

New infection rate of bovine mammary glands after application of an internal teat seal at dry-off

Volker Krömker^{1*}, Nils Th. Grabowski² and Julia Friedrich¹

¹ Faculty II, Department of Microbiology, University of Applied Sciences and Arts Hannover, Heisterbergallee 12, 30453 Hannover, Germany

² Institute for Food Quality and Food Safety, University of Veterinary Medicine Hannover, Foundation, Bischofsholer Damm 15, 30173 Hannover, Germany

Received 10 July 2013; accepted for publication 3 October 2013

The internal teat sealant OrbeSeal[®] (Zoetis, Berlin, Germany) is intended to prevent new intramammary infection (IMI) throughout the dry period. The aim of this field study was to determine new infection rates in udder quarters applied exclusively with the sealant at dry-off and untreated ones (control group). For that, the new infection rate of udder quarters treated with the sealant was evaluated in a split-udder design (front right and rear left udder quarters treated, the other ones untreated) in 128 cows from nine dairy farms in North Germany that were healthy in all four udder quarters (i.e. <100 000 cells/ml, bacteriologically negative). After calving, duplicate quarter milk samples were collected twice at DIM (days in milk) 5–12 and 7 d later for culture. The new infection rates for treated and untreated groups were 3·4 and 10·5%, respectively. The results showed that applying a standardised study design with very few confounders under field conditions, using internal teat sealants reduced the new infection rate on quarter level to a third of that of the control group. Thus, internal teat sealants represent a viable option for dry-off treatment of udder-healthy cows.

Keywords: Teat sealant, mastitis, new infection rate, dry period.

The udder health situation is strongly connected with the economic success of a dairy farm and depends on the amount of quarters infected. Self-cure, therapy or culling may reduce the amount of infected quarters; many more factors tend to increase the rate of new intramammary infection (IMI), particularly during the beginning and the end of the dry period, i.e. involution and colostrigenesis (Bradley & Green, 2001). For more than 60 years, drying animals off using antibiotics has been one of the main actions in combating mastitis, reducing new infection rate (NIR) from 30 to 60% (untreated cows) down to 0–15% (treated cows; Berry, 2000). Recently, this routine has been criticised for political reasons, considering increasing resistances towards antibiotics in human patients and only moderate therapeutic success, but no corresponding studies have been conducted (Krömker, 2012). Moreover, epidemiological surveys showed a shift in the mastitis-producing flora in the rest of the cow (i.e. from cow-associated toward environmental pathogens) so therapy during the dry period becomes less important than preventive measures to reduce the risk of new

infections (Krömker et al. 2011). Therapeutical alternatives include vaccination and the application of teat sealants. Although known for decades (Meaney, 1977), teat sealants represent relatively new ways to combat mastitis in the dry period and have been promoted in Germany since 2003.

Teats are sealed by introducing a stable substance which is supposed to avoid the penetration of pathogens; as it does not contain any antibiotics which eventually could stop iatrogenic infections, special attention must be paid to an aseptic application. With OrbeSeal[®] (Zoetis, Berlin, Germany) the German market has been relying on a similar bismuth-based preparation that has a high degree of biocompatibility and lasts inside the teat canal and teat cistern throughout the entire dry period. While it was initially used in selective dry-off therapy (i.e. antibiotic treatment for infected quarters and teat sealing for uninfected ones) now a combined application of long-lasting antibiotics plus teat sealant has come into practice (Redetzky & Hamann, 2003). Halasa et al. (2009) compared several management practices for the dry period, among applying teat sealants. Four studies were included and this meta-analysis confirmed a 'significant protection against new IMI'. Several studies focused on the protection conveyed by teat sealants, but only few have been published so far regarding the efficacy of

*For correspondence; e-mail: volker.kroemker@hs-hannover.de

commercial bismuth-based internal teat sealants to prevent new infections in comparison with untreated udder quarters (e.g. Woolford et al. 1998; Berry & Hillerton, 2002; Huxley et al. 2002; Krömker et al. 2010; Petrovski et al. 2011). This field study describes the experiences with an internal teat sealant made of bismuth subnitrate suspended in paraffin (OrbeSeal® Zoetis, Berlin, Germany) (ITS) in nine dairy farms in Northern Germany. In contrast to other studies that dealt with OrbeSeal®, this one is based on a very strict selection of animals which permitted the evaluation of any effects on the new infection rate in a way almost devoid of other animal-rated variables (previous udder diseases, teat and udder morphology etc.). The aim of this study was to determine the NIR using ITS and the rate of clinical mastitis cases during the first 100 d of the subsequent lactation in healthy udder quarters at dry-off.

Material and methods

Selection of dairy farms

The field studies were carried out between March and October 2012 on nine dairy farms located in the German states of North-Rhine Westphalia, Lower Saxony and Saxony-Anhalt. All selected farms managed a mean of 411 ± 353 dairy cows (min. 98 to max. 1022). Yield over 305 d varied between 8900 and 11 832 kg (mean 10252 ± 1072 kg) with a fat content of $4.4 \pm 0.1\%$ and a protein content of $3.4 \pm 0.1\%$; the mean intercalving interval was 401 ± 16 d. Bulk tank somatic cell count at the beginning of the study ranged from 98 000 to 264 000 cells/ml (mean $179\,000 \pm 47\,000$ cells/ml). The management's willingness to participate, the breed (German Holstein), and the participation in the German Dairy Herd Improvement were the inclusion criteria for this experiment. It was also mandatory that, of all clinical mastitis cases, less than 5% were due to *Staphylococcus aureus* infections. All of them performed a twice-a-day milking routine and an abrupt dry-off.

Selection of animals

Trial animals had to be scheduled for dry-off during the survey with a planned dry period of 42 ± 4 d. Individual cow milk samples had to present a geometric mean cell count of $< 50\,000$ cells/ml milk during the previous three dairy herd improvement tests and a negative record of clinical mastitis in the present lactation.

Number of cows

The tested hypothesis was that treated quarters would have a lower new infection rate (4%) than untreated ones (12%). Based on a one-sided χ^2 test with type I error $\alpha = 0.05$ and type II error $\beta = 0.10$, a total of 194 quarter pairs were

Table 1. Bacteriological distribution post partum in quarters treated with OrbeSeal® ($n = 256$, OS) and in control quarters ($n = 256$, C)

Bacteriological result	OS, n (%)	C, n (%)
No growth	247 (96.5)	229 (89.5)
<i>Staphylococcus aureus</i>	0	3 (1)
Coagulase-negative staphylococci	3 (1)	6 (2)
<i>Streptococcus uberis</i>	3 (1)	5 (2)
Coliforms/ <i>Escherichia coli</i>	2 (1)	3 (1)
Enterococci	0	2 (1)
Other	1 (0.5)	8 (3)

needed. If 10–20% of cows drop out of the trial post admission, approximately 120 cows were needed.

A total of $n = 128$ out of 155 preselected dairy cows fulfilled the criteria for inclusion and were rated as 'normally secreting' (i.e. cell count $< 100\,000$ cells/ml and bacteriologically negative) on all four quarters before dry-off between March and June 2012. Dry-off treatment for these cows was performed using a 'split-udder design', i.e. the front right-side and the rear left-side quarters of the animals were treated with OrbeSeal® (ITS), while the opposite quarters were left untreated as control. The 27 excluded cows had 39 infected quarters (31 infected quarters 7 d prior to dry-off excluded 20 cows and 8 infected quarters excluded belatedly 7 cows at dry-off).

Sampling

Quarter foremilk samples were aseptically collected in duplicate at four points in time for cyto-bacteriological analysis: (i) within 7 d prior to dry-off, (ii) at dry-off, (iii) within 5–12 d in milk (DIM) and (iv) 7 d later. Milk samples were transferred immediately after sampling to the microbiological laboratory at the University of Applied Sciences, Hannover and submitted to analysis within 2 h after arrival. Storage time until sample processing was a maximum of 8 h. Sampling as described above was used just before dry-off treatment.

Treatment

At dry-off, teat tips of treated and untreated quarters alike were aseptically prepared. Then, ITS was applied to the corresponding udder quarter using hygienic procedures. The blunt cannula of the injector was introduced into the teat canals only as deep as necessary for a safe administration of the drug. The product administration was done by one of the authors, so that farmer or herdspersons were blinded towards treated and untreated quarters.

Laboratory procedures

Milk samples were cultured according to NMC recommendations (NMC, 1999) as cited by the German Veterinary Association (GVA, 2009).

Table 2. Final mixed logistic regression model for new infections

Effect	Estimate	SE	t Value	Pr> t	Odds Ratio	95% CI
Intercept	3.600	0.509	7.079	0.000	36.6	13.477–99.401
Treatment, no vs. yes	–1.338	0.533	–2.510	0.01	0.262	0.092–0.748
Parity 1 2+ reference	–0.435	0.625	–0.696	0.487	0.647	0.190–2.210
Quarter position rear	–0.117	0.327	–0.358	0.721	0.890	0.468–1.690
Parity (1) × treatment (0)	0.299	0.731	0.409	0.682	1.349	0.321–5.675

The somatic cell count of quarter foremilk samples was determined by flow cytometry with the Somascope Smart (DeltaInstruments B.V., Drachten, The Netherlands).

Definitions

A milk sample was termed 'contaminated' if more than 3 bacterial species were isolated. IMI was assessed if in duplicate samples > 500 cfu/ml of the same bacterial species were cultured, and 1–3 bacterial species could be isolated. Quarters with only one bacteriologically positive sample were termed 'non-infected'. Quarters were termed 'newly infected' if they presented positive results for a pathogen that, previously, was not isolated in those quarters. Animals were examined periodically (twice a day by the milkers) for clinical mastitis until DIM 100.

Statistical analysis

Data were collected and analysed using Excel 2000 (Microsoft Corporation) and SPSS (SPSS 20.0, Chicago, USA). The udder quarter was recognised as the statistical unit. The untreated opposed quarter served as a paired control. New infection (NI) was analysed using a mixed model logistic regression analysis. For the regression model, the linear predictor was given by

$$\text{Logit(NI)} = \text{Lactation} + \text{Front/hind quarter} \\ + \text{Treatment} + \text{Herd(random)} + e$$

where 'lactation' was divided into two categories: '1' and '2+'; 'front/hind quarter' was two categories; and 'treatment' was indicated as 'internal teat sealant' or 'no treatment'. A random herd effect was included in the model. These factors were included as a fixed effect and 'e' was a binomial error term. A backward stepwise regression analysis was performed. For all outcome variables, treatment was compared with the control group using significance tests with a 5% significance level (*P* value). The homogeneity of treatment groups was given due to the paired study design.

Results

A total of *n* = 256 quarter pairs in *n* = 128 cows was used for the analysis. All quarters displayed low cell counts (i.e. < 100 000 cells/ml) and were culture-negative at dry-off. The percentage of new infections differed significantly between

treatment groups: 3.5% (9/256) after internal teat sealant and 10.5% (27/256) after no treatment (*P* < 0.001) (Table 1). Of the new infections, 25% were caused by coagulase-negative staphylococci (CNS), 22% by *Streptococcus uberis* and 14% by *Enterobacteriaceae*, irrespective of treatment protocol.

Within the first 100 d after calving, four clinical mastitic quarters were detected, two in each group, thus being non-significant. No clinical case of mastitis occurred during the first 30 d after parturition. The logistic regression proved that ITS was able to protect the udder quarters from new infection during the dry period in a more efficient way than leaving them untreated (OR = 0.262 CI 0.092–0.748; Table 2). Random farm effect was not significant (*P* = 0.44).

Discussion

The periparturient period is a time when external factors can easily affect udder health; they are not only multiple but also vary greatly between herds and locations, leading to changing stages of immunocompetence facing a highly variable array of pathogens (Suriyasathaporn et al. 2000). Internal teat sealants are one way to support udder health by minimising the risk of pathogens passing through teat canals. The present trial intended to determine the level of efficacy that these sealants achieve under strict conditions.

Methods

As mentioned already, reports on the efficacy of teat sealants are scarce, and study designs varied greatly, making the results difficult to compare. None, however, evaluated the influence of internal teat sealants on 'truly udder-healthy' animals [i.e. cell counts below 100 000 cells/ml and no pathogens (not even minor pathogens) encountered, both in all quarter foremilk samples of a given udder]. By using a split-udder design and including only healthy udders, this approach minimised the influence of animal-related confounders (Krömker et al. 2010). Any possible underestimation of the observed effects may, in the case of split-udder designs, be due to an eventual non-interdependence of the quarters (Lam et al. 1996). However, as all four quarters were healthy, this issue appears negligible. Still, not matching quarters with treatment groups randomly may possibly create a bias, eventually by teat condition impairments due to hyperkeratosis that tend to affect specific quarters because of e.g. positioning problems at milking.

However, animals had to be free from clinical changes in the quarter morphology in order to participate in this study, and so the risk of this error was judged to be low.

Udder pathogens

As can be seen in Table 1, opportunistic (CNS) and environmental pathogens (*Str. uberis* and coliforms) were the most prominent new infection agents. These results are generally similar to other study results describing the distribution of mastitis pathogens in Northern and Central Germany (Tenhagen et al. 2006; Grabowski et al. 2012) but some studies also show presence of coryneform bacteria.

In general terms, the few studies dealing with ITS found that sealing uninfected teats internally provides a comparable (or even better) protection to the udder than found with antibiotic treatment at drying-off. They calculated NIR between 2.0 and 25.5% for ITS and between 16 and 25% for long-term antibiotics. Cell count thresholds were usually higher (200 000 cells/ml or below) than in the present study (Woolford et al. 1998; Berry & Hillerton, 2002; Huxley et al. 2002), or challenged sealed teats artificially (Petrovski et al. 2011), so a comparison is possible only on very basic terms.

With a NIR of 3.5% for ITS-treated and of 10.5% for untreated quarters, the present study follows the general tendency observed in others. However, the effect was investigated under very strict conditions (only truly udder-healthy animals, split-udder design). The multivariate analysis that also considered some factors (quarter position, lactation number, herd effect) did not reveal any significant influence on NIR. Therefore, applying the teat sealant in healthy udder quarters alone already provided a protective and significant effect on NIR. The results achieved with this model show that additional systemic factors (which had to be left unaddressed in this paper) clearly influence the incidence rate for a new infection to occur. The model was focused on few variables because its objective was merely to quantify the influence of determining variables (those that were studied here, and in comparison between exposed and non-exposed ones) on the dependent ones, rather than to predict dependent variables.

The array of pathogens and the efficacy of ITS against them also varied. It is thought that some infections with environmental pathogens were even more likely to be avoided when teats were sealed rather than when they were treated with antimicrobials. For Woolford et al. (1998) teat sealing led to a marked reduction of new infections with environmental streptococci and coliforms. Huxley et al. (2002) detected significant differences only for quarters with *Escherichia coli* while environmental streptococci were not considered in that study. According to Berry & Hillerton (2002) the percentage of quarters infected by *Str. uberis* did not vary, however. Although significantly fewer infections were recorded for the teat-sealed group, the incidence of clinical mastitis within the first 100 d after calving did

not differ. So the type of pathogen could be a potential confounder. In fact, infections by coliforms were recorded less frequently than those with other pathogens. The present results did not support this observation, meaning that the reduction in new infections took place independently of the type of pathogen, at least in the case of the pathogens isolated in this study. Still, the influence by the exposure to pathogens as a confounder naturally depends on the selection of animals (definition of udder health and new infection), the housing system (increased exposure to environmental pathogens), climate differences among farm locations and management differences from farm to farm (Pieper et al. 2013).

With these results, the present survey supports the use of ITS in uninfected quarters at dry-off. Therefore, using ITS as the only dry-off treatment will be successful in those animals with uninfected quarters and no clinical mastitis while using careful application hygiene and hygienic husbandry conditions at the time of dry-off and in the dry period. Teat sealing should be accompanied by periodical monitoring during the dry period in order to recognise milk dropping or the development of clinical mastitis.

We are thankful to the dairy farmers who supported this study. We certify that there is no conflict of interest and disclose any financial or personal relationships with persons or organisations which could have influenced our work.

References

- Berry EA 2000 Dry cow strategies and results from a selective dry cow trial. *Cattle Practice* **8** 255–258
- Berry EA & Hillerton JE 2002 The effect of an intramammary teat seal on new intramammary infections. *Journal of Dairy Science* **85** 2512–2520
- Bradley AJ & Green MJ 2001 An investigation of the impact of intramammary antibiotic dry cow therapy on clinical coliform mastitis. *Journal of Dairy Science* **84** 1632–1639
- German Veterinary Association (GVA) 2009 *Leitlinien zur Entnahme von Milchproben unter antiseptischen Bedingungen und Isolierung und Identifizierung von Mastitiserregern [Guidelines for Aseptic Milk Sampling and Guidelines to Isolate and Identify Mastitis Pathogens]*. 1st edition. Gießen, Germany: German Veterinary Association
- Grabowski NT, Ahlfeld B & Klein G 2012 Udder health from the point of view of a diagnosis laboratory. *Journal of Food Quality and Food Safety* **63** 86–91
- Halasa T, Østerås O, Hogeveen H, van Werven T & Nielsen M 2009 Meta-analysis of dry cow management for dairy cattle, part 1: protection against new intramammary infections. *Journal of Dairy Science* **92** 3134–3149
- Huxley JN, Green MJ, Green LE & Bradley AJ 2002 Evaluation of the efficacy of an internal teat sealer during the dry period. *Journal of Dairy Science* **85** 551–561
- Krömker V 2012 Strategies against mastitis due to cow-associated pathogens as a herd problem. *Journal of Food Quality and Food Safety* **63** 61–64
- Krömker V, Pfannenschmidt F & Friedrich J 2010 Neuinfektionsrate der Milchdrüsen von Milchkühen in der Trockenperiode nach Anwendung eines internen Zitzenversieglers zum Trockenstellen [New infection rate of bovine mammary glands after application of an internal teat seal at dry off]. *Berliner und Münchener Tierärztliche Wochenschrift* **123** 215–225

- Krömker V, Friedrich J, Kleen JL, Klocke D, Paduch JH, Zinke C, Haverkamp H, Taffe B, Koch A, Reinecke F & Volling O 2011 Mastitis control on herd level. *Der Praktische Tierarzt* **92** 4–35
- Lam TJ, DeJong MC, Schukken YH & Brand A 1996 Mathematical modeling to estimate efficacy of postmilking teat disinfection in split-udder trials of dairy cows. *Journal of Dairy Science* **79** 62–70
- Meaney WJ 1977 Effect of a dry period teat seal on bovine udder infection. *Irish Journal of Agricultural Research* **16** 293–299
- National Mastitis Council 1999 *Laboratory Handbook on Bovine Mastitis*. 360 Revised edition. Madison, WI, USA: National Mastitis Council Inc
- Petrovski KR, Caicedo-Caldas A, Williamson NB, Lopez-Villalobos N, Grinberg A, Parkinson TJ, Tucker IG 2011 Efficacy of a novel internal dry period teat sealant containing 0.5% chlorhexidine against experimental challenge with *Streptococcus uberis* in dairy cattle. *Journal of Dairy Science* **94** 3366–3375
- Pieper J, Hoedemaker M, Krömker V 2013 Significance of the dry period for the development and prevention of new infections of the bovine mammary gland. *Tierärztl Prax* **41** (G) 1–9
- Redetzky R & Hamann J 2003 Possibilities and limitations of teat sealers for prevention of mastitis. *Der Praktische Tierarzt* **84** 530–538
- Suriyasathaporn W, Heuer C, Nordhuizen-Stassen EN & Schukken YH 2000 Hyperketonaemia and the impairment of udder defense: a review. *Veterinary Research* **31** 397–412
- Tenhagen BA, Köster G, Wallmann J & Heuwieser W 2006 Prevalence of mastitis pathogens and their resistance against antimicrobial agents in dairy cows in Brandenburg, Germany. *Journal of Dairy Science* **89** 2542–2551
- Woolford MW, Williamson JH, Day AM & Copeman PJA 1998 The prophylactic effect of a teat sealer on bovine mastitis during the dry period and the following lactation. *New Zealand Veterinary Journal* **46** 12–19