

# Phthalates in neonatal health: friend or foe?

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Exposure to environmental chemicals has adverse effects on the health and survival of humans. Emerging evidence supports the idea that exposure to endocrine-disrupting compounds (EDCs) can perturb an individual's physiological set point and as a result increase his/her propensity toward several diseases. The purpose of this review is to provide an update on di-(2-ethylhexyl) phthalate, the primary plasticizer found in plastic medical devices used in neonatal intensive care units, its effects on the fetus and newborn, epidemiological studies, pharmacokinetics, toxicity and epigenetic implications. We searched the PubMed databases to identify relevant studies. Phthalates are known EDCs that primarily are used to improve the flexibility of polyvinyl chloride plastic products and are called plasticizers in lay terms. Neonates and infants are particularly vulnerable to the effects of phthalates, beginning with maternal exposure and placental transfer during gestation and during infancy following birth. In line with the developmental origins of adult disease, a focus on the effects of environmental chemicals *in utero* or early childhood on the genesis of adult diseases through epigenome modulation is timely and important. The epigenetic effects of phthalates have not been fully elucidated, but accumulating evidence suggests that they may be associated with adverse health effects, some of which may be heritable. Phthalate exposure during pregnancy and the perinatal period is particularly worrisome in health-care settings. Although the clinical significance of phthalate exposure has been difficult to assess with epidemiologic studies, the evidence that physiological changes occur due to exposure to phthalates is growing and points toward the need for more investigation at a molecular, specifically epigenetic level.

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## Introduction

The first American newborn intensive care unit (NICU) was opened at Yale Hospital in New Haven, Connecticut in 1960.<sup>1</sup> The advent of specialized care units and associated support personnel as well as medical advances in neonatal care have greatly reduced infant mortality rate; for example, 95% of infants with birth weights of 1000 g now survive compared with 5% one hundred years ago.<sup>2</sup> Increased neonatal exposure to plasticizers by means of medical devices, tubing and the hospital environment has accompanied the medical and technological advances in care. Many studies have examined exposures to plasticizers in this and other patient populations in attempts to determine whether they are safe or not.<sup>3–5</sup>

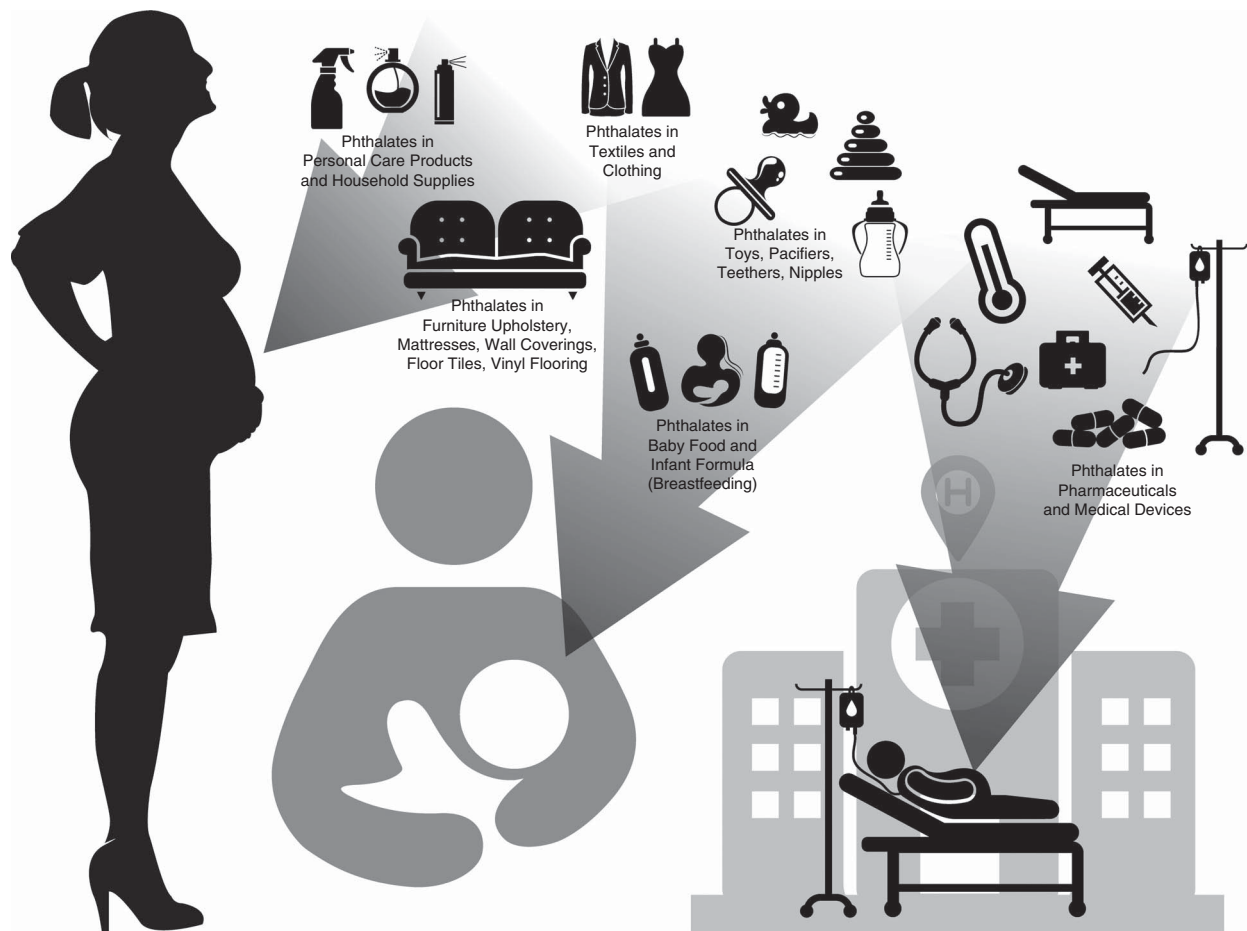
While plasticizers have enabled advances in medical products, exposure to plasticizers may play an important role in the etiology of several diseases.<sup>6,7</sup> Plasticizers are classified as endocrine-disrupting compounds (EDCs) by the National Institutes of Health, the World Health Organization and other agencies. EDCs also include pesticides and other chemicals that may enter the environment and result in cancers, birth defects or other developmental disorders.<sup>8,9</sup> EDCs were initially investigated for estrogenic properties, but later it became

apparent that androgenic and anti-androgenic properties were also significant.<sup>10</sup> Neonatal exposures to plasticizers may occur *in utero*, in a neonatal care unit, from breast milk and from household products. This review is focused on the common plasticizer di-(2-ethylhexyl) phthalate (DEHP) in relation to each of these pathways for neonatal exposures and subsequent effects (Fig. 1).

Polyvinyl chloride (PVC) or plastic may contain 30–50% of DEHP.<sup>11</sup> In addition to DEHP, low molecular weight phthalates such as dimethyl phthalate (DMP), dibutyl phthalate (DBP) and diethyl phthalate (DEP) have been incorporated into cosmetics, fragrance, adhesives, inks, pharmaceuticals and other personal care products. Di-*n*-butyl phthalate (DnBP) is used in certain adhesives. High molecular weight and branching alkyl chain phthalates, primarily butyl-benzyl phthalate (BBP), di-*n*-octyl phthalate and DEHP are added to food packaging, building materials, paints, toys, building materials, medical devices, textiles and clothing, automobiles and many other articles.<sup>12</sup> Thus, phthalates are ubiquitous in our environment such that humans, livestock and wild animals are exposed continuously.<sup>12</sup> Human exposure occurs via ingestion, inhalation, intravenous (IV) delivery during medical procedures and dermal absorption.<sup>13</sup> Yet, published data concerning sources of phthalates are incomplete, in part because their inclusion in products does not have to be disclosed.<sup>12</sup>

DEHP has historically been the preferred plasticizer for PVC products, particularly those used for medical applications, because it imparts flexibility, transparency, strength and

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**Fig. 1.** Neonate's exposure to commonly used products containing phthalates especially di(2-ethylhexyl) phthalate: neonates are exposed to phthalates via breast feeding, pharmaceuticals, medical devices or cribs in the health-care unit as well as dietary or day to day household supplies.

weldability, and also is compatible with various sterilization processes.<sup>14</sup> Plasticizers alter the rigid mechanical properties of PVC by embedding themselves between polymeric chains, spacing them apart to make the plastic soft and flexible.<sup>15</sup> Leaching into liquids and migration to the environment occurs throughout the period of use because DEHP is not chemically bound to PVC; more precisely, it is not covalently bound.<sup>16</sup> Chronic exposure to environmental DEHP has been a source of concern and research for over 40 years.<sup>17,18</sup>

Furthermore, exposures to plasticizers may have effects beyond acute or chronic toxicity. Alterations in the phenotype due to environmental exposures during critical periods of development *in utero* and early life are regulated through epigenetic gene programming in various tissues, and may adversely affect later life. Epigenetics is an important mechanism in the ability of the human host to respond to environmental challenges with either disease or healthy outcomes.<sup>19</sup> The term 'epigenetic' is a broad descriptor of heritable processes independent of changes in the DNA sequence, including the necessary phenomena of genomic imprinting and X chromosome inactivation. In addition, modifications to histone or non-coding RNA [e.g. microRNA

(mRNA)] can occur in response to environmental signals, such as diet and stress, and modulate gene expression and protein production. The more widely studied plastic component, bisphenol A, is another endocrine disruptor that has largely been abandoned by the food packaging industry.<sup>20</sup> While endocrine disruptors have been widely studied, it is evident that epigenetic investigations are just now emerging to explain the effects of endocrine disruptors in humans at various stages of life.<sup>8</sup>

Interestingly, epigenetic modifications may play a central role in gene reprogramming, and carry over to subsequent generations. Epigenetic transgenerational effects occur when the phenotype is altered due to an environmental exposure or stressor and passed along to subsequent generations.<sup>21</sup> Although the fetal programming hypothesis proposes that environmental stimuli act during the critical periods of development and may permanently alter the structure and function of the fetus, the mechanism(s) underlying such observations are not well elucidated. By understanding the mechanism(s) underlying the fetal origin of adult disease due to endocrine disruptors, specifically plasticizers, policy makers as well as health-care professionals can make this issue a high

health-care priority and provide appropriate treatment for people at high risk for these chronic diseases more effectively.

## Methods

To review this topic, we undertook a comprehensive search of the PubMed (<http://www.ncbi.nlm.nih.gov/pubmed>) databases using 'epigenetics,' 'DNA methylation,' 'histone modification,' 'neonate,' 'DOHAD' and/or 'DEHP' or 'phthalate' as key terms. We also combined these search terms with others in which we added '*in utero* exposure' and 'endocrine disruptors.' We also considered review articles as well as references within articles found in our literature search. We excluded articles which were not written in English. The PubMed search covered all articles up to 12 May 2015.

## Toxicities of and guidelines for DEHP

Many animal studies have demonstrated the toxicity of DEHP in liver and testes as well as other tissues.<sup>22</sup> DEHP is carcinogenic, fetogenic and teratogenic in rats and mice.<sup>23</sup> DEHP exhibits very low acute oral toxicity, with LD50 values in rabbits and rats around 30–34 g/kg. However, when administered intravenously to rats, the acute LD50 is as low as 200 mg/kg.<sup>24</sup>

Because of health concerns, the U.S. Environmental Protection Agency and the European Food Safety Authority have established oral intake limit values for various phthalates: the Reference Dose (RfD) and the tolerable daily intake (TDI), respectively. The RfD's for DEHP, BBP and DBP are 0.02, 0.2, and 0.1 mg/kg/day and the TDI's for DEHP and DBP are 0.05, 0.5 and 0.01 mg/kg/day, respectively.<sup>25</sup> The Environmental Protection Agency has established a maximum contaminant level for DEHP of 0.006 mg/l (6 ppb) for drinking water.<sup>26</sup> Other government agencies and organizations have issued regulations or guidelines regarding DEHP, summarized by the National Toxicology Program Report on Carcinogens (Table 1).<sup>27</sup>

## Pharmacokinetics of DEHP

Because several routes of exposure are possible with different absorption, distribution, metabolism and elimination characteristics, many studies have focused on measuring urinary metabolites of phthalates as the final common means of clearance from the body (Table 2). Xenobiotics may be completely or partially rendered water soluble by first pass metabolism and glucuronidation, and eliminated in the urine.<sup>28</sup> The parent phthalate may also undergo metabolic transformation via enzymatic processes in the liver. Because clearing enzyme activity is lower at birth than adulthood, health risks may be increased in neonates. Developmental changes in the liver and kidneys throughout pre- and postnatal growth modify the pharmacokinetic clearance of xenobiotics in ways that are not well described. In children, phthalate metabolite levels in urine have been characterized in various populations.<sup>29</sup> The highest daily phthalate intake was found to be DEHP in a

study of 431 Danish children between 3 and 6 years of age (median: 4.42 µg/day/kg body weight) and BBP the lowest (median: 0.49 µg/day/kg).<sup>29</sup>

DEHP is rapidly metabolized regardless of the route of exposure, although in a healthy adult, only 67% of the DEHP dose was excreted in the urine as five major metabolites in the first 24 h and 3.8% in the next 24 h.<sup>30</sup> Mono-2-ethylhexyl phthalate (MEHP) is only a minor metabolite due to further oxidative reactions and also has the shortest half-life, so that studies which only measure urinary MEHP levels do not reflect the extent of DEHP exposure. With their longer half-lives, oxidized DEHP/MEHP metabolites may accumulate in the body with continuous and prolonged daily exposure in the NICU. Koch *et al.* concluded that 'almost all neonates in medical care exceed the TDI and the RfD. Maximum DEHP exposures of neonates exceed the TDI and RfD by a factor of 100.'<sup>30</sup> The metabolism of DEHP involves hydrolysis by lipase to MEHP and further oxidation by liver enzymes, which together are referred to as phase I reactions in detoxification that result in molecular modification.<sup>31</sup> MEHP has a half-life of 5 h in adults and is further metabolized by different oxidative reactions into products listed in Table 2 which have half-lives from 10 to 24 h in adults. Because the neonatal liver is not fully functional, much longer elimination half-lives might be expected. The P450 cytochrome families (CYP) are the enzymes responsible for biotransformation of most xenobiotics. However, some enzymes are normally expressed at high levels during fetal life but are silenced after birth; some are expressed at constant levels throughout gestation and the postnatal period; and some are not expressed at all or at low levels in the fetal liver with activity occurring late in pregnancy or after birth and increasing later in life.<sup>32</sup> As a result, neonatal metabolism and elimination of phthalates is poorly understood.

In addition, phase I biotransformation of DEHP and MEHP in different human organs was investigated using recombinant human CYP isoforms.<sup>33</sup> The oxidative metabolism of MEHP, 5OH-MEHP, 5oxo-MEHP, 5cx-MEPP and phthalic acid was primarily catalyzed by CYP2C9\*1. Dealkylation to phthalic acid was catalyzed by CYP3A4. In addition, the investigators found that CYP2C9 polymorphisms demonstrated changes in substrate specificity and catalytic activity to MEHP, suggesting potentially important variation in human adverse effects of DEHP. The finding of CYP2C9 polymorphisms is consistent with other studies.<sup>34,35</sup> In order to assess interspecies and interindividual differences in DEHP metabolism, Ito *et al.* assessed the activities of four enzymes that metabolize DEHP in the livers of 38 human subjects and eight mice.<sup>36</sup> Uridine 5'-diphospho-glucuronosyltransferase activity was widely variable among human subjects, but generally lower than mice. The Ito group concluded that interindividual differences of DEHP metabolism in humans may be greater than the difference between humans and mice.<sup>36</sup>

The CYP2C family comprises about 20% of the P450 enzymes in the adult liver, and CYP2C9 is the predominant enzyme.<sup>31</sup> Levels of CYP2C9 are very low in early fetal

**Table 1.** U.S. government regulations regarding DEHP and other guidelines<sup>27</sup>

Agency	Regulation or guideline
Consumer Product Safety Commission	A voluntary standard provides that pacifiers, rattles, and teethers shall not intentionally contain DEHP It is unlawful to manufacture, sell, distribute or import any children's toy or child-care article that contains DEHP at concentrations of >0.1%
Environmental Protection Agency	
Clean Air Act	
National Emissions Standards for Hazardous Air Pollutants	DEHP listed as hazardous air pollutant
New Source Performance Standards	Manufacture of DEHP is subject to certain provisions for the control of volatile organic compound emissions
Clean Water Act	
Effluent Guidelines	Phthalate esters are listed as toxic pollutants
Water Quality Criteria	Based on fish or shellfish and water consumption = 1.2 µg/l; based on fish or shellfish consumption only = 2.2 µg/l
Comprehensive Environmental Response, Compensation, and Liability Act	Reportable quantity: 100 lb
Emergency Planning and Community Right-to-Know Act	
Toxic Release Inventory	DEHP subject to reporting requirements
Resource Conservation and Recovery Act	
Listed hazardous waste	Waste code for DEHP = U028 Listed as a hazardous constituent of waste
Safe Drinking Water Act	Maximum contaminant level = 0.006 mg/l
Food and Drug Administration	21 CFR 177 limits use of DEHP in basic components of single and repeated use food contact surfaces
Occupational Safety and Health Administration	Permissible exposure limit = 5 mg/m <sup>3</sup>
American Conference of Governmental Industrial Hygienists	Threshold limit value – time-weighted average = 5 mg/m <sup>3</sup>
National Institute for Occupational Safety and Health	Immediately dangerous to life and health limit = 5000 mg/m <sup>3</sup> Recommended exposure limit (time-weighted average workday) = 5 mg/m <sup>3</sup> Short-term exposure limit = 10 mg/m <sup>3</sup> Listed as a potential occupational carcinogen

CFR, code of federal regulations; DEHP, di(2-ethylhexyl) phthalate.

development, but between 25 and 40 weeks they achieve levels about 10% of adult values. CYP3A4 is the primary hepatic CYP in adulthood, accounting for 30% of the P450 system, and is involved in the biotransformation of over 75 medications. Its expression is very low at birth but increases to adult levels by 1 year of age. As neonates do not have fully active enzyme metabolism systems, it might be expected that DEHP would accumulate upon continued exposure, as in the NICU. Therefore, toxicity is possible even though the exposure level may be under regulatory limits.

Phase II biotransformations include conjugation with water-soluble moieties (glucuronidation) in order to facilitate rapid elimination from the body by the kidneys. Depending on the phthalate, metabolites may be partially glucuronidated and excreted through the urine and feces.<sup>37</sup> Glucuronidation enzymes are not fully active until after birth.<sup>38</sup> Once solubilized by hepatic glucuronidation or other processes, xenobiotics are

excreted proportionally to the glomerular filtration rate (GFR). In term neonates, the GFR is 2–4 ml/min/1.73 m<sup>2</sup>, which doubles by 1 week of age.<sup>39</sup> The GFR in healthy young adults is about 125 ml/min/1.73 m<sup>2</sup>.<sup>40</sup> The neonate's liver and kidneys function much less effectively than at older ages.

Because phthalates are ubiquitous in the environment including laboratory sampling and procedural plastics, and the fact that monoesters (primary metabolites of DEHP) are also observed, secondary DEHP metabolites in urine have been suggested to be more accurate and sensitive biomarkers of exposure than the monoester MEHP.<sup>37</sup> However, there are several limitations to this approach. DEHP-containing urine collection devices may contaminate urinary samples with MEHP, studies showing high values for this metabolite should be carefully interpreted. Mono-(2-ethyl-5-carboxypentyl) phthalate is by far the chief secondary DEHP metabolite in neonates.<sup>30</sup> DEHP metabolite concentrations may vary among

**Table 2.** Phthalates and their metabolites (Goodman et al. 2014)<sup>10,43,107,114</sup>

Parent compound	Primary metabolite	Secondary metabolite
Low molecular weight phthalates		
DMP (dimethyl phthalate)	MMP (monomethyl phthalate)	
DEP (diethyl phthalate)	MEP (monoethyl phthalate)	
DPB (di- <i>n</i> -pentyl phthalate)	MnPeP (mono- <i>n</i> -pentyl phthalate)	
BBzP (or BBP) (butyl-benzyl phthalate)	MBzP (monobenzyl phthalate)	
DEEP (or DBP) (di- <i>n</i> -butyl phthalate)	MBP (or MnBP) (mono- <i>n</i> -butyl phthalate)	M CPP (or 3cx-MPP) (mono-(3-carboxypropyl) phthalate) 3OH-MnBP (3OH-mono- <i>n</i> -butyl phthalate)
DiBP (di-isobutyl phthalate)	MiBP (mono-isobutyl phthalate)	2OH-MiBP (2OH-mono-methylpropyl phthalate)
High molecular weight phthalates		
DEHP (di(2-ethylhexyl) phthalate)	MEHP (mono(2-ethylhexyl) phthalate)	MEHHP (or 5OH-MEHP) (mono-(2-ethyl-5-hydroxyhexyl) phthalate) MEOHP (or 5oxo-MEHP) (mono-(2-ethyl-5-oxohexyl) phthalate) MCMHP (or 5cx-MEPP) (mono-(2-carboxymethylhexyl) phthalate) MECPP (mono-(2-ethyl-5-carboxypentyl) phthalate)
DnOP (di- <i>n</i> -octyl phthalate)	MnOP (mono- <i>n</i> -octyl phthalate)	
DiNP (di-isononyl phthalate)	MiNP (mono-isononyl phthalate)	OH-MiNP (7OH-mono-methyloctyl phthalate) oxo-MiNP (7oxo-mono-methyloctyl phthalate) cx-MiDP (7carboxy-mono-methylheptyl phthalate)
DiDP (di-iso-decyl phthalate)		OH-MiDP (6OH-mono-propyl-heptyl phthalate) oxo-MiDP (6oxo-mono-propyl-heptyl phthalate) cx-MiDP (mono-(2,7-methyl-7-carboxy-heptyl) phthalate)

infants. Fluctuations in fluid intake and losses may result in more or less dilute urine volumes among newborns. In addition, a point-in-time urine sample may not reflect the possible diurnal variations in intake and elimination, nor account for differing half-lives of metabolites.<sup>41</sup> Adjusting for urinary dilution by incorporating a creatinine correction is a subject of debate.<sup>42</sup>

Recently, a two-phase study measured 21 urinary DEHP metabolites in mothers and newborns, using non-PVC urine collection bags in a hospital that took steps to minimize exposure of plasticizers to its patients.<sup>43</sup> Newborn metabolite levels were lower in first urine than later urine, and the metabolite pattern for newborns was markedly different than mothers. In newborn urine, the carboxylated metabolites of high molecular weight phthalates DEHP, DiNP and DiDP were found at much higher concentrations than the monoesters of the low molecular weight phthalates compared with pregnant women. The authors concluded that the presence of phthalate metabolites in the first urine of newborns implicated placental transfer.

In summary, evidence to date concerning the pharmacokinetics of phthalates from humans suggests that intersubjective variability among humans is greater than in rodents; hepatic and renal elimination of phthalates may be markedly different at birth and during infancy than during later life due to organ immaturity; accumulation of DEHP in neonates is not known

because studies have focused on analyzing urinary metabolites. Insufficient pharmacokinetic data is available to describe the distribution, metabolism and excretion of phthalates in neonates and infants. Data that characterizes serum levels of phthalates and metabolites in conjunction with excretion should be developed, and steady state serum levels should be assessed for potential toxicity.

#### Iatrogenic exposure to phthalates in neonatal care units

The neonatal care unit environment provides many opportunities for phthalate exposure, for example, IV tubing, feeding and suctioning tubing, plastic containers of IV fluids and medications, ventilation tubing and supplies, surfaces of floors, walls, furniture, blood product containers and infusion systems. DEHP exposures are highest for medical procedures, and PVC medical tubing contains up to 80% DEHP. Leaching rates of DEHP have been studied for banked blood and plasma and are a function of length and temperature of storage.<sup>44</sup> The highest estimated exposure for blood transfusion in neonates was 22.6 mg/kg. Concomitant use of medical products in the neonatal intensive care unit includes hemodialysis, pheresis, nasogastric or IV feeding, extracorporeal membrane oxygenation, IV medication and infusion supplies, and respirator use. On a mg/kg basis, neonates likely receive a much greater dose of DEHP than adults do, and exceed tolerable daily dose recommendations.<sup>45</sup>



Another possible phthalate exposure occurs via IV fat emulsion. IV fat emulsion is a caloric source often provided to neonates in intensive care, and studies have shown that leaching of DEHP from IV tubing into the IV fat emulsion is significant at room temperature and increases with ambient temperatures as might be observed in neonatal incubators.<sup>46,47</sup> Leaching assays of DEHP from PVC IV infusion lines for six different fat emulsion products available in France suggested that the choice of fat emulsion product may be important in minimizing DEHP exposure in the NICU.<sup>48</sup> Some IV perfusion lines have been co-extruded with outer PVC layers and an inner polyurethane (PU) or polyethylene (PE) layer in order to prevent DEHP leaching.<sup>49,50</sup> There was no difference between PVC and PVC/PU lines, while PVC/PE lines leached about half the DEHP of PVC lines. Leaching was proportional to the length of the tubing. Calafat *et al.* studied multiple urine samples from six premature neonates for DEHP metabolites.<sup>51</sup> Even though the levels for the metabolites varied widely among the six newborns, the geometric mean MEHP concentration of 100 ng/ml was considerably higher than for children in the general U.S. population (3.43 ng/ml in 2000).<sup>52</sup> Green *et al.* measured MEHP urinary levels in 54 neonates admitted to an intensive care unit and classified them into three exposure levels based on the estimated amount of exposure to PVC materials: low, medium and high.<sup>44</sup> Median MEHP levels were 4, 28 and 86 ng/ml for the low-, medium- and high-DEHP exposure groups, respectively. After adjustment for institution and sex, MEHP levels were five times greater in the high exposure group compared with the neonates in the low exposure group. In a follow-up report, the investigators measured two additional metabolites of DEHP: MEHHP and MEOHP.<sup>53</sup> Comparing the three exposure level groups using all three metabolites strengthened the association between amount of DEHP exposure with the result that the two additional metabolites in the high-DEHP group's urine was 13–14 times that of infants in the low-DEHP group.

In another study where 58 full term and 67 preterm infants were followed from birth until 14 months, with nine sequential urinary samples obtained,<sup>54</sup> metabolites of BBP, DiNP and DEHP were 5–50 times higher at 7 days and 1 month in preterm compared with full term infants. Median hazard quotients were estimated from TDI recommendations, and at 7 days over 80% of preterm infants exceeded the anti-androgenic threshold, while about 30% of full term and older preterm infants exceeded the threshold during the entire first year of life.

Another avenue of exposure is extracorporeal membrane oxygenation (ECMO) therapy. Karle *et al.* found that plasma DEHP concentrations were greater early in the course of ECMO therapy compared with control, and that most patients cleared DEHP from their plasma before ECMO was discontinued.<sup>55</sup> The mean highest concentration observed at any time was  $8.3 \pm 5.7$  µg/ml or 2 mg/kg. (The RfD for DEHP is 0.02 mg/kg/day.) However, the weakness of the study was that metabolites were not assessed in plasma or urine. These results point to leaching of DEHP from ECMO apparatus but

the undetectable levels in control patients suggest the assay was not sensitive.

Most *in vivo* studies addressing the effects of phthalates utilized exposure levels in the hundreds of mg/kg/day. However, human exposures to phthalates are estimated to be in the µg/kg/day range, and exposure to infants and children are estimated up to three to five-fold higher. In neonatal exchange transfusion, replacement transfusion, and ECMO, DEHP exposure was 1.8 mg/kg/exchange (0.8–3.3), 0.3 mg/kg/transfusion (0.14–0.72) and 2 mg/kg, respectively.<sup>55–57</sup> Levels in children with these exposures exceed the no observed effect level in animal studies. The effects of phthalate exposure in laboratory animals using comparable human exposure doses remain to be studied.

### Exposure to phthalates via breastfeeding and medications

Breastfeeding is an additional means of phthalate exposure for neonates and infants. Breast milk from 62 healthy mothers living in Italy was tested for a number of phthalate metabolites, and mono-isobutyl phthalate and MEHP were found in all samples.<sup>58</sup> MEHP concentrations were two to three times that of urinary samples from the general U.S. population.<sup>52</sup> Mono-*n*-butyl phthalate (MBP) and monobenzyl phthalate (MBzP) were found in 64.5 and 43.5% of samples, respectively. These findings are comparable with that of other countries. Others have also demonstrated that neonates can be exposed to significant amounts of phthalates via breast milk.<sup>59,60</sup> These studies strongly suggest the presence of phthalates in breast milk and establish breast milk as an exposure route to neonates. A follow-up study on these neonates would be informative for possible effects in the neonates. Therefore, future studies should consider prospective outcome studies of newborns, in addition to measurement of presence of phthalate in breast milk. Mothers who are breastfeeding and given phthalate-containing medications may unknowingly pass the phthalates to neonates via breastfeeding. Kelley *et al.* studied the use of polymers in medications and supplements marketed in the United States and Canada based on labeling information.<sup>61</sup> Six prescription drug products included DBP and 45 utilized DEP. Phthalates were found in 75 prescription drug products. Nonprescription and supplement products also included many polymers. Recently, the Food and Drug Administration issued a non-binding guidance document for the pharmaceutical industry urging removal of DBP or DEHP from excipient formulations in medications, but packaging material was exempted from the guidance.<sup>62</sup>

### Exposure to phthalates by direct contact with household products

Another potential source of exposure is by contact with household products. Infants and toddlers are vulnerable because they exhibit more hand-to-mouth activity and consume greater food as a percent of their body weight. A Canadian study evaluated 252 personal care products including 98 baby care products

collected at retail stores in 2007.<sup>63</sup> Of 18 phthalates assessed by GC-MS analysis, DEP, DMP, di-isobutyl phthalate, DnBP and DEHP were detected.

Self-reported use of personal care lotion, cosmetic and cologne/perfume were associated with the greatest increases in urinary phthalates, although the magnitude varied by product.<sup>64</sup> Women using cologne/perfume had monoethyl phthalate (MEP) concentrations 167% higher than non-users. Sathyanarayana *et al.* measured nine phthalate metabolites in 163 infants born from 2000 to 2005.<sup>65</sup> All had been exposed to baby lotion, powder and/or shampoo in the previous 24 h. Metabolites were found in most (81%) infants and higher associations were found in those 8 months of age or less, suggesting sources of exposure may differ before infants begin to crawl and hand-to-mouth activities increase. A limitation of these studies includes a lack of prospective follow-up studies which could have demonstrated the effect of these phthalates. Therefore, future studies should be concentrated on more mechanistic as well as molecular level investigations to discover the strength of association, identify the target molecule or biomarkers for the exposures.

### ***In utero* exposure to phthalates**

Phthalate exposure during pregnancy significantly increases the odds of delivering preterm, which is the leading cause of neonatal mortality.<sup>4</sup> A case-control study in Boston involved 130 mothers who delivered before 37 weeks of gestation and 352 randomly selected mothers who delivered at or after 37 weeks.<sup>4</sup> Maternal levels of DEHP metabolites were associated with increased odds of preterm birth: for summed DEHP metabolites, the odds ratio (OR) of preterm birth was 1.33 (95% CI, 1.04–1.70) and for spontaneous preterm birth the OR was 1.63 (CI, 1.15–2.13). The OR for women in the top quartile was four times greater than women in the bottom quartile, demonstrating a dose-response effect on preterm birth. Another study of 311 African-American and Dominican women from New York City assessed personal air and spot urine samples for DEHP, and also found that gestational age decreased with increasing DEHP exposure.<sup>66</sup> Amniotic fluid contains fetal urine which is swallowed by the fetus which in turn is reabsorbed; thus, amniotic fluid may be representative of phthalate exposure *in utero*. For this reason, Silva *et al.* identified three phthalate metabolites: MEP in 39% of samples, MBP in 93%, and MEHP in 24% in amniotic fluid samples from 54 donors.<sup>67</sup> *In utero* exposure to DEHP or its major metabolite, MEHP, was noted through umbilical cord blood samples in 88% of 84 consecutive newborns in Italy.<sup>68</sup> MEHP-positive newborns showed a lower gestational age compared with MEHP-negative infants. Phthalates were also measured in the cord blood of 207 Chinese women, 33 of whom had preterm delivery before 37 weeks.<sup>69</sup> This study suggested that prenatal exposure to phthalates except dicyclohexyl phthalate (DCHP) is associated with younger gestational age and preterm delivery. Phthalates including dibutyl phthalate (DEHP), DEP, di-n-hexyl phthalate (DNHP), BBP, DNP, DBP, DCHP, DEHP, dipentyl phthalate (DPP) and bis(2-n-butoxyethyl)phthalate (DBEP) also adversely

affected fetal growth parameters via gestational age reduction and preterm delivery with a significant gender effect among various phthalate metabolites.

Furthermore, combined multiple exposures to EDCs are also common during pregnancy.<sup>70</sup> Bisphenol A and other phenols, parabens and phthalates were detected in the urine of 174 of 200 healthy Danish women. Risk assessments that do not account for multiple simultaneous daily exposures may underestimate the total risk burden of EDCs even when individual exposure levels are low, as additive effects have not been studied.

Other investigators have reported no association or a decreased odd of preterm birth with DEHP metabolite exposure.<sup>71–73</sup> These and other studies have notable limitations, such as collecting only single urine samples, self-reported recall of last menstrual period to calculate gestational age at birth and/or small sample sizes.

Several recent reports suggest studying epigenetic modifications associated with gestational age and preterm birth.<sup>74,75</sup> To our knowledge there is no published mechanistic study correlating DEHP or any other phthalates with human epigenetics and pre-term birth or gestational age. We recently showed that MEHP induced epigenetic changes in a human first trimester placental cell line.<sup>76</sup>

### **Effects on the reproduction system**

Phthalate exposure may affect male offspring to a greater degree than female. A systematic review<sup>77</sup> of epidemiological and experimental animal literature examined the relationship between phthalate exposure and adverse female reproductive health outcomes. The authors concluded that phthalates may not significantly affect the female reproductive system of women exposed to low levels. On the other hand, the safety of exposures to higher levels of phthalates or in conjunction with other endocrine disruptors is not known.

A recent review of molecular mechanisms of action of EDCs indicates the complexity of ligand-activated nuclear receptor transcription in the inappropriate modulation of hormone receptors.<sup>78</sup> There are numerous potential targets for EDC disruption of hormone signaling, and some EDCs may have binding affinities or interactions with hormone receptors besides estrogen receptors, leading to complexities in understanding the entirety of their effects. However, evidence of epigenetic regulation are mostly unrecognized.

Short- and long-term antiandrogenic effects of DEHP after *in utero* exposure have been demonstrated in several animal models as well as humans. Exposure to a wide range of DEHP doses in pregnant rats resulted in increased volumes of Leydig cells in adult testes, but dose-dependent reductions in testosterone production were observed.<sup>79</sup> Increased rather than decreased expression of steroidogenesis-related genes was noted, so the cause of adult testosterone expression in rats exposed to higher doses of DEHP remains unknown. Bustamante-Montes *et al.* studied the association between maternal phthalate exposure and male offspring anthropometric and genital measurements in 74 male newborns from women who provided urine samples at their last

prenatal visit.<sup>80</sup> Significant inverse associations were observed between prenatal total phthalate exposure and the distance from the anus to anterior base of the penis, penile width and stretched length. Specifically, prenatal exposure to MEHP was associated with a reduction in the stretched length of the penis. Albert *et al.* reported the number of studies of phthalate effects on the testis of animals and humans, noting 25 human studies compared with 265 studies in mice, rats and marmosets.<sup>10</sup>

It has been proposed that male reproductive disorders such as cryptorchidism, hypospadias, infertility and testicular cancer are linked to a condition known as testicular dysgenesis syndrome (TDS) resulting from EDC exposure *in utero*.<sup>13,81</sup> Several epidemiological studies from different countries suggest a possible effect of environmental EDCs in TDS pathogenesis including phthalates.<sup>82</sup> In addition, TDS induced in mice with maternal exposure to DEHP demonstrated a relative increase in global DNA methylation and increased DNA methyltransferase expression, suggesting that epigenetic modifications may play a role in TDS.<sup>83</sup> DNA methyltransferase expression returned to normal levels in adulthood, although DEHP exposure was discontinued at birth.<sup>84</sup> However, testosterone suppression did not recover. Other reports have corroborated the antiandrogenic effects of fetal phthalate exposure.<sup>78</sup> Gonadal development was studied in four generations of rats whose mothers were gavaged during pregnancy with DEHP.<sup>85</sup> Cryptorchidism was noted in the F1 and F2 offspring but not F3 or F4, while conception rates were 50, 75 and 100%, respectively, for F1, F2, and F3 and F4 offspring. DNA methyltransferase expression was upregulated with each successive generation. Two studies of male infants in the United States and Japan observed decreased anogenital distance with maternal DEHP exposure.<sup>86,87</sup> These findings were consistent with a study of male rat pups exposed to phthalates *in utero*.<sup>88</sup> In the latter study, DEHP and DBP maternal exposure decreased testosterone production and insulin-like peptide 3 (*insl3*) gene expression as well as *cyp11a*. The peptide *insl3* facilitates the first phase of testicular descent, while *cyp11a* is a rate-limiting enzyme responsible for conversion of cholesterol to pregnenolone in steroidogenesis. In addition, the Howdeshell study noted that fetal/neonatal mortality was significantly increased particularly when DBP and DEHP were combined. In a case-control study in France, the OR of male genital malformation in infants with parental exposure to pesticides was 4.41 (95% CI, 1.21–16.00).<sup>89</sup> Results of a recent large prospective cohort study of pregnant women and their adolescent daughters suggested that antenatal exposure to phthalates may be associated with long-term effects on reproductive development, including a significantly increased uterine volume and a potentially protective effect against polycystic ovarian syndrome in girls.<sup>90</sup> Recently, Mouritsen *et al.* carried out a longitudinal study where children were examined every 6 months for serum levels of dehydroepiandrosterone (DHEA),  $\Delta$ 4-androstenedione, testosterone, and urinary morning excretion of 14 phthalate metabolites to determine whether urinary phthalate levels are associated with

circulating androgen levels and age at puberty.<sup>91</sup> Girls in puberty exhibited lower serum levels of adrenal androgens in those who excreted the highest amount of MBP and DEHP metabolites; these girls were most exposed to DBP and DEHP. In contrast, boys in the high MBP excretion group developed pubic hair almost a year earlier than boys in the low group. Testosterone level at age 13 was higher in boys who excreted the highest amount of MBzP, while DHEA levels were lower.

### Epigenetic studies of EDCs and phthalates

Epidemiological studies suggest that scientists should investigate the effects of phthalates at the molecular mechanistic level in *in vitro* or *in vivo*. To date, very few studies have examined epigenetic effects of phthalates in humans or even in experimental animals or cells. Besides previously described antiandrogenic effects of phthalates, anti-aldosterogenic effects of DEHP exposure have been observed in the decreased expression of mineralocorticoid receptor (MR) in adult Leydig cells of rats which may be epigenetically mediated.<sup>92</sup> A follow-up study of *in utero* DEHP exposure in rats found significant sex-specific long-term effects in steroid levels in response to DEHP exposure at concentrations close to or higher than human exposures observed with hemodialysis or total parenteral nutrition in infants.<sup>93</sup> The results suggest that *in utero* DEHP exposure reduces both adrenal aldosterone synthesis and MR expression in Leydig cells. The authors note that ‘this is the first evidence showing that *in utero* exposure to DEHP has cardiovascular and behavioral effects in the adult male offspring.’ These investigators recently performed global gene expression analysis of pre-pubertal and adult rat adrenal glands following long-term *in utero* exposure to DEHP.<sup>94</sup> They also observed postnatal alteration in DNA methylation in nuclear receptor genes in Leydig cells, including estrogen receptor  $\beta$ , thyroid receptor  $\beta$ , peroxisome proliferator-activated receptor  $\alpha$  and the mineralocorticoid receptor.<sup>95</sup> *In utero* exposure to DEHP resulted in long-term activation of the adrenal gland and reduced aldosterone synthesis. In addition to adverse reproductive effect, DEHP also increased lipid import, cholesterol biosynthesis and cholesterol storage. Treatment of human breast cancer cells (MCF7) with BBP resulted in demethylation of estrogen receptor-1 (ESR1) promoter-associated CpG islands, suggesting that altered *ESR1* mRNA expression by BBP is related to DNA hypomethylation in the promoter region of the receptor gene.<sup>96</sup> Maternal exposure to DEHP increased DNA methylation in the mouse testis with increased expression of DNA methyltransferases leading to TDS.<sup>84</sup> Another recent study was designed to assess the effects of DEHP on DNA methylation of imprinting genes in germ cells from fetal and adult mice.<sup>97</sup> DEHP exposure significantly reduced the percentage of methylated CpG sites in *Igf2r* and *Peg3* differentially methylated regions in germ cells, particularly in postpartum day 21 oocytes. The modification was inherited in offspring.

As people are exposed to various phthalates other than DEHP, it is possible that some may antagonize or amplify the effect of DEHP in actual exposure situations. However, DEHP



or MEHP may act in other ways as suggested by several studies.<sup>98–103</sup> A recent report suggested apparent biphasic effects of pollutants with high fat high sucrose diet in an obesity model.<sup>104</sup> Mannikam *et al.* studied a mixture of plastic constituents (bisphenol-A, DEHP and DBP) on pubertal and endocrine abnormalities in F1 and F3 generation male and female rats, and found results supporting transgenerational inheritance of adult onset disease.<sup>105</sup> Doses used were considered high in relation to human exposures.

One well-known early example of the developmental origins of health and disease (DOHaD) concept related to xenobiotic exposure is linked to epigenetic theory. In brief, the first synthetic estrogenic drug, diethylstilbestrol (DES) was given to an estimated 10 million mothers from 1940 to 1971, and increased the risk for breast cancer in both mothers and their daughters when they were older than 40 years.<sup>106</sup> In this review, Newbold noted that DES also was linked to vaginal adenocarcinoma in a few adolescent daughters, as well as frequent reproductive problems in 90–95% of daughters. Male offspring also developed a range of male reproductive tract problems.<sup>106</sup> Newbold argues that the transgenerational as well as delayed effect of diethylstilbestrol is explained by epigenetic programming, and that grandchildren of the original mothers should be studied today. This early tragedy suggests that the epigenome is extremely vulnerable to disruption by environmental factors including estrogenic chemicals such as phthalates during prenatal and perinatal life.

Even though EDC exposure during critical periods of early development and risk of chronic diseases later in life has been reported in a number of recent studies, the mechanistic knowledge is still in infancy. Studies of the epigenetic effects of DEHP are still warranted. Given the potential reversibility of environmentally induced epigenetic modifications, mechanisms underlying the lifelong as well as transgenerational consequences of perinatal endocrine disruption may ultimately lead to the development of efficient diagnostic tools and therapeutic approaches for the prevention and mitigation of adverse effects of EDCs, in particular DEHP.

## Discussion

An expert panel critically reviewed available studies and issued their report in 2006 concerning the reproductive and developmental toxicity of DEHP.<sup>107</sup> This report indicated minimal concern for the general population; some concern for male children older than 1 year; concern for infants <1 year; serious concerns for intensively medically treated infants; and some concern for effects of DEHP on male children exposed to general population levels during pregnancy. Finally, the panel identified a number of specific data needs for further investigation: sexual development in adolescents previously exposed to ECMO treatment; studies with larger numbers of neonates relating the nature of procedures to exposure levels using measures of internal dose with multiple metabolites; additional studies on prenatal, perinatal and postnatal exposure

on developmental reproductive toxicity. In addition, the panel suggested that physiologically based pharmacokinetic studies remain one of the most important data needs.

In addition, clinicians should maintain awareness of phthalate-containing products used in the medical care of pregnant and lactating women, and in the environment of neonatal intensive care settings. We speculate that substituting non-DEHP medical products for PVC products wherever possible will reduce exposure levels. We recommend if IV fat emulsions are used, selection of a soybean-based product will reduce the exposure, also avoid excessive warming and agitation during their storage and use. Avoid using vinyl flooring and avoid foods packaged or stored in plastics if alternatives (DEHP-free PVC, non-PVC or bio-based plastics) are available.<sup>108</sup> However, these recommendations should be strengthened with more research evidence about these indoor environmental exposures. Some resources have been made available to help clinicians identify alternatives to PVC medical devices for the NICU and other patients.<sup>109–111</sup>

In conclusion, several studies have demonstrated clear differences in the ways that mice, rats, primates, and humans respond to phthalate exposures. Most of the *in vivo* studies addressing the effects of phthalates utilized exposure levels in the hundreds of mg/kg/day. However, human exposures to phthalates are estimated to be in the µg/kg/day range and exposures to infants and children are estimated up to three to five-fold higher. Additional experiments with a more realistic human exposure dose in laboratory animals are warranted. In addition, humans are exposed to several mixtures of phthalates and other EDCs besides DEHP, and it is possible that other compounds may dampen or amplify the effect of DEHP in actual exposure situations. Such studies should be considered by investigators. A recent critical review of endocrine disruption on the human testis to phthalates from fetal life to adulthood provides several caveats.<sup>10</sup> The authors state ‘it cannot be excluded that transgenerational effects of phthalates and/or epigenetic changes exist in humans.’ Yet it does appear that the effects persist in adulthood. Accumulating evidence indicates the involvement of epigenetic regulation which can be inherited. Thus, the evidence to date supports the DOHaD hypothesis. In 2002 the European Union Scientific Committee on Medicinal Products and Medical Devices published an opinion that the net advantages of using DEHP in medical devices outweighed the disadvantages.<sup>112</sup> However, continued concern by the European Union about DEHP safety resulted in the identification of potential alternative plasticizers to DEHP for use in PVC medical products.<sup>113</sup> More mechanistic studies are warranted to support the epigenetic hypothesis and increase awareness of potential transgenerational effects.

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## Conflicts of Interest

None.

## References

- Gluck L. *Conceptualization and Initiation of a Neonatal Intensive Care Nursery in 1960*. 1992, National Institutes of Health: Bethesda, MD.
- Jorgensen A. Born in the USA – the history of neonatology in the United States: a century of caring, Abbot Nutrition Health Institute, 2010. Retrieved 8 January 2014 from <http://anhi.org/articles/the-history-of-neonatology-in-the-united-states-a-century-of-caring>.
- Mallow EB, Fox MA. Phthalates and critically ill neonates: device-related exposures and non-endocrine toxic risks. *J Perinatol*. 2014; 34, 892–897.
- Ferguson KK, McElrath TF, Meeker JD. Environmental phthalate exposure and preterm birth. *JAMA Pediatr*. 2014; 168, 61–67.
- Shen Q, Shi H, Zhang Y, Cao Y. Dietary intake and phthalates body burden in boys and girls. *Arch Public Health*. 2015; 73, 5.
- Anway MD, Rekow SS, Skinner MK. Transgenerational epigenetic programming of the embryonic testis transcriptome. *Genomics*. 2008; 91, 30–40.
- Perera F, Herbstman J. Prenatal environmental exposures, epigenetics, and disease. *Reprod Toxicol*. 2011; 31, 363–373.
- Waring RH, Harris RM, Mitchell SC. In utero exposure to carcinogens: epigenetics, developmental disruption and consequences in later life. *Maturitas*. 2016; 86, 59–63.
- Woodruff TJ. Making it real – the environmental burden of disease. What does it take to make people pay attention to the environment and health? *J Clin Endocrinol Metab*. 2015; 100, 1241–1244.
- Albert O, Jégou B. A critical assessment of the endocrine susceptibility of the human testis to phthalates from fetal life to adulthood. *Hum Reprod Update*. 2014; 20, 231–249.
- Berge A, Cladiere M, Gasperi J, et al. Meta-analysis of environmental contamination by phthalates. *Environ Sci Pollut Res*. 2013; 20, 8057–8076.
- Schettler T. Human exposure to phthalates via consumer products. *Int J Androl*. 2006; 29, 134–139, discussion 181–185.
- Meeker JD, Sathyanarayana S, Swan SH. Phthalates and other additives in plastics: human exposure and associated health outcomes. *Philos Trans R Soc Lond B Biol Sci*. 2009; 364, 2097–2113.
- Latini G, Ferri M, Chiellini F. Materials degradation in PVC medical devices, DEHP leaching and neonatal outcomes. *Curr Med Chem*. 2010; 17, 2979–2989.
- Trasande L, Attina TM. Association of exposure to di-2-ethylhexylphthalate replacements with increased blood pressure in children and adolescents. *Hypertension*. 2015; 66, 301–308.
- Mankidy R, Wiseman S, Ma H, Giesy JP. Biological impact of phthalates. *Toxicol Lett*. 2013; 217, 50–58.
- Jaeger RJ, Rubin RJ. Contamination of blood stored in plastic packs. *Lancet*. 1970; 2, 151.
- Mayer FL, Stalling DL, Johnson JL. Phthalate esters as environmental contaminants. *Nature*. 1972; 238, 411–413.
- Rozek LS, Dolinoy DC, Sartor MA, Omenn GS. Epigenetics: relevance and implications for public health. *Annu Rev Public Health*. 2014; 35, 105–122.
- Singh S, Li SS. Epigenetic effects of environmental chemicals bisphenol a and phthalates. *Int J Mol Sci*. 2012; 13, 10143–10153.
- Skinner MK, Manikkam M, Guerrero-Bosagna C. Epigenetic transgenerational actions of environmental factors in disease etiology. *Trends Endocrinol Metab*. 2010; 21, 214–222.
- Agency for Toxic Substances and Disease Registry. *Toxicological Profile for Di(2-Ethylhexyl) Phthalate*. 2002. Agency for Toxic Substances and Disease Registry: Atlanta, GA.
- Tomita I, Nakamura Y, Yagi Y, Tutikawa K. Teratogenicity/fetotoxicity of DEHP in mice. *Environ Health Perspect*. 1982; 45, 71–75.
- Rubin RJ, Ness PM. What price progress? An update on vinyl plastic bags. *Transfusion (Paris)*. 1989; 29, 358–361.
- Scientific Panel on Food Additives, Flavours, Processing Aids and Materials in Contact with Food. (Packaging) EFFIA. Bis (2-ethylhexyl)phthalate (DEHP) for use in food contact materials. *EFSA J*. 2005; 243, 1–20. doi:10.2903/j.efsa.2005.243.
- EPA. Basic information about di(2-ethylhexyl) phthalate in drinking water. US Environmental Protection Agency, 2014. Retrieved 23 June 2014 from [http://water.epa.gov/drink/contaminants/basicinformation/di\\_2-ethylhexyl\\_phthalate.cfm](http://water.epa.gov/drink/contaminants/basicinformation/di_2-ethylhexyl_phthalate.cfm).
- NTP (National Toxicology Program). *Report on Carcinogens*, 13th edn, 2014. National Toxicology Program: Research Triangle Park, NC.
- Nachman RM, Hartle JC, Lees PSJ, Groopman JD. Early life metabolism of bisphenol A: a systematic review of the literature. *Curr Environ Health Rep*. 2014; 1, 90–100.
- Beko G, Weschler CJ, Langer S, et al. Children's phthalate intakes and resultant cumulative exposures estimated from urine compared with estimates from dust ingestion, inhalation and dermal absorption in their homes and daycare centers. *PLoS One*. 2013; 8, e62442.
- Koch HM, Preuss R, Angerer J. Di(2-ethylhexyl)phthalate (DEHP): human metabolism and internal exposure – an update and latest results. *Int J Androl*. 2006; 29, 155–165, discussion 181–185.
- Cuzzolin L. Drug metabolizing enzymes in the perinatal and neonatal period: differences in the expression and activity. *Curr Drug Metab*. 2013; 14, 167–173.
- Dotta A, Chukhlantseva N. Ontogeny and drug metabolism in newborns. *J Matern Fetal Neonatal Med*. 2012; 25(Suppl. 4), 83–84.
- Choi K, Joo H, Campbell JL, Jr, et al. In vitro metabolism of di(2-ethylhexyl) phthalate (DEHP) by various tissues and cytochrome P450s of human and rat. *Toxicol In Vitro*. 2012; 26, 315–322.
- Van Overmeire B, Touw D, Schepens PJ, Kearns GL, van den Anker JN. Ibuprofen pharmacokinetics in preterm infants with patent ductus arteriosus. *Clin Pharmacol Ther*. 2001; 70, 336–343.
- Sharma PK, Garg SK, Narang A. Pharmacokinetics of oral ibuprofen in premature infants. *J Clin Pharmacol*. 2003; 43, 968–973.
- Ito Y, Kamijima M, Hasegawa C, et al. Species and inter-individual differences in metabolic capacity of di(2-ethylhexyl) phthalate (DEHP) between human and mouse livers. *Environ Health Prev Med*. 2014; 19, 117–125.
- Koch HM, Rossbach B, Drexler H, Angerer J. Internal exposure of the general population to DEHP and other phthalates – determination of secondary and primary phthalate monoester metabolites in urine. *Environ Res*. 2003; 93, 177–185.

38. Ligi I, Boubred F, Grandvullemin I, Simeoni U. The neonatal kidney: implications for drug metabolism and elimination. *Curr Drug Metab.* 2013; 14, 174–177.
39. Chen N, Aleksa K, Woodland C, Rieder M, Koren G. Ontogeny of drug elimination by the human kidney. *Pediatr Nephrol.* 2006; 21, 160–168.
40. Levey AS, Inker LA, Coresh J. GFR estimation: from physiology to public health. *Am J Kidney Dis.* 2014; 63, 820–834.
41. Lorber M, Koch HM, Angerer J. A critical evaluation of the creatinine correction approach: can it underestimate intakes of phthalates? A case study with di-2-ethylhexyl phthalate. *J Expo Sci Environ Epidemiol.* 2011; 21, 576–586.
42. Lorber M, Angerer J, Koch HM. A simple pharmacokinetic model to characterize exposure of Americans to Di-2-ethylhexyl phthalate. *J Expos Sci Environ Epidemiol.* 2009; 20, 38–53.
43. Enke U, Schlessner E, Palmke C, Seyfarth L, Koch HM. Phthalate exposure in pregnant women and newborns – the urinary metabolite excretion pattern differs distinctly. *Int J Hyg Environ Health.* 2013; 216, 735–742.
44. Green R, Hauser R, Calafat AM, et al. Use of di(2-ethylhexyl) phthalate-containing medical products and urinary levels of mono (2-ethylhexyl) phthalate in neonatal intensive care unit infants. *Environ Health Perspect.* 2005; 113, 1222–1225.
45. Latini G, De Felice C, Verrotti A. Plasticizers, infant nutrition and reproductive health. *Reprod Toxicol.* 2004; 19, 27–33.
46. Loff S, Kabs F, Subotic U, et al. Kinetics of diethylhexyl-phthalate extraction from polyvinylchloride-infusion lines. *JPEN J Parenter Enteral Nutr.* 2002; 26, 305–309.
47. Rose RJ, Priston MJ, Rigby-Jones AE, Sneyd JR. The effect of temperature on di(2-ethylhexyl) phthalate leaching from PVC infusion sets exposed to lipid emulsions. *Anaesthesia.* 2012; 67, 514–520.
48. Bagel S, Dessaigne B, Bourdeaux D, et al. Influence of lipid type on bis (2-ethylhexyl)phthalate (DEHP) leaching from infusion line sets in parenteral nutrition. *JPEN J Parenter Enteral Nutr.* 2011; 35, 770–775.
49. Loff S, Subotic U, Reinicke F, Wischmann H, Brade J. Extraction of di-ethylhexyl-phthalate from perfusion lines of various material, length and brand by lipid emulsions. *J Pediatr Gastroenterol Nutr.* 2004; 39, 341–345.
50. Bourdeaux D, Sautou-Miranda V, Bagel-Boithias S, Boyer A, Chopineau J. Analysis by liquid chromatography and infrared spectrometry of di(2-ethylhexyl)phthalate released by multilayer infusion tubing. *J Pharm Biomed Anal.* 2004; 35, 57–64.
51. Calafat AM, Needham LL, Silva MJ, Lambert G. Exposure to di-(2-ethylhexyl) phthalate among premature neonates in a neonatal intensive care unit. *Pediatrics.* 2004; 113, e429–e434.
52. Centers for Disease Control. *Second National Report on Human Exposure to Environmental Chemicals.* 2003. Centers for Disease Control and Prevention: Atlanta, GA.
53. Weuve J, Sanchez BN, Calafat AM, et al. Exposure to phthalates in neonatal intensive care unit infants: urinary concentrations of monoesters and oxidative metabolites. *Environ Health Perspect.* 2006; 114, 1424–1431.
54. Frederiksen H, Kuiri-Hanninen T, Main KM, Dunkel L, Sankilampi U. A longitudinal study of urinary phthalate excretion in 58 full-term and 67 preterm infants from birth through 14 months. *Environ Health Perspect.* 2014; 122, 998–1005. doi:10.1289/ehp.1307569.
55. Karle VA, Short BL, Martin GR, et al. Extracorporeal membrane oxygenation exposes infants to the plasticizer, di(2-ethylhexyl) phthalate. *Crit Care Med.* 1997; 25, 696–703.
56. Sjoberg P, Bondesson U, Sedin G, Gustafsson J. Dispositions of di- and mono-(2-ethylhexyl) phthalate in newborn infants subjected to exchange transfusions. *Eur J Clin Invest.* 1985; 15, 430–436.
57. Sjoberg PO, Bondesson UG, Sedin EG, Gustafsson JP. Exposure of newborn infants to plasticizers. Plasma levels of di-(2-ethylhexyl) phthalate and mono-(2-ethylhexyl) phthalate during exchange transfusion. *Transfusion (Paris).* 1985; 25, 424–428.
58. Latini G, Wittassek M, Del Vecchio A, et al. Lactational exposure to phthalates in Southern Italy. *Environ Int.* 2009; 35, 236–239.
59. Calafat AM, Slakman AR, Silva MJ, Herbert AR, Needham LL. Automated solid phase extraction and quantitative analysis of human milk for 13 phthalate metabolites. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2004; 805, 49–56.
60. Mortensen GK, Main KM, Andersson AM, Leffers H, Skakkebaek NE. Determination of phthalate monoesters in human milk, consumer milk, and infant formula by tandem mass spectrometry (LC-MS-MS). *Anal Bioanal Chem.* 2005; 382, 1084–1092.
61. Kelley KE, Hernandez-Diaz S, Chaplin EL, Hauser R, Mitchell AA. Identification of phthalates in medications and dietary supplement formulations in the United States and Canada. *Environ Health Perspect.* 2012; 120, 379–384.
62. FDA. Guidance for industry: limiting the use of certain phthalates as excipients in CDER-regulated products. US Department of Health and Human Services, 2012. Retrieved 22 June 2014 from <http://www.fda.gov/downloads/Drugs/GuidanceComplianceRegulatoryInformation/Guidances/UCM294086.pdf?source=govdelivery>.
63. Koniecki D, Wang R, Moody RP, Zhu J. Phthalates in cosmetic and personal care products: concentrations and possible dermal exposure. *Environ Res.* 2011; 111, 329–336.
64. Braun JM, Just AC, Williams PL, et al. Personal care product use and urinary phthalate metabolite and paraben concentrations during pregnancy among women from a fertility clinic. *J Expo Sci Environ Epidemiol.* 2014; 24, 459–466.
65. Sathyanarayana S, Karr CJ, Lozano P, et al. Baby care products: possible sources of infant phthalate exposure. *Pediatrics.* 2008; 121, e260–e268.
66. Whyatt RM, Adibi JJ, Calafat AM, et al. Prenatal di(2-ethylhexyl) phthalate exposure and length of gestation among an inner-city cohort. *Pediatrics.* 2009; 124, e1213–e1220.
67. Silva MJ, Reidy JA, Herbert AR, et al. Detection of phthalate metabolites in human amniotic fluid. *Bull Environ Contam Toxicol.* 2004; 72, 1226–1231.
68. Latini G, De Felice C, Presta G, et al. In utero exposure to di-(2-ethylhexyl)phthalate and duration of human pregnancy. *Environ Health Perspect.* 2003; 111, 1783–1785.
69. Huang Y, Li J, Garcia JM, et al. Phthalate levels in cord blood are associated with preterm delivery and fetal growth parameters in Chinese women. *PLoS One.* 2014; 13, e87430.
70. Tefre de Renzy-Martin K, Frederiksen H, Christensen JS, et al. Current exposure of 200 pregnant Danish women to phthalates, parabens and phenols. *Reproduction.* 2014; 147, 443–453.

71. Adibi JJ, Hauser R, Williams PL, *et al.* Maternal urinary metabolites of di-(2-ethylhexyl) phthalate in relation to the timing of labor in a US multicenter pregnancy cohort study. *Am J Epidemiol.* 2009; 169, 1015–1024.
72. Wolff MS, Engel SM, Berkowitz GS, *et al.* Prenatal phenol and phthalate exposures and birth outcomes. *Environ Health Perspect.* 2008; 116, 1092–1097.
73. Suzuki Y, Niwa M, Yoshinaga J, *et al.* Prenatal exposure to phthalate esters and PAHs and birth outcomes. *Environ Int.* 2010; 36, 699–704.
74. Parets SE, Bedient CE, Menon R, Smith AK. Preterm birth and its long-term effects: methylation to mechanisms. *Biology (Basel).* 2014; 3, 498–513.
75. Gao F, Zhang J, Jiang P, *et al.* Marked methylation changes in intestinal genes during the perinatal period of preterm neonates. *BMC Genomics.* 2014; 15, 716.
76. Meruvu S, Zhang J, Bedi YS, Choudhury M. Mono-(2-ethylhexyl) phthalate induces apoptosis through miR-16 in human first trimester placental cell line HTR-8/SVneo. *Toxicol In Vitro.* 2016; 31, 35–42.
77. Kay VR, Chambers C, Foster WG. Reproductive and developmental effects of phthalate diesters in females. *Crit Rev Toxicol.* 2013; 43, 200–219.
78. Yoon K, Kwack SJ, Kim HS, Lee BM. Estrogenic endocrine-disrupting chemicals: molecular mechanisms of actions on putative human diseases. *J Toxicol Environ Health B Crit Rev.* 2014; 17, 127–174.
79. Culty M, Thuillier R, Li W, *et al.* In utero exposure to di-(2-ethylhexyl) phthalate exerts both short-term and long-lasting suppressive effects on testosterone production in the rat. *Biol Reprod.* 2008; 78, 1018–1028.
80. Bustamante-Montes LP, Hernandez-Valero MA, Flores-Pimentel D, *et al.* Prenatal exposure to phthalates is associated with decreased anogenital distance and penile size in male newborns. *J Dev Orig Health Dis.* 2013; 4, 300–306.
81. Fisher JS, Macpherson S, Marchetti N, Sharpe RM. Human ‘testicular dysgenesis syndrome’: a possible model using in-utero exposure of the rat to dibutyl phthalate. *Hum Reprod.* 2003; 18, 1383–1394.
82. Olesen IA, Sonne SB, Hoei-Hansen CE, Rajpert-De Meyts E, Skakkebaek NE. Environment, testicular dysgenesis and carcinoma in situ testis. *Best Pract Res Clin Endocrinol Metab.* 2007; 21, 462–478.
83. Wu S, Zhu J, Li Y, *et al.* Dynamic epigenetic changes involved in testicular toxicity induced by di-2-(ethylhexyl) phthalate in mice. *Basic Clin Pharmacol Toxicol.* 2010; 106, 118–123.
84. Wu S, Zhu J, Li Y, *et al.* Dynamic effect of di-2-(ethylhexyl) phthalate on testicular toxicity: epigenetic changes and their impact on gene expression. *Int J Toxicol.* 2010; 29, 193–200.
85. Chen J, Wu S, Wen S, *et al.* The mechanism of environmental endocrine disruptors (DEHP) induces epigenetic transgenerational inheritance of cryptorchidism. *PLoS One.* 2015; 10, e0126403.
86. Swan SH, Main KM, Liu F, *et al.* Decrease in anogenital distance among male infants with prenatal phthalate exposure. *Environ Health Perspect.* 2005; 113, 1056–1061.
87. Suzuki Y, Yoshinaga J, Mizumoto Y, Serizawa S, Shiraishi H. Foetal exposure to phthalate esters and anogenital distance in male newborns. *Int J Androl.* 2012; 35, 236–244.
88. Howdeshell KL, Furr J, Lambright CR, *et al.* Cumulative effects of dibutyl phthalate and diethylhexyl phthalate on male rat reproductive tract development: altered fetal steroid hormones and genes. *Toxicol Sci.* 2007; 99, 190–202.
89. Gaspari L, Paris F, Jandel C, *et al.* Prenatal environmental risk factors for genital malformations in a population of 1442 French male newborns: a nested case-control study. *Hum Reprod.* 2011; 26, 3155–3162.
90. Hart R, Doherty DA, Frederiksen H, *et al.* The influence of antenatal exposure to phthalates on subsequent female reproductive development in adolescence: a pilot study. *Reproduction.* 2013; 147, 379–390. doi:10.1530/REP-13-0331.
91. Mouritsen A, Frederiksen H, Sorensen K, *et al.* Urinary phthalates from 168 girls and boys measured twice a year during a 5-year period: associations with adrenal androgen levels and puberty. *J Clin Endocrinol Metab.* 2013; 98, 3755–3764.
92. Martinez-Arguelles DB, Culty M, Zirkin BR, Papadopoulos V. In utero exposure to di-(2-ethylhexyl) phthalate decreases mineralocorticoid receptor expression in the adult testis. *Endocrinology.* 2009; 150, 5575–5585.
93. Martinez-Arguelles DB, Guichard T, Culty M, Zirkin BR, Papadopoulos V. In utero exposure to the antiandrogen di-(2-ethylhexyl) phthalate decreases adrenal aldosterone production in the adult rat. *Biol Reprod.* 2011; 85, 51–61.
94. Martinez-Arguelles D, Campioli E, Lienhart C, *et al.* In utero exposure to the endocrine disruptor di-(2-ethylhexyl) phthalate induces long-term changes in gene expression in the adult male adrenal gland. *Endocrinology.* 2014; 155, 1667–1678. doi:10.1210/en.2013-1921.
95. Martinez-Arguelles DB, Campioli E, Culty M, Zirkin BR, Papadopoulos V. Fetal origin of endocrine dysfunction in the adult: the phthalate model. *J Steroid Biochem Mol Biol.* 2013; 137, 5–17.
96. Kang SC, Lee BM. DNA methylation of estrogen receptor alpha gene by phthalates. *J Toxicol Environ Health A.* 2005; 68, 1995–2003.
97. Li L, Zhang T, Qin XS, *et al.* Exposure to diethylhexyl phthalate (DEHP) results in a heritable modification of imprint genes DNA methylation in mouse oocytes. *Molecular Biology Reports.* 2014; 41, 1227–1235.
98. Gupta RK, Singh JM, Leslie TC, *et al.* Di-(2-ethylhexyl) phthalate and mono-(2-ethylhexyl) phthalate inhibit growth and reduce estradiol levels of antral follicles in vitro. *Toxicol Appl Pharmacol.* 2010; 242, 224–230.
99. Thomas JA, Curto KA, Thomas MJ. MEHP/DEHP: gonadal toxicity and effects on rodent accessory sex organs. *Environ Health Perspect.* 1982; 45, 85–88.
100. Erkekoglu P, Rachidi W, Yuzugullu OG, *et al.* Evaluation of cytotoxicity and oxidative DNA damaging effects of di(2-ethylhexyl)-phthalate (DEHP) and mono(2-ethylhexyl)-phthalate (MEHP) on MA-10 Leydig cells and protection by selenium. *Toxicol Appl Pharmacol.* 2010; 248, 52–62.
101. Rose ML, Rivera CA, Bradford BU, *et al.* Kupffer cell oxidant production is central to the mechanism of peroxisome proliferators. *Carcinogenesis.* 1999; 20, 27–33.
102. Wan X, Zhu Y, Ma X, *et al.* Effect of DEHP and its metabolite MEHP on in vitro rat follicular development. *Wei Sheng Yan Jiu.* 2010; 39, 268–270, 274.



103. Sjoberg P, Bondesson U, Gray TJ, Ploen L. Effects of di-(2-ethylhexyl) phthalate and five of its metabolites on rat testis in vivo and in vitro. *Acta Pharmacol Toxicol (Copenh)*. 1986; 58, 225–233.
104. Naville D, Labaronne E, Vega N, *et al*. Metabolic outcome of female mice exposed to a mixture of low-dose pollutants in a diet-induced obesity model. *PLoS One*. 2015; 10, e0124015.
105. Manikkam M, Tracey R, Guerrero-Bosagna C, Skinner MK. Plastics derived endocrine disruptors (BPA, DEHP and DBP) induce epigenetic transgenerational inheritance of obesity, reproductive disease and sperm epimutations. *PLoS One*. 2013; 8, e55387.
106. Newbold RR. Prenatal exposure to diethylstilbestrol and long-term impact on the breast and reproductive tract in humans and mice. *J Dev Orig Health Dis*. 2012; 3, 73–82.
107. Kavlock R, Barr D, Boekelheide K, *et al*. NTP-CERHR expert panel update on the reproductive and developmental toxicity of di(2-ethylhexyl) phthalate. *Reprod Toxicol*. 2006; 22, 291–399.
108. Braun JM, Sathyanarayana S, Hauser R. Phthalate exposure and children's health. *Curr Opin Pediatr*. 2013; 25, 247–254.
109. Health Care Without Harm. Alternatives to polyvinyl chloride (PVC) medical devices for the Neonatal Intensive Care Unit (NICU), Reston, VA, 2006. Health Care Without Harm. Retrieved 1 July 2014 from <https://noharm-uscanada.org/documents/alternatives-polyvinyl-chloride-pvc-medical-devices-neonatal-intensive-care-unit-nicu>.
110. Health Care Without Harm. Alternatives to polyvinyl chloride (PVC) and di(2-ethylhexyl) phthalate (DEHP) medical devices, Reston, VA, 2008. Health Care Without Harm. Retrieved 1 July 2014 from <https://noharm-uscanada.org/documents/alternatives-polyvinyl-chloride-pvc-and-di2-ethylhexyl-phthalate-dehp-medical-devices>.
111. Pediatric Environmental Health Specialty Units. Resources for health professionals. Pediatric Environmental Health Specialty Units, Washington, DC, March 2014. Retrieved 2 July 2014 from [http://www.pehsu.net/\\_Phthalates\\_and\\_Bisphenol\\_A\\_Advisory.html](http://www.pehsu.net/_Phthalates_and_Bisphenol_A_Advisory.html)
112. The Scientific Committee on medicinal products and medical devices. Opinion on medical devices containing DEHP plasticised PVC; neonates and other groups possibly at risk from DEHP toxicity, 2002. Retrieved 17 September 2014 from [http://ec.europa.eu/health/ph\\_risk/committees/scmp/documents/out43\\_en.pdf](http://ec.europa.eu/health/ph_risk/committees/scmp/documents/out43_en.pdf).
113. Scientific Committee on Emerging and Newly Identified Health Risks (SCENIHR). Preliminary report on the safety of medical devices containing DEHP-plasticized PVC or other plasticizers on neonates and other groups possibly at risk. Scientific Committee on Emerging and Newly Identified Health Risks, European Commission, Brussels and Luxembourg, 2013. Retrieved 18 June 2014 from [http://ec.europa.eu/health/scientific\\_committees/consultations/public\\_consultations/scenih\\_r\\_cons\\_05\\_en.htm](http://ec.europa.eu/health/scientific_committees/consultations/public_consultations/scenih_r_cons_05_en.htm).
114. Goodman M, Lakind JS, Mattison DR. Do phthalates act as obesogens in humans? A systematic review of the epidemiological literature. *Crit Rev Toxicol*. 2014; 44, 151–175.