# Cerebral microvascular endothelium and the pathogenesis of neurodegenerative diseases

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Diseases of the central nervous system (CNS) pose a significant health challenge, but despite their diversity, they share many common features and mechanisms. For example, endothelial dysfunction has been implicated as a crucial event in the development of several CNS disorders, such as Alzheimer disease, Parkinson disease, amyotrophic lateral sclerosis, multiple sclerosis, human immunodeficiency virus (HIV)-1-associated neurocognitive disorder and traumatic brain injury. Breakdown of the blood-brain barrier (BBB) as a result of disruption of tight junctions and transporters, leads to increased leukocyte transmigration and is an early event in the pathology of these disorders. The brain endothelium is highly reactive because it serves as both a source of, and a target for, inflammatory proteins and reactive oxygen species. BBB breakdown thus leads to neuroinflammation and oxidative stress, which are implicated in the pathogenesis of CNS disease. Furthermore, the physiology and pathophysiology of endothelial cells are closely linked to the functioning of their mitochondria, and mitochondrial dysfunction is another important mediator of disease pathology in the brain. The high concentration of mitochondria in cerebrovascular endothelial cells might account for the sensitivity of the BBB to oxidant stressors. Here, we discuss how greater understanding of the role of BBB function could lead to new therapeutic approaches for diseases of the CNS that target the dynamic properties of brain endothelial cells. ere

In 1966, the eminent cell biologist Lord Florey described vascular endothelial cells as a 'sheet of nucleated cellophane' (Ref. 1). Using light microscopy, the technology available at the time, vascular endothelium appeared as a uniform,

passive barrier interposed between circulating blood and the underlying arterial wall (Ref. 1). With the advent of modern vascular biology, and the ability to isolate and culture endothelial cells, an image of a metabolically active, highly

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> 1 Accession information: doi:10.1017/S1462399411001918; Vol. 13; e19; June 2011 © Cambridge University Press 2011

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synthetic cell that is a key participant in inflammatory, immune and endocrine processes has emerged (Refs 2, 3). Endothelial cells display heterogeneity in morphology, molecular components and functional output (Ref. 4). Studies using gene expression profiling have demonstrated tissue-specific expression patterns in microvascular endothelial cells (Ref. 5). The brain endothelium is a highly specialised endothelial cell type that has an active role in the physiology and pathology of the brain (Refs 6, 7, 8).

The concept of a blood-brain barrier (BBB) evolved from the early work of the German scientist Paul Ehrlich, who in 1885 showed that intravascular dyes that freely permeate peripheral tissue are unable to penetrate the brain (Ref. 9). It is now recognised that the BBB is not an impermeable wall, but rather an active, dynamic interface that permits selective entry of solutes into the central nervous system (CNS) (Refs 6, 8). In 1967, Reese and Karnovsky demonstrated that the cerebral endothelium is the anatomical site of the BBB (Ref. 10). Brain endothelia possess structural, biochemical and functional characteristics that are adapted for this purpose. The presence of highly organised tight and adherens junctions between adjacent endothelial cells, as well as the relative paucity of vesicular transport, restricts the passage of non-lipid-soluble substances from the blood to the brain. The presence of enzymes such as 3,4dihydroxyphenylalanine (DOPA) decarboxylase and monoamine oxidase on the surface of brain endothelial cells prevents unregulated entry of intravascular amines into the CNS (Refs 11, 12). The cerebral endothelium actively controls the passage of polar solutes into the CNS through energy-dependent, carrier-mediated systems that transport hexose sugars, amino acids, monocarboxylic acids, nucleosides and vitamins (Refs 6, 8, 13, 14, 15, 16, 17).

The confines of the cranial space, as well as the need to preserve the functional relationships between elements of the CNS, stress the importance of the BBB in the maintenance of the brain milieu. The haemodynamic communication between neurons and the cerebrovasculature is necessary to efficiently couple cerebral blood flow (CBF) to neuronal activation (Ref. 18). The cerebral endothelium is a key component of the neurovascular unit, an emerging concept that emphasises the interactions among glial, neuronal and vascular

elements (Refs 6, 7, 19, 20) (Fig. 1). Pericytes are integral members of the neurovascular unit nesi (Ref. 21). These cells are important players in vascular-mediated events in the CNS. However, a discussion of pericytes is beyond the scope of this review; the reader is referred to informative publications on this important and understudied cell type (Refs 6, 21, 22). Homeostatic signalling within the neurovascular unit is crucial for normal brain function. Dysfunctional cell-cell signalling in the neurovascular unit is increasingly implicated as a characteristic

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feature of CNS diseases (Ref. 7). Several excellent papers have addressed the importance of the neurovascular unit for brain health (Refs 23, 24, 25, 26); the role of the brain endothelial cell in the development of diseases

#### Markers of endothelial dysfunction and **CNS** disease

of the CNS is the focus of this review.

r endothelium and The activated endothelial cell in various clinical states has become a major target of human therapeutics (Refs 27, 28, 29). A dysfunctional endothelium contributes significantly to systemic vascular disorders such as atherosclerosis (Ref. 30). Increasingly, endothelial dysfunction has been implicated as a link DC between neurological dysfunction and vascular disease and as a crucial player in the Ũ development of CNS disorders, including Alzheimer disease (AD), Parkinson disease (PD), amyotrophic lateral sclerosis (ALS), multiple U U sclerosis (MS), human immunodeficiency virus (HIV)-1-associated neurocognitive disorder and traumatic brain injury (TBI) (Refs 6, 31, 32, 33, 34, 35, 36). Vascular risk factors, including cholesterol, homocysteine and hypertension, have been shown to be important or relevant for the development of neurodegenerative diseases such as AD (Ref. 35). Using post-occlusive reactive hyperaemia and spectral analysis of skin vasomotion as markers of endothelial function, a recent study documents peripheral endothelial dysfunction in unmedicated patients with schizophrenia (Ref. 37). Markers of endothelial injury or dysfunction are demonstrable in several neurodegenerative disorders, as discussed below.

#### **Endothelial microparticles**

Endothelial microparticles (EMPs) are submicrometre-sized vesicles shed from plasma S

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Figure 1. The neurovascular unit. The neurovascular unit (NVU) consists of astrocytes, pericytes, neurons and the endothelium. Endothelial cells are linked by tight junctions and encircled by the basal lamina. The figure illustrates the close relationships among the NVU, microglia and neurons. Note that in the figure only the capillary vessel is shown with its adjacent pericyte and nearby microglia and neurons. The factors released from endothelial cells in this vessel could impact all these cell types. In larger smooth muscle containing venules and arterioles, smooth muscle cells might also be affected (not shown). The following is a partial listing of the secretory products released by highly synthetic endothelial cells (Ref. 96). Anti- and procoagulation proteins: antithrombin III, factor V, heparin sulfate, plasminogen activator inhibitor-1, plasminogen activator inhibitor-2, platelet-activating factor, prostacyclin, thrombomodulin, thromboxane A2, thromboplastin, von Willebrand factor. Extracellular matrix proteins: collagen I-XVIII, fibronectin, laminin, proteoglycans. Growth factors: endothelium-derived growth factor, fibroblast growth factor, granulocyte colony-stimulating factor, granulocyte macrophage colony-stimulating factor, insulin-like growth factor, platelet-derived growth factor, transforming growth factor beta. Inflammatory mediators: cell adhesion molecules, interleukin-1, interleukin-6, interleukin-8, leukotriene B4, leukotriene C4, leukotriene D4, leukotriene E4, major histocompatibility complex class II, monocyte chemoattractant protein-1, monocyte chemoattractant protein-2. Proteases: matrix metalloproteinases, thrombin, tissue plasminogen activator. Vasoconstrictor/dilation factors: endothelin, endothelium-derived contracting factor, free radicals, leukotrienes, thromboxane, A2/prostaglandin F2a, endothelium-derived hyperpolarising factor, nitric oxide, prostacyclin/prostaglandin E2.

membranes in response to cell activation, injury or apoptosis. EMPs are an emerging marker of endothelial dysfunction and are also considered to have a major biological role in inflammation, vascular injury, angiogenesis and thrombosis (Ref. 38). EMPs are known to be increased in acute coronary syndromes, severe hypertension with end organ damage, and thrombotic thrombocytopenic purpura, which are all conditions associated with endothelial injury (Ref. 39). Several publications implicate EMPs in the development of several CNS disorders (Ref. 40). A study examining the cerebrospinal

erebral microvascular endothelium and the pathogenesis of fluid (CSF) and plasma of patients with severe TBI shows increased generation of EMPs in the CSF of patients with TBI compared with controls (Ref. 33). Sustained generation of these microparticles in TBI patients is associated with a poor clinical outcome, suggesting that EMPs drive deleterious processes that contribute to TBI-induced functional decline (Ref. 33).

The release of EMPs has also been associated with endothelial dysfunction in patients with MS (Refs 41, 42, 43). A study examining the plasma from 50 patients with MS (30 in exacerbation and 20 in remission) and from 48

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controls showed that endothelial dysfunction, demonstrated by shedding of EMPs that express platelet-endothelial cell adhesion molecule-1 (CD31), is a consistent feature during exacerbation of MS. Furthermore, EMPs from activated cerebral endothelial cells in MS promote the inflammatory reaction characteristic of MS and enhance transmigration of activated leukocytes into the brain (Ref. 43).

#### Blood-based biomarkers of microvascular pathology

Several well-defined markers of endothelial injury are detected in CNS disease states. In schizophrenia, markers of endothelial dysfunction (von Willebrand factor and C-reactive protein) are correlated with the severity of clinical symptoms in attack-like schizophrenia (Ref. 44). Elevated levels of soluble adhesion molecules, including intracellular adhesion molecule-1 (ICAM-1) and vascular cell adhesion molecule-1 (VCAM-1), have been reported as markers of BBB damage in MS and AD (Refs 45, 46). Also, endothelialderived vasoactive proteins, including endothelin and atrial natriuretic peptide, are significantly altered in mild AD and in mild cognitive impairment (Ref. 46), which suggests that these proteins could be useful in disease staging and diagnosis.

#### **Endothelial progenitor cells**

An emerging marker of vascular function that has been little explored in neurodegenerative disease is the endothelial progenitor cell (EPC). Recent evidence has shown that vascular function not only depends on cells that reside within the vessel wall, but also appears to be significantly modulated by circulating cells derived from the bone marrow. A specific subset of these stem cells, termed EPCs, has been shown to promote vascular repair and improve endothelial function (Refs 47, 48). Since their original discovery in 1997 (Ref. 49), circulating EPCs have emerged as an important biomarker for cardiovascular disease (Ref. 50). Evidence is accumulating that EPCs contribute substantially to preservation of a structurally and functionally intact endothelium (Ref. 48). A decrease in the number of EPCs is a predictor of morbidity and mortality in cardiovascular diseases (Refs 50, 51). A recent study documents a decrease in EPCs in patients with AD (Ref. 52). Further research into EPCs in CNS disorders is warranted.

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Data from EMPs, blood-based proteins and EPCs support the idea that endothelial cells are injured in CNS diseases. At the gene level, endothelial dysfunction is suggested by transcriptional profiling, which shows low expression of the homeobox gene MEOX2 (also known as GAX) in brain endothelial cells in AD (Ref. 53). In mice, deletion of this gene results in vascular regression and altered amyloid beta (A $\beta$ ) transport (Ref. 53). The consequences of endothelial dysfunction for BBB brain permeability, transport and BBB neurodegenerative disease pathogenesis are described below.

#### Implications of BBB dysfunction in **CNS** disease

Pathological changes in BBB permeability Brain capillaries exhibit a number of anatomical biochemical characteristics and seemingly unique to membranes known to regulate water and electrolyte permeability such as renal proximal tubules, including the presence of tight junctions and a high mitochondrial content (Ref. 54). The major components of brain endothelial junctions are occludin, claudins, VEcadherin, zona occludens proteins ZO-1 to ZO-3 (also known as TJP1-TJP3) and junctionalassociated molecules (Refs 6, 55) (Fig. 2). Although tight junctions serve as the main anatomic barrier, restricting paracellular traffic of hydrophilic molecules, lipophilic compounds move freely along their concentration gradients and enter the CNS. BBB breakdown as a result of disruption of tight junctions, which might initiate or contribute to a vicious circle of disease resulting in progressive synaptic and neuronal dysfunction and loss, has been demonstrated in MS, ALS and HIV-1-associated erel neurocognitive disorder (Refs 6, 55, 56).

#### Multiple sclerosis

Endothelial cell dysfunction might contribute to the pathogenesis of MS (Ref. 36). MS is a demyelinating disease characterised bv breakdown of the BBB and accumulation of inflammatory cell infiltrates in the CNS. In MS, finger-like projections of demyelination extend into the white matter alongside blood vessels (Ref. 57). In experimental autoimmune encephalomyelitis (EAE), an animal model of MS, there is a significant relocalisation of the tight junction protein ZO-1, which precedes



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Sunctional proteins create a barrier between adjacent brain endotnellal c Expert Reviews in Molecular Medicine © 2011 Cambridge University Press

**Figure 2. Junctional proteins create a barrier between adjacent brain endothelial cells.** Main microvascular endothelial cells form the primary anatomical barrier between the blood and the brain. The endothelium is relatively impermeable to polar molecules as a result of tight and adherens junctions between cells. The junctional complexes are composed of several proteins. The primary proteins that form the tight junctions are the claudins (3, 5 and 12), occludin, zona occludens proteins ZO-1, ZO-2 and ZO-3, and junctional adhesion molecule-1 (JAM-1). Adherens junctions are primarily formed by VE cadherin, which is connected to actin within the cells.

overt clinical disease and correlates with sites of inflammatory cell accumulation (Ref. 58). Modifications in the integrity and organisation of junctional molecules and BBB permeability are thought to contribute to the pathogenesis of MS (Refs 59, 60). Dysregulation of junctional permits molecular complexes progressive immune cell entry into the CNS. Also, under normal physiological conditions, resting leukocytes do not cross the BBB in significant numbers because nonactivated leukocytes do not avidly bind to or penetrate the healthy BBB. In MS, increased expression of several chemokines and chemokine receptors on reactive cerebral endothelial cells facilitates transmigration of leukocytes across the BBB (Refs 36, 61, 62). Dysregulation of the BBB and transendothelial

migration of activated leukocytes are among the earliest cerebrovascular abnormalities seen in MS and parallel the release of inflammatory cytokines or chemokines in this disease (Ref. 61).

#### Amyotrophic lateral sclerosis

ALS is a chronic neurodegenerative disorder of motor neurons in the brain, brainstem and spinal cord that results in progressive paralysis and death (Ref. 63). A breach of the BBB has been suggested in ALS because of the increased levels of albumin, IgG and complement proteins found in the CSF and spinal cord in ALS patients and in animal models of the disease (Refs 56, 64). Mouse models of ALS, which express mutant forms of the antioxidant enzyme superoxide dismutase-1 (SOD1), show

vascular abnormalities before motor neuron degeneration (Refs 34, 63, 65). Specifically, there is a reduction in the level of tight junction proteins ZO-1, occludin and claudin-5 (Refs 56, 65). Also, Prussian Blue staining, an indicator of haemosiderin deposition and microhaemorrhages, in presymptomatic SOD1 mutants suggests that barrier breakdown is an early event in disease progression (Ref. 34). Endothelial damage accumulates before motor degeneration and neurovascular neuron inflammation in the SOD1 mutant, supporting a primary role for endothelial dysfunction in disease initiation (Refs 56, 65).

#### HIV-1-associated neurocognitive disorder

HIV-1-associated neurocognitive disorder is the most common and clinically important CNS complication of late HIV-1 infection (Ref. 66). BBB-regulated cellular trafficking is crucial for the development of CNS pathology caused by HIV, as well as other neuroinflammatory conditions, including meningitis and encephalitis (Ref. 6). Exposure of the BBB to the HIV-1 virus or virus-infected monocytes affects several aspects of BBB function as well as brain endothelial cell protein expression (Ref. 67). HIV-1 virus or virus-infected monocytes activate brain endothelial cells, enabling leukocyte passage across the BBB through highly regulated processes such as diapedesis and adsorptive endocytosis (Ref. 32). High levels of soluble CD40 ligand (CD40L) are found in CSF and plasma, whereas CD40, a receptor for CD40L, is highly expressed in the brain endothelial cells of patients with HIV-1 encephalitis (Ref. 68). Exposure of human brain cells to CD40L upregulates endothelial expression of leukocyte adhesion molecules (ICAM-1, VCAM-1), which causes a fourfold increase in monocyte adhesion to, and migration across, brain endothelial cells in an in vitro BBB model (Ref. 68). Brain endothelial cells also secrete neuroinflammatory molecules when stimulated by HIV-1 or HIV-derived proteins Tat (Ref. 69). A proteomic gp120 and examination of human brain microvascular endothelial cells exposed to HIV-1-infected macrophages shows that over 200 proteins in brain endothelial cells are upregulated (Ref. 70). Proteins in a wide range of functional categories are affected, including voltage-gated ion channel, heat shock, transport, cytoskeletal, regulatory

and calcium-binding proteins. Also, HIV-1 virions induce transcriptional upregulation and expression of proinflammatory genes and the transcription factor STAT1 in brain endothelial cells (Ref. 69). Overall, evidence that the BBB is impaired during progressive HIV-1 disease and the emergence of cognitive impairments during HIV-1-associated cognitive disorder support a major role for brain endothelial cells in establishing and maintaining the virus within the CNS.

#### Pathological changes in BBB transport

The high energy and nutrient demands of the brain, as well as the need to tightly regulate the electrolyte milieu for optimal neuronal function, highlight the importance of BBB transport systems. Specific carrier-mediated transport systems facilitate transport of nutrients such as hexose sugars (glucose, galactose), neutral, basic and acidic amino acids, monocarboxylic acids (lactate, pyruvate, ketone bodies), nucleosides (adenosine, guanosine, uridine), purines (adenine, guanine) and amines (choline) (Refs 6, 9, 13, 14, 15, 16, 17) (Fig. 3). The concentration gradients for nutrients are generally in the direction from the blood to the brain. These are regulated by brain metabolic needs and by the concentrations of substrates in plasma (Ref. 6). The complexity of the BBB is further indicated by the polarity of cerebral endothelium with regard to transport, with differential localisation of carriers in luminal and abluminal plasma membranes (Ref. 71). Accumulating evidence suggests that these transport systems are dysregulated in CNS disease and, indeed, might significantly contribute to disease initiation and progression.

#### Alzheimer disease

Because of the importance of BBB transporters for the maintenance of the CNS milieu, abnormalities in brain endothelial cell function that result in altered transporter activities have consequences for brain function and disease development. In AD, a consistent finding evident early in the disease process is region-specific decrease in glucose utilisation (Ref. 72). Analysis of positron emission tomography scans from patients with mild cognitive impairment shows diminished glucose uptake in brain areas susceptible to the development of AD (Refs 73, 74). It is likely that the decreased expression of GLUT1, the primary

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Figure 3. Carrier-mediated transporters of the blood-brain barrier. The high energy and nutrient demands of the brain, as well as the need to tightly regulate the electrolyte milieu for optimal neuronal function, highlight the importance of blood-brain barrier transport systems. Specific carrier-mediated transport systems facilitate transport of nutrients such as hexose sugars, neutral, basic and acidic amino acids and monocarboxylic acids, purines and amines. Transporters and prototypical substrates are shown. Glucose 1 transporter (GLUT1) (glucose); equilibrative nucleoside transporters (ENTs) (purines); monocarboxylic acid transporter 1 (MCT1) (lactate); organic cation transporters (OCTs) (choline).

glucose transporter at the BBB, documented in AD patients contributes to this energy deficit (Ref. 75).

Also important in the pathogenesis of AD is the Aß protein. The amyloid cascade hypothesis of AD postulates a central role for this protein in disease pathogenesis (Ref. 76). Levels of  $A\beta$  in the brain are regulated by active processes at the BBB. Individuals with late-onset AD do not have increased production of  $A\beta$  and are thought to have increased AB accumulation as a result of its faulty clearance (Ref. 6). The receptor for advanced glycation end products (RAGE) is the major influx transporter for  $A\beta$ across the BBB. Low-density lipoprotein receptor-related protein 1 (LRP1) is a major efflux transporter for  $A\beta$  across the BBB (Ref. 77). LRP1 is a member of the LDL receptor family and acts as a multifunctional scavenger and signalling receptor (Ref. 78). Binding of  $A\beta$ to LRP1 at the abluminal side of the BBB

initiates AB clearance from brain to blood by transcytosis across the BBB (Ref 77). The importance of LRP for AB clearance is highlighted by several studies in transgenic AD mice and AD patients. In animals deficient in the homeobox gene MEOX2, impaired Aβ efflux is caused by reduced LRP levels (Ref. 53). In AD patients and AD mouse models, a non-AB clearing phenotype results from downregulation of LRP1 (Ref. 79). Aβ deposition in AD individuals with cerebral amyloid angiopathy is controlled through the LRP1 receptor (Ref. 79). Finally, the Dutch mutant form of  $A\beta$  is poorly cleared from the brain because of low-affinity binding of LRP to AB (Ref. 80). In AD and transgenic models, RAGE expression has been shown to increase, whereas the number of LRP receptors decreases (Ref. 81). In mouse models of AD, dense plaques develop initially on blood vessels. It is believed that plaques are generated on blood vessels because of deficient AB

Accession information: doi:10.1017/S1462399411001918; Vol. 13; e19; June 2011 © Cambridge University Press 2011 clearance across the BBB (Ref. 82). Reduced expression of LRP1 has been associated with positive staining of cerebral vessels for AB during normal ageing in rodent and nonhuman primates, and in individuals with AD (Ref. 83).

RAGE-dependent BBB transport of circulating A $\beta$  results in expression of proinflammatory cytokines in the brain and production of endothelin-1, which causes decreased CBF. Systemic administration of a recombinant truncated form of RAGE reduced the accumulation of AB in the brain parenchyma of transgenic APP mice (Ref. 80). Finally, inflammatory proteins have been shown to impair LRP activity and increase RAGE, thereby promoting increased A $\beta$  in the AD brain (Ref. 84).

#### Parkinson disease

PD is a neurodegenerative disorder that is characterised by resting tremor, rigidity and involves bradykinesia and progressive degeneration of dopaminergic neurons in the substantia nigra leading to dopamine deficiency (Ref. 85). The BBB expresses transporters that affect both PD drug therapy and dopaminergic neuronal function. L-DOPA, but not dopamine, is transported across the BBB in humans by the L1 facilitative transporter. After transport across the BBB, L-DOPA is converted to dopamine at nerve terminals that contain DOPA decarboxylase (Ref. 86). Impairment of BBB function has been implicated in the pathogenesis of PD (Refs 87, 88). In PD individuals, capillaries are mis-shapen and normal contacts between nigral neurons and capillaries are lost at an early stage of disease. Capillary basement membrane thickening and collagen accumulation have also been shown in PD (Ref. 89). Focal disruption of the BBB induces in vivo degeneration of dopaminergic neurons, which is a characteristic feature of PD (Ref. 87). In an animal model of PD, administration of 1-methyl-4-phenyl-1,2,3,6tetrahydropyridine results in motor deficits and loss of dopaminergic neurons, and is temporally associated with a significant reduction in mRNA for the BBB L-type amino acid transporter for tyrosine, a precursor for dopamine, which enters the brain through the L-type amino acid transporter (Ref. 90).

It has also been suggested that absorption or metabolism of putative PD toxins and/or their faulty elimination across the BBB has a role in the pathogenesis of PD (Ref. 6). The ATP-

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binding cassette superfamily of transporter () genes regulates the bioavailability of xenobiotics. nesi The multidrug transporter of P-glycoprotein (Pgp) is the best characterised member of this family (Ref. 91). Low activity of the P-gp efflux Ū transporter at the BBB in the midbrain of σ individuals at risk for PD supports a mechanistic role for this transporter in the retention of putative PD toxins. Imaging studies have demonstrated impairment in the ability of BBB efflux pumps to remove neurotoxins in patients with PD (Ref. 92). Furthermore, inflammation has also been shown to reduce the activity of P-gp (Ref. 93). The importance of Pgp in the pathogenesis of PD is derived from gene polymorphism studies. Some polymorphisms might be protective and reduce risk, whereas others might predispose carriers to the damaging effects of pesticides, leading to a PD phenotype (Refs 91, 94, 95). This transporter endothe system is a significant modulator of susceptibility to PD among male ethnic Chinese who are more than 60 years old (Ref. 91).

#### Brain endothelial cells as regulators of neuroinflammation and oxidative stress in neurodegenerative disease

J Neuroinflammation and oxidative stress are both implicated in the pathogenesis of Ū neurodegenerative disease. The brain Ú, endothelium is a highly dynamic, reactive interface that serves as both a source of, and a target for, inflammatory proteins and reactive oxygen species (ROS) (Ref. 96). Using laser Ü capture microdissection, a recent study documents that endothelial samples showed differences in gene expression between schizophrenics and controls. Functional profiling revealed that the genes affected are related to inflammatory processes (Ref. 97). Vascularproducts derived of а dysfunctional endothelium could result in neuronal injury in neurodegenerative disease states (Ref. 98). Thus, beyond overt disruptions in BBB permeability or transport functions, abnormal endothelial function can affect the CNS milieu and neuronal viability or survival. For example, in the AD brain, an injured or altered brain endothelial cell releases a large number of bioactive molecules, some of which are directly toxic to neurons (Ref. 99). Because of the intimate communication among cells in the neurovascular unit, cell products endothelial have important

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paracrine effects on astrocytes and microglia (Refs 6, 20). In this regard, the neurotoxic protease thrombin, which is synthesised and released by brain endothelial cells in AD (Ref. 100), has significant proinflammatory effects on both microglia and astrocytes. In microglia, thrombin increases transcription of inflammation-associated genes tumour necrosis factor alpha (TNF $\alpha$ ) and inducible nitric oxide synthase (Ref. 101). In astrocytes, activation of the thrombin receptor (PAR-1) leads to increased expression of matrix metalloproteinase 9 (MMP-9) (Ref. 102). Thus, vascular-derived thrombin might directly injure neurons or affect neuronal viability indirectly by activation of microglia and astrocytes in the neurovascular unit.

#### Neuroinflammation: a key role for matrix metalloproteinases in the inflammatory cycle

The cerebromicrocirculation is a rich source of cytokines and chemokines. In neurodegenerative diseases, a myriad of inflammatory-associated proteins have been documented (Refs 6, 68, 69, 84, 96, 103, 104, 105). Rather than list these factors, we focus on MMPs, central players in the inflammatory response, in order to explore how their expression and effects on BBB function might contribute to pathological events in the diseased brain.

The major group of enzymes controlling basement membrane and extracellular matrix turnover are the gelatinases (MMP-2 and MMP-9), collagenases, stromelysins and membranetype MMPs. MMPs are usually secreted in an inactive proenzyme form and can be activated extracellularly by other proteinases, including active MMPs and some tissue inhibitors of metalloproteinases (TIMPs) (Refs 103, 106) (Table 1). Several physiological mechanisms prevent uncontrolled MMP activation. One of these is binding to MMPs by TIMPs. By inhibition of MMP activity, TIMPs contribute to the regulation of ECM remodelling associated with various physiological and pathological conditions (Refs 107, 108). TIMPs can also aid in the activation of MMPs. For example, TIMP-2 forms a trimolecular complex with the pro-form of MMP-2 and membrane type (MT)-MMP-1, thereby activating MMP-2 at the cell surface (Ref. 103). MMPs can cleave virtually all structural ECM molecules. In addition to providing structural scaffolding, ECM molecules

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act as binding reservoirs for various growth () factors and cytokines (Ref. 109). MMPs increase **B**S the permeability of the BBB by attacking the ECM, basal lamina and tight junctional proteins, e pathogen leading to neuroinflammation (Ref. 103). MMPs are expressed by all cell types of the neurovascular unit, including endothelial cells (Refs 103, 106, 110, 111); however, the expression of MMPs by brain endothelial cells is relatively unexplored.

Inflammatory cytokines can regulate expression of MMPs, and these enzymes can, in turn, affect the level and activity of inflammatory proteins. The major proinflammatory cytokines interleukin 1 $\beta$  (IL-1 $\beta$ ), TNF $\alpha$ , interferon- $\gamma$ (IFN $\gamma$ ) and IL-6 are important regulators of MMP and TIMP expression (Refs 106, 112). All these cytokines have been shown to be upregulated in neurodegenerative disease. In this regard, the inflammatory protease thrombin, which is upregulated in AD and other brain diseases (Refs 100, 113), causes a time- and Ö dose-dependent decrease in the attachment of endothelial cells to basement membrane components, with a concomitant increase in Φ MMP-2 activation in endothelial cells (Ref. 114). Exposure of cultured brain endothelial cells to both IL-1 $\beta$  and TNF $\alpha$  causes a shift in the expression of TIMP-1 and TIMP-3; TIMP-1 expression is upregulated, whereas levels of TIMP-3 decrease (Ref. 106). This imbalance in Ū TIMP expression has implications for the regulation of MMPs in brain microvasculature (Ref. 106). TNF $\alpha$  and IL-1 $\beta$  also induce the ΰ transcription of MMP-3 and MMP-9, which are both involved in acute and chronic neuroinflammation (Ref. 103). MT-MMP-1, a cell surface MMP, acts as a broad-spectrum ECM proteinase and can regulate activation of other MMPs. Because MT-MMP-1 is efficiently inhibited by TIMP-3 but not by TIMP-1, the 'reprogramming' of the TIMP-1–TIMP-3 balance during brain endothelial cell activation by inflammatory proteins could result in a local increase in MT-MMP-1 activity, with consequences for microvascular basement membrane proteolysis.

MMPs contribute to the propagation and regulation of neuroinflammatory responses to injury. The development of tissue damage that follows transient ischaemia suggests the crucial interplay between MMPs and mediators of neuroinflammation (Refs 115, 116). MMPs are

https://doi.org/10.1017/S1462399411001918 Published online by Cambridge University Press

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MMP/TIMP	Function/effects	Increased levels in disease	Decreased levels in disease
MMP-2 (Gelatinase A)	Disrupts BBB, angiogenesis, neurogenesis Causes apoptosis	Microvessels (AD)	CSF (AD)
MMP-9 (Gelatinase B)	Disrupts BBB, angiogenesis, neurogenesis Causes apoptosis	CSF (MS) Plasma (AD) Microvessels (AD) Brain (ALS) Serum (ALS, MS) Plasma (AD)	CSF (ALS) Plasma (AD) CSF (MS)
MMP-3 (Stromelysin-1)	Disrupts ECM, BBB, angiogenesis Causes apoptosis Activates microglia		
MT-MMP-1 (MMP-14)	Membrane-bound MMP Broad-spectrum ECM proteinase Activates MMP-2		
TIMP-1	Inhibits MMP-9	CSF (AD, PD, ALS) Microvessels (AD) Brain (PD)	
TIMP-2	Inhibits MMP-2 Activates MMP-2	CSF (AD) Correlated with MMP-9 (AD)	Correlated with MMP-9 (AD)
TIMP-3	Remodels EBM Inhibits MMP-2, MMP-9 and MT-MMP-1 Inhibits angiogenesis Causes apoptosis		
Abbreviations: AD, cerebrospinal fluid; 1β, interleukin-1 be Parkinson disease; <sup>a</sup> See Refs 103, 124	Alzheimer disease; ALS, amyotroph EBM, endothelial basement membr eta; MMP, matrix metalloproteinase; TIMP, tissue inhibitor of metallopro 1, 158, 159, 160, 161.	nic lateral sclerosis; BBB, blo rane; EC, endothelial cell; EC MT-MMP-1, membrane-type teinases; TNFα, tumour neci	ood-brain barrier; CSF, CM, extracellular matrix; IL e metalloproteinase-1; PD rosis factor alpha.

involved in the processing of proinflammatory cytokines, such as IL-1β, into biologically active forms through a caspase-1-independent pathway (Ref. 117). MMPs and TIMPs can contribute to immunoregulatory processes by affecting cytokine and cytokine receptor turnover (Ref. 109). For example, MMPs can recombinant soluble activate proTNFa (Ref. 118). In addition to causing the activation and release of cytokines and growth factors, MMPs can also cleave their cell surface receptors (Ref. 119).

The relevance of MMPs in the development of injury in several chronic neurodegenerative disorders, including AD, PD, ALS, MS and HIV-1-associated neurocognitive disorder (Refs 103, 110), is supported by studies documenting altered MMP and TIMP levels, relative to controls, in the CSF, brain-derived microvessels and plasma. It is important to note that there are some contradictory data in the literature regarding changes in the levels of MMPs and TIMPs in the CSF and plasma in neurodegenerative diseases (see Table 1 for a

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more detailed breakdown of MMP and TIMP changes in various neurodegenerative diseases).

#### MMPs regulate lymphocyte and monocyte transmigration across the BBB

MS is a disease of the CNS that is characterised by multifocal infiltration of T cells from the systemic immune system across the BBB (Ref. 36). MMPs disrupt the BBB and facilitate the remodelling of the ECM, they are therefore heavily implicated in MS because changes in the barrier function of brain microvessels allow T cells to invade the CNS more readily (Ref. 120). In EAE, inhibitors of MMPs slow down the progression of symptoms (Ref. 103). This indicates that MMPfacilitated endothelial cell dysfunction might be part of the mechanism by which leukocytes penetrate the brain tissue and damage the myelin sheath of white matter neurons in MS.

Exposure of human brain microvascular endothelial cells to HIV-infected monocytes results in decreased expression of tight junction proteins (Ref. 121). Alterations of protein expression in tight junctions are associated with increased endothelial permeability and increased transendothelial migration of HIV-infected monocytes across an in vitro model of the BBB (Ref. 122). The HIV-induced decrease in the expression of junctional adhesion molecule-1 (JAM-1), occludens and ZO-1 is restored by inhibition of MMP activity (Ref. 122). This suggests that downregulation of MMP activity constitutes a novel mechanism for protection against HIV-induced BBB disruption (Ref. 122). The increased permeability of human microvascular endothelial cell monolayers to monocytes caused by HIV-1 infection is inhibited by TIMP-1 and TIMP-2, suggesting that MMP-2 and MMP-9 are primary mediators of enhanced permeability (Ref. 123).

#### MMPs in vascular inflammation and direct neurotoxic effects in AD and PD

In addition to altered levels of MMPs in CSF and plasma, MMPs have been localised to intraparenchymal lesions found in neurodegenerative diseases (Ref. 103). This implies that in addition to their known function degrading ECM, MMPs might serve as mediators of apoptotic or necrotic cell death.

The cerebral microvasculature could be central to a destructive cycle of events where inflammatory factors stimulate MMP expression,

of which in turn promotes the release inflammatory mediators. Brain microvessels derived from patients with AD release IL-6, TNF $\alpha$ , IL-8, IL-1 $\beta$ , thrombin, and MMP-2 and MMP-9 (Refs 100, 104, 105, 124). Through the effects of MMPs on cytokines and stimulation of MMPs by inflammatory proteins, MMPs could be key amplifiers of local inflammation. The close proximity of blood vessels to AD lesions suggests intimate communication between the neurovascular unit and neurons (Ref. 125). In PD, mutation or overexpression of  $\alpha$ -synuclein protein has a pivotal role in the pathogenesis of PD (Ref. 126).  $\alpha$ -Synuclein-stimulated microglia release MMPs that subsequently activate the thrombin receptor, amplifying microglial inflammatory signals in an autocrine manner (Ref. 126). Similar pathways might operate in brain endothelial cells because MMPs, thrombin and  $\alpha$ -synuclein have all been identified in these cells (Refs 100, 124, 127).

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Finally, MMPs might have a role in 0 neurodegenerative disease by both direct and indirect neurotoxic mechanisms. MMP-3 has been implicated as a neurotoxic factor that Ð contributes of directly to the death dopaminergic neurons in PD (Ref. 128). In AD, Π MMP-9 toxicity in A $\beta$ -mediated animal models DC is supported by experiments where inhibition of MMPs improves Aβ-mediated cognitive Ũ impairment and neurotoxicity in mice (Ref. 129). Ū Recent evidence indicates that protein aggregation, and in particular the formation of 5 toxic protein oligomers, is a key mechanism in synucleinopathies such as PD (Ref. 130). In vitro models of pathological  $\alpha$ -synuclein aggregation indicate that MMP-1 and MMP-3 can influence the pathogenesis of PD by the generation of aggregation-enhancing specific  $\alpha$ -synuclein Ð fragments that result from limited proteolysis (Ref. 130). Φ

#### Mitochondria as mediators of oxidative stress

The human brain, although it constitutes only 2% of total body weight, accounts for 20% of oxygen consumption in the body, reflecting its high rate of metabolic activity and the high level of ATP consumption by neurons (Ref. 131). Mitochondria have a crucial role in the supply of energy to the brain. The mitochondrial hypothesis of neurodegeneration postulates that defects in mitochondrial metabolism lead to

mitochondrial dysfunction and eventually to depletion of cellular ATP, increased free radical generation, calcium dysregulation and. potentially, defective mitochondrial fission or fusion (Ref. 132). Mitochondrial membrane lipids are highly susceptible to ROS, especially the long-chain polyunsaturated fatty acid components (Ref. 133). Inner mitochondrial membrane proteins such as complexes I, II and III are themselves directly susceptible to effects of oxidative stress, leading to membrane depolarisation and subsequently, impaired mitochondrial function (Ref. 133). Disorders of mitochondrial function, caused by mutations in mitochondrial DNA, proteins or the presence of mitochondrial inhibitors, appear to have a role in several neurodegenerative diseases (Ref. 132).

## Cerebrovascular mitochondria, ageing and oxidative stress

Diseases of the CNS are for the most part ageassociated disorders (Ref. 134). It is well mitochondrial established that DNA accumulates mutations with ageing (Ref. 133). A decrease in the mitochondrial antioxidant MnSOD (superoxide dismutase) has also been found in the cerebrovasculature with increasing age (Ref. 135). The physiology and pathophysiology of endothelial cells are closely related to the functioning of mitochondria, and mitochondrial dysfunction could be an important mediator of vascular lesions (Refs 136, 137). Endothelial mitochondria might serve as sensors of the local environment (i.e. oxygen concentrations) (Ref. 138). This is especially important for cerebrovascular endothelial cells because of the interactions between endothelia, astrocytes and neurons within the neurovascular unit. Also, because of the highly metabolic nature of the BBB and the numerous transport systems it supports, brain endothelial cells extensive have an mitochondrial network and, relative to other endothelial cell types, an increased mitochondrial mass (Ref. 54). Many injuries or insults thought to initiate BBB dysfunction result in ROS, resulting in oxidative damage, tight junction disruption and activation of MMPs (Refs 139, 140). Exposure of brain endothelial cells to oxidised lipids increases nitric oxide and ROS production and stimulates translocation of the proapoptotic protein Bax (Refs 141, 142). Activation of the protease pathway involving

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Bax, mitochondrial cytochrome *c* and caspase in cerebral endothelial cells is attenuated by the antioxidant resveratrol (Ref. 142). The brain vasculature appears to be especially sensitive to oxidative stress. This sensitivity might in part be due to higher levels of NAD(P)H-oxidase in brain endothelial cells compared with endothelial cells in peripheral vessels (Ref. 143). In this regard, a recent study shows that the inflammatory protein cysteine and glycine-rich protein 1 (CRP) evokes NAD(P)H-oxidasedependent functional derangements in endothelial cells derived from the brain but not from the aorta (Ref. 143). The higher concentration of mitochondria in cerebrovascular endothelial cells, relative to other endothelial cell types (Ref. 54), might also account for the sensitivity of the BBB to oxidant stressors.

# Mitochondrial dysfunction in sporadic neurodegenerative diseases

Cerebral vasculopathy is a central aspect of most, if all, inherited mitochondrial not diseases. end Although these are rare diseases, this demonstrates that disorders whose cause is known to be mitochondrial in origin result quite specifically in cerebrovascular pathology. There increasing evidence that acquired is mitochondrial defects promote а similar vasculopathy in sporadically occurring disorders Ũ such as PD, AD and ALS (Refs 74, 144) (Fig. 4). Π The literature suggests pathogenic links among mitochondrial malfunction, ROS and chronic inflammatory diseases (Ref. 145). Genes Ü associated with autosomal recessive PD, including Parkin (PARK2), PINK1 (PARK6) and DJ-1 (PARK7), all encode proteins with functional effects on mitochondria (Refs 131, 146). Both PINK1 and Parkin are crucial in the removal of damaged mitochondria by mitophagy, whereas DJ-1 is localised to Φ mitochondria and has a role in oxidative stress protection (Ref. 131). Approximately 90% of cellular ROS production is attributable to mitochondria (Ref. 133). Oxidative stress has been consistently linked to neurodegenerative (Ref. 147). ROS production diseases in mitochondria can be induced by blocking complex I, a key enzyme of the respiratory chain, resulting in superoxide production. Administration of rotenone, a complex I inhibitor, to experimental animals results in loss of mitochondrial membrane potential, robust



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**Figure 4. Proposed blood-brain barrier mitochondrial dysfunction in CNS diseases.** In CNS disease states, the mitochondria become dysfunctional and produce large amounts of ROS. AD is characterised by the formation of  $A\beta$  plaques.  $A\beta$  disrupts the electron transport chain and calcium homeostasis, leading to ROS production in mitochondria. ALS is thought to be caused by damage or mutation to SOD1. Mutant SOD1 results in reduced clearance of mitochondrial-produced ROS. In HIV-1-associated neurocognitive disorder, HIV-1-infected cells produce the TAT protein. This viral-encoded protein induces apoptosis in cells by causing proapoptotic proteins to translocate to the mitochondrial cell membrane, leading to mitochondrial dysfunction, the release of ROS and eventual cell death. Genes associated with autosomal recessive PD encode proteins with functional effects on mitochondria. These genes are crucial in the removal of damaged mitochondrial dysfunction and eventual ROS production. Abbreviations:  $A\beta$ , amyloid beta protein; AD, Alzheimer disease; ALS, amyotrophic lateral sclerosis; CNS, central nervous system; HIV-1, human immunodeficiency virus-1; PD, Parkinson disease; ROS, reactive oxygen species; SOD1, superoxide dismutase-1.

ROS production, and PD-like symptoms and pathology (Ref. 148). Exposure of experimental animals to high levels of manganese results in neurodegenerative changes that are similar to those described in PD. Manganese induces direct injurv to mitochondria brain in endothelial cells (Ref. 149). Altered mitochondrial function probably underlies the ensuing impairment in energy metabolism, redox status and compromised BBB function. The central role of mitochondrial ROS-mediated signalling in endothelial cells implies that

mitochondrial dysfunction might lead directly to vascular pathology.

Oxidative stress, vascular abnormalities and metabolic energy defects are early features thought to have a crucial role in AD pathogenesis (Ref. 150). Mitochondria, as both a major source of ROS and energy-generating organelles, are probable contributors to the disease process. Studies of the BBB in AD patients have shown decreased mitochondrial content (Ref. 151). Studies of  $A\beta$ -peptideinduced cerebral endothelial cell death have

Accession information: doi:10.1017/S1462399411001918; Vol. 13; e19; June 2011 © Cambridge University Press 2011 demonstrated mitochondrial dysfunction and caspase activation (Ref. 152). AB disrupts the integrity of nuclear and mitochondrial DNA, activates caspase-8 and caspase-3, and causes apoptosis in brain endothelial cells (Ref. 152). Furthermore, Aβ-induced cell death can be prevented by zVAD-fmk, a broad-spectrum caspase inhibitor, or by the antioxidant N-acetylcysteine (Ref. 152).

In ALS, morphological and functional defects in mitochondria are found in both human ALS patients and ALS mice overexpressing mutant (Refs 153, 154). In SOD1 mice, SOD1 mitochondrial degeneration precedes disease symptoms, arguing for a causal role in disease progression. In these mouse models, BBB abnormalities, such as reduced numbers of tight junction proteins, also occur before motor neuron degeneration. SOD1 mice show dysfunctional mitochondria with reduced ATP production, oxidative phosphorylation and calcium buffering capacity (Ref. 34). Although SOD1 is a cytoplasmic protein, a segment of this protein partitions into the mitochondria. Mitochondrial accumulation of misfolded mutant SOD1 has been proposed as one possible trigger of cell death in this disease. A recent study shows that in ALS mice, mutant SOD1 damages mitochondria by promoting conformational changes in the apoptosis protein Bcl-2 (Ref. 155).

Taken together, these studies highlight the role of vascular oxidative stress and BBB mitochondria and suggest that strategies aimed at improving mitochondrial function or ROS scavenging are of potential clinical relevance for a wide variety of CNS diseases.

#### **Clinical implications**

Diseases of the nervous system pose a significant health challenge, affecting one in three Americans at some point in life, with a cost estimated by the Alzheimer's Association to exceed \$500 billion per year. Although these diseases are a diverse group, they share many features and common mechanisms. Increasingly, the brain vasculature appears to be an active player in the initiation and progression of these disorders. Indeed, with over 400 miles of capillary length in the brain and the intimate communication among cells of the neurovascular unit, a key role for the brain endothelium in mediating brain disease processes is not surprising (Ref. The 6).

endothelium is a common target for all cardiovascular risk factors, and functional impairment of vascular endothelial cells occurs

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long before overt disease. Neuroinflammation and oxidative stress, invariant features of neurodegenerative diseases, are associated with endothelial dysfunction. If therapeutic strategies to combat cardiovascular disease prove useful in treating or preventing neurodegenerative disorders, novel and effective therapies for devastating CNS disorders could be possible in the near future.

#### **Research in progress and outstanding** research questions

mand Although clinical manifestations of neurodegenerative disorders have long been described, identification of the cellular, biochemical and molecular bases for CNS disease is more recent. Many gaps remain in our knowledge and understanding of both the interplay of genetics and environment and the chronology of disease evolution. The specialised nature of BBB endothelium has evolved to protect the brain from injury; thus, even minor disturbances of endothelial function have important consequences for neuronal function. Despite the importance of brain endothelial cell function for CNS health, the idea that the cerebral vasculature could be an active participant in the pathogenesis of CNS disease is evolving concept. Because the an vascular–neuronal axis is still relatively unexplored, there is great potential for new discoveries in the areas of new disease markers and animal models.

To understand the contribution of brain endothelial function to CNS disease pathogenesis, neurovascular-derived markers such as EMPs, bloodborne proteins or EPCs could be used to investigate neurovascular pathophysiology in these diseases and could 0 highlight mechanisms that might be common among disparate brain diseases. The use of neurovascular-derived markers for the diagnosis and monitoring of neurodegenerative disorders could provide new insights into the disease processes in the brain. There is a concerted research effort to develop peripheral, accessible markers for neurodegenerative diseases. This is crucial for the development of noninvasive approaches that would allow in-office screening protocols as well as early disease diagnosis. The use of BBB endothelial biomarkers in the staging

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of CNS disorders, especially EPCs, is in its infancy and warrants further study.

The finding that vascular-based processes are contributory to CNS diseases opens up new possibilities for the development of unique CNS disease animal models. Currently, most animal models of CNS disease focus on the overexpression of specific proteins that are thought to have a role in disease pathogenesis. Although considerable useful information has been derived from these studies, there are limitations to this approach. Results are skewed to processes that involve those proteins and are based on a priori assumptions that these proteins or processes are the most relevant for disease progression. The development of new animal models where the inciting stimulus is 'vascular injury' could produce animals that show defects in proteins or signalling cascades not previously identified. This unbiased approach could identify new targets for therapy. It should be borne in mind that because endothelial injury is increasingly identified in CNS diseases, efforts will need to focus on whether these vascular changes drive the pathological events in the brain or are secondary bystander effects. Also, a role for vascular or endothelial abnormalities in the development of CNS diseases is an emerging field. It is therefore difficult to determine from the literature how 'common' these findings are, because until recently they had not been explored. Thus, the significance of these findings is open to discussion.

Finally, when considering the cerebromicrovasculature and diseases of the CNS, the BBB has traditionally been considered a hindrance for disease treatment. Indeed, an intact BBB is a major obstacle for the delivery of drugs into the CNS parenchyma. Approximately 98% of small-molecule drugs and all largemolecule neurotherapeutics are normally excluded from the brain (Refs 156, 157). However, the location of the cerebrovascular endothelium as an interface that is readily accessible to intravascular therapeutics, suggests that the BBB itself could be a target for both novel and existing drugs that might otherwise have been overlooked as neurotherapeutics because of their inability to cross the BBB. Understanding the functional BBB could lead to new therapeutic approaches for diseases of the CNS that target the dynamic properties of brain endothelial cells.

#### Acknowledgements and funding

This work was supported in part by grants from the National Institutes of Health (AG15964, AG020569 and AG028367). Dr. Grammas is the recipient of the Shirley and Mildred Garrison Chair in Aging. Dr. Miller is the recipient of the CH Foundation Chair. The authors gratefully acknowledge the assistance of Terri Stahl and Lauren Wyshywaniuk and appreciate the helpful suggestions of the peer reviewers.

#### References

- 1 Florey, Lord. (1966) The endothelial cell. British Medical Journal 2, 487-490
- 2 Gimbrone, M.A., Cotran, R.S. and Folkman, J. (1974) Human vascular endothelial cells in culture. Growth and DNA synthesis. Journal of Cell Biology 60, 673-684
- 3 Nachman, R.L. and Jaffe, E.A. (2004) Endothelial cell culture: beginnings of modern vascular biology. Journal of Clinical Investigation 114, 1037-1040
- 4 Aird, W.C. (2003) Endothelial cell heterogeneity. Critical Care Medicine 31, 5221-5230
- 5 Chi, J.T. et al. (2003) Endothelial cell diversity revealed by global expression profiling.
   Proceedings of the National Academy of Sciences of the United States of America 100, 10623-10628
- 6 Zlokovic, B.V. (2008) The blood–brain barrier in health and chronic neurodegenerative disorders. Neuron 57, 178-201
- 7 Guo, S. and Lo, E.H. (2009) Dysfunctional cell–cell signaling in the neurovascular unit as a paradigm for central nervous system disease. Stroke 40, S4-S7
- 8 Pardridge, W.M. (1983) Brain metabolism: a perspective from the blood–brain barrier. Physiological Reviews 63, 1481-1535
- 9 Ribatti, D. et al. (2006) Development of the blood–brain barrier: a historical point of view. The Anatomical Record: Neuroanatomist 289B, 3-8
- 10 Reese, T.S. and Karnovsky, M.J. (1967) Fine structural localization of a blood–brain barrier to exogenous peroxidase. Journal of Cell Biology 34, 207-217
- 11 Hardebo, J.E. et al. (1979) Studies on the enzymatic blood-brain barrier: qualitative measurements of DOPA decarboxylase in the wall of microvessels as related to the parenchyma in various CNS regions. Acta Physiologica Scandinavica 105, 453-460
- 12 Kalaria, R.N. and Harik, S.I. (1987) Blood-brain barrier monoamine oxidase: enzyme characterization in cerebral microvessels

### expert reviews

molecular medicine

and other tissues from six mammalian species, including human. Journal of Neurochemistry 49, 856-864

- 13 Gerhart, D.Z. (1997) Expression of monocarboxylate transporter MCT1 by brain endothelium and glia in adult and suckling rats. American Journal of Physiology (London) 273, E207-E213
- 14 Smith, Q.R. and Stoll, J. (1998) Blood-brain barrier amino acid transport. In Introduction to the Blood–Brain Barrier (Pardridge W.M., ed), pp. 188-197, Cambridge University Press, Cambridge, UK
- 15 Tsuji, A. (2005) Small molecular drug transfer across the blood–brain barrier via carrier-mediated transport system. NeuroRx®: The Journal of the American Society for Experimental Therapeutics 2, 54-62
- 16 Ohtsuki, S. and Terasaki, T. (2007) Contribution of carrier-mediated transport systems to the bloodbrain barrier as a supporting and protecting interface for the brain; importance for CNS drug discovery and development. Pharmacology Research 24, 1745-1758
- 17 Spector, R. and Johanson, C.E. (2007) Vitamin transport and homeostasis in mammalian brain: focus on Vitamins B and E. Journal of Neurochemistry 103, 425-438
- 18 Attwell, D. et al. (2010) Glial and neuronal control of brain blood flow. Nature 468, 232-243
- 19 Park, J.A. et al. (2003) Coordinated interaction of the vascular and nervous systems: from molecule- to cell-based approaches. Biochemical and Biophysical Research Communications 311, 247-253
- 20 Lok, J. et al. (2007) Cell-cell signaling in the neurovascular unit. Neurochemical Research 32, 2032-2045
- 21 Lai, C.H., Kuo, K.H. (2005) The critical component to establish in vitro BBB model: pericyte. Brain Research Reviews 50, 258-265
- 22 Krueger, M. and Bechmann, I. (2010) CNS pericytes: concepts, misconceptions, and a way out. Glia 58, 1-10
- 23 Iadecola, C. (2004) Neurovascular regulation in the normal brain and in Alzheimer's disease. Nature Reviews Neuroscience 5, 347-360
- 24 Benarroch, E. (2007) Neurovascular unit dysfunction: a vascular component of Alzheimer's disease? Neurology 68, 1730-1732
- 25 Zlokovic, B.V. (2005) Neurovascular mechanisms of Alzheimer's neurodegeneration. Trends in Neuroscience 28, 202-208

- 26 Zacchigna, S., Lambrechts, D. and Carmeliet, P. (2008) Neurovascular signaling defects in neurodegeneration. Nature Reviews Neuroscience 9, 169-181
- 27 Schnitzer, J.E. (1998) Vascular targeting as a strategy for cancer. New England Journal of Medicine 339, 472-474
- 28 Versari, D. et al. (2009) Endothelial dysfunction as a target for prevention of cardiovascular disease. Diabetes Care 32, S314-S321
- 29 Tang, E.H. and Vanhoutte, P.M. (2010) Endothelial dysfunction: a strategic target in the treatment of hypertension? Pflugers Archiv: European Journal of Physiology 459, 99501004
- 30 Gimbrone, M.A. et al. (1997) Hemodynamics, endothelial gene expression and atherogenesis.
   Annals of the New York Academy of Science 811, 1-10
- 31 Barcia, C. et al. (2005) Changes in vascularization in substantia nigra pars compacta of monkeys rendered parkinsonian. Journal of Neural Transmission 112, 1237-1248
- 32 Banks, W.A., Ercal, N. and Price, T.O. (2006) The blood–brain barrier in neuroAIDS. Current HIV Research 4, 259-266
- 33 Morel, N. et al. (2008) Generation of procoagulant microparticles in cerebrospinal fluid and peripheral blood after traumatic brain injury. Journal of Trauma 64, 698-704
- 34 Zhong, Z. et al. (2008) ALS-causing SOD1 mutants generate vascular changes prior to motor neuron degeneration. Nature Neuroscience 11, 420-422
- 35 Altman, R. and Rutledge, J.C. (2010) The vascular contribution to Alzheimer's disease. Clinical Science (London) 119, 407-421
- 36 Alexander, J.S. et al. (2011) Multiple sclerosis and cerebral endothelial dysfunction: mechanisms. Pathophysiology 18, 3-12
- 37 Israel, A.K. et al. (2011) Peripheral endothelial dysfunction in patients suffering from acute schizophrenia: a potential marker for cardiovascular morbidity? Schizophrenia Research 128, 44-50
- 38 Chironi, G.N. et al. (2009) Endothelial microparticles in diseases. Cell and Tissue Research 335, 143-151
- 39 Boulanger, C.M. (2010) Microparticles, vascular function and hypertension. Current Opinion in Nephrology and Hypertension 19, 177-180
- 40 Jung, K.H. et al. (2009) Circulating endothelial microparticles as a marker of cerebrovascular disease. Annals of Neurology 66, 191-199

- 41 Minagar, A. et al. (2001) Elevated plasma endothelial microparticles in multiple sclerosis. Neurology 56, 1319-1324
- 42 Jy, W. et al. (2004) Endothelial microparticles (EMP) bind and activate monocytes: elevated EMPmonocyte conjugates in multiple sclerosis. Frontiers in Bioscience 9, 3137-3144
- 43 Jimenez, J. et al. (2005) Elevated endothelial microparticle-monocyte complexes induced by multiple sclerosis plasma and the inhibitory effects of interferon-beta 1b on release of endothelial microparticles, formation and transendothelial migration of monocyte-endothelial microparticle complexes. Multiple Sclerosis 11, 310-315
- 44 Shcherbakova, I.V. et al. (2005) Markers of endothelial dysfunction in attack-like schizophrenia. Zhurnal Nevrologii I Psikhiatri Imeni S. S. Korsakova 105, 43-46
- 45 Ukkonen, M. et al. (2007) Cell surface adhesion molecules and cytokine profiles in primary progressive multiple sclerosis. Multiple Sclerosis 13, 701–701
- 46 Ewers, M., Mielke, M.M. and Hampel, H. (2010) Blood-based biomarkers of microvascular pathology in Alzheimer's disease. Experimental Gerontology 45, 75-79
- 47 Dzau, V.J. et al. (2005) Therapeutic potential of endothelial progenitor cells in cardiovascular diseases. Hypertension 46, 7-18
- 48 Van Craenenbroeck, E.M. and Conraads, V.M. (2010) Endothelial progenitor cells in vascular health: focus on lifestyle. Microvascular Research 79, 184-192
- 49 Asahara, T. et al. (1997) Isolation of putative progenitor endothelial cells for angiogenesis. Science 275, 964-967
- 50 Möbius-Winkler, S. et al. (2009) Endothelial progenitor cells: implications for cardiovascular disease. Cytometry 75A, 25-37
- 51 Umemura, T. and Higashi, Y. (2008) Endothelial progenitor cells: therapeutic target for cardiovascular diseases. Journal of Pharmacological Science 108, 1-6
- 52 Lee, S.T. et al. (2009) Reduced circulating angiogenic cells in Alzheimer disease. Neurology 72, 1853-1863
- 53 Wu, Z. et al. (2005) Role of MEOX2 homeobox gene in neurovascular dysfunction in Alzheimer's disease. Nature Medicine 11, 959-965
- 54 Oldendorf, W.H., Cornford, M.E. and Brown, W.J. (1977) The apparent work capability of the blood-brain barrier: a study of the mitochondrial content of capillary endothelial cells in brain and

other tissues of the rat. Annals of Neurology 1, 409-417

- 55 Förster, C. (2008) Tight junctions and the modulation of barrier function in disease. Histochemistry and Cell Biology 130, 55-70
- 56 Arhart, R.W. (2010) A possible haemodynamic mechanism for amyotrophic lateral sclerosis. Medical Hypotheses 75, 341-346
- 57 Weir, B. (2010) Multiple sclerosis a vascular etiology? Canadian Journal of Neurological Science 37, 745-757
- 58 Bennett, J. et al. (2010) Blood–brain barrier disruption and enhanced vascular permeability in the multiple sclerosis model EAE. Journal of Immunology 229, 180-191
- 59 Plumb, J. et al. (2002) Abnormal endothelial tight junctions in active lesions and normal-appearing white matter in multiple sclerosis. Brain Pathology 12, 154-169
- 60 Kirk, J. et al. (2003) Tight junctional abnormality in multiple sclerosis white matter affects all calibers of vessel and is associated with blood–brain barrier leakage and active demyelination. Journal of Pathology 201, 319-327
- 61 Minagar, A. and Alexander, J.S. (2003) Blood–brain barrier disruption in multiple sclerosis. Multiple Sclerosis 9, 540-549
- 62 Holman, D.W., Klein, R.S. and Ransohoff, R.M. (2011) The blood–brain barrier, chemokines and multiple sclerosis. Biochimcia et Biophysica Acta 1812, 220-230
- 63 Garbuzova-Davis, S. et al. (2007) Evidence of compromised blood-spinal cord barrier in early and late symptomatic SOD1 mice modeling ALS. PLoS ONE 2, e1205
- 64 Engelhardt, J.I. and Appel, S.H. (1990) IgG reactivity in the spinal cord and motor cortex in amyotrophic lateral sclerosis. Archives of Neurology 47, 1210-1216
- 65 Nicaise, C. et al. (2009) Impaired blood-brain and blood-spinal cord barriers in mutant SOD1-linked ALS rat. Brain Research 1301, 152-162
- 66 Ances, B.M. and Clifford, D.B. (2008) HIVassociated neurocognitive disorders and the impact of combination antiretroviral therapies. Current Neurology and Neuroscience Reports 8, 455-461
- 67 Kanmogne, G.D. et al. (2007) HIV-1 gp120 compromises blood-brain barrier integrity and enhances monocyte migration across blood-brain barrier: implication for viral neuropathogenesis. Journal of Cerebral Blood Flow and Metabolism 27, 123-134

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Accession information: doi:10.1017/S1462399411001918; Vol. 13; e19; June 2011 © Cambridge University Press 2011

- 68 Ramirez, S.H. et al. (2010) Dyad of CD40/CD40
  ligand fosters neuroinflammation at the
  blood-brain barrier and is regulated via JNK
  signaling: implications for HIV-1 encephalitis.
  Journal of Neuroscience 30, 9454-9464
- 69 Yang, B. et al. (2008) HIV-1 gp120 induces cytokine expression, leukocyte adhesion, and transmigration across the blood–brain barrier: modulatory effects of STAT1 signaling. Microvascular Research 77, 212-219
- 70 Ricardo-Dukelow, M. et al. (2007) HIV-1 infected monocyte-derived macrophages affect the human brain microvascular endothelial cell proteome: new insights into blood-brain barrier dysfunction for HIV-1-associated dementia. Journal of Neuroimmunology 185, 37-46
- 71 Shen, S. and Zhang, W. (2010) ABC transporters and drug efflux at the blood–brain barrier. Reviews in Neuroscience 21, 29-53
- 72 Ferreira, I.L. et al. (2010) Multiple defects in energy metabolism in Alzheimer's disease. Current Drug Targets 11, 1193-1206
- 73 Chetelat, G. et al. (2003) Mild cognitive impairment: can FDG-PET predict who is to rapidly convert to Alzheimer's disease? Neurology 60, 1374-1377
- 74 Gibson, G.E. and Shi, Q. (2010) A mitocentric view of Alzheimer's disease suggests multi-faceted treatments. Journal of Alzheimer's Disease 20, S591-S607
- 75 Kalaria, R.N. and Harik, S.I. (1989) Reduced glucose transporter at the blood–brain barrier in cerebral cortex in Alzheimer disease. Journal of Neurochemistry 53, 1083
- 76 Pimplikar, S.W. (2009) Reassessing the amyloid cascade hypothesis of Alzheimer's disease.
   International Journal of Biochemistry and Cell Biology 41, 1261-1268
- 77 Deane, R., Bell, R.D., Sagare, A., and Zlokovic, B.V. (2009) Clearance of amyloid-β peptide across the blood–brain barrier: implications for therapies in Alzheimer's disease. CNS and Neurological Disorders Drug Targets 8, 16-30
- 78 Deane, R. et al. (2004) RAGE (yin) versus LRP (yang) balance regulates Alzheimer amyloid betapeptide clearance through transport across the blood–brain barrier. Stroke 35, 2628-2631
- 79 Bell, R.D. et al. (2009) SRF and myocardin regulate LRP-mediated amyloid-β clearance in brain vascular cells. Nature Cell Biology 11, 143-153
- 80 Deane, R. et al. (2003) RAGE mediates amyloid-β peptide transport across the blood–brain barrier and accumulation in brain. Nature Medicine 9, 907-913

- 81 Yan, S.D. et al. (1996) RAGE and amyloid-beta peptide neurotoxicity in Alzheimer's disease. Nature 382, 685-691
- 82 Kumar-Singh, S. et al. (2005) Dense-core plaques in Tg2576 and PSAPP mouse models of Alzheimer's disease are centered on vessel walls. American Journal of Pathology 167, 527-543
- 83 Shibata, M. et al. (2000) Clearance of Alzheimer's amyloid β(1-40) peptide from brain by LDL receptor-related protein-1 at the blood-brain barrier. Journal of Clinical Investigation 106, 1489-1499
- 84 Jaeger, L.B. (2009) Lipopolysaccharide alters the blood–brain barrier transport of amyloid beta protein: a mechanism for inflammation in the progression of Alzheimer's disease. Brain, Behavior and Immunology 23, 507-517
- 85 Qian, L., Flood, P.M. and Hong, J.S. (2010) Neuroinflammation is a key player in Parkinson's disease and a prime target for therapy. Journal of Neural Transmission 117, 971-999
- 86 Hawkins, R.A. et al. (2006) Structure of the blood–brain barrier and its role in the transport of amino acids. Journal of Nutrition 136, 218S-226S
- 87 Carvey, P.M. et al. (2005) 6-Hydroxydopamineinduced alterations in blood–brain barrier permeability. European Journal of Neuroscience 22, 1158-1168
- 88 Kortekaas, R. et al. (2005) Blood–brain barrier dysfunction in parkinsonian midbrain *in vivo*. Annals of Neurology 57, 176-179
- 89 Farkas, E. et al. (2000) Pathological features of cerebral cortical capillaries are double in Alzheimer's disease and Parkinson's disease. Acta Neuropathologica (Berlin) 100, 395-402
- 90 Ohtsuki, S. et al. (2010) Reduction of L-type amino acid transporter 1 mRNA expression in brain capillaries in a mouse model of Parkinson's disease. Biological and Pharmaceutical Bulletin 33, 1250-1252
- 91 Lee, C.G. et al. (2004) *MDR*1, the blood–brain barrier transporter, is associated with Parkinson's disease in ethnic Chinese. Journal of Medical Genetics 41, e60
- 92 Bartels, A.L. et al. (2008) Decreased blood-brain barrier P-glycoprotein function in the progression of Parkinson's disease, SP and MSA. Journal of Neural Transmission 115, 1001-1009
- 93 Hartz, A.M.S. et al. (2003) Rapid modulation of Pglycoprotein-mediated transport at the blood–brain barrier by tumor necrosis factor- $\alpha$  and lipopolysaccharide. Molecular Pharmacology 69, 462-470

erebra

- 94 Funke, C. et al. (2009) Genetic analysis of coding SNPs in blood-brain barrier transporter MDR1 in European Parkinson's disease patients. Journal of Neural Transmission 116, 443-450
- 95 Westerlund, M. et al. (2009) Association of polymorphism in the ABCB1 gene with Parkinson's disease. Parkinsonism and Related Disorders 15, 422-424
- 96 Grammas, P. (2011) Neurovascular dysfunction, inflammation and endothelial activation: implications for the pathogenesis of Alzheimer's disease. Journal of Neuroinflammation 25, 8-26
- 97 Harris, L.W. et al. (2008) The cerebral microvasculature in schizophrenia: a laser capture microdissection study. PloS ONE 3, e3964
- 98 Grammas, P., Moore, P. and Weigel, P.H. (1999) Microvessels from Alzheimer's disease brain kill neurons in vitro. American Journal of Pathology 154, 337-342
- 99 Grammas, P. (2000) A damaged microcirculation contributes to neuronal cell death in Alzheimer's disease. Neurobiology of Aging 21, 199-205
- 100 Yin, X. et al. (2010) Brain endothelial cells synthesize neurotoxic thrombin in Alzheimer's disease. American Journal of Pathology 176, 1600-1606
- 101 Huang, C. et al. (2008) JAK2-STAT3 signaling pathway mediates thrombin-induced proinflammatory actions of microglia in vitro. Journal of Neuroimmunology 204, 118-125
- 102 Choi, M.S. et al. (2008) Activation of proteaseactivated receptor1 mediates induction of matrix metalloproteinase-9 by thrombin in rat primary astrocytes. Brain Research Bulletin 76, 368-375
- 103 Rosenberg, G.A. (2009) Matrix metalloproteinases and their multiple roles in neurodegenerative diseases. Lancet Neurology 8, 205-216
- 104 Grammas, P. and Ovase, R. (2001) Inflammatory factors are elevated in brain microvessels in Alzheimer's disease. Neurobiology of Aging 22, 837-842
- 105 Grammas, P., Samany, P.G. and Thirumangalakudi, L. (2006) Thrombin and inflammatory proteins are elevated in Alzheimer's disease microvessels: implications for disease pathogenesis. Journal of Alzheimer's Disease 9, 51-58
- 106 Bugno, M. et al. (1999) Reprogramming of TIMP-1 and TIMP-3 expression profiles in brain microvascular endothelial cells and astrocytes in response to proinflammatory cytokines. FEBS Journal 448, 9-14
- 107 Vilalta, A. et al. (2010) Matrix metalloproteinases in neurological brain lesions: a new therapeutic target? Reviews in Neurology 51, 95-107

- 108 Zhang, H. et al. (2010) Matrix metalloproteinases and neurotrauma: evolving roles in injury and reparative processes. The Neuroscientist 16, 156-170
- 109 Sternlicht, M.D. and Werb, Z. (2001) How matrix metalloproteinases regulate cell behavior. Annual Review of Cellular and Developmental Biology 17, 463-516
- 110 Visse, R. and Nagase, H. (2003) Matrix metalloproteinases and tissue inhibitors of metalloproteinases. Circulation Research 92, 827-839
- 111 del Zoppo, G.J. (2010) The neurovascular unit, matrix proteases, and innate inflammation. Annals of the New York Academy of Sciences 1207, 46-49
- 112 Ries, C. and Petrides, P.E. (1995) Cytokine regulation of matrix metalloproteinase activity and its regulatory dysfunction in disease. Biology Chemistry Hoppe-Seyler 376, 345-355
- 113 Sokolova, E. and Reiser, G. (2008) Prothrombin/ thrombin and the thrombin receptors PAR-1 and PAR-4 in the brain: localization, expression and participation in neurodegenerative diseases. Thrombosis and Haemostasis 100, 576-581
- 114 Maragoudakis, M.E., Tsopanoglou, N.E. and Andriopoulou, P. (2002) Mechanism of thrombininduced angiogenesis. Biochemical Society Transactions 20, 173-177
- 115 Cunningham, L.A., Wetzel, M. and Rosenberg, G.A. (2005) Multiple roles for MMPs and TIMPs in cerebral ischemia. Glia 50, 329-339
- 116 Amantea, D. et al. (2009) Post-ischemic brain damage: pathophysiology and role of inflammatory mediators. FEBS Journal 276, 13-26
- 117 Schönbeck, U., Mach, F. and Libby, P. (1998) Generation of biologically active IL-1 beta by matrix metalloproteinases: a novel caspase-1 independent pathway of IL-1 beta processing. Journal of Immunology 161, 3340-3346
- 118 Woessner, J.F. and Nagase, H. (2000) Matrix Metalloproteinases and TIMPs, Oxford University Press, New York
- 119 Sheu, B.C. et al. (2001) A novel role of metalloproteinase in cancer-mediated immunosuppression. Cancer Research 61, 237-242
- 120 Rosenberg, G.A. (2002) Matrix metalloproteinases and neuroinflammation in multiple sclerosis. The Neuroscientist 8, 586-595
- 121 Persidsky, Y. et al. (2006) Rho-mediated regulation of tight junctions during monocyte migration across the blood-brain barrier in HIV-1 encephalitis (HIVE). Blood 107, 4770-4780

- 122 Huang, W. et al. (2009) PPARalpha and PPARgamma attenuate HIV-induced dysregulation of tight junction proteins by modulation of matrix metalloproteinase and protease activities. FASEB Journal 23, 1596-1606
- 123 Dhawan, S. et al. (1995) HIV-1 infection alters monocyte interactions with human microvascular endothelial cells. Journal of Immunology 154, 422-432
- 124 Thirumangalakudi, L. et al. (2006) Angiogenic proteins are expressed by brain blood vessels in Alzheimer's disease. Journal of Alzheimer's Disease 10, 111-118
- 125 Miyakawa, T. et al. (1982) The relationship between senile plaques and cerebral blood vessels in Alzheimer's disease and senile dementia.
  Morphological mechanism of senile plaque production. Virchows Archives (Cellular Pathology) 40, 121-129
- 126 Lee, E.J. et al. (2010) Alpha-synuclein activates microglia by inducing the expression of matrix metalloproteinases and the subsequent activation of protease-activated receptor-1. Journal of Immunology 185, 615-623
- 127 Tamo, W. et al. (2002) Expression of alphasynuclein, the precursor of non-amyloid beta component of Alzheimer's disease amyloid, in human cerebral blood vessels. Neuroscience Letters 326, 5-8
- 128 Kim, S.T. et al. (2010) Matrix metalloproteinase-3 contributes to vulnerability of the nigral dopaminergic neurons. Neurochemistry International 56, 161-167
- 129 Mizoguchi, H. et al. (2009) Matrix metalloproteinase-9 inhibition improves amyloidβ-mediated cognitive impairment and neurotoxicity in mice. Journal of Pharmacology and Experimental Therapeutics 331, 14-22
- 130 Levin, J. et al. (2009) Increased alpha-synuclein aggregation following limited cleavage by certain matrix metalloproteinases. Experimental Neurology 214, 201-208
- 131 Pimenta de Castro, I., Martins, L.M. and Tufi, R.(2010) Mitochondrial quality control and neurological disease: an emerging connection. Expert Reviews in Molecular Medicine 12, e12
- 132 Reeve, A.K., Krishnan, K.J. and Turnbull, D. (2008) Age related mitochondrial degenerative disorders in humans. Biotechnology Journal 3, 750-756
- 133 Müller, W.E. et al. (2010) Mitochondrial dysfunction: Common final pathway in brain aging and Alzheimer's disease-Therapeutic aspects. Molecular Neurobiology 41, 159-171

- 134 Sakar, D. and Fisher, P.D. (2006) Molecular mechanisms of aging-associated inflammation. Cancer Letters 236, 3-23
- 135 Tripathy, D. et al. (2010) Cerebrovascular expression of proteins related to inflammation, oxidative stress and neurotoxicity is altered with aging. Journal of Neuroinflammation 7, 63
- 136 Victor, V.M. and Rocha, M. (2007) Targeting antioxidants to mitochondria: potential new therapeutic strategy for cardiovascular diseases. Current Pharmaceutical Design 13, 845-863
- 137 Puddu, P. et al. (2009) The emerging role cardiovascular risk factor-induced mitochondrial dysfunction in atherogenesis. Journal of Biomedical Science 16, 112-121
- 138 Davidson, S.M. and Duchen, M.R. (2007) Endothelial mitochondria: contributing to vascular function and disease. Circulation Research 100, 1128-1141
- 139 Kolev, K. et al. (2003) Matrix metalloproteinase-9 expression in post-hypoxic human brain capillary endothelial cells: H<sub>2</sub>O<sub>2</sub> as a trigger and NF-kappaB as a signal transducer. Thrombosis and Haemostasis 90, 528-537
- 140 Pu, P.B., Lu, J. and Moochhala, S. (2009) Involvement of ROS in BBB dysfunction. Free Radical Research 43, 348-364
- 141 Hamdheydari, L. et al. (2003) Oxidized LDLs affect nitric oxide and radical generation in brain endothelial cells. Biochemical Biophysical Research Communications 311, 486-490
- 142 Chang, H.C. et al. (2010) Resveratrol attenuates oxidized LDL-evoked Lox-1 signaling and consequently protects against apoptotic insults to cerebrovascular endothelial cells. Journal of Cerebral Blood Flow and Metabolism 3, 842–54
- 143 Closhen, D. et al. (2010) CRP-induced levels of oxidative stress are higher in brain than aortic endothelial cells. Cytokine 50, 117-120
- 144 Martin, L.J. (2010) Mitochondrial pathobiology in Parkinson's disease and amyotrophic lateral sclerosis. Journal of Alzheimer's disease 20, S335-S356
- 145 Fernandez-Checa, J.C. et al. (2010) Oxidative stress and altered mitochondrial function in neurodegenerative diseases: lessons from mouse models. CNS Neurological Disorders and Drug Targets 9, 439-454
- 146 Lopez, G. and Sidransky, E. (2010) Autosomal recessive mutations in the development of Parkinson's disease. Biomarkers in Medicine 4, 713-721

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Accession information: doi:10.1017/S1462399411001918; Vol. 13; e19; June 2011 © Cambridge University Press 2011

- 147 Casetta, I., Govoni, V. and Granieri, E. (2005) Oxidative stress, antioxidants and neurodegenerative diseases. Current Pharmacologic Design 11, 2033-2052
- 148 Tanner, C.M. et al. (2011) Rotenone, paraquat and Parkinson's disease. Environmental Health Perspectives January 26 [Epub ahead of print]
- 149 dos Santos, A.P. et al. (2010) Rat brain endothelial cells are a target of manganese toxicity. Brain Research 1326, 152-161
- 150 Aliev, G. et al. (2003) Role of vascular hypoperfusion-induced oxidative stress and mitochondria failure in the pathogenesis of Alzheimer disease. Neurotoxicity Research 5, 491-504
- 151 Claudio, L. (1996) Ultrastructural features of the blood–brain barrier in biopsy tissue from Alzheimer's disease patients. Acta Neuropathologica 91, 6-14
- 152 Xu, J. et al. (2001) Amyloid beta peptide-induced cerebral endothelial cell death involves mitochondrial dysfunction and caspase activation. Journal of Cerebral Blood Flow and Metabolism 21, 702-710
- 153 Wong, P.C. et al. (1995) An adverse property of a familial ALS-SOD1 mutation causes motor neuron disease characterized by vacuolar degeneration of mitochondria. Neuron 14, 1105-1116
- 154 Kong, J. and Xu, Z. (1998) Massive mitochondrial degeneration in motor neurons triggers the onset of

amyotrophic lateral sclerosis in mice expressing a mutant SOD1. Journal of Neuroscience 18, 32410–3250

- 155 Pedrini, S. et al. (2010) ALS-linked mutant SOD1 damages mitochondria by promoting conformational changes in Bcl-2. Human Molecular Genetics 19, 2974-2986
- 156 Pardridge, W.M. (2005) The blood-brain barrier and neurotherapeutics. NeuroRx®: The Journal of the American Society for Experimental Therapeutics 2, 1-2
- 157 Agarwal, S. et al. (2011) Delivery of molecularly targeted therapy to malignant glioma, a disease of the whole brain. Expert Reviews in Molecular Medicine 13, e17
- 158 Lorenzl, S. et al. (2003) Tissue inhibitors of matrix metalloproteinases in neuromuscular disease. Journal of the Neurological Sciences 207, 71-76
- 159 Renaud, S. and Leppert, D. (2007) Matrix metalloproteinases in neuromuscular disease. Muscle and Nerve 36, 1-13
- 160 Horstmann, S. et al. (2010) Matrix metalloproteinases in peripheral blood in cerebrospinal fluid in patients with Alzheimer's disease. International Psychogeriatrics 22, 966-972
- 161 Sastre-Garriga, J. et al. (2004) Decreased MMP-9 production in primary progressive multiple sclerosis patients. Multiple Sclerosis 10, 376-380

#### Further reading, resources and contacts

- Neuwelt, E. et al. (2008) Strategies to advance translational research into brain barriers. Lancet Neurology 7, 84–96
- This article summarises findings and recommendations from a meeting convened to discuss the role of blood-brain and blood-CSF barriers in inflammation, injury, tumours and neurodegenerative diseases.

Zlokovic, B. (2010) Neurodegeneration and the neurovascular unit. Nature Medicine 16, 1370-1371

This letter to the Editor presents a succinct discussion of the key points underlying the 'vascular hypothesis' of neurodegeneration.

Grammas, P. et al. (2008) Neurodegeneration and the brain barriers. www.ibbsoc.org/PDFs/Neurodegeneration.pdf

- This online article focuses on the state of brain barriers research from the basic science perspective and summarises recommendations for advancing the field.
- The website of International Brain Barriers Society (IBBS) has useful information and meeting links for those interested in biological barriers in the CNS:

www.ibbsoc.org.

Accession information: doi:10.1017/S1462399411001918; Vol. 13; e19; June 2011 © Cambridge University Press 2011

http://www.expertreviews.org/

#### Features associated with this article

#### Figures

Figure 1. The neurovascular unit.

Figure 2. Junctional proteins create a barrier between adjacent brain endothelial cells.

Figure 3. Carrier-mediated transporters of the blood-brain barrier.

Figure 4. Proposed blood-brain barrier mitochondrial dysfunction in CNS diseases.

Table

Table 1. Matrix metalloproteinases and tissue inhibitors of metalloproteinases involved in neurodegenerative disorders.

#### Citation details for this article

Paula Grammas, Joseph Martinez and Bradley Miller (2011) Cerebral microvascular endothelium and the pathogenesis of neurodegenerative diseases. Expert Rev. Mol. Med. Vol. 13, e19, June 2011, doi:10.1017/S1462399411001918

neurodegenerative diseases Cerebral microvascular endothelium and the pathogenesis of