

Density-dependent regulation of fecundity in *Syngamus trachea* infrapopulations in semi-naturally occurring ring-necked pheasants (*Phasianus colchicus*) and wild Carrion Crows (*Corvus corone*)

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SUMMARY

Previous work has highlighted increased opportunities for the transmission of *Syngamus trachea* within pheasant release pens, due in part to high levels of environmental contamination around communal areas. Despite this, the distribution of adult worms within their definitive hosts is not significantly different from predicted distributions under Taylor's power law. Therefore, density-dependent processes are probably acting to regulate *S. trachea* population dynamics. Patterns of nematode fecundity were investigated in a semi-naturally occurring population of ring-necked pheasants (*Phasianus colchicus*) and a wild population of carrion crows (*Corvus corone*). Worm length was a reliable indicator of nematode fecundity, and a negative association between mean worm length and mean worm burden was identified within both the species. The stunting of worms at greater parasite densities was present in both immunologically naïve and previously exposed pheasants, so is unlikely to be a function of age-dependent acquired immunity. Interestingly, the effect of parasite crowding in the crow population explained more of the variation in mean worm length, apparently driven by a greater mean worm burden when compared with pheasants. The findings of the present study suggest that fecundity is a function of parasite density, i.e. parasite-mediated competition and not host-mediated heterogeneities in immunocompetence.

Key words: *Syngamus trachea*, density dependence, pheasant, crow, fecundity, worm length, immunity.

INTRODUCTION

One recurring theme within parasite ecology is the relative stability of parasite populations in domestic and wild animal hosts (Anderson and May, 1978; Tompkins and Hudson, 1999), which suggests that some form of regulatory mechanism must be ensuring population stability. The majority of these mechanisms are driven by parasite density, i.e. are a function of parasite burden within individual hosts; thus acting on infra-populations as opposed to populations as a whole. Indeed, density-dependent regulatory mechanisms act on many aspects of the parasite lifecycle, such as parasite establishment, growth, fecundity, development and maturation times, and adult survival (Walker *et al.* 2009). Growth and fecundity for instance, being the two most common aspects of the life cycle regulated by such mechanisms in helminth populations (Tompkins and Hudson, 1999), are particularly important at regulating the abundance of the 'free-living', infectious stages within the environment, and therefore determining the extent of future infections. These density-dependent mechanisms are

important for regulating and stabilizing transmission dynamics, and therefore the parasite–host relationship.

Despite knowledge of the existence of such regulation, the mechanisms underlying density dependence are poorly understood, as it is difficult to disentangle host and parasite responses to increasing parasite challenge (Paterson and Viney, 2002). Host immune responses have been demonstrated to reduce establishment, survivability and fecundity of parasitic nematodes, and it is hypothesized that innate and adaptive immune responses, whose response to infection increases with increasing parasite density, are responsible for the manifestation of density dependence (Paterson and Viney, 2002). Similarly, intraspecific competition for space and resources once inside the host has also been implicated as a driver of density-dependent regulation. Indeed, Michael and Dunby (1989), hypothesized that parasite-mediated competition was responsible for *Trichuris muris* establishment in the mouse, owing to the finite carrying-capacity of the caecum.

Syngamus trachea is a parasitic nematode occurring in a wide range of avian hosts (Lewis, 1928; Morgan and Clapham, 1934; Goble and Kutz, 1945). The non-specific nature of this parasite makes it possible to study differences in host-mediated responses to a natural challenge of *S. trachea*. In a previous

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paper by Gethings *et al.* (2015), we highlighted the fact that despite increased opportunities for transmission of *S. trachea* within pheasant release pens, relatively low numbers of adult worms are consistently recovered upon post-mortem investigation. This is often the case in experimental infections using large numbers of infective larvae (Olivier, 1944; Guilford and Herrick, 1954). Despite these previous studies finding relationships between *S. trachea* establishment and host immunity, no such work has been conducted in semi-naturally occurring pheasant populations using natural infections of *S. trachea*. The aims of the present study were to determine firstly, whether worm length is a good indicator of fecundity within *S. trachea* populations, and secondly, to determine whether fecundity is impaired in response to increasing worm burden in two host species.

MATERIALS AND METHOD

Study sites

The two study sites were selected due to their large size and annual occurrence of clinical Syngamosis. Site 1 was located approximately at grid reference ST 97502 39837 and consisted of seven release pens. Site 2 was situated approximately at grid reference SU 17769 30326 and similarly consisted of seven release pens. Site 2 undertook rigorous corvid control as part of a game management programme with the use of Larsen traps, whereas site 1 used traps sporadically.

Carcass recovery

Male and female ring-necked pheasants (*Phasianus colchicus*) were recovered from two pheasant estates in the South West of England from January to November 2015. All birds were either obtained during the shooting season or were found dead on the estates at various times of the year. Crows (*Corvus carone*) were opportunistically sampled throughout the season, as the sites were undertaking Corvid control via the use of Larsen traps. Crows and Rooks are known to be commonly infected with *S. trachea*, and any density-dependent effects would likely be more apparent as worm burdens tend to be larger than in pheasants. Crows were primarily recovered from site 2, as their corvid control programme was more consistent. Age was roughly estimated by the presence/absence and size of the bursa of fabricius, which has usually atrophied by 6 months (Williams and Newton, 1969); however, no formal analysis of the effects of age on either parasite burdens or length was undertaken during this study.

Adult worm recovery

Adult *S. trachea* worms were recovered from the trachea of pheasants and crows. Adult worms are sexually dioecious, and show marked sexual

dimorphism from 7 to 9 days post infection (dpi) with female worms being considerably larger than males; female length = ≥ 13 mm, male length = > 4 mm at 14 dpi (Fernando *et al.* 1971). The trachea was first resected from the underlying connective tissue and transected slightly above the proximal bifurcation of the bronchi. The trachea was then incised longitudinally through the tracheal cartilage and the worms recovered using fine-tipped forceps. Adult worms were distinguished from juvenile (L4) worms by observation under a microscope at varying magnifications in order to detect the presence of fertilized ova. Adult worms from both species were assessed according to Lewis (1928), which confirmed that these worms were indeed *S. trachea* and that the worms were identical between species justifying the between-species comparisons.

Worm length and fecundity

Fernando *et al.* (1971) conducted in-depth pathogenetic examinations detailing adult worm length at various stages of development, and determined the number of dpi to the production of fertilized ova. Female *S. trachea* worms are fertile by 14 dpi, with minimum female length at the adult stage generally averaging 10–15 mm. Once fertile, Guilford and Herrick (1954) found there is no relationship between dpi and female worm length, so we concluded that the number of dpi was not a significant confounding factor within this study. As several authors have demonstrated that worm length is significantly correlated with worm fecundity (Michael and Dunby, 1989; Stear *et al.* 1997; Stear and Bishop, 1999; Tompkins and Hudson, 1999; Walker *et al.* 2009), the same principle was applied in this study. A total of 157 female worms were recovered using systematic sampling from ten randomly selected pheasants and crows in order to estimate the effect of length on the number of eggs per worm. Although adult worms were recovered from 130 birds, *in utero* eggs were counted in female worms recovered from every seventh ($130/20 = 6.5$ rounded to 7) bird, providing it was infected. Female worm was measured using a digital calliper (accuracy to 0.01 mm) and the number of eggs was counted using a stereomicroscope. Genital tubes were liberated from female worms and each egg was counted *in situ*. To ensure egg viability, eggs were recovered from each worm and maintained in the laboratory at 24 °C (Wehr, 1937). Eggs were cultured to the infective stage (L3) and manually hatched by applying light pressure between two cover slips. Male worms were not measured during this study (Fig. 1).

Condition of the trachea

It has been demonstrated that prolonged infections with *S. trachea* result in the formation of hyperplastic tracheal cartilage in which the adult male worms are

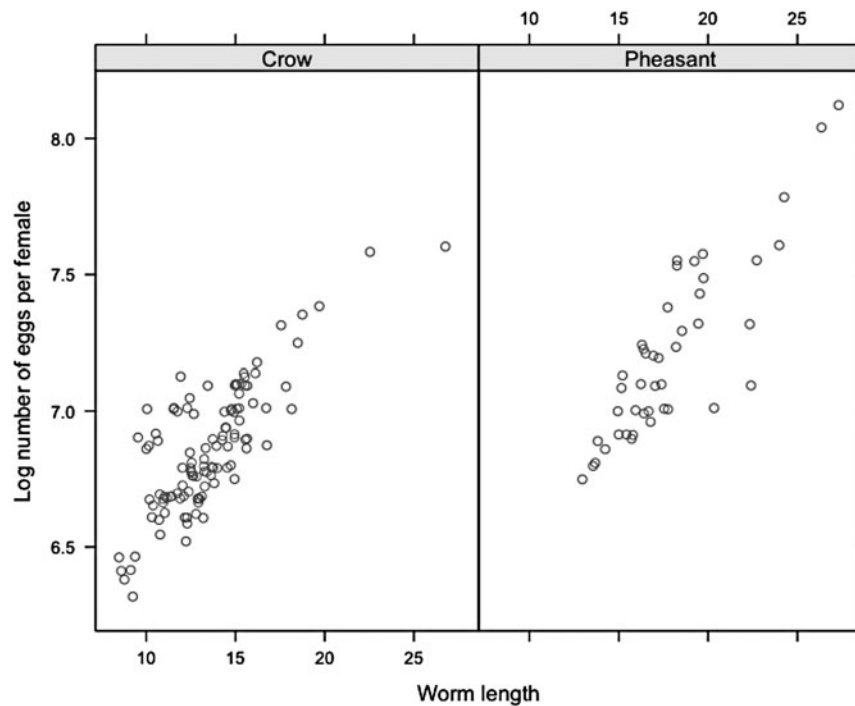


Fig. 1. Relationship between worm length and the number of eggs per worm (fecundity) for both species.

deeply embedded (Clapham, 1935). These nodules begin to form between 26 and 37 dpi and generally remain indefinitely; meaning previous exposure and duration of current infection can be determined. These nodules form at the point of attachment, so if no worms are found within these nodules, it can be concluded that the bird has been exposed to *S. trachea* previously, thus conferring some degree of immunity. To assess whether previous exposure influenced mean worm length or mean worm burden in subsequent infections, pheasant tracheas were examined for the presence of nodules. These nodules do not form at the point of attachment in corvids, so previous exposure cannot be determined. Therefore, crows were excluded from this part of analysis.

Retrospective data analysis. Guilford and Herrick (1954) experimentally infected pheasants with *S. trachea* larvae and counted the number of worms upon post-mortem examination, measured their length and noted the presence/absence of nodules. These data ($n = 32$) were combined with data recovered in this study ($n = 21$) to evaluate whether physiological evidence of previous exposure influenced parasite intensity and/or parasite length.

Statistical analysis

All data were analysed using the R statistical package for Macintosh. Differences in the mean number of worms and mean worm length between species were assessed using Welch's *t*-test for unequal samples. Unless stated otherwise, all regression analyses were conducted using generalized linear models with

negative binomial error distributions and log link functions (*glm.nb* available in the MASS package). Data were assessed for negative-binomial model fit by comparing with models with Poisson error distributions by maximum likelihood using the *pchisq* function in R. Significance levels were calculated using χ^2 tests from the deviance explained by each factor and pseudo R^2 values were calculated for each model ($1 - \text{residual deviance}/\text{null deviance}$). The effect of parasite intensity on mean parasite length was analysed using log-transformed mean counts for parasite intensity in the 37 pheasants and 92 crows sampled (Tompkins and Hudson, 1999; Ives, 2015). Non-constant error variance was assessed using the Breusch–Pagan test (B–P test) and 'length' data were transformed to the appropriate power transformation ($y^{0.02}$). The B–P test was then conducted on the transformed data, which confirmed constant variance [$\chi^2 = 0.30$, D.F. = 1, $P = 0.57$]. In order to ensure that worm length was a reliable indicator of fecundity, the effect of parasite length on the number of eggs per adult female was assessed using raw count data of 157 adult female worms recovered from ten pheasants and ten crows. In order to determine the minimum parasite density at which negative effects are observable, iterative backwards-stepwise deletion of the highest parasite densities was conducted until the regression was no longer significant at the $P \leq 0.05$ level.

RESULTS

The trachea of 37 pheasants and 92 crows were recovered and examined for the presence of adult *S. trachea* worms, of which 1314 pairs were recovered.

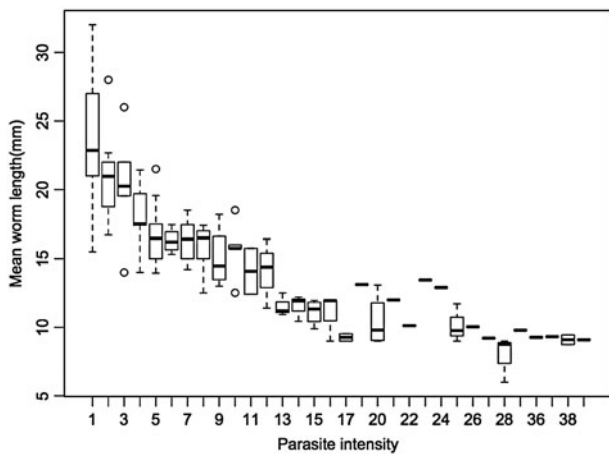


Fig. 2. Mean worm length (mm) for varying parasite intensities ($n = 1314$ worms across 129 hosts).

Parasite fecundity

Model comparisons suggested the negative binomial model best fit the data ($\chi^2 = 3281.27$, D.F. = 1, $P \leq 0.001$) so data were analysed with negative binomial error distributions. *In utero* egg counts were performed on 157 pooled adult female worms recovered from ten crows (113 worms) and ten pheasants (44 worms), with an average (\pm S.E.M.) of 1066 (± 41.5) eggs per worm. Worm length (coef = 0.06, deviance = 590.47, D.F. = 1, $P \leq 0.001$), after accounting for within-host variation in worm burden (coef = -0.05, deviance = 7.77, D.F. = 1, $P = 0.006$), was significantly correlated with the number of eggs per female worm; explaining 79% of the deviance (residual deviance = 157.05, D.F. = 154, $P \leq 0.001$).

Worm length and parasite intensity

A total of 286 and 1028 adult *S. trachea* pairs were recovered from the trachea of 37 pheasants and 92 crows respectively. Mean worm length was significantly correlated with mean parasite density for pheasants and crows, with a significant reduction in mean worm length at higher parasite densities ($F_{1,127} = 393.3$, $R^2 = 0.759$, $P \leq 0.001$) (Fig. 2). For individual species, there was a stronger effect of mean worm burden on mean worm length for crows ($F_{1,90} = 340.2$, $R^2 = 0.79$, $P \leq 0.001$) than for pheasants ($F_{1,35} = 64.21$, $R^2 = 0.64$, $P \leq 0.001$). Stepwise data-point deletion of the highest parasite densities revealed that density-dependent effects begin to manifest above four worms per bird for pheasants, and two worms per bird for crows, with the regression model not reaching the significance level of $P < 0.05$ below these thresholds.

Presence of nodules, mean worm length and number of adult worms

Crows were excluded from this part of analysis, so the results are not reported. Retrospective analysis

of the Guilford and Herrick (1954) data, and trachea condition in the present study revealed that pheasants with hyperplastic tracheal nodules had fewer adult worms present in the trachea (mean \pm S.E.M. = 5.26 ± 1.01 worms per bird) than birds without nodules (mean \pm S.E.M. = 11.12 ± 1.68 worms per bird) ($n = 51$, $t_{46} = 2.97$, $P = 0.004$). Female worms in birds with the evidence of previous exposure were also longer (mean \pm S.E.M. = $16.5 \text{ mm} \pm 1.51$) when compared with worms in birds that had no evidence of previous exposure (mean \pm S.E.M. = $13.01 \text{ mm} \pm 0.53$) ($n = 51$, $t_{21.3} = -2.10$, $P = 0.04$) (Fig. 3).

Mean worm length and mean worm burden between species

The mean number of adult worms per trachea differed significantly between species, with crows having a mean worm burden of 11.17 (\pm S.E.M. = 0.10) and pheasants having an average of 7.72 (\pm S.E.M. = 1.39) worms per trachea ($t^{72.14} = 2.02$, $P = 0.04$). Similarly, mean worm length differed significantly between species, with pheasants having a mean worm length of 17.97 mm (\pm S.E.M. = 0.85) and crows having a mean worm length of 15.55 mm (\pm S.E.M. = 0.55) ($t^{66.58} = 2.34$, $P = 0.02$).

DISCUSSION

Density-dependent reductions in worm size and fecundity have been reported in a large number of studies (Michael and Dunby, 1989; Stear *et al.* 1997; Stear and Bishop, 1999; Tompkins and Hudson, 1999; Walker *et al.* 2009), and density-dependent reductions in worm length, but not necessarily fecundity, are reported in the vast majority of nematode species studied (Mossinger and Wenk, 1986; Szalai and Dick, 1989; Sinniah and Subramaniam, 1991; Skorping *et al.* 1991; Marcogliese, 1997; Irvine *et al.* 2001; Richards and Lewis, 2001; Dezfuli *et al.* 2002). A vast majority of studies concerning density dependence have been laboratory-based experimental infections, which do not accurately represent conditions facing free-living wild animal populations in terms of parasite load and encounter rates. The present study provides reliable information concerning apparent density-dependent regulation of fecundity in both an intensively-managed pheasant population, and a free-living wild population of corvids. Although the fact that immune status is responsible for regulating the establishment of *S. trachea* in ring-necked pheasants is not novel, this is first mention of both parasite and host-mediated factors regulating *S. trachea* populations in any bird species.

In agreement with previous studies (Olivier, 1944; Guilford and Herrick, 1954), immune function appears to be predominantly responsible for the

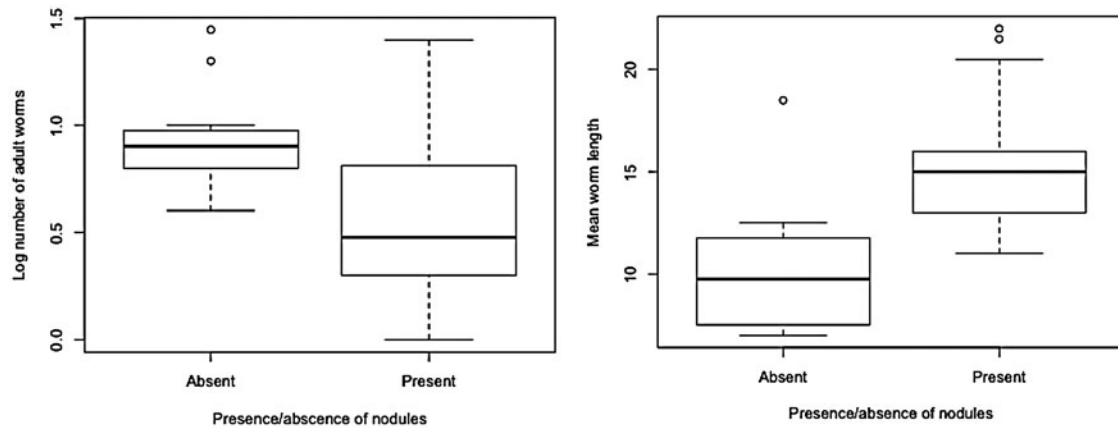


Fig. 3. Relationship between the presence of nodules and mean worm burden and mean worm length in pheasants.

establishment of *S. trachea* within the ring-necked pheasant; however, other factors such as exposure rates and larval viability were not taken into account in this study. This is demonstrated by a reduction in parasite abundance in birds that had evidence of previous exposure. There were, however, a greater number of adult worms in crows, which suggests that worm establishment is not constrained by size or length of the trachea, and therefore overall host size, however, is perhaps a function of host immunity. Indeed, Olivier (1944) found that *S. trachea* establishment was dose dependent. He found that the number of worms establishing was inversely proportional to the size of the infective dose, and attributed this to the strength of the immune response (Olivier, 1944). This result is in stark contrast to the findings of Michael and Dunby (1989), who found that *T. muris* establishment in the murine host is believed to be regulated by density-dependent intraspecific competition, owing to the finite space in the caecum. It is unlikely however that *S. trachea* establishment is regulated in a similar manner as more worms have been found in crows with a shorter trachea. This apparent immune-mediated inhibition on worm establishment has also been identified for *S. trachea* in chickens, with a lower mean worm burden generally identified in older, previously exposed chickens (Crawford, 1940). If establishment was merely a result of parasite-mediated competition, worm establishment, and therefore burden, would be similar in both immunological naïve and previously exposed birds (Luong *et al.* 2011).

One reason to explain the trend for higher worm abundance in crows is acquired-immunity. Pheasants are known to develop moderate immunity to *S. trachea*; however, no such work has been conducted in wild crow populations. Being a known reservoir for *S. trachea*, it may be that crows have a higher parasite threshold for the stimulation of an immune response or they do not develop significant immunity to subsequent infections. Indeed, pheasants appear to be more susceptible to infection

early on in the rearing process, whereas *S. trachea* adults have been recovered from crows of varying ages (personal unpublished data). Further work is however required in order to determine whether wild crows develop any immunity to *S. trachea*.

Although density-dependent reduction of worm fecundity was present in both species, the fact that the effect of crowding on mean worm length was more profound within the crow population is interesting. Mean worm burden explained 82% of the variation in mean worm length in crows, compared with 64% in pheasants, with crows having a tendency for a greater mean worm burden when compared with pheasants. Even so, the fact that worms tended to be shorter in crows, in response to higher mean worm burdens, suggests that these effects are indeed density-dependent. The negative association between worm length and worm burden was present in both species, and appears to be a result of parasite-mediated competition, for either space or resources. Indeed, these effects were even observed in pheasants with no history of previous exposure. Similarly, as there were a vast number of birds of different ages, it is unlikely that age-dependent acquired-immunity was responsible for the manifestation of density dependence within these birds, as the effects were identified in juveniles, as well as adult birds, with little to no acquired immunity. Conversely, Paterson and Viney (2002), observed the absence of density-dependent mechanisms at regulating survivability and fecundity of *Strongyloides ratti* infra-populations in immunocompromised hosts. These mechanisms were however, present in later primary infections, suggesting that host-mediated heterogeneities in immunocompetence are regulating population dynamics before intraspecific competition for space and nutrients ever occurs in experimentally infected rats (Paterson and Viney, 2002). Alternatively, worm length has been shown to be related to levels of local parasite-specific immunoglobulin A (Stear *et al.* 1997). These responses are however, often absent in immunologically naïve animals, and only

generally manifest in animals that have been previously exposed (Craig *et al.* 2014) so it is unlikely to be occurring within these study populations.

The parasite threshold for the manifestation of density dependence within this study was low compared with other studies. For instance, the threshold for density-dependent reductions in fecundity in the caecal nematode, *Heterakis gallinarum*, in pheasants is 96 worms (Tompkins and Hudson, 1999). Similarly, this threshold for *Trichostrongylus colubriformis* in sheep is around 3000 worms per host (Dobson *et al.* 1990). It is generally believed that density-dependent effects are of greater importance for parasites that are large compared with their host (Poulin and Morand, 2000). Indeed, *S. trachea* adults can grow up to ~33 mm in length in an 80–100 mm long trachea (Crow). In comparison, mean worm length of *H. gallinarum* adults in the caecae of pheasants is about 9.64 mm (± 0.11) (Tompkins and Hudson, 1999), in caecae ranging from 240.11 for male and 213.84 for female pheasants, respectively. Similarly, *Pterygodermatites peromysci*, a nematode parasite of mice, is regulated by tight density-dependent restrictions on the number and length of adult worms in the small intestine (Luong *et al.* 2011). Similarly to *S. trachea*, *P. peromysci* can grow up to 33 mm in a 250 mm mouse intestine (Luong *et al.* 2011).

The findings of the present study are in agreement with previous work that pheasants do indeed develop immunity to *S. trachea* (Olivier, 1944; Guilford and Herrick, 1954). However, nematode length and fecundity appear to be a function of parasite density, and therefore parasite-mediated competition and not host-mediated heterogeneities in immunocompetence.

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REFERENCES

- Anderson, R. M. and May, R. M. (1978). Regulation and stability of the host-parasite population interactions. *Journal of Animal Ecology* **47**, 219–247.
- Clapham, P. A. (1935). On nodules occasioned by gapeworm in pheasants. *Journal of Helminthology* **13**, 9–12.
- Craig, N. M., Smith, D. W., Pate, J. A., Morrison, I. W. and Knight, P. A. (2014). Local cytokine transcription in naïve and previously infected

- sheep and lambs following challenge with *Teladorsagia circumcincta*. *BMC Veterinary Research* **10**. doi: 10.1186/1746-6148-10-87.
- Crawford, M. (1940). Infection of adult fowls with *Syngamus trachealis*. *Indian Journal of Veterinary Science and Animal Husbandry* **10**, 293–294.
- Dezfuli, B. S., Volponi, S., Beltrami, I. and Poulin, R. (2002). Intra- and interspecific density-dependent effects on growth in helminth parasites of the cormorant, *Phalacrocorax carbo sinensis*. *Parasitology* **124**, 537–544.
- Dobson, A. P., Waller, P. J. and Donald, A. D. (1990). Population dynamics of *Trichostrongylus colubriformis* in sheep: the effect of infection rate on the establishment of infective larvae and parasite fecundity. *International Journal for Parasitology* **20**, 347–352.
- Fernando, M. A., Stockdale, P. H. G. and Remmler, O. (1971). The route of migration, development and pathogenesis of *Syngamus trachea* (Motagu, 1811), Chapin 1925, in Pheasants. *Journal of Parasitology* **57**, 107–116.
- Gettings, O. J., Sage, R. B. and Leather, S. R. (2015). Spatial distribution of the infectious stages of the nematode *Syngamus trachea* within release pens in the South West of England: potential density dependence? *Veterinary Parasitology* **212**, 267–274. doi: 10.1016/j.vetpar.2015.07.016.
- Goble, F. C. and Kutz, H. L. (1945). Notes on the Gapeworms (Nematoda: Syngamidae) of Galliform and Passeriform birds in New York State. *Journal of Parasitology* **31**, 394–400.
- Guilford, H. G. and Herrick, C. A. (1954). The effect of gapeworm disease in pheasants. *Transactions of the Wisconsin Academy of Sciences Arts and Letters* **43**, 25–50.
- Irvine, R. J., Stien, A., Dallas, J. F., Halvorsen, O., Langvatn, R. and Albon, S. D. (2001). Contrasting regulation of fecundity in two abomasal nematodes of Svalbard reindeer (*Rangifer tarandus platyrhynchus*). *Parasitology* **122**, 673–681.
- Ives, A. R. (2015). For testing the significance of regression coefficients, go ahead and log-transform count data. *Methods in Ecology and Evolution* **6**, 828–835.
- Lewis, A. (1928). Observations on the morphology of *Syngamus trachea* from some wild and domestic birds. *Journal of Helminthology* **6**, 99–112.
- Luong, L. T., Vigliotti, B. A. and Hudson, P. J. (2011). Strong density-dependent competition and acquired immunity constrain parasite establishment: implications for parasite aggregation. *International Journal for Parasitology* **41**, 505–511.
- Marcogliese, D. J. (1997). Fecundity of sealworm (*Pseudoterranova decipiens*) infecting grey seals (*Halichoerus grypus*) in the Gulf of St. Lawrence, Canada: lack of density-dependent effects. *International Journal for Parasitology* **27**, 1401–1409.
- Michael, E. and Dunby, D. A. P. (1989). Density dependence in establishment, growth and worm fecundity in intestinal helminthiasis: the population biology of *Trichuris muris* (Nematoda) infection in CBA/Ca mice. *Parasitology* **98**, 451–458.
- Morgan, D. O. and Clapham, P. A. (1934). Some observations on Gape-worm in Poultry and Game Birds. *Journal of Helminthology* **12**, 63–70.
- Mossinger, J. and Wenk, P. (1986). Fecundity of *Litosomoides carinii* (Nematoda, Filarioidea) *in vivo* and *in vitro*. *Parasitology Research* **72**, 121–131.
- Olivier, L. (1944). Acquired resistance in chickens, turkeys, and ring-necked pheasants to the gapeworm, *Syngamus trachea*. *Journal of Parasitology* **30**, 64–76.
- Paterson, S. and Viney, M. E. (2002). Host immune responses are necessary for density dependence in nematode infections. *Parasitology* **125**, 283–292.
- Poulin, R. and Morand, S. (2000). Parasite body size and interspecific variation in levels of aggregation among nematodes. *Journal of Parasitology* **86**, 642–647.
- Richards, D. T. and Lewis, J. W. (2001). Fecundity and egg output by *Toxocara canis* in the red fox, *Vulpes vulpes*. *Journal of Helminthology* **75**, 157–164.
- Sinniah, B. and Subramaniam, K. (1991). Factors influencing the egg production of *Ascaris lumbricoides*: relationship to weight, length and diameter of worms. *Journal of Helminthology* **65**, 141–147.
- Skorping, A., Read, A. F. and Keymer, A. E. (1991). Life history covariation in intestinal nematodes of mammals. *Oikos* **60**, 365–372.
- Stear, M. J. and Bishop, S. C. (1999). The curvilinear relationship between worm length and fecundity of *Teladorsagia circumcincta*. *International Journal for Parasitology* **29**, 777–780.
- Stear, M. J., Bairden, K., Duncan, J. L., Holmes, P. H., McKellar, Q. A., Park, M., Strain, S., Murray, M., Bishop, S. C. and Gettinby, G. (1997). How hosts control worms. *Nature* **389**, 27.
- Szalai, A. J. and Dick, T. A. (1989). Differences in numbers and inequalities in mass and fecundity during the egg-producing period for *Raphidascaris acus* (Nematoda: Anisakidae). *Parasitology* **98**, 489–495.

Tompkins, D. M. and Hudson, P. J. (1999). Regulation of nematode fecundity in the ring-necked pheasant (*Phasianus colchicus*): not just density dependence. *Parasitology* **118**, 417–423.

Walker, M., Hall, A., Anderson, R. M. and Basanez, M. G. (2009). Density-dependent effects on the weight of female *Ascaris lumbricoides* infections of humans and its impact on patterns of egg production. *Parasites and Vectors* **2**.

Wehr, E. E. (1937). Observations on the development of poultry gape-worm *Syngamus trachea*. *Transactions of the American Microscopical Society* **56**, 72–78.

Williams, I. C. and Newton, I. (1969). Intestinal helminths of the bullfinch *pyrrhula pyrrhula* (L.), in southern England. *Journal of the Helminthological Society of Washington* **36**, 76–82.