, Euceraphis punctipennis (Zetterstedt) Acyrthosiphon pisum (Harris), Sitobion avenae (Fabricius) Titanosiphon bellicosum Nevsky	Serbia Serbia, Greece Iran
Aphidius hortensis Marshall Aphidius matricariae Haliday	Liosomaphis berberidis (Kaltenbach) Aphis gossypii Glover, Brachycaudus helichrysi (Kaltenbach), Myzus persicae (Sulzer)	Serbia, Russia Serbia, Greece, Russia
Aphidius phalangomyzi Starý **Aphidius ramithyrus (Smith)	Macrosiphoniella oblonga (Mordvilko) Pleotrichophorus cf. wasatchii (Knowlton), Pleotrichophorus spp.	Serbia United States of America, Mexico
*Aphidius arvensis Starý *Aphidius erysimi Starý	Coloradoa artemisiae (del Guercio) Pseudobrevicoryne erysimi Holman	Czech Republic, Serbia Czech Republic
*Aphidius adelocarinus (Smith) *Aphidius viaticus Sedlag *Aphidius rosaphidis (Smith)	Microsiphoniella artemisiae Boyer de Fonscolombe Pleotrichophorus glandulosus (Kaltenbach) Chaetosiphon fragaefolii (Cockerell)	United States of America Germany, Serbia United States of America
Outgroup Ephedrus persicae Froggatt	Myzus persicae (Sulzer)	Serbia, France, Iran, Japan, Czech Republic, United States of America
Toxares deltiger (Haliday)	Unknown	Turkey
Note: Full data for all chacies are	Note: Eall date for all energies are multished in Kavallieretos <i>et al (2004)</i> and are available from the authors	

Note: Full data for all species are published in Kavallieratos *et al.* (2004) and are available from the authors. *Previously placed in *Lysaphidus*.

Table 2. Character matrix used in cladistic analysis of *Aphidius* species.

	1	2	3	4	5	6	7	8	9	10	11	12
A. absinthii	0	1	1	1	1	1	0	0	0	1	1	1
A. artemisicola	1	1	1	1	1	1	1	0	1	0	1	1
A. aquilus	0	1	0	1	1	1	1	0,1	0	0	0	0
A. ervi	0	0	1	0	0	1	0	0	0	1	1	0
A. iranicus	1	2	1	1	1	0	0	0	1	0	1	0
A. hortensis	0	1	0	1	1	1	0	0	0	0	1	0
A. matricariae	1	1	0	0	0	1	0	0	0	1	1	0
A. phalangomyzi	0	0	1	1	1	1	0	0	0	1	1	0
*A. ramithyrus	0	1	0	1	1	1	1	1	0	0	1	0
*A. arvensis	1	2	0	1	1	1	0	1	0,1	0	1	1
*A. erysimi	1	2	0	0	0	1	0	1	0	0	0	0
*A. adelocarinus	1	1	0	1	1	0	0	1	2	0	1	1
*A. viaticus	0	1	1	1	1	1	1	1	0	1	1	0
*A. rosaphidis	0	1	1	1	1	0	1	1	0	0	1	1
E. persicae	0	0	1	0	0	1	0	0	0	0	0	1
T. deltiger	0	0	0	0	0	1	0	0	0	0	0	0

Note: Ephedrus persicae and Toxares deltiger were used as outgroups.

A. iranicus (Fig. 2) vs. F_1 as long as F_2 in A. artemisicola), the number of segments of labial palps (one-segmented in A. iranicus (Fig. 1) vs. two-segmented in A. artemisicola), the ratio between length of the stigma and length of the distal abscissa of R1 (stigma as long as distal abscissa of R1 in A. iranicus (Fig. 5) vs. 2.20 times as long in A. artemisicola), and coloration of the head (brown in A. iranicus vs. orange in A. artemisicola) and mesosoma (brown in A. iranicus vs. mostly yellow-orange in A. artemisicola).

Aphidius iranicus also resembles A. absinthii Marshall but differs clearly from it in the range of the tentorial index (0.35-0.40 in A. iranicus vs. 0.45–0.60 in A. absinthii), the ratio between malar space and longitudinal eye diameter (0.20 in A. iranicus vs. 0.25-0.33 in A. absinthii), the number of long setae on the clypeus (4-6 in A. iranicus vs. 7–13 in A. absinthii), the number of antennal segments (13-14 in A. iranicus vs. 15–17 in A. absinthii), the ratio between length of the stigma and length of the distal abscissa of R1 (stigma as long as distal abscissa of R1 in A. iranicus vs. 1.50-3.00 times as long in A. absinthii), the number of segments of maxillary and labial palps (three-segmented maxillary and one-segmented labial palps in A. iranicus vs. four-segmented maxillary and two-segmented labial palps in A. absinthii), and the ratio between length and width of the

petiole at the spiracles (2.70–2.80 in *A. iranicus* vs. 3.00–3.50 in *A. absinthii*).

Description

Female

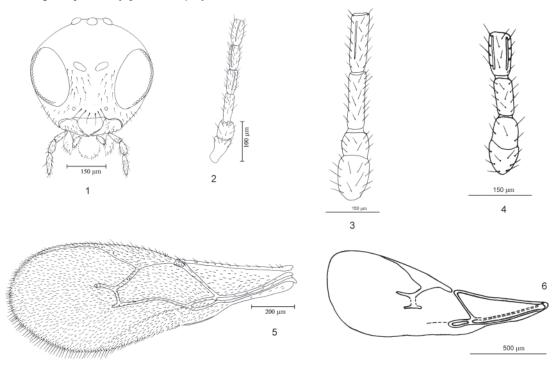
Head (Fig. 1) wider than mesosoma at tegulae (ratio between width of head and width of mesoscutum, 1.40–1.45). Frons, vertex, and occipital area with sparse setae, face with narrow, bare band surrounded laterally by moderately long, regular, sparse setae. Tentorial index 0.35-0.40. Malar space equal to 0.20 of longitudinal eye diameter. Eyes oval, converging toward clypeus, with few short setae in posterior margin. Clypeus rounded, with 4-6 long setae. Antennae 13–14-segmented, moderately thickened at apex, with semierect and adpressed setae about as long as half of segment diameter. Scape and pedicel subglobular. Flagellar segments slender, with sparse, semierected setae. F_1 about 3.0 times as long as its maximum width, 1.1-1.2 times as long as F_2 and 1.3-1.4times as long as F_3 . $F_{1, 2, 3}$ with 0, 1–2, and 1–2 longitudinal placodes, respectively (Fig. 2). Maxillary palp 3-segmented (Fig. 1), apical segment distinctly longer than median and basal segments. Labial palp 1-segmented (Fig. 1).

Mesosoma

Mesonotum (Fig. 15) with notaulices in ascendant portion of anterolateral area, erased dorsally and outlined by 1–2 rows of long

^{*}Previously placed in Lysaphidus.

Figs. 1–6. 1, *Aphidius iranicus*, holotype \mathfrak{P} : anterior aspect of head and maxillary and labial palps; 2, *Aphidius iranicus*, holotype \mathfrak{P} : flagellomeres 1–4; 3, *Aphidius absinthii* \mathfrak{P} : flagellomeres 1 and 2; 4, **Aphidius erysimi* \mathfrak{P} : flagellomeres 1 and 2; 5, *Aphidius iranicus*, holotype \mathfrak{P} : forewing; 6, **Aphidius ramythirus* \mathfrak{P} : forewing. (*, previously placed in *Lysaphidus*.)



sparse setae, which extend near to scutellum. Scutellum usually with 5–6 long setae laterally. Forewing (Fig. 5) stigma triangular, 2.70–3.00 times as long as its width, equal to or slightly longer than R1. Distal abscissa of R1 1.50 times as long as 3/Rs. Posterior margin of forewing with long setae. Propodeum (Fig. 9) with incomplete central areola bordered by two divergent posterocentral carinae extending transversely to the spiracles. Anterocentral carinae weakly developed, visible only in upper part, where united. Upper areola with 2–3 long setae laterally and lower areola with single seta in lower part. Upper areola about 1.50 times as long as lower areola. Hind femur and tibia with semierected, sparse setae, more dense near tibial spur.

Metasoma

Petiole (= metasomal tergum 1) (Fig. 12) parallel-sided, 2.70–2.80 times as long as its width at spiracle, anterolateral area with 6–8 costulae. Dorsal surface of petiole with fine rugosities and 6 long, erected lateromedial setae on basal portion. Genitalia: Ovipositor sheath (Fig. 13) short, distally truncated, with concave

line at dorsal margin; ventral line straight. First valvula strongly convex ventrally and second valvula slightly concave (Fig. 13).

Coloration

Head brown, face and gena yellowish brown, mouthparts yellow, antenna uniformly brown. Mesosoma brown. Wings hyaline, slightly infumate; basal vein brown, stigma, R1, and radial veins yellow. Forewing surface covered with black, regular, short setae. Legs yellowish to brown. Petiole yellowish to brown; rest of metasoma and genitalia brown.

Body length 1.5–1.7 mm.

Male

Unknown.

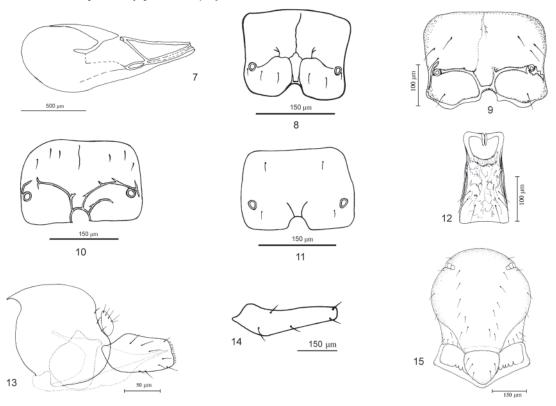
Aphid mummy

Shiny white.

Host

Titanosiphon bellicosum Nevsky on Artemisia absinthium L.

Figs. 7–15. 7, Aphidius aquilus \mathfrak{P} : forewing; 8, *Aphidius rosaphidis \mathfrak{P} : dorsal aspect of propodeum; 9, Aphidius iranicus, holotype \mathfrak{P} : dorsal aspect of propodeum; 10, Aphidius artemisicola \mathfrak{P} : dorsal aspect of propodeum; 11, *Aphidius adelocarinus \mathfrak{P} : dorsal aspect of propodeum; 12, Aphidius iranicus, holotype \mathfrak{P} : dorsal aspect of ovipositor sheath; 14, *Aphidius adelocarinus \mathfrak{P} : lateral aspect of ovipositor sheath; 15, Aphidius iranicus, holotype \mathfrak{P} : dorsal aspect of mesonotum. (*, previously placed in Lysaphidus.)



Distribution Iran.

Phylogenetic inference

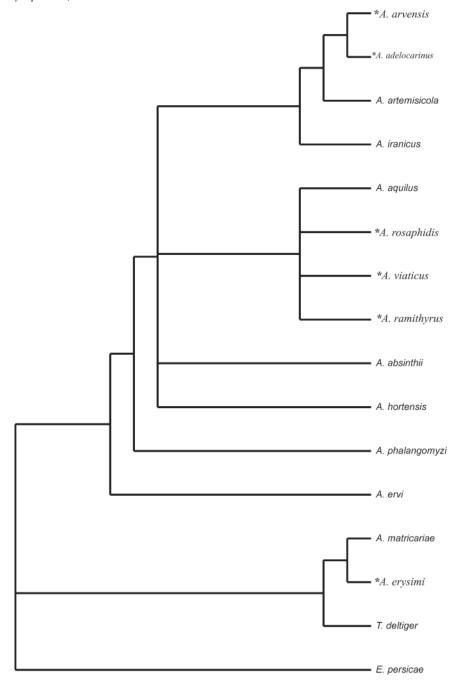
The consensus tree (Fig. 16) resulted from an unweighted parsimony analysis obtained from the six most parsimonious trees (tree length 34, consistency index 0.412, retention index 0.600, and rescaled consistency index 0.247).

Discussion

Tizado and Núñez-Pérez (1994) described the subgenus *Tremblayia* based on *Aphidius artemisicola* from the *Titanosiphon artemisiae* (Koch) – *Artemisia campestris* L. subsp. *glutinosa* (Gay *ex* Besser) Batt. association in Spain. The most important diagnostic character was "...propodeum with incomplete areola that is distinct as two divergent carinae in the lower

portion...". After checking the paratype and topotypes we found that the propodeal areola in most specimens is not complete but has a clear indication of two upper carinae. Also, one topotype male has a complete propodeal areola. However, this character state is common in some small Aphidius species, probably because of allometry (Gärdenfors 1986). For example, some specimens of A. urticae Haliday emerged from the Schizaphis scirpi (Passerini) - Typha latifolia L. association also have an incomplete propodeal areola. Furthermore, some smaller A. absinthii specimens reared from the Macrosiphoniella spp. – Artemisia spp. association and A. hortensis specimens reared from the Liosomaphis berberidis (Kaltenbach) - Berberis vulgaris L. association have an incomplete propodeal areola, whereas larger ones have a complete pentagonal areola. Aphidius iranicus has an incomplete propodeal areola but a clear indication of two upper carinae. Although the

Fig. 16. Strict consensus tree of fourteen *Aphidius* taxa based on unweighted parsimony analysis of the six most parsimonious trees (tree length 34, consistency index 0.412, retention index 0.600, and rescaled consistency index 0.247). *Ephedrus persicae* and *Toxares deltiger* were used as outgroups. (*, previously placed in *Lysaphidus*.)



propodeal areola tends to be incomplete in parasitoids of small aphid hosts, as in the case of *A. artemisicola*, we expect to find additional

A. iranicus specimens with a developed propodeal areola. We believe that there are no reliable characters justifying taxonomic separation of

the subgenus *Tremblayia* and thus recognize *Tremblayia*, **syn. nov.** as a new synonym of *Aphidius*, not a subgenus.

Aphidius artemisicola has been found in associations with Artemisia species in Spain, Andorra, and Serbia (Tizado and Núñez-Pérez 1994; Tomanović et al. 2003; Kavallieratos et al. 2004). Aphidius iranicus was found in association with Titanosiphon bellicosum on Artemisia absinthium in Iran.

Lysaphidus species parasitize mainly small aphids (e.g., species of Titanosiphon Nevsky, Coloradoa Wilson, Microsiphoniella Hille Ris Lambers, and Lipaphis Mordvilko) associated with Artemisia spp. and related plants of Asteraceae (Smith 1944; Starý 1960b; Pike et al. 1997). Owing to the small size of their aphid hosts, reductions in morphology occur. For example, Lysaphidus species have a one- or twosegmented labial palp, a reduced M+m-cu vein (only a small part is developed under the r-m vein), and an undeveloped propodeal areola in some species (L. arvensis Starý, L. erysimi Starý, and L. adelocarinus Smith) or even significantly reduced lower carinae (L. adelocarinus and L. arvensis). Aphidius species have two- or three-segmented labial palps, but A. iranicus has a one-segmented labial palp, like some Lysaphidus species (L. arvensis, L. erysimi). Also, forewing vein M+m-cu is usually developed in Aphidius species but is reduced in small Aphidius aquilus specimens. The propodeal areola is usually developed in all Aphidius species except A. iranicus, A. artemisicola (in certain cases, developed), and some small specimens of A. absinthii, A. hortensis, and A. urticae.

Lysaphidus arvensis, L. adelocarinus, Aphidius artemisicola, and A. iranicus form a clade (Fig. 16) sharing the following two synapomorphies: a small number of longitudinal placodes on F₁ and F₂ and a clearly concave dorsal margin of the ovipositor sheath. Aphidius aquilus, L. rosaphidis Smith, L. viaticus Sedlag, and L. ramithyrus Smith form another clade characterized by a two-segmented labial palp, a small number of longitudinal placodes on F₁ and F2, and forewing vein R1 less than half the length of the forewing stigma as synapomorphies. Both clades are composed of species that parasitize small aphid hosts. The position of Toxares deltiger as sister group to the clade of A. matricariae + L. erysimi is probably due to the small number of species analyzed or the

insufficient number of characters used. *Toxares deltiger* represents one of the basal lineages within Aphidiinae. It is characterized by several plesiomorphies, such as the braconid type of wing venation with seven closed cells in the forewing, a large number of longitudinal placodes on flagellomeres 1 and 2, and a short and triangular ovipositor sheath.

After bootstrap analysis with 1000 replicates, all Aphidius and Lysaphidus species studied form a polytomic tree (Fig. 16). Our tree is weakly supported because we analyzed closely related taxa with a restricted number of reliable characters for species separation. On the basis of the morphological characters examined, there is no clear distinction between Aphidius and Lysaphidus, and the tree does not support the generic status of Lysaphidus. Lysaphidus, syn. nov. is here synonymized with Aphidius, and the following new or revised combinations are proposed: Aphidius adelocarinus Smith, comb. rev., A. ramythirus Smith, comb. rev., A. rosaphidis Smith, comb. rev., A. viaticus (Sedlag), comb. nov., A. arvensis (Starý), comb. nov., and A. erysimi (Starý), comb. nov.

We did not revise the following species attributed to *Lysaphidus* by various authors: *L. multiarticulatus* Ashmead (Smith 1944); *L. pleotrichophori* Takada, *L. matsuyamensis* Takada, and *L. callipterinellae* Takada (Takada 1966); *L. macrosiphoniella* Tamili and Raychaudhuri and *L. quadrii* Shujauddin (Raychaudhuri 1990); *L. santolinae* Michelena Saval and Sanchis Segovia (Michelena Saval and Sanchis Segovia 1995); and *L. kunmingensis* Wang and Dong (Wang and Dong 1997). These taxa are problematic and need further taxonomic treatment.

Acknowledgments

We thank Dr. V. Rezaei for his help in the collection of the material and Dr. Jorge Tizado for the loan of the paratype and topotype specimens of *Aphidius artemisicola*. The research was partially supported by the grants 143006B (The Ministry of Science and Environmental Protection of the Republic of Serbia) and S5007102 (Grant Agency, Academy of Sciences of the Czech Republic) and by the Entomology Institute project Z50070508 (Academy of Sciences of the Czech Republic).

References

- Eastop, V.F., and van Emden, H.F. 1972. The insect material. *In Aphid technology. Edited by H.F.* van Emden. Academic Press, London. pp. 1–45.
- Gahan, A.B. 1926. Some braconid and chalcid flies from Formosa, parasitic on aphids. Proceedings of the United States National Museum, 70(8): 1–7.
- Gärdenfors, U. 1986. Taxonomic and biological revision of Palaearctic *Ephedrus* (Haliday) (Hymenoptera: Braconidae: Aphidiinae). Entomologica Scandinavica Supplement, **27**: 1–95.
- Huber, J.T., and Sharkey, M.J. 1993. Structure. In Hymenoptera of the world: an identification guide to families. Edited by H. Goulet and J.T. Huber. Agriculture Canada Publication 1894/E, Ottawa, Ontario. pp. 13–59.
- Kambhampati, S., and Mackauer, M. 1988. Intraand interspecific morphological variation in some *Aphidius* species (Hymenoptera: Aphidiidae) parasitic on the pea aphid in North America. Annals of the Entomological Society of America, **81**(6): 1010–1016.
- Kambhampati, S., Volkl, W., and Mackauer, M. 2000. Phylogenetic relationships among genera of Aphidiinae (Hymenoptera: Braconidae) based on DNA sequence of the mitochondrial 16S rRNA gene. Systematic Entomology, 25: 437–445.
- Kavallieratos, N.G., Lykouressis, D.P., Sarlis, G.P., Stathas, G.J., Sanchis Segovia, A., and Athanassiou, C.G. 2001. The Aphidiinae (Hymenoptera: Ichneumonoidea: Braconidae) of Greece. Phytoparasitica, 29: 306–340.
- Kavallieratos, N.G., Tomanović, Ž., Starý, P., Athanassiou, C.G., Sarlis, G.P., Petrović, O., Niketić, M., and Veroniki, M.A. 2004. A survey of aphid parasitoids (Hymenoptera: Braconidae: Aphidiinae) of southeastern Europe and their aphidplant associations. Applied Entomology and Zoology, 39: 527–563.
- Kavallieratos, N.G., Tomanović, Ž., Starý, P., Athanassiou, C.G., Fasseas, C., Petrović, O., Stanisavljević, Lj.Z., and Anagnou, M.V. 2005. *Praon* Haliday (Hymenoptera: Aphidiinae) of southeastern Europe: key, host range and phylogenetic relationship. Zoologischer Anzeiger, 243: 181–209.
- Klausnitzer, B. 1968. Zur Kenntnis der Entomofauna von Tanacetum vulgare L. und Artemisia vulgaris L. Wissenschaftliches Zeitschrift der technischen Universität Dresden, 17: 19–21.
- Mackauer, M. 1961. Die Gattungen der Familie Aphidiidae und ihre verwandtschaftliche Zuordnung (Hymenoptera: Ichneumonoidea). Beiträge zur Entomologie, 11: 792–803.
- Michelena Saval, J., and Sanchis Segovia, A. 1995. El genero *Lysaphidus* (Hymenoptera: Braconidae) en la Peninsula Iberica. Graellsia, **51**: 23–26.
- Müller, C.B., Adeiaanse, I.C.T., Belshaw, R., and Godfray, H.C.J. 1999. The structure of an aphid-

- parasitoid community. Journal of Animal Ecology, **68**: 346–370.
- Page, R.D.M. 1996. TreeView: an application to display phylogenetic trees on personal computers. Computer Applications in the Biosciences, 12: 357–358.
- Pike, K.S., Starý, P., Allison, D., Graf, G., Boydston, L., and Miller, T. 1997. Parasitoids (Hymenoptera, Braconidae, Aphidiinae) of aphids on big sagebrush (*Artemisia tridentata* Nuttall) and prairie sage (*Artemisia ludoviciana* Nuttall) in Washington state. Proceedings of the Entomological Society of Washington, 99(1): 143–155.
- Pungerl, N.B. 1986. Morphometric and electrophoretic study of *Aphidius* species (Hymenoptera: Aphidiidae) reared from a variety of aphid hosts. Systematic Entomology, **11**: 327–354.
- Quicke, D.L.J., and Belshaw, R. 1999. Incongruence between morphological data sets: an example from the evolution of endoparasitism among parasitic wasps (Hymenoptera: Braconidae). Systematic Biology, **48**: 436–454.
- Quicke, D.L.J., and van Achterberg, C. 1990. Phylogeny of the subfamilies of the family Braconidae (Hymenoptera: Ichneumonoidea). Zoologische Verhandelingen, **258**: 1–95.
- Raychaudhuri, D. 1990. Aphidiids (Hymenoptera) of northeast India. Indira Publishing House, Michigan.
- Raychaudhuri, D.N., Poddar, S.C., and Raychaudhuri, D. 1982. Study of the genus *Aphidius* (Hymenoptera: Aphidiidae) of India. Entomon, 7(1): 11–22.
- Sanchis, A., Latorre, A., Gonzalez-Candelas, F., and Michelena, J.M. 2000. An 18S rDNA based molecular phylogeny of Aphidiinae (Hymenoptera: Braconidae). Molecular Phylogenetics and Evolution, 14: 180–194.
- Shi, M., and Chen, X.X. 2005. Molecular phylogeny of the Aphidiinae (Hymenoptera: Braconidae) based on DNA sequences of 16S rRNA, 18S rDNA and ATPase 6 genes. European Journal of Entomology, **102**(2): 133–138.
- Smith, C.F. 1944. The Aphidiinae of North America. Ohio State University Contributions in Zoology and Entomology, 6: 1–144.
- Smith, P.T., Kambhampati, S., Völkl, W., and Mackauer, M. 1999. A phylogeny of aphid parasitoids (Hymenoptera: Braconidae: Aphidiinae) inferred from mitochondrial NADH 1 dehydrogenase gene sequence. Molecular Phylogenetics and Evolution, 11: 236–245.
- Starý, P. 1960a. The generic classification of the family Aphidiidae (Hymenoptera). Acta Societatis Entomologicae Czechosloveniae, 57: 238–252.
- Starý, P. 1960b. The aphidiid genus Lysaphidus Smith C.F. in Europe (Hymenoptera, Aphidiidae). Bulletin Entomologique de Pologne, 30(18): 357–366.

Starý, P. 1973. A review of the Aphidius species (Hymenoptera: Aphidiidae) of Europe. Annotationes Zoologicae et Botanica Bratislava, 84: 1–85.

- Starý, P. 1975. A checklist of the Far East Asian Aphidiidae. Beiträge zur Entomologie, Berlin, 25(1): 53–76.
- Starý, P. 1976. External female genitalia of the Aphidiidae (Hymenoptera). Acta Entomologica Bohemoslovaca, 73: 102–112.
- Starý, P. 1979. Aphid parasites (Hymenoptera, Aphidiidae) of the Central Asian area. Transactions of the Czechoslovak Academy of Sciences, Series of Mathematical and Natural Sciences, 89(3): 1–116.
- Starý, P. 1981. On the strategy, tactics and trends of host specificity evolution in aphid parasitoids (Hymenoptera: Aphidiidae). Acta Entomologica Bohemoslovaca, 78: 65–75.
- Starý, P. 2006. Aphid parasitoids of the Czech Republic (Hymenoptera: Braconidae: Aphidiinae). Academia, Prague.
- Starý, P., Naumann-Etienne, K., and Remaudière, G. 1998. A review and tritrophic associations of aphid parasitoids (Hymenoptera, Braconidae, Aphidiinae) of Pakistan. Parasitica, 54: 3–21.
- Starý, P., Remaudière, G., González, D., and Shahrokhi, S. 2000. A review and host associations of aphid parasitoids (Hymenoptera: Braconidae: Aphidiinae) of Iran. Parasitica, **56**: 15–41.
- Swofford, D.L. 2003. PAUP*: Phylogenetic Analysis Using Parsimony (*and Other Methods). Version 4 (10b). Sinauer Associates, Inc., Sunderland, Massachusetts.

Takada, H. 1966. Three new species of the genus Lysaphidius Smith from Japan (Hymenoptera: Aphidiidae). Insecta Matsumurana, 28: 127–132.

- Takada, H. 1968. Aphidiidae of Japan (Hymenoptera). Insecta Matsumurana, 30: 67–124.
- Takada, H. 1979. Aphidiidae (Hymenoptera) from Mongolia. Folia Entomologica Hungarica, 32(1): 189–202.
- Tizado, E.J., and Núñez-Pérez, E. 1994. *Aphidius* (*Tremblayia*) *artemisicola*, a new subgenus and species of Aphidiinae from Spain (Hymenoptera: Braconidae). Graellsia, **50**: 25–27.
- Tomanović, Ž., Kavallieratos, N.G., Starý, P., Athanassiou, C.G., Žikić, V., Petrović-Obradović, O., and Sarlis, G.P. 2003. *Aphidius* Nees aphid parasitoids (Hymenoptera: Braconidae: Aphidiinae) in Serbia and Montenegro: tritrophic associations and key. Acta Entomologica Serbica, **8**(1–2): 15–39.
- Trdan, S. 2002. Evaluation of morphological and genetic variability of populations of economically important thrips species (Thysanoptera) in Slovenia. Doctoral Dissertation, Department of Agronomy, Biotechnical Faculty, University of Ljubljana, Ljubljana.
- Unruh, T.R., White, W., Gonzalez, D., and Woolley, J.B. 1989. Genetic relationships among seventeen *Aphidius* (Hymenoptera: Aphidiidae) populations, including six species. Annals of the Entomological Society of America, **82**: 754–768.
- Wang, Y., and Dong, D. 1997. Two new species of *Lysaphidus* and *Pauesia* (Hymenoptera: Aphidiidae) from Yunnan, China. Journal of Southwest Agricultural University, **19**: 213–216.