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Influence of temperature on the development and reproduction of *Cinara cedri* (Hemiptera: Aphidoidea: Lachninae)

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Abstract

Temperature is one of the main factors affecting insect growth, development and reproduction. The effects of temperatures (10, 15, 20, 25 and 30°C) on the development and reproduction of *Cinara cedri* Mimeur (Hemiptera: Aphidoidea: Lachnidae) fed on *Cedrus deodara* (Roxb.) G. Don were evaluated in this study. With the increase of temperature from 10 to 30°C, the development duration at different development stages gradually shortened. There was a significant positive correlation between the developmental rates and temperature, following a quadratic regression model. The lower developmental threshold temperature (*C*) and effective accumulated temperatures (*K*) for completing a generation were 4.13°C and 263.4 degree-days, respectively. The highest fecundity was observed at 20°C with 25.74 first-instar nymphs/female. Both the highest intrinsic rate of increase (*r*, 0.11 ± 0.03) and net reproduction rate (R_0 , 19.06 ± 2.05) were observed at 20°C, whereas the lowest values of *r* (0.05 ± 0.01) at 10°C and R_0 (5.78 ± 0.88) at 30°C were observed. The results suggest that temperature significantly affects the biology of *C. cedri* and the optimal temperature for its development is 20°C.

Introduction

Cinara cedri Mimeur, 1936 (Hemiptera: Aphidoidea: Lachnidae) is an important pest of Cedrus deodara, C. libani, C. brevifolia and C. atlantica (Fabre et al., 1988; Yu and Wang, 2014; Binazzi et al., 2015). At present, C. cedri are widely distributed all over the world, including Europe (Spain, Italy, France, Britain, etc.), Asia (China, Iran and Turkey), the United States, Argentina and Morocco (Blackman and Eastop, 2013; Huang et al., 2017; Şükran and Mustafa, 2019). In China, C. cedri is an important invasive pest, with reports of damage caused by this species in Beijing, Shandong, Henan and Shaanxi provinces (Yu and Wang, 2014; Tian, 2018; Zhang, 2016). The adults and nymphs of C. cedri cause damage by sucking the sap from the 1-2 years old shoots and leaves of cedar trees (Lin and Lei, 2018; Şükran and Mustafa, 2019). A large amount of honeydew secreted by C. cedri was accumulated on the shoots and leaves of cedar trees, which caused the stomata and lenticels to be blocked. In the period of peak abundance of C. cedri occurrence, sooty mold can also occur, which has a serious impact on the normal growth and development of cedar trees (Zhang, 2016; Tian, 2018). At present, only a few studies on C. cedri biological characteristics are available (Yu and Wang, 2014; Tian, 2018; Şükran and Mustafa, 2019). It was observed in Beijing, Henan and Shandong provinces of China in both spring and autumn seasons, but was more serious in spring (Yu and Wang, 2014; Huang et al., 2017; Tian, 2018). In addition, the populations overwintered as eggs with nymphs hatching during the first week of April in the Isparta region in Turkey. The adult stage was reached through four nymphal instars over 7 to 10 days (Sükran and Mustafa, 2019).

Temperature is one of the most important factors influencing most biological processes of insects, including their growth, development and reproduction (Clissold and Simpson, 2015; Cao *et al.*, 2018; Wang *et al.*, 2020). Generally, the developmental and reproductive rates of insects differ with temperature (Fand *et al.*, 2015; Krechemer and Foerster, 2015; Shi *et al.*, 2017). The time spent in developmental stages of an insect at a constant temperature can be used to generate developmental rate curves, suitable for modeling and predicting development time as a function of temperature (Southwood, 1966; Zou *et al.*, 2018; Hirotsugu and Norio, 2019). These models can also be used to calculate the developmental threshold temperature, optimum temperature and thermal accumulation of insect, contributing to models to predict the geographic distribution and phenology in a region (Riahi *et al.*, 2013; Guo *et al.*, 2016). In addition, life tables can be used to evaluate the comprehensive effects of temperature on the development, survival and reproduction of insects (Michèle *et al.*, 2003; Farhadi *et al.*, 2011; Ren *et al.*, 2016). The effect of temperature on the growth and development of *C*.

cedri is not fully studied. Consequently, the effects of temperature on development and reproduction of *C. cedri* on *C. deodara* were evaluated in the laboratory in this study.

Materials and method

Insects

C. cedri adults were collected from *Cedrus deodara* (Roxb.) G. Don in the campus of Shandong agricultural university in Tai'an Shandong province, China and were used to establish a laboratory colony. The aphid adults reared in a Petri dish (12 cm diameter) with a branch of *C. deodara* (1 cm diameter, 10 cm length). The Petri dishes were kept in an incubator (PQX-330A-3H) at a constant temperature and relative humidity ($25 \pm 0.5^{\circ}$ C, $70 \pm 5\%$ RH) and photoperiod (16:8, L:D).

Observation on life history of C. cedri

From November 2018 to March 2020, three cedar plants that were harmed by *C. cedri* were randomly selected on the campus of Shandong Agricultural University to investigate and record the aphid morphology and development state of *C. cedri* once a week. According to the survey data, the life history chart of *C. cedri* was drawn.

Development and reproduction at constant temperatures

A branch segment of C. deodara was placed in a Petri dish (12 cm diameter), and the branch was wrapped in mud at both ends to reduce water loss. Five Petri dishes were prepared, and a group of 50-100 female adult aphids were placed in each Petri dish and used to produce offspring at a constant temperature 25°C for 4 h. After 4 h, 450 first instars were placed in 15 Petri dishes on average with a branch segment of C. deodara, 30 first instars in each Petri dish. The first-instar nymph was reared and observed at five different incubators with five constant temperatures, 10 ± 0.5 , 15 ± 0.5 , 20 ± 0.5 , 25 ± 0.5 and $30 \pm 0.5^{\circ}$ C, $70 \pm 5\%$ RH and a 16:8 (L:D) h photoperiod, and each temperature treatment group contained three Petri dishes. Each developmental stage from first-instar nymph to adult was observed daily, and the developmental times, prenatal period, birth period, postpartum period, molt, death and births of C. cedri were recorded until the adult aphid died.

Life tables

A life table was constructed from the observed survival and fecundity rates for individuals. Parameters at constant temperatures were calculated and analyzed using the methods of Chi (1988, 2012), including the age-stage survival, intrinsic rate of increase (r), the net reproductive rate (R0), the finite rate of increase (λ) and the generation time (T). The calculation formulae were as follows:

$$1 = \sum_{x=1}^{w} l_x \times m_x \times e^{-r(x+1)}$$
(1)

$$R_0 = \sum_{x=1}^{w} l_x \times m_x \tag{2}$$



Figure 1. Life history of C. cedri on C. deodara.

$$\lambda = e^r \tag{3}$$

$$T = (\ln R_0)/r \tag{4}$$

where *w* represents the maximum age; *x* represents the age; l_x represents the survival rate of a newborn first instar at age *x* and m_x represents the average number of offspring produced by each female aphid at age *x*.

Statistical analysis

The developmental time, prenatal period, birth period, postpartum period and fecundity at different temperatures were compared using one-way analysis of variance followed by Tukey's HSD tests to obtain the significance of differences (P < 0.05). Linear regression, quadratic regression and logistic regression curve models were used to fit the relationship between development rate and temperature to select the best result. All statistical analyses were performed using software SPSS 19.0.

The effective accumulated temperature (K) and lower developmental threshold temperature (C) were calculated from the following formulae (Qiu *et al.*, 2012; Cao *et al.*, 2018):

$$K = \frac{n \sum V^2 \sum^T - \sum^V \sum VT}{n \sum V^2 - \left(\sum V\right)^2}$$
(5)

$$C = \frac{\sum V^2 \sum^T - \sum^V \sum VT}{n \sum V^2 - (\sum V)^2}$$
(6)

where V (1/developmental duration) is the developmental rate in days at temperature T (°C), K is the required heat units in degreedays (DD) for effective accumulation and C (°C) is the minimum developmental threshold.

Results

Life history of C. cedri

C. cedri has a holocyclic life cycle on *C. deodara* (fig. 1). It overwinters on the needles of *C. deodara* by fertilized eggs. The eggs developed into apterous fundatrix in mid-March of the following year.

Table 1. Developmental duration of C. cedri at five constant temperatures fed on C. deodara

Temperature (°C)	First instar	Second instar	Third instar	Fourth instar	Adult aphid	Life cycle
10	7.72 ± 0.08a	7.52 ± 0.06a	7.71±0.08a	7.47 ± 0.10a	36.30 ± 0.53a	66.10 ± 0.56a
15	5.87 ± 0.12b	5.44 ± 0.13b	6.80 ± 0.11b	6.09 ± 0.10b	22.40 ± 0.58b	46.12 ± 0.57b
20	4.39 ± 0.10c	3.88 ± 0.72c	3.65 ± 0.10c	4.55 ± 0.11c	20.31 ± 0.53c	37.11 ± 0.63c
25	3.40 ± 0.10d	2.78 ± 0.77e	3.65 ± 0.14c	4.09 ± 0.12d	17.12 ± 0.48d	30.92 ± 0.60d
30	2.84 ± 0.05e	3.18 ± 0.87d	3.16 ± 0.07d	4.09 ± 0.12d	12.19 ± 0.53e	25.63 ± 0.64e

Mean ± SE (days), with the same column the values with different letters are significantly different (one-way analysis of variance followed by Tukey's HSD tests, P<0.05).

Table 2. Relationship between temperature and developmental rate of C. cedri

Life stage	Regression equation	R ²	Р	C (°C)	K (DD)
First-instar nymph	$V = 0.0002T^2 + 0.0056T - 0.055$	0.999	0.01	4.13	65.3
Second-instar nymph	$V = -0.0003T^2 + 0.0219T - 0.062$	0.984	0.01	5.83	54.6
Third-instar nymph	$V = -0.0001T^2 - 0.0138T - 0.0017$	0.992	0.008	6.50	56.5
Fourth-instar nymph	$V = -0.0003T^2 + 0.0161T - 0.002$	0.972	0.01	8.95	51.3
Adult	$V = 0.00007T^2 - 0.00008T + 0.0248$	0.950	0.05	5.65	263.4

V is the developmental rate and T is the temperature (°C), C for the lower thermal threshold for development (°C) and K is the effective accumulative temperature (DD).

The number of aphids increased sharply after the fundatrix developed into adult aphids and began to reproduce fundatrigenia parthenogenetically. In mid-April, the fundatrigenia produced alate and apterous viviparous female aphids, but the number of alate viviparous female was small. With the increase of temperature, the proportion of alate viviparous female aphids increased gradually. The aphids reproduced parthenogenetically several generations and began to over-summer in late-July. The population began to increase in mid-September. The aphids continued to reproduce parthenogenetically until the emergence of the sexuales in mid-November. After mating, sexuales laid fertilized eggs on cedar needles to overwinter. The newly laid eggs are brownish yellow, and then gradually turn to black.

Effects of temperature on the development of C. cedri

Over the range of temperatures tested, *C. cedri* fed on the branch of *C. deodara* successfully developed from first-instar nymph to adult. With the increase of environmental temperature from 10 to 30°C, the developmental duration was significantly shortened. The developmental times from 10 to 30°C for first-instar nymph ranged from 7.71 to 2.84 days ($F_{4,289} = 512.25$, P < 0.001), for second-instar nymph ranged from 7.52 to 3.18 days ($F_{4,287} = 529.84$, P < 0.001), for third-instar nymph ranged from 7.47 to 4.09 days ($F_{4,282} = 184.70$, P < 0.001), for adult aphid ranged from 36.30 to 12.19 days ($F_{4,133} = 314.76$, P < 0.001), for entire developmental stage from 66.10 to 25.63 days ($F_{4,132} = 747.59$, P < 0.001), respectively (table 1).

Relationship between the developmental rate and temperature

There was a significant positive correlation between the developmental rates and temperature, which was in accordance with the quadratic regression model (table 2). Among the different developmental stages of *C. cedri*, the lowest value for the lower thermal threshold for development (*C*) was 4.13° C for the first-instar nymph, and the highest *C* value was 8.95° C for the fourth-instar nymph. The effective accumulative temperature value (*K*) was 521.2 DD for *C. cedri* to complete the whole generation (table 2).

Effects of temperature on the survival and fecundity of C. cedri

Generally, C. cedri showed similar patterns of age-specific fecundity on C. deodara at five different temperatures (fig. 2a-e). After the beginning of the birth period, the estimated number of firstinstar nymphs laid per day peaked at different days. Female adult mortality was initially low, but survival began to decline sharply after day 39 at 10°C, day 28 at 15°C, day 26 at 20°C, day 21 at 25°C and day 17 at 30°C. In addition, the survival times of C. cedri nymph decreased with the increase of temperature, and the highest survival rate of C. cedri nymph occurred at 20°C (fig. 3a-e). Different temperatures significantly affected the prenatal period ($F_{4,130} = 24.11$, P < 0.001), birth period ($F_{4,130} =$ 148.98, P < 0.001), postpartum period ($F_{4,130} = 18.10$, P < 0.001) and fecundity $(F_{4,130} = 168.23, P < 0.001)$ of female C. cedri. Prenatal period, birth period, postpartum period decreased progressively as temperature increased. Prenatal period, birth period and postpartum period from 30 to 10°C were 2.22 to 5.67, 7.81 to 24.56 and 2.22 to 5.90 days, respectively (table 3). Female adult of C. cedri showed different fecundity performances at different temperatures, and the fecundity of C. cedri reached the maximum at 20°C, which was 25.74 first-instar nymphs/female.

Effects of temperature on the life table parameters of C. cedri

Temperature significantly affected the population parameters of *C. cedri* (table 4). With the increase of environmental temperature, the generation time (*T*) was shortened from 48.56 ± 3.15 at 10°C to 20.09 ± 2.76 at 30°C. The value of net reproductive rate (R_0), intrinsic rate of increase (r), finite rate of increase (λ) increases first and then decreases with the increase of



Figure 2. Age-specific survival (lx) and fecundity (mx) of C. cedri on C. deodara at five different temperatures ((a) 10°C; (b) 15°C; (c) 20°C; (d) 25°C; (e) 30°C).



Figure 3. Age-specific survival of C. cedri nymphs on C. deodara at five different temperatures ((a) 10°C; (b) 15°C; (c) 20°C; (d) 25°C; (e) 30°C).

temperature, and reached a maximum value at 20°C (19.06 \pm 2.05, 0.11 \pm 0.03 and 1.12 \pm 0.02, respectively).

Discussion

Temperature, as an important ecological factor, has a significant impact on the development duration, fecundity and population dynamics of insects (Campbell *et al.*, 1974; Özder and Ozgur, 2013; Cao *et al.*, 2018). A large number studies have shown that the developmental duration of insects shortens with the increase of temperature over the appropriate temperature range (Cui *et al.*, 2018; Abu Alloush, 2019). In this study, developmental duration of each developmental stage of *C. cedri* was shortened as the temperature increased from 10 to 30°C. However, *C. cedri* could complete its whole growth and development process at 30°C. This may be because the maximum temperature in the design

of this experiment has not yet reached the temperature that limits the development of *C. cedri*. Although the appropriate temperature will promote the growth and development of insects, the development of insects will be delayed, stagnated or the insects may die if the temperature exceeds the developmental temperature (Davidson, 1994). A piece of white paper $(1 \text{ m} \times 1 \text{ m})$ was hung under the branches of *C. deodara* in August 2019 (average temperature 32°C). After a week of observation, it was found that there was no honeydew on the white paper indicating that the aphid activity was less. This may be due to the high temperature, coupled with increased rainfall, which causes *C. cedri* dormant or diapause. Therefore, the tolerance of *C. cedri* to high temperature needs further study.

In addition, the temperature can significantly affect the longevity of female adult aphids of *C. cedri*. With the increase of temperature, the longevity of *C. cedri* decreases gradually, the

Table 3. Longevity and fecundity	y of C. cedri females at different ter	nperatures fed on C. deodard
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		Longevity (days)		
Temperature (°C)	Prenatal period	Birth period	Postpartum period	Fecundity (first-instar nymph/female)
10	5.67 ± 0.26a	24.50 ± 0.60a	5.90 ± 0.41a	13.22 ± 0.54c
15	4.20 ± 0.28b	14.40 ± 0.41b	3.86 ± 0.39b	20.26 ± 0.86b
20	3.30 ± 0.31bc	13.23 ± 0.55bc	3.81 ± 0.40b	25.74 ± 0.96a
25	3.08 ± 0.34cd	11.40 ± 0.66c	2.60 ± 0.36bc	17.48 ± 0.80bc
30	2.22 ± 0.21d	7.81 ± 0.39d	2.22 ± 0.22c	8.69 ± 0.39d

Mean ± SE with the same column the values with different letters are significantly different (one-way analysis of variance followed by Tukey's HSD tests, P<0.05).

Table 4. Life-table parameters of	of <i>C. cedri</i> at fi	ve constant temperatures f	ed on C. deodara
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Temperature (°C)	Net reproductive rate (R_0)	Intrinsic rate of increase (r)	Generation time (T)	Finite rate of increase (λ)
10	9.58 ± 1.11	0.05 ± 0.01	48.56 ± 3.15	1.05 ± 0.02
15	12.51 ± 1.51	0.07 ± 0.01	38.78 ± 2.68	1.07 ± 0.01
20	19.06 ± 2.05	0.11 ± 0.03	26.46 ± 2.35	1.12 ± 0.02
25	12.06 ± 1.26	0.10 ± 0.02	25.48 ± 1.89	1.10 ± 0.02
30	5.78 ± 0.88	0.08 ± 0.01	20.09 ± 2.76	1.09 ± 0.01

longevity at 10°C is 36.30 days, and the longevity at 30°C is only 12.19 days. Guo (2012) found that the adult aphid longevity of *Aphid gosypii* Glover decreased from 11.25 to 3.00 days with increasing temperature in the range of 18–30°C. Durak and Borowiak-Sobkowiak (2013) found that the adult aphid longevity of *Cinara cupressi* was also related to temperature, and the adult aphid longevity was 40.4 days at 25°C and 25.1 days at 28°C.

There are some differences in the developmental threshold temperature of different species of aphids, but most of them are around 4°C (Shao *et al.*, 2017). In this study, the results showed that the developmental threshold temperature of first-instar nymph, second-instar nymph, third-instar nymph, fourth-instar nymph and adult aphid is 4.13, 5.83, 6.50, 8.95 and 5.65°C, respectively. The eggs of *C. cedri* have hatched by the middle of March in Tai'an, Shandong province. The temperature in the middle of March is $6-16^{\circ}$ C, which is close to the developmental threshold temperature of *C. cedri*. Combined with the annual average temperature in Shandong province, the theoretical generation of *C. cedri* is estimated to be 7–8 generations per year. However, the generations of aphids are affected by climate change under field conditions (Beetge and Krüger, 2019).

The best indicator to measure the relationship between insect population growth and the temperature is intrinsic growth rate, which can generally reflect the effects of temperature on insect development, reproduction and survival. Typically, the intrinsic growth rate of aphid population is the highest in the temperature range of 25–30°C (Gilbert and Raworth, 1996; Özder and Ozgur, 2013), such as *Schizolachnus pineti* in 24–36°C (Holopainen and Kainulainen, 2004) and *Cinara pinea* in 21°C (Shao *et al.*, 2017). In this study, the intrinsic growth rate of *C. cedri* is the highest at the temperature of 20°C. The results of this study are consistent with maximum populations occurring in the cooler months of spring.

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Conflict of interest. The authors declare that they have no conflict of interest.

Ethical standards. This paper does not contain any studies with human participants or animals performed by any of the authors.

References

- Abu Alloush AH (2019) Developmental duration and predation rate of the coccidophagous coccinellid *Rhyzobius lophanthae* (Blaisdell) (Coleoptera: Coccinellidae) on *Aspidiotus nerii* Bouche. B. *Entomological Research* 109, 1–5.
- Beetge L and Krüger K (2019) Drought and heat waves associated with climate change affect performance of the potato aphid *Macrosiphum euphorbiae*. Scientific Reports 9, 3645.
- Binazzi F, Sabbatini PG and Roversi PF (2015) First record in Cyprus of *Cinara (Cinara) cedri* Mimeur (Aphididae: Lachninae) on *Cedrus brevifolia* (Hooker Fil.) Henry. *Redia-Giornale Di Zoologia* **98**, 151–154.
- Blackman RL and Eastop VF (2013) Aphids on the world's plants: an online identification and information guide. Available at http://www.aphidson-worldsplants.info/.
- Campbell A, Frazer BD, Gilbert N, Gutierrez AP and Mackauer M (1974) Temperature requirements of some aphids and their parasites. *Journal of Applied Ecology* 11, 431–438.
- Cao Y, Li C, Yang WJ, Meng YL, Wang LJ, Shang BZ and Gao YL (2018) Effects of temperature on the development and reproduction of *Thrips hawaiiensis* (Thysanoptera: Thripidae). *Journal of Economic Entomology* 11, 755–760.
- Chi H (1988) Life-table analysis incorporating both sexes and variable development rates among individuals. *Environmental Entomology* 17, 26–34.
- Chi H (2012) TWOSEX-MSChart: a computer program for the age stage, twosex life table analysis. Available at http://140.120.197.173/Ecology/.
- Clissold FJ and Simpson SJ (2015) Temperature, food quality and life history traits of herbivorous insects. *Current Opinion in Insect Science* 11, 63–70.
- Cui J, Zhu SY, Bi R, Xu W, Gao Y and Shi SS (2018) Effect of temperature on the development, survival, and fecundity of *Heliothis viriplaca* (Lepidoptera: Noctuidae). *Journal of Economic Entomology* 111, 1940–1946.

- Davidson J (1994) On the relationship between temperature and rate of development of insects at constant temperature. *Journal of Animal Ecology* 13, 26–38.
- Durak R and Borowiak-Sobkowiak B (2013) Influence of temperature on the biological parameters of the anholocyclic species *Cinara tujafilina* (Hemiptera: Aphidoidea). *Central European Journal of Biology* 8, 570–577.
- Fabre JP, Chalon A, Chizky J and Robert E (1988) Possibilités d'infestation par les pucerons: *Cedrobium laportei* Remaudière, *Cinara cedri* Mimeur (Homoptera, Lachnidae), chez le genre *Cedrus. Annales des Sciences Forestières* 45, 125–140.
- Fand BB, Sul NT, Bal SK, Minhas PS and Córdoba-Aguilar A (2015) Temperature impacts the development and survival of common cutworm (*Spodoptera litura*): simulation and visualization of potential population growth in India under warmer temperatures through life cycle modelling and spatial mapping. *PLoS One* **10**, e0124682.
- Farhadi R, Allahyari H and Chi H (2011) Life table and predation capacity of *Hippodamia variegate* (Coleoptera: Coccinellidae) feeding on *Aphis fabae* (Hemiptera: Aphididae). *Biological Control* 59, 83–89.
- Gilbert NE and Raworth DA (1996) Insects and temperature a general theory. *The Canadian Entomologist* **128**, 1–13.
- **Guo R** (2012) Studies on biological characteristics and ecology population of Aphid gosypii Glover from host Lycium barbarum L. in Qaidam (M.S. thesis). Qinghai University.
- Guo TT, Men XY, Yu Y, Chen H, Zhou XH, Zhuang QY, Wang ZY and Li LL (2016) Effects of temperature on the development and reproduction of *Athetis dissimilis* (Lepidoptera: Noctuidae). *Acta Entomologica Sinica* 59, 865–870 (in Chinese).
- Hirotsugu T and Norio K (2019) Forecasting method of persimmon fruit moth, Stathmopoda masinissa using the effective accumulated temperature calculation simulation of the JPP-NET. Annual Report of the Kansai Plant Protection Society 61, 155–156.
- Holopainen JK and Kainulainen P (2004) Reproductive capacity of the grey pine aphid and allocation response of Scots pine seedlings across temperature gradients: a test of hypotheses predicting outcomes of global warming. *Canadian Journal of Forest Research* 34, 94–102.
- Huang BS, Wu D, Sun LZ and Du CZ (2017) The harm and control technology of *Cinara cedri* Mimeur. *Henan Forestry Science and Technology* **37**, 45– 47 (in Chinese).
- Krechemer FS and Foerster LA (2015) Temperature effects on the development and reproduction of three *Trichogramma* (Hymenoptera: Trichogrammatidae) species reared on *Trichoplusia ni* (Lepidoptera: Noctuidae) eggs. *Journal of Insect Science* 15, 90.
- Lin J and Lei W (2018) The occurrence and prevention of *Cinara cedri* in Shangzhou District, Shangluo City, China. *Modern Agricultural Science and Technology* 16, 112–113 (in Chinese).

- Michèle R, Brodeur J and Cloutier C (2003) Effect of temperature on intrinsic rates of natural increase (r_m) of a coccinellid and its spider mite prey. *Biocontrol* **48**, 57–72.
- Özder N and Ozgur S (2013) The effects of temperature for development time, fecundity and reproduction on some ornamental aphid species. *Journal of Central European Agriculture* 14, 149–157.
- Qiu B, Zhou ZS, Luo SP and Xu ZF (2012) Effect of temperature on development, survival, and fecundity of *Microplitis manilae* (Hymenoptera: Braconidae). *Environmental Entomology* **41**, 657–664.
- Ren JL, Tu XB, Ge J, Zhao L and Zhang ZH (2016) Influence of temperature on the development, reproduction, and life table of *Calliptamus italicus* (L.) (Orthoptera: Acridoidea). *Journal of Asia-Pacific Entomology* 19, 203–207.
- Riahi E, Shishehbor P, Nemati AR and Saeidi Z (2013) Temperature effects on development and life table parameters of *Tetranychus urticae* (Acari: Tetranychidae). *Journal of Agriculture, Science and Technology* 15, 661–672.
- Shao D H, Duan L Q and Duan J P (2017) Population dynamics, effective accumulative temperatures and control method of *Cinara formosana*. *Journal of Northeast Forestry University* 45, 84–93 (in Chinese).
- Shi PJ, Gadi VPR, Chen L and Ge F (2017) Comparison of thermal performance equations in describing temperature-dependent developmental rates of insects: (II) two thermodynamic models. *Annals of the Entomological Society of America* 110, 113–120.
- Southwood TRE (1966) Ecological Methods with Particular Reference to the Study of Insect Populations. London, Methuen, pp. 60–65.
- Şükran O and Mustafa A (2019) Distribution, biology, morphology and damage of *Cinara cedri* Mimeur, 1936 (Hemiptera: Aphididae) in the Isparta Regional Forest Directorate. *Forestist* 69, 1–10.
- Tian T (2018) Studies on basic biological and ecological characteristics of *Cinara cedri* Mimeur (M.S. thesis). Shandong Agricultural University.
- Wang L, Walter GH and Furlong MJ (2020) Temperature, deltamethrinresistance status and performance measures of *Plutella xylostella*: complex responses of insects to environmental variables. *Ecological Entomology* 45, 345–354.
- Yu GY and Wang H (2014) First record of *Cinara cedri* Mimeur (Hemiptera, Aphididae, Lachninae) on *Cedrus deodara* in Beijing, China. *Journal of Environmental Entomology* 36, 260–264 (in Chinese).
- Zhang XF (2016) The problems and countermeasures in the prevention and control of *Cinara cedri*. *Chinese Horticulture Abstracts* 32, 118–179 (in Chinese).
- Zou Z, Xi J, Liu G, Song S, Xin T and Xia B (2018) Effect of temperature on development and reproduction of the carmine spider mite, *Tetranychus cinnabarinus* (Acari: Tetranychiae), fed on cassava leaves. *Experimental and Applied Acarology* 74(4), 383–394.