

Making inferences about non-detection observations to improve occurrence predictions in Venezuelan Psittacidae

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Summary

The global decline in psittacid populations highlights the need for monitoring programmes that allow us to estimate the level of confidence that can be placed in a non-detection observation in order to assess changes in range status. We used the detection/non-detection records for 26 psittacid species detected during the first national bird monitoring programme in Venezuela carried out in 2010 by the Neotropical Biodiversity Mapping Initiative. We fitted occupancy models and evaluate the suitability of the data to explain the lack of detections given the current sampling effort, and the expected occurrence probabilities due to environmental conditions (conditional probability of occurrence; Ψ_{CONDL}). We were able to fit reliable models for 13 of the 26 species detected. For Green-rumped Parrotlet *Forpus passerinus*, Blue-headed Parrot *Pionus menstruus*, and Orange-winged Amazon *Amazona amazonica*, the probability of detection (p) under the current sampling effort was too low (< 0.2) in areas where environmental conditions would imply high Ψ_{CONDL} (> 0.3). This suggests that sampling effort should be increased to generate reliable estimations of occurrence. In contrast, for Scarlet Macaw *Ara macao*, Yellow-crowned Amazon *Amazona ochrocephala*, Orange-chinned Parakeet *Brotogeris jugularis* and Brown-throated Parakeet *Eupsittula pertinax* the model estimated high p (> 0.3) and low Ψ_{CONDL} (< 0.2), suggesting that the species are reliably detected and better models could be obtained by including other predictive variables related to temporal use of resources and habitat heterogeneity. To improve the effectiveness of parrot monitoring programme in Neotropical countries, we suggest increasing the sampling effort, developing several surveys per year, and including variables related with temporal use of resources and habitat heterogeneity.

Keywords: bird survey, detection histories, distribution models, monitoring, occupancy models, parrot conservation

Introduction

Among birds, Psittacidae is a family of major conservation concern. Nearly half of the 344 species in the world (Snyder *et al.* 2000) are considered threatened under the IUCN Red List criteria, including nine already 'Extinct in the Wild' due high levels of habitat loss and trapping for illegal trade (BirdLife International 2015).

Although research on the ecology and behaviour of psittacids has increased in the last decade, there is an urgent need to determine trends in distribution at larger scales (Martin *et al.* 2014). The few monitoring programmes for this family are usually focused on one, often globally threatened species, or are limited to narrow geographic areas (but see Hille 2014). Status assessment of range and population trends for psittacids are usually based on static representation of species distribution (range maps; Snyder *et al.* 2000). However, range maps could provide a misleading interpretation of trends because they lack estimates of uncertainty in under- or over-prediction and assume homogeneous probability of occurrence across the range (Peterson *et al.* 2011). Data on presence and absence are required in order to track population changes, but most studies are limited in time and resources, and many apparent absences are in fact lack of detections (Kéry & Schmidt 2008). Standardised survey methods and appropriate analytical tools allow for the inclusion of heterogeneous detection probabilities, and the calculation of reliable estimates of presence or absence and their associated uncertainty (MacKenzie *et al.* 2002). Occupancy models have been widely used for monitoring avian populations (Baumgardt *et al.* 2014). These models use repeated detection and non-detection data (detection histories) at each location to jointly estimate the probability of presence (Ψ), and the probability of detection (p ; MacKenzie *et al.* 2002). Indeed, the most common application of detectability estimates is to determine whether a species is, in fact, present at a given site when not detected, but fewer case studies provides evaluations of the level of confidence that can be placed in a particular non-detection observation (Wintle *et al.* 2011). The accuracy with which a non-detection could be interpreted as a true absence, may have direct implications in our ability to confidently interpret a current species distribution and hence, our capacity to monitor temporal and geographical changes (Garrard *et al.* 2014).

In Venezuela, as in several other Neotropical countries, the high diversity of psittacids is combined with an increasing rate of land transformation (Rodríguez *et al.* 2010), illegal wildlife trade (Sánchez-Mercado *et al.* 2020), and limited resources for monitoring and conservation effort (Rodríguez 2014). The IUCN reports declining regional trends for 34 of the 50 Psittacidae species occurring in the country (Table 1), and six species are already under some threat category in the Venezuelan Red Data Book (Rodríguez *et al.* 2015). However, a more detailed national assessment requires the evaluation of current status and trends for the whole family. As a first step, a national bird monitoring programme was carried out in 2010 as a component of the Neotropical Biodiversity Mapping Initiative (NeoMaps; Rodríguez *et al.* 2012, Ferrer-Paris *et al.* 2013). This programme provides an important source of detection and non-detection records, ideal for fitting occupancy models allowing the establishment of a baseline to evaluate temporal and spatial changes in distributions (Ferrer-Paris *et al.* 2014, Berkunsky *et al.* 2015). A previous analysis using NeoMaps data showed that even the most widespread psittacids from the *Amazona* genus could be experiencing negative changes in their distribution (Ferrer-Paris *et al.* 2014). Here, we used the NeoMaps data available for all psittacid species in Venezuela to provide a complete description of the geographic distributions of most species occurring in the country. We used NeoMaps detection/non-detection records to fit occupancy models and evaluated the uncertainty and reliability of the resulting predictions for presence probabilities, and the suitability of the data to explain the lack of detections across the survey sites. Furthermore, we provide recommendations for improving future surveys to monitoring distribution changes in the country and any other Neotropical country.

Table 1. Psittacidae species reported for Venezuela. Distribution description, conservation categories, and population trend for each species according IUCN is shown. The number of NeoMaps sampling sites overlapping with the expected distribution of the species according to the available range maps from BirdLife is shown. The number of sites with detections, the ratio of current/expected detections and the number of detections for each Venezuelan psittacid species reported in Global Biodiversity Facility (GBIF) in 2010 is shown. The total number of sites sampled was 1,350.

Species	Common name	Endemism	Distribution range category for Venezuela	Conservation category	Population trend	Number of sites sampled	Number of sites with detections	Ratio Current/Expected detection	Detections GBIF 2010
<i>Amazona amazonica</i>	Orange-winged Amazon	No	Widespread	LC	Decreasing	715	97	14	78
<i>Amazona autumnalis</i>	Red-lored Amazon	No	Restricted	LC	Decreasing	50	0	0	2
<i>Amazona barbadensis</i>	Yellow-shouldered Amazon	Almost	Restricted	VU	Decreasing	85	5	6	9
<i>Amazona bodini</i>	Northern Festive Amazon	Almost	Restricted	NT	Decreasing	102	0	0	0
<i>Amazona dufresniana</i>	Blue-cheeked Amazon	No	Restricted	NT	Decreasing	109	0	0	16
<i>Amazona farinosa</i>	Southern Mealy Amazon	No	Widespread	NT	Decreasing	482	17	4	23
<i>Amazona mercenarius</i>	Scaly-naped Amazon	No	Restricted	LC	Decreasing	103	0	0	1
<i>Amazona ochrocephala</i>	Yellow-crowned Amazon	No	Widespread	LC	Decreasing	1,191	143	12	118
<i>Ara ararauna</i>	Blue-and-yellow Macaw	No	Widespread	LC	Decreasing	112	0	0	6
<i>Ara chloropterus</i>	Red-and-green Macaw	No	Widespread	LC	Decreasing	1,045	13	1	57
<i>Ara macao</i>	Scarlet Macaw	No	Widespread	LC	Decreasing	607	8	1	21
<i>Ara militaris</i>	Military Macaw	No	Restricted	VU	Decreasing	35	8	23	10
<i>Ara severus</i>	Chestnut-fronted Macaw	No	Widespread	LC	Stable	642	61	10	29
<i>Aratinga solstitialis</i>	Sun Parakeet	No	Restricted	EN*	Decreasing	0	0	0	0
<i>Bolborhynchus lineola</i>	Barred Parakeet	No	Restricted	LC	Stable	96	0	0	4
<i>Brotogeris chrysopterus</i>	Golden-winged Parakeet	No	Widespread	LC	Decreasing	454	2	0	5
<i>Brotogeris cyanoptera</i>	Cobalt-winged Parakeet	No	Restricted	LC	Stable	50	0	0	0
<i>Brotogeris jugularis</i>	Orange-chinned Parakeet	No	Widespread	LC	Stable	557	20	4	98

Table 1. Continued.

Species	Common name	Endemism	Distribution range category for Venezuela	Conservation category	Population trend	Number of sites sampled	Number of sites with detections	Ratio Current/Expected detection	Detections GBIF 2010
<i>Deropterus accipitrinus</i>	Red-fan Parrot	No	Widespread	LC	Decreasing	308	0	0	13
<i>Diopsittaca nobilis</i>	Northern Red-shouldered Macaw	No	Widespread	LC	Stable	361	2	1	43
<i>Eupsittula pertinax</i>	Brown-throated Parakeet	No	Widespread	LC	Increasing	1,180	190	16	234
<i>Forpus conspicillatus</i>	Spectacled Parrotlet	No	Restricted	LC	Increasing	0	0	0	0
<i>Forpus modestus</i>	Dusky-billed Parrotlet	No	Restricted	LC	Decreasing	102	0	0	2
<i>Forpus passerinus</i>	Green-rumped Parrotlet	No	Widespread	LC	Stable	1,060	47	4	174
<i>Hapalopsittaca amazonina</i>	Rusty-faced Parrot	No	Restricted	VU C2a(i)	Decreasing	43	0	0	0
<i>Nannopsittaca panychlora</i>	Tepui Parrotlet	Almost	Restricted	LC	Stable	146	1	1	35
<i>Orthopsittaca manilata</i>	Red-bellied Macaw	No	Widespread	LC	Stable	407	5	1	9
<i>Pionites melanocephala</i>	Black-headed Parrot	No	Widespread	LC	Stable	354	5	1	38
<i>Pionus chalcopterus</i>	Bronze-winged Parrot	No	Restricted	LC	Decreasing	38	0	0	11
<i>Pionus fuscus</i>	Dusky Parrot	No	Widespread	LC	Decreasing	283	0	0	4
<i>Pionus menstruus</i>	Blue-headed Parrot	No	Widespread	LC	Decreasing	798	47	6	90
<i>Pionus seniloides</i>	White-capped Parrot	No	Restricted	LC	Decreasing	128	0	0	0
<i>Pionus sordidus</i>	Red-billed Parrot	No	Widespread	LC	Decreasing	162	0	0	8
<i>Thectocercus acuticaudatus</i>	Blue-crowned Parakeet	No	Widespread	LC	Decreasing	575	0	0	6
<i>Psittacara leucophthalmus</i>	White-eyed Parakeet	No	Widespread	LC	Decreasing	411	7	2	2
<i>Psittacara wagleri</i>	Scarlet-fronted Parakeet	No	Widespread	NT	Decreasing	215	4	2	42
<i>Pyrilia barrabandi</i>	Orange-cheeked Parrot	No	Widespread	NT	Stable	50	2	4	0
<i>Pyrilia caica</i>	Caica Parrot	No	Widespread	NT	Decreasing	299	0	0	5

Table 1. Continued.

Species	Common name	Endemism	Distribution range category for Venezuela	Conservation category	Population trend	Number of sites sampled	Number of sites with detections	Ratio Current/Expected detection	Detections GBIF 2010
<i>Pyrrhura pyrrhura</i>	Saffron-headed Parrot	No	Widespread	NT	Decreasing	150	0	0	11
<i>Pyrrhura caeruleiceps</i>	Perija Parakeet	No	Restricted	EN	Decreasing	7	0	0	0
<i>Pyrrhura egregia</i>	Fiery-shouldered Parakeet	Almost	Restricted	LC	Decreasing	152	3	2	16
<i>Pyrrhura emma</i>	Venezuelan Parakeet	Endemic	Restricted	LC	Decreasing	107	0	0	0
<i>Pyrrhura hoematotis</i>	Blood-eared Parakeet	Endemic	Restricted	LC	Decreasing	85	1	1	19
<i>Pyrrhura melanura</i>	Maroon-tailed Parakeet	No	Widespread	LC	Decreasing	50	3	6	0
<i>Pyrrhura picta</i>	Painted Parakeet	No	Widespread	LC	Decreasing	317	10	3	28
<i>Pyrrhura rhodocephala</i>	Rose-headed Parakeet	Endemic	Restricted	LC	Stable	160	1	1	12
<i>Touit batavica</i>	Lilac-tailed Parrotlet	No	Widespread	LC	Decreasing	183	0	0	6
<i>Touit dilectissima</i>	Blue-fronted Parrotlet	No	Restricted	LC	Stable	87	0	0	1
<i>Touit huetii</i>	Scarlet-shouldered Parrotlet	No	Widespread	VU	Stable	213	0	0	2
<i>Touit purpurata</i>	Sapphire-rumped Parrotlet	No	Widespread	LC	Stable	222	0	0	13

Methods

Study species

A total of 50 species from 19 genera of psittacids occur in Venezuela, although the presence of Sun Parakeet *Aratinga solstitialis* has only been confirmed in the disputed territory of Guyana Esequiba (Hilty 2003, Rojas-Suárez pers. comm.). Seven species are endemic or almost endemic to Venezuela (*Amazona barbadensis*, *Amazona bodini*, *Nannopsittaca panychlora*, *Pyrrhura egregia*, *P. emma*, *P. hoematotis* and *P. rhodocephala*). Fourteen further species have restricted distribution in the country (Table 1; Hilty 2003).

Range maps for all these species were obtained from BirdLife (BirdLife International 2008) and clipped to the region between 0–13°N and 59–73°W which includes all of Venezuela and neighbouring regions. Within this polygon we retrieved 21,860 presence records for all 50 psittacid species from the Global Biodiversity Information Facility (GBIF Occurrence Download <http://doi.org/10.15468/dl.ofmi8y>, 10 June 2016).

Field survey

The NeoMaps bird survey was performed between March and April 2010 by a team composed of seven expert ornithologists and several field assistants (methods fully described in Rodríguez *et al.* 2012). The sampling universe consisted of 170 half-degree cells defined in the Venezuelan Biodiversity Grid, which cover over half of the country, but do not include the southern forest regions (Figure 1). Twenty-seven cells were selected using a stratified sampling design based on environmental and biogeographical variables.

Standardised field sampling protocols for birds were implemented along a 40-km roadside transect within each cell. Two surveys were performed during two consecutive days in each transect: on the first day, 3-min point counts were performed at 50 stops, 800 m apart. On the second day, cumulative species lists were recorded at a selection of 10 stops sampled for 9 min each, divided into three consecutive 3-min periods. Total sampling effort was 108 hours of bird surveys (Rodríguez *et al.* 2012).

For this analysis we built detection histories for each psittacid species recorded by NeoMaps. We considered each stop as a 'site' (*i*; 1,350 sites, 50 stops across 27 transects), and each timed survey period of 3 min as a 'observation' (*j*), with duration $d = 3$ min. For the first day survey, detections were recorded as '1' and lack of detections as '0'. For the cumulative list of the second day the detection history was filled with '0' until the first detection, and with null values (N) afterward. Thus, valid detection histories for the second day are 1NN, 01N, 001 and 000, or NNN if the site was not visited on the second day (Ferrer-Paris *et al.* 2013). Time of day was used as an observation covariate. Sites covariates were extracted from the spatial location of each site.

Site covariates

Ranges of psittacid species are often described in terms of elevation, aridity and vegetation cover (Hilty 2003). Taking this into consideration, we searched for site covariates that could describe the environmental conditions during sampling and decided to use time series of remotely sensed data (Kerr *et al.* 2001). In order to obtain representative data on climatic and vegetation conditions at the time of the survey, we matched the location and date of each observations with time-series of environmental variables derived from the Moderate Resolution Radio Spectrometer (MODIS) sensors in Terra-Satellites and queried them using the global MODIS Subsetting Tool (Land Processes Distributed Active Archive Center (LP DAAC 2014), and the Climate Hazards Group InfraRed Precipitation with Station data archive (CHIRPS version 2.0; Funk *et al.* 2015). We calculated the representative value of the variable for the year prior to the sampling time (approx. March 2009–March 2010).

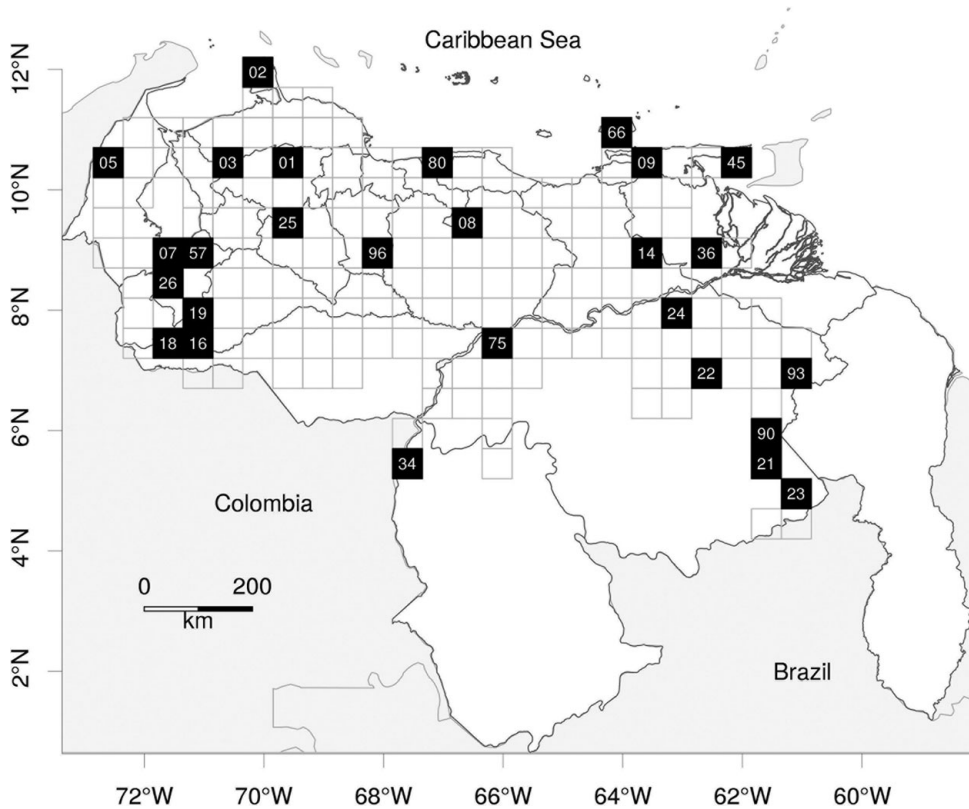


Figure 1. Sampling universe consisted in 170 half-degree cells defined in the Venezuelan Biodiversity Grid. Numbers indicate NeoMaps' cells code visited by survey teams in 2010.

We considered that the effect of elevation could be properly described by the annual mean temperature, thus we used the Land Surface Temperature with Daily Cycle (LST; MOD11A2, version 6, 1-km spatial resolution) as a measure of local temperature during day time (Wan *et al.* 2015). We used the annual mean value of the Enhanced Vegetation Index (EVI; MOD13Q1, version 5, 250 m resolution) as proxy for vegetation cover, because it measures the chlorophyll concentration across all vegetation components (Didan 2015). We used total annual precipitation and total annual potential evapotranspiration as proxies for water balance. We used the CHIRPS precipitation data (PREC; version 2, 1-km resolution) and the Potential Evapo-transpiration (PET; MOD16A2, version 6, 1-km resolution; Running *et al.* 2017).

Occupancy models

We used a single-season occupancy model based on zero-inflated binomial models (MacKenzie *et al.* 2006) to estimate the probability of occurrence for species detected in the surveys (Ψ). The occupancy state (z_i) of site i was modelled as $z_i \sim \text{Bernoulli}(\Psi_i)$, while the observation process was modelled as $y_{ij}|z_i \sim \text{Bernoulli}(z_i * p_{ij})$ in which p_{ij} represented site and occasion specific detection probability. Covariates of Ψ_i (site covariates) and p_{ij} (observation covariates) were modelled using the logit link (Fiske and Chandler 2011).

We fitted eight models representing different combinations of covariates for probability of detection and probability of occurrence. First, we considered models with constant probability of

detection ($p(\cdot)$), and others that assumed detection changed linearly with time of the day in hours ($p(h)$). Regarding probability of occurrence, we defined a null model with constant probability ($\Psi(\cdot)$) and alternative models considering the effects of vegetation ($\Psi(V)$, using second degree polynomials of mean EVI), climatic ($\Psi(C)$, second degree polynomials of mean LST and total PREC and PET), or both vegetation and climatic covariates ($\Psi(VC)$). The models were fitted with data from sampling regions that had at least one detection or that overlapped with the expected distribution of the species according to the available range maps from BirdLife (BirdLife International 2008) and GBIF presence records for 2010 (GBIF 2018).

We evaluated the individual performance of each model using the corrected Akaike Information Criterion (AICc; Burnham and Anderson 2002). Then we used the model with the best performance for each species to explain the lack of detections across the survey sites. For the sites without detections, we calculated the conditional probability of occurrence given that the species was not detected (MacKenzie *et al.* 2006). This probability (Ψ_{CONDNL}), considers two components: whether sampling effort was enough to detect the species at least once, conditional on its presence ($p^* = 1 - \text{Prod}(1 - p)$), and the unconditional probability of occurrence given the values of the site covariates (Ψ'). We used the *unmarked*, *raster*, and *AICcmodavg* packages of R to fit the models (Fiske and Chandler 2011).

We visualised the spatial distribution of the unconditional probability of occurrence (Ψ') for the whole country for the species with more than 15 detections, based on the model with the highest support for each species (Table 2) and values of the vegetation and climatic covariates. We used the *predict* function of *unmarked* package (Fiske and Chandler 2011) and a raster stack of predictive variables at a resolution of 1 km.

Results

NeoMaps sampling in 2010 detected 26 of the 50 species of psittacids present in Venezuela (Table 1). The most detected species were Brown-throated Parakeet *Eupsittula pertinax* (190 detections), Yellow-crowned Amazon *Amazona ochrocephala* (143 detections) and Orange-winged Amazon *Amazona amazonica* (97). For six species, NeoMaps sampling provided more presence records than GBIF data for the year 2010, including two detections of Orange-cheeked Parrot *Pyrrhura barrabandi* and three detections for Maroon-tailed Parakeet *Pyrrhura melanura* (Table 1).

NeoMaps also provided detections outside the BirdLife distribution ranges for six species (*Amazona amazonica*, *Amazona farinosa*, *Ara militaris*, *Diopsittaca nobilis*, *Orthopsittaca manilata*, and *Pionus menstruus*). The ratio between actual and expected detections was usually lower than 10%, except for *Amazona amazonica*, *A. ochrocephala*, Chestnut-fronted Macaw *Ara severus* and *E. pertinax*, with values between 11% and 15% (Table 1). Detailed methods and results are shown in Appendix S1 in the online supplementary material.

Model fitting

The number of models fitted to each species was limited due to non-convergence or unrealistic estimates of coefficients. For the 12 species with less than five detections one or two models could be fitted. The four species with more detections also had several candidate models: *Eupsittula pertinax* (eight models), Orange-chinned Parakeet *Amazona amazonica*, *Brotogeris jugularis* and Blue-headed Parrot *Pionus menstruus* (six) and Green-rumped Parrotlet *Forpus passerines* (five), the rest of the species had three or four models fitted (Table 2).

Although we were able to fit models for 25 out of 26 detected species, we discarded the models for 12 species with less than five detections due to obvious over-fitting in probabilities of presence or detection. Among the 13 remaining species with reliable models, for five of them the model with the lowest AICc considered constant detectability (Table 2). For Scarlet Macaw *Ara macao*, *Amazona ochrocephala* and *Eupsittula pertinax* the model suggested constant high probability of detection (> 0.3 ; Figure 2a), while for *Amazona farinosa* and *Pyrrhura picta*, constant low probability (< 0.2 ; Figure 2a). For White-eyed Parakeet *Psittacara leucophthalmus*, Black-headed

Table 2. Top performing occupancy models for 13 psittacid species with at least one detection during NeoMaps surveys. The sampling size used to fit each model is shown as the total number of sites within species range sampled during NeoMaps surveys, as well as the number of sites where each species was detected is indicated (detections). AIC_c = corrected Akaike Information Criterion. ΔAIC_c = the difference between the AIC for the i_{th} model and the lowest AIC among all the models. AIC_w = relative weight from the differences in values of AIC_c . LL = 2log likelihood. The model with the best performance by species is in bold.

Specie	Model	Sampling size	Number of sites with detections	AIC_c	ΔAIC_c	AIC_w	LL
<i>Amazona amazonica</i>	p(h)Ψ(VC)	900	97	553.79	0	0.999	-265.75
	p(h)Ψ(C)	900	97	567.44	13.64	0.001	-274.62
	p(.)Ψ(VC)	900	97	673.87	120.07	0	-326.81
	p(.)Ψ(C)	900	97	697.20	143.41	0	-340.52
	p(.)Ψ(V)	900	97	712.80	159.01	0	-352.38
	Null	900	97	726.60	172.81	0	-361.29
<i>Amazona barbadensis</i>	p(h)Ψ(.)	150	5	55.07	0	0.571	-24.45
	Null	150	5	55.64	0.57	0.429	-25.78
<i>Amazona farinosa</i>	p(.)Ψ(VC)	550	17	158.55	0	0.94	-69.07
	p(.)Ψ(V)	550	17	164.08	5.54	0.059	-78.01
	p(.)Ψ(C)	550	17	171.35	12.8	0.002	-77.54
<i>Amazona ochrocephala</i>	p(.)Ψ(VC)	1,250	143	888.48	0	1	-434.15
	p(.)Ψ(C)	1,250	143	916.99	28.52	0	-450.44
	p(.)Ψ(V)	1,250	143	987.57	99.1	0	-489.77
	Null	1,250	143	1042.13	153.65	0	-519.06
<i>Ara chloropterus</i>	p(h)Ψ(.)	1,050	13	149.49	0	0.964	-71.74
	Null	1,050	13	156.08	6.58	0.036	-76.03
<i>Ara macao</i>	p(.)Ψ(C)	600	8	79.23	0	0.999	-31.49
	p(.)Ψ(V)	600	8	93.84	14.61	0.001	-42.89
	Null	600	8	101.00	21.77	0	-48.49
	p(h)Ψ(.)	600	8	102.79	23.57	0	-48.38
<i>Ara militaris</i>	p(h)Ψ(V)	150	8	71.81	0	0.365	-30.7
	p(.)Ψ(V)	150	8	72.39	0.58	0.273	-32.06
	p(h)Ψ(.)	150	8	72.61	0.8	0.245	-33.22
	Null	150	8	74.10	2.29	0.116	-35.01
<i>Ara severus</i>	p(.)Ψ(V)	700	61	450.52	0	1	-221.23
	Null	700	61	482.05	31.52	0	-239.02

Table 2. Continued.

Species	Model	Sampling size	Number of sites with detections	AICc	Δ AICc	AICw	LL
<i>Psittacara leucophthalmus</i>	p(h)Ψ(.)	450	7	80.87	0	0.712	-37.41
	Null	450	7	82.69	1.81	0.288	-39.33
<i>Eupsittula pertinax</i>	p(.)Ψ(VC)	1,300	190	1177.36	0	0.666	-578.59
	p(h)Ψ(VC)	1,300	190	1178.85	1.5	0.315	-578.32
	p(.)Ψ(C)	1,300	190	1185.12	7.76	0.014	-584.5
	p(h)Ψ(C)	1,300	190	1186.87	9.52	0.006	-584.37
	p(.)Ψ(V)	1,300	190	1284.34	106.98	0	-638.15
	p(h)Ψ(V)	1,300	190	1286.35	108.99	0	-638.15
	Null	1,300	190	1293.65	116.29	0	-644.82
	p(h)Ψ(.)	1,300	190	1295.62	118.27	0	-644.8
<i>Psittacara wagleri</i>	p(h)Ψ(.)	250	4	47.11	0	1	-20.51
<i>Brotogeris chrysopterus</i>	Null	450	2	31.54	0	0.734	-13.76
	p(h)Ψ(.)	450	2	33.56	2.03	0.266	-13.76
<i>Brotogeris jugularis</i>	p(h)Ψ(C)	650	20	174.63	0	0.551	-78.17
	p(.)Ψ(C)	650	20	175.43	0.8	0.369	-79.6
	p(.)Ψ(V)	650	20	179.17	4.54	0.057	-85.55
	p(h)Ψ(V)	650	20	180.90	6.27	0.024	-85.4
	Null	650	20	199.99	25.36	0	-97.99
	p(h)Ψ(.)	650	20	200.78	26.15	0	-97.37
<i>Diopsittaca nobilis</i>	Null	450	2	31.56	0	1	-13.76
<i>Forpus passerinus</i>	p(h)Ψ(C)	1,050	47	399.47	0	1	-190.65
<i>Forpus passerinus</i>	Null	1,050	47	441.40	41.93	0	-218.69
	p(h)Ψ(.)	1,050	47	442.01	42.53	0	-217.99
	p(.)Ψ(V)	1,050	47	443.34	43.87	0	-217.65
	p(h)Ψ(V)	1,050	47	444.10	44.63	0	-217.02
	Null	150	1	17.04	0	1	-6.48
<i>Orthopsittaca manilata</i>	p(h)Ψ(.)	450	5	63.46	0	0.522	-28.7
	Null	450	5	63.64	0.17	0.478	-29.8
<i>Pyrilia barrabandi</i>	Null	50	2	22.75	0	0.744	-9.25
	p(h)Ψ(.)	50	2	24.89	2.14	0.256	-9.18

Table 2. Continued.

Specie	Model	Sampling size	Number of sites with detections	AICc	Δ AICc	AICw	LL
<i>Pionites melanocephala</i>	p(h)Ψ(.)	350	5	60.64	0	1	-27.28
<i>Pionus menstruus</i>	p(h)Ψ(VC)	850	47	361.18	0	0.782	-169.43
	p(h)Ψ(.)	850	47	364.38	3.2	0.158	-179.18
	p(h)Ψ(V)	850	47	366.30	5.12	0.06	-178.11
	p(.)Ψ(VC)	850	47	401.83	40.65	0	-190.79
	p(.)Ψ(V)	850	47	415.45	54.27	0	-203.7
	Null	850	47	419.05	57.87	0	-207.52
<i>Pyrrhura egregia</i>	p(.)Ψ(V)	150	3	32.51	0	0.46	-12.12
	Null	150	3	33.49	0.98	0.282	-14.71
	p(h)Ψ(V)	150	3	34.65	2.14	0.158	-12.12
	p(h)Ψ(.)	150	3	35.58	3.06	0.1	-14.71
<i>Pyrrhura hoematotis</i>	Null	100	1	15.32	0	1	-5.6
<i>Pyrrhura melanura</i>	p(h)Ψ(.)	50	3	31.24	0	1	-12.36
<i>Pyrrhura picta</i>	p(.)Ψ(V)	350	10	89.88	0	0.724	-40.88
	p(h)Ψ(V)	350	10	91.82	1.94	0.275	-40.82
	Null	350	10	104.15	14.26	0.001	-50.06
	p(h)Ψ(.)	350	10	106.02	16.14	0	-49.98

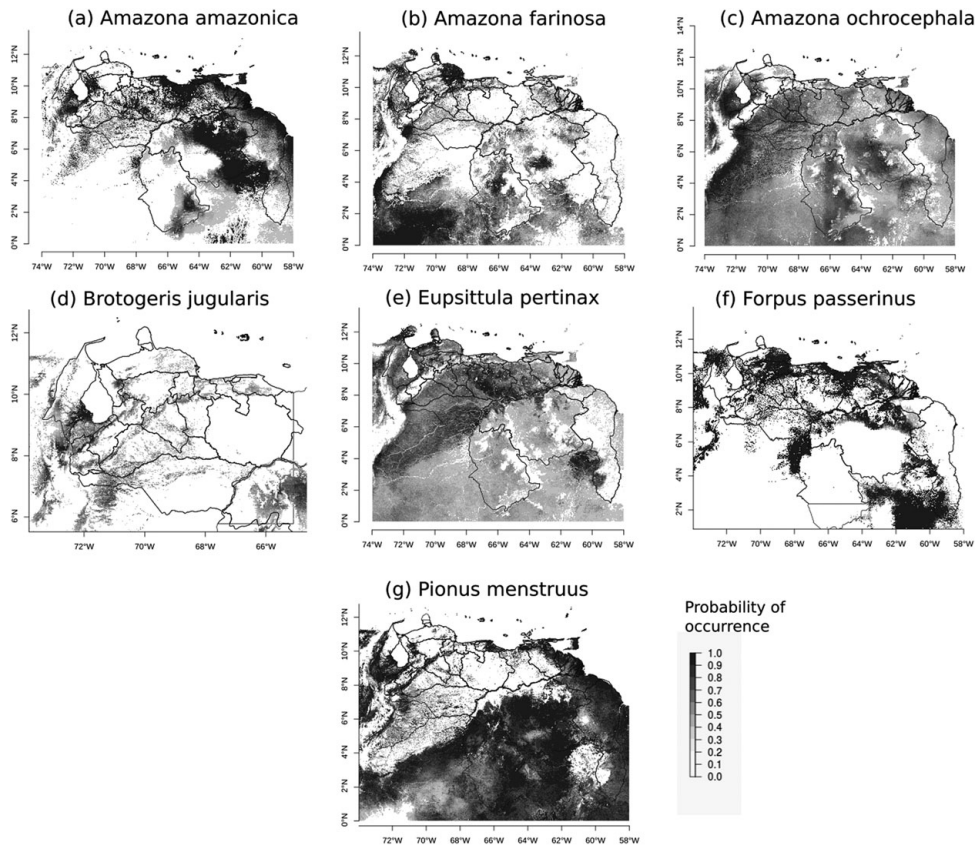


Figure 2. Spatial prediction of the (unconditional) probability of occurrence for the whole country based on the model with highest support for each species (Table 2) and the values of the vegetation and climatic covariates. Darker colours indicate higher probabilities. a) *Amazona amazonica*; b) *Amazona farinosa*, c) *Amazona ochrocephala*; d) *Brotogeris jugularis*; e) *Eupsittula pertinax*; f) *Forpus passerinus*; g) *Pionus menstruus*.

Parrot *Pionites melanocephala*, Military Macaw *Ara militaris*, Yellow-shouldered Amazon *Amazonas barbadensis*, Blue-headed Parrot and Orange-winged Amazon *Amazona amazonica* the probability of detection was low with important variation across the time, but for *Brotogeris jugularis* the detection was also variable but higher (Figure 2a).

For three species, the models with constant probability of occurrence ($p(h)\psi(\cdot)$) had the lowest AICc. Models including vegetation covariates either assuming constant detectability ($p(\cdot)\psi(V)$) or not ($p(h)\psi(V)$) had the lowest AICc for two species, *Ara militaris* and Painted Parakeet *Pyrrhura picta*. However due the low number of detections (< 10) we were not able to perform spatial prediction of unconditional probability of occurrence for these species.

Models including both climatic and vegetation covariates either assuming constant detectability ($p(\cdot)\psi(VC)$) or not ($p(h)\psi(VC)$), were selected for five species (Table 2). The spatial prediction of unconditional probability of occurrence for *Amazona amazonica*; Figure 2a) showed a widespread distribution, with the highest values in the most forested and humid part of the country, in the east (south and north of the Orinoco river) and in the west (the Maracaibo Lake basin). For Southern

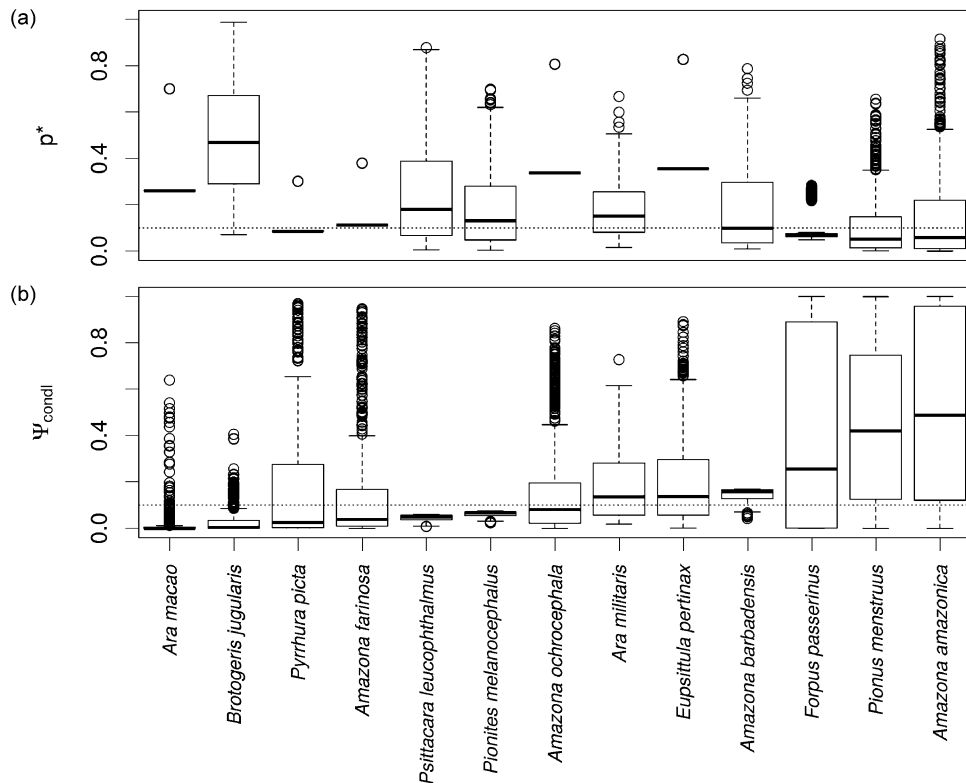


Figure 3. Model predictions. a) Detection probability (p^*) = Sampling effort required to detect the species at least once conditional on its presence. b) Conditional probability of presence given that the species was not detected (Ψ_{CONDNL}).

Mealy Amazon *Amazona farinosa*; Figure 2b) the highest probabilities of occurrence were in the dry areas along the coast, north of the country. Yellow-crowned Amazon *Amazona ochrocephala*; Figure 2c) and Brown-throated Parakeet *Eupsittula pertinax*; Figure 2e), show a widespread distribution across the country, only excluded from the Venezuelan Andes (Figure 2c,e). However, for *E. pertinax*, higher probabilities were predicted in the central flood plains, and the north-west, characterised by high temperature and lower vegetation cover (Figure 2e), while for *A. ochrocephala* higher probabilities were focused on more humid areas with moderate forest cover.

Models including only climatic covariates had the lowest AICc for two species, Orange-chinned Parakeet *Brotogeris jugularis* and Green-rumped Parrotlet *Forpus passerinus*. *B. jugularis* showed a restricted distribution focused on the western part of the country, in the Maracaibo Lake basin (Figure 2d), while *F. passerinus*, Figure 2f) had the highest probabilities in the dry areas across the coastal north.

Conditional probability of occurrence

For most species a great proportion of sites which lacked detections had low conditional probabilities of occurrence ($\Psi_{\text{CONDNL}} < 0.2$). For *Ara macao*, *Amazona ochrocephala*, *Brotogeris jugularis* and *Eupsittula pertinax* the model estimated extremely high probabilities of detection (Figure 3a) and low probabilities of occurrence (Figure 3b), suggesting that the species are reliably detected were they are present. For

Forpus passerinus, *Pionus menstrus* (Blue-headed Parrot), and *Amazona amazonica* the situation was the opposite, suggesting that the species is often present but seldom detected (Figure 3a,b).

Discussion

The observed declines in psittacid populations across Venezuela highlights the need for monitoring programmes that can reliably detect occurrence in a cost-effective and logistically feasible manner. The first national bird monitoring program in Venezuela developed by NeoMaps is an important step to achieve this aim by providing confident detection and non-detection records for 92% of psittacid species in the country (48 species with sites sampled). With this data we were able to fit occupancy models for 50% of Venezuelan psittacid species, which provide: 1) more reliable data on species absences, 2) better understanding of the importance of factors affecting psittacid occurrence, and 3) improving sampling strategy to get more confident occurrence probabilities.

Reliability of species absence

For highly mobile species such as parrots, the lack of detections is likely to be a combination of insufficient sampling effort and true absences during the time of survey (Ferrer-Paris *et al.* 2014). For at least nine of the missing species, the lack of detection is most probably due to low sampling effort within their distribution (less than four transects, 200 sampling sites). For *Aratinga solstitialis* and Spectacled Parrotlet *Forpus conspiciellatus*, no single NeoMaps survey locality matched the expected distribution (Table 2). For eight further species the expected distribution overlapped with survey localities along a single transect (less than 50 sampling sites; *Amazona autumnalis*, *Ara militaris*, *Brotopogon cyanoptera*, *Hapalopsittaca amazonina*, *Pionus chalcopterus*, *Pyrrhura barrabandi*, *Pyrrhura caeruleiceps* and *Pyrrhura melanura*). All these species require targeted sampling in order to monitor their populations in the future. The most notable “absences” are those of Red-fan Parrot *Deroptyus accipitrinus*, Caica Parrot *Pyrrhura caica* and Dusky Parrot *Pionus fuscus*, which were expected in six or seven transects (more than 300 sampling sites) south of the Orinoco river, but were never detected. For these species the presence records in GBIF during 2010 were also scarce (< 8; Table 2). Several species of Neotropical psittacids perform seasonal movements following availability of food as well as adapting to novel foods in modified environments (Juniper and Park 1998). So in those cases of non-detections in spite of the high sampling effort, taking into account seasonal movements during the sampling design would likely improve the detection probability. Evidence from Río Manu in Peru shown a three-fold decline in the number of large macaws encountered during the dry season compared with the rainy season, which coincides with a sharp decline in plant energy production of the forest during the dry season (Renton 2002).

For the species with low probability of detections (Figure 3a), we were able to provide useful insights to improve occurrence predictions by analysing the components of conditional probability of occurrence in those sites where the species was expected to occur, but was not detected. For example for three species (*Forpus passerinus*, *Pionus menstrus*, and *Amazona amazonica*), the probability of detection taking into account the current sampling effort was too low (< 0.2) in areas where environmental conditions would imply high probabilities of presence (Figure 3a,b). This suggests that sampling effort should be increased to generate reliable estimations of occurrence. Evidence from parrot communities in northern Bolivia suggest that detection probability significantly improves with a larger sampling effort (nine weeks; Berkunsky *et al.* 2015). Recent empirical studies suggest that the detectability rate is significantly correlated with sighting frequency, and less conspicuous psittacids may require longer observation session in order to register the presence of some species (Rodrigues *et al.* 2012).

Factors affecting species occurrence

Factors related to vegetation, climatic conditions or both were important in explaining distributions of 10 species (Table 2; Figure 2). We used remotely sensed data to better represent the vegetation and climatic conditions prior to NeoMaps' sampling period, which were heavily influenced by one episode of the 'El Niño-Southern Oscillation' (ENSO; May 2009 and April 2010), resulting in a severe drought, diminished water bodies and overall drier vegetation throughout the country, which in turn could have affected the probability of detection or occurrence of some species (Lentino and Portas 1994, Hilty 2003). A low number of detections and subsequent low probability of occurrence of some widespread species in the Orinoco floodplains (llanos) could thus be explained by this extended drought and its consequences on vegetation growth and resource availability (Ferrer-Paris *et al.* 2013).

We were able to predict spatial distribution for seven species (Figure 2), which reflects in general the expected distribution for these species, but for *Amazona farinosa* (Figure 2b) the model also predicts high probabilities in the north-central region (Falcón and Lara states) where the species is absent, probably due to biogeographic constraints or the influence of additional variables not included in our model.

The low number of detections of several species often resulted in the selection of null and constant models, with low predictive power. However, for three species (*Brotogeris jugularis*, *Amazona ochrocephala* and *Eupsittula pertinax*; Figure 3a) with high probability of detection, better models could be obtained by including other predictive variables. This is similar to other studies where, for example, the distribution of food resources, improved the model predictions for both specialist and generalist parrot species in the Brazilian cerrado (De Araújo *et al.* 2014). Topographic variables like slope and orientation also determined the distribution pattern of Mexican psittacid species, because they account for complexity in the landscape, especially at local scales (Plasencia-Vázquez *et al.* 2014).

Improving sampling strategy

Our study indicates that important changes in the sampling design and modeling approach are necessary to improve occurrence predictions in those sites where a species is expected to occur but is not detected. Given the low ratio of actual/expected detections (~10%), we suggest that more effective parrot monitoring programmes require: a) increasing sampling effort to improve estimates of probability of occurrence for all psittacids in Venezuela. This implies adding more sampling localities and days, and optimising survey time to those periods of the day when detection probabilities are higher; b) implementing additional surveys per year to improve estimates of seasonal patterns; c) including variables related with temporal use of resources and habitat heterogeneity in the survey and models, and d) alternatively, combining records from systematic surveys with other sources of data (collections, literature, GBIF) which could significantly increase the sample size allowing fitting of more informative models (Ferrer-Paris *et al.* 2014).

Supplementary Materials

To view supplementary material for this article, please visit <http://dx.doi.org/10.1017/S0959270919000522>.

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