cambridge.org/par

Research Article

Cite this article: Hiller T, Honner B, Page RA, Tschapka M (2018). Leg structure explains host site preference in bat flies (Diptera: Streblidae) parasitizing neotropical bats (Chiroptera: Phyllostomidae). *Parasitology* **145**, 1475–1482. https://doi.org/10.1017/S0031182018000318

Received: 8 November 2017 Revised: 2 January 2018 Accepted: 8 February 2018 First published online: 22 March 2018

Key words:

Bat flies; co-occurrence; ecomorphology; parasite interactions; phyllostomidae

Author for correspondence:

Thomas Hiller, E-mail: thomas.hiller@alumni. uni-ulm.de

Leg structure explains host site preference in bat flies (Diptera: Streblidae) parasitizing neotropical bats (Chiroptera: Phyllostomidae)

Thomas Hiller^{1,2}, Benjamin Honner¹, Rachel A. Page² and Marco Tschapka^{1,2}

¹Institute of Evolutionary Ecology and Conservation Genomics, University of Ulm, Albert-Einstein-Allee 11, Ulm 89081, Germany and ²Smithsonian Tropical Research Institute, Apartado 0843-03092, Balboa, Panamá

Abstract

Bat flies (Streblidae) are diverse, obligate blood-feeding insects and probably the most conspicuous ectoparasites of bats. They show preferences for specific body regions on their host bat, which are reflected in behavioural characteristics. In this study, we corroborate the categorization of bat flies into three ecomorphological groups, focusing only on differences in hind leg morphology. As no detailed phylogeny of bat flies is available, it remains uncertain whether these morphological differences reflect the evolutionary history of bat flies or show convergent adaptations for the host habitat type. We show that the division of the host bat into three distinct habitats contributes to the avoidance of interspecific competition of bat fly species. Finally, we found evidence for density-dependent competition between species belonging to the same ecomorphological group.

Introduction

Approximately half of all animal species, from nearly all orders (Poulin and Morand, 2000), show a facultative or obligatory parasitic lifestyle (Windsor, 1998). Traditionally these are divided into two groups: microparasites, i.e. single-celled organisms, and macroparasites, which primarily include a variety of endoparasitic helminths and ectoparasitic insects (Morand *et al.* 2006). Every host individual provides a discontinuous habitat patch that may support a discrete community of parasites (Bush *et al.* 1997; Gotelli and Rohde, 2002). Resource partitioning between parasitic species reflects strategies to avoid competition on a host (Schoener, 1974; Mouillot *et al.* 2003) and is often correlated with ecomorphological and/or behavioral adaptations, for example to an efficient use of particular locations on the host (Bush *et al.* 1997; Linhares and Komeno, 2000; Hsu *et al.* 2002; ter Hofstede *et al.* 2004; Tello *et al.* 2008).

In this context, bats are an interesting study system, as they offer at least two distinctly different habitat surfaces, the furry body and the furless wing membranes (Wenzel et al. 1966; ter Hofstede et al. 2004). In addition, many bats are hosts for a variety of ectoparasites, such as bat flies, which are obligate parasites and highly host-specific (Wenzel et al. 1966; Wenzel, 1976; Dick and Patterson, 2007). Bat flies are generally assigned to two families, the monophyletic Nycteribiidae and the paraphyletic Streblidae (Dittmar et al. 2015). In response to bat species richness, both parasite families are most diverse in tropical regions, with Nycteribiidae being most speciose in the Old World and Streblidae in the New World (Dick and Miller, 2010; Graciolli, 2010). The majority of the neotropical Streblidae are parasites of the highly diverse leaf-nosed bats (Phyllostomidae) (Wenzel et al. 1966; Wenzel, 1976; Dick and Gettinger, 2005). Due to high host specificity, bat flies form distinct parasite communities on each host species. Such a community may include 2-5 species, with each parasite species showing preferences for a specific body area on the host (Wenzel, 1976; ter Hofstede et al. 2004; Dick and Gettinger, 2005; Tello et al. 2008; Patterson et al. 2009). Without quantifying Wenzel et al. (1966) pointed out when several bat fly species are occurring on the same host, they are usually morphologically different types, presumably specialized for living on different body regions of the host. Based on the body regions from which the parasites were primarily collected, ter Hofstede et al. (2004) distinguished two distinct groups, one associated with the wing membranes and the other associated with the fur of the host bat. Dick (2005) included behavioural observations and morphological characteristics to define three ecomorphological groups, assigning each to a niche on the bat host: some bat fly species live almost exclusively on the wing membranes ('wing crawlers'), others stay mainly on the furred body, with some moving on the fur ('fur runner') and some moving through it ('fur swimmer'). While tarsal claws are well developed in all three groups and important for attachment (Dick and Patterson, 2006), overall leg morphology is also crucial for efficient movement of insects and therefore also for bat flies (Full and Tu, 1991).

Leg morphology is directly correlated to habitat preferences in anoles, lizards and spiders, and may be a key factor in mechanisms of coexistence (Losos and Sinervo, 1989; Beuttell and Losos, 1999; Lapinski *et al.* 2015). In recent years, various studies have focused on the co-occurrence of bat flies, revealing positive correlations in abundance among sympatric

© Cambridge University Press 2018



streblid species on a given host bat, indicating niche partitioning between bat fly species parasitizing the same host species (Tello *et al.* 2008; Patterson *et al.* 2009; Presley, 2010).

The morphological characterization of the three behavioural groups is based on 12 characteristics (head, thorax, fore-, midand hindleg) (Dick, 2005). In this study we attempt to corroborate the categorization of streblid bat fly species into these groups using a reduced set of characteristics, focusing only on the morphology of the hind leg. Using only the hind leg offers a highly reproducible approach and provides measurements, which are easily comparable between ecomorphological groups. Further, we offer new implications for the inclusion of bat fly niche partitioning in models of species co-occurrence and species competition.

Material and methods

All streblid flies were collected within a large-scale bat diversity assessment in central Panama between 2013 and 2015. Bats were captured at 18 different locations with mist nets (Ecotone, $6 \times 2.5 \text{ m}^2$, 16 mm mesh and 2×70 denier) set at ground level, checked every 15 min. After capture, bats were held in clean, soft cotton bags for up to 1 h until processing at an improvised field camp at the capture location. Wings, dorsal and ventral pelage of every bat individual were thoroughly examined for the presence of streblid bat flies. Bat flies were collected with entomological forceps and stored in individual vials containing 96% ethanol. We further recorded standard body measurements, age, sex and reproductive status as well as further samples, totalling the handling time of each bat to 10-15 min. In the laboratory, all collected bat flies were counted and identified to species level (Wenzel et al. 1966; Wenzel, 1976; Guerrero, 1993, 1994a, b, 1995a, b, 1996, 1997, 1998). Based on the literature and prior personal observations, all species were assigned to one of the

following three behaviourally distinct groups: *wing crawler* for bat fly species living on the wing membranes, *fur runner* for species moving along the surface of the host fur, and *fur swimmer* for species moving through the host fur (Dick, 2005).

The main differences between these three groups are the preference of a particular site on the bat host as well as their movement behaviour within these niches. To investigate if these three designated groups correlate with morphological characteristics, we focused on the structure of the hind legs, as these are crucial for movement on the host and qualitative observations of structural differences are mentioned in the literature (ter Hofstede *et al.* 2004; Dick, 2005). Insect legs allow standardized measurements of the clearly separable leg sections: femur, tibia and tarsi. Additionally, the thickness of the femur provides more information as it can give indications on the muscular force of the leg, characterizing insect locomotion (Graham, 1985).

Under a stereomicroscope, we used a scalpel to separate the right hind legs of 110 bat flies (Streblidae) from 20 species, collected from 12 species of phyllostomid bats (Table 1). We focused on streblid species that commonly co-occur on their host bats, covering 10 genera of Streblidae with species of different size and host associations. We photographed the separated legs, and with the software ImageJ [version 1.50i (Rasband, 1997–2016)], measured in pixels the length of the femur, tibia and tarsi, including the terminal claws, as well as the width of the femur at half distance (Fig. 1). To account for size differences between bat fly species and degree of magnification, we used ratios for our analyses and standardized all length measurements by dividing them by the total leg length. We standardized the femur width by dividing it by the femur length.

We conducted all statistical analyses in R v3.3.1 (R Development Core Team, 2016). We tested the individual leg measurements for differences between the three bat fly groups with Kruskal–Wallis rank sum tests [kruskalmc(), R-package

Table 1. Bat fly species included for the analysis, with associated species and assigned behavioral group

Bat fly species (<i>n</i>)	Bat host species	Behavioural group	Source	
Aspidoptera phyllostomatis (7)	Artibeus jamaicensis	Wing crawler	Dick, 2005	
Megistopoda aranea (7)	A. jamaicensis	Fur runner	Dick, 2005	
Mastoptera guimaraesi (5)	Phyllostomus hastatus	Wing crawler	Dick, 2005	
Metelasmus pseudopterus (5)	A. jamaicensis	Fur swimmer	Dick, 2005	
Neotrichobius stenopterus (5)	Dermanura phaoetis & D. watsoni	Fur runner	ter Hofstede et al. 2004, personal observation	
Paratrichobius dunni (5)	Uroderma bilobatum	Fur runner	Dick, 2005	
Paratrichobius longicrus (5)	Artibeus lituratus	Fur runner	ter Hofstede et al. 2004, personal observation	
Speiseria ambigua (5)	Carollia perspicillata	Fur runner	Dick, 2005	
Strebla christinae (5)	Phylloderma stenops	Fur swimmer	personal observation	
Strebla galindoi (5)	Tonatia saurophila	Fur swimmer	personal observation	
Strebla guajiro (5)	C. perspicillata	Fur swimmer	Dick, 2005	
Strebla hertigi (5)	Phyllostomus discolor	Fur swimmer	Dick, 2005	
Strebla mirabilis (5)	Phyllostomus hastatus	Fur swimmer	Dick, 2005	
Strebla wiedemanni (5)	Desmodus rotundus	Fur swimmer	Dick, 2005	
Trichobioides perspicillatus (5)	P. discolor	Wing crawler	Dick, 2005	
Trichobius costalimai (5)	P. discolor	Wing crawler	Dick, 2005	
Trichobius dugesioides (5)	T. cirrhosis	Wing crawler	Dick, 2005	
Trichobius joblingi (7)	C. perspicillata	Wing crawler	Dick, 2005	
Trichobius longipes (5)	P. hastatus	Wing crawler	Dick, 2005	
Trichobius parasiticus (5)	D. rotundus	Wing crawler	Dick, 2005	

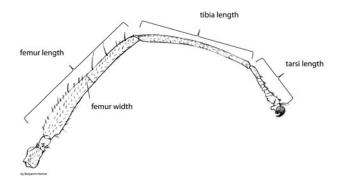


Fig. 1. Schematic depiction of a streblid bat fly leg, indicating the measurements for analysis.

pgirmess, Giraudoux, 2016]. Using nonmetric multidimensional scaling [metaMDS(), R-package vegan, Oksanen *et al.* 2016], we visualized the distance matrix based on the leg measurements (Bray–Curtis dissimilarity) and tested for significant differences among the observed bat fly groups using permutational multivariate analysis of variance [PERMANOVA, adonis(), 1000 permutations, R-package vegan].

As we were focusing only on possible negative correlations and Kendall's Tau is a more accurate test to use with smaller sample sizes, we further analysed the co-occurrence of bat fly species on their respective host species (Table 2, Supplementary Table S2) with one-tailed correlations [cor.test(), method = Kendall] for each bat fly species pair. To avoid large-scale and location-specific characteristics influencing the occurrence of bat fly species in our dataset, for this analysis we only analysed bat flies from host individuals captured at locations where we collected all focal bat fly species for a particular bat species. We

Table 2. Results for co-occurrence analyses (Kendall correlation) of bat fly species on their specific host bat species (N = number of pairs analysed)

Species combination	п	z-value	tau	P value
A. jamaicensis				
M. pseudopterus – A. phyllostomatis	40	-1.368	-0.206	0.086
M. pseudopterus – M. aranea	69	-0.642	-0.075	0.261
A. phyllostomatis – M. aranea	90	-5.357	-0.515	<0.001
C. perspicillata				
S. guajiro – S. ambigua	38	-5.261	-0.822	<0.001
S. guajiro – T. joblingi	136	-0.694	-0.053	0.244
S. ambigua – T. joblingi	140	-1.143	-0.086	0.127
D. rotundus				
S. wiedemanni – T. parasiticus	17	-1.221	-0.244	0.111
P. discolor				
S. hertigi – T. perspicillatus	25	-0.288	-0.050	0.387
S. hertigi – T. costalimai	21	0.997	0.185	0.841
T. perspicillatus – T. costalimai	25	0.459	0.072	0.677
P. hastatus				
S. mirabilis – M. guimaraesi	17	-1.458	-0.294	0.072
S. mirabilis – T. longipes	17	-0.088	-0.018	0.465
M. guimaraesi – T. longipes	18	-1.341	-0.254	0.090

also excluded double-zero records from our pairwise correlations (Tello *et al.* 2008).

Results

The different bat flies showed characteristic movement patterns on their host bat, especially when trying to evade capture. Wing crawlers moved slowly across the bats' wing membranes, often scuttling sideways like crabs, and attached strongly to the wing membrane when collected. Bat flies using the furry body parts of the host bats moved quickly, typically showing one of two modes of escape. Fur runners pushed through the fur to the surface, moving quickly to areas around the ears or the ventral axillary regions. Fur swimmers, in contrast, were mostly found in the long fur of the bats' neck, only noticeable when slightly blowing into the fur or by the wave-like fur movement when moving through the hairs, like velociraptors through high grass in the movie Jurassic Park (Spielberg, 1997). Unlike wing crawlers and fur runners, fur swimmers possess a ctenidium, or a necklace of spines on their ventral side, which allows them to cling to the bat hairs, sometimes so strongly that their legs or even heads would be ripped off when pulling them out of the fur with forceps. Based on these behavioural observations we can clearly distinguish these three groups of bat flies on phyllostomid bats.

Comparing the relative leg dimensions revealed significant differences in all four measurements between the three groups of bat flies (Fig. 2, Supplementary Table S1). Wing crawlers had a significantly shorter relative femur length than fur runners and fur swimmers (Kruskal–Wallis test, $\chi^2 = 42.8$, P < 0.001), whereas the latter two did not differ from each other. Tibia length differed significantly between all three groups (Kruskal–Wallis test, $\chi^2 =$ 73.0, P < 0.05) and was longest in fur runners and shortest in fur swimmers. Fur runners had significantly shorter relative tarsi length than the other two groups (Kruskal–Wallis, $\chi^2 =$ 66.0, P < 0.001). The relative width of the femur differed significantly between all three groups, with fur runners having the thinnest and wing crawlers the broadest femur (Kruskal–Wallis test, $\chi^2 = 96.0$, P < 0.001).

A non-metric multidimensional scaling (NMDS, final stress = 0.035, linear fit $r^2 = 0.996$) analysis on these leg measurements show a clear separation of all three groups (Fig. 3). A dissimilarity analysis shows that the three groups significantly differ from each other (Adonis, F = 480.92, $R^2 = 0.90$, P < 0.001).

With regard to species co-occurrence, we found negative interactions only in two species pairs [*Aspidoptera phyllostomatis vs Megistopoda aranea* (z = -5.357, t = -0.515, P < 0.001) and *Strebla guajiro vs Speiseria ambigua* (z = -5.261, t = -0.822, P < 0.001)], whereas the remaining 11 pairs showed no or positive density correlations (Table 2, Fig. 4).

Discussion

Our behavioural observations of 20 species of bat flies are consistent with reports in the literature (Wenzel *et al.* 1966; ter Hofstede *et al.* 2004; Dick, 2005) and allowed us to clearly distinguish three groups of bat flies based on movement patterns and niche use on their respective host bat. The occurrence of these three bat fly groups on single host individuals allow the suggestion of niche separation as a mechanism to avoid competition between co-occurring bat fly species (Wenzel *et al.* 1966; ter Hofstede *et al.* 2004; Tello *et al.* 2008; Presley, 2010). These three groups were first categorized as distinct ecomorphological groups by Dick (2005) who termed them as 'fur runners', 'fur swimmers' and 'wing crawlers'. Our behavioural observations of streblid bat flies parasitizing phyllostomid bats in Panama are consistent with these ecomorphological categorizations.

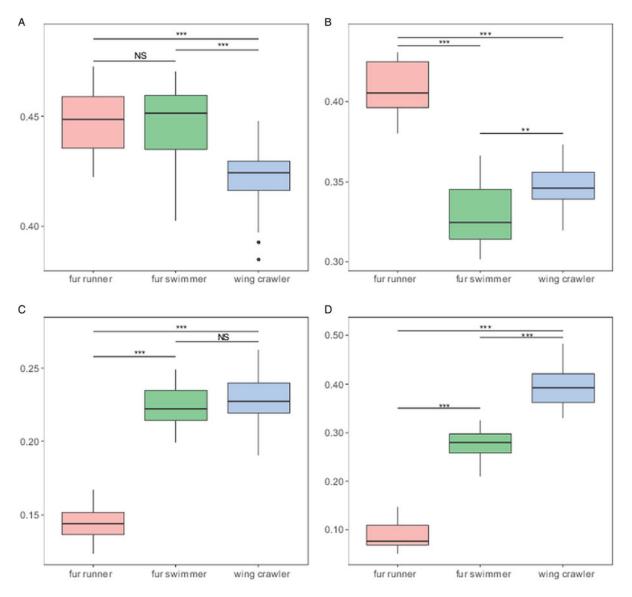


Fig. 2. Differences in relative (A) femur, (B) tibia and (C) tarsi length as well as (D) relative femur width for the three ecomorphologically adapted groups of bat flies (NS = not significant, ** = P < 0.1, *** = P < 0.01).

Our study fortifies the evidence for a clear morphological separation of these three groups of bat flies. The assignment of bat fly species to these three ecomorphological groups, based only on the hind-leg morphology, is consistent with the categorization based on 12 morphological measurements shown by Dick (2005).

Fur runners have long and thin legs which allow them to move quickly over the surface of the fur of their host bat, while wing crawlers have short and sturdy legs with massive femurs and pronounced tarsi and terminal claws that facilitate the attachment to the hairless and exposed wing membranes (Wenzel *et al.* 1966; Dick, 2005; Dick and Patterson, 2006). Fur swimmers combine characteristics of the other two groups, namely long and strong legs that allow them to move quickly within the fur of their host bats (Wenzel *et al.* 1966; Dick, 2005).

Whether these morphological adaptations have evolved several times independently, or whether the distinct ecomorphological groups represent phylogenetic units is still unclear, as no concise phylogeny of New World Streblidae exists (Dick and Patterson, 2006; Dittmar *et al.* 2006, 2015). Five of the currently recognized eleven subfamilies of Phyllostomidae are parasitized by members of all three ecomorphological groups, whereas flies of two groups can be found in four more subfamilies (Wenzel *et al.* 1966; Wenzel, 1976; Cirranello *et al.* 2016; Rojas *et al.* 2016). This

strongly indicates a shared origin for these three groups, as convergent developments are possible in some cases, but highly unlikely for all of them. This coincides well with the evidence for a long history of association between bat flies and bats, with the currently oldest fossil record of bat flies, a specimen preserved in amber (*Enischnomya stegosoma*, Nycterophiliinae), dating back to the upper Eocene to middle Miocene (Schlee, 1990; Iturralde-Vinent and MacPhee, 1996; Poinar and Brown, 2012), while the separation of Phyllostomidae and Mormoopidae is estimated to have occurred in the late Eocene and the diversification of Phyllostomidae is assumed to have started in the Oligocene, around 30 ma ago (Rojas *et al.* 2016).

On extant neotropical bat species we frequently find several co-occurring bat fly species, but generally only one representative species for each ecomorphological group (Wenzel, 1976; Fritz, 1983; Linhares and Komeno, 2000; Dick and Miller, 2010). The abundance of one bat fly species is often positively correlated with the abundance of other species inhabiting the same host individual (Wenzel *et al.* 1966; Dick and Patterson, 2006; Presley, 2007, 2010; Tello *et al.* 2008). This contrasts apparently other co-occurrence studies suggesting density compensation, an inverse relationship between population density and species diversity generated by competition between different species

1479

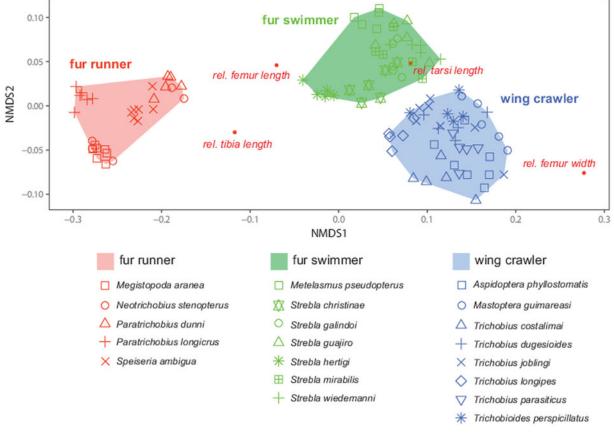


Fig. 3. Non-metric multidimensional scaling (NMDS) showing significant separation of three ecomorphological groups (Adonis, F = 484.08, $R^2 = 0.90$, P < 0.001). Red asterisks represent weighted averages for the corresponding leg measurements.

(Dick and Patterson, 2006; Presley, 2007, 2010; Tello et al. 2008). The results of our density correlations are consistent with these findings, showing no significant negative correlation for 14 of 16 bat fly species interactions on six bat species, indicating strong niche differentiation. In choice experiments bat flies (Trichobius joblingi) preferred clean bats over bats already infested with individuals of the same parasite species, whereas no preference could be detected when offering them clean bats and bats infested with a bat fly species (Speiseria ambigua) occupying another niche on the host bat (Dick and Dick, 2006). This indicates that intraspecific competition might be less influential than interspecific competition, whereas interspecific competition might be largely synonymous to 'inter-ecomorphological group' competition. The threshold for the existence of competition effects might also depend on host size and therefore habitat size or food availability for the parasites. The two medium-to-large-sized bat species of the genus Phyllostomus, P. discolor and P. hastatus, were the only records in our data set exhibiting the co-occurrence of two bat fly species of the same ecomorphological group. In both cases, we could not find any negative interactions between the two species of wing crawlers. Similar results were obtained by Presley (2007) for two parasites with the same habitat preference found on the large, 60 cm wingspan Noctilio leporinus in Paraguay, showing positive correlations in both occurrence and abundance.

On two host bat species we observed negative density correlations between two bat fly species. The first observed significant negative correlation between *S. guajiro vs S. ambigua* on *Carollia perspicillata*, was also reported by Tello *et al.* (2008). These two bat flies belong to the ecomorphological group of fur swimmer and fur runner, respectively, using both the furry body area of the host bat (Fritz, 1983; Tello *et al.* 2008). This behaviour may cause resource competition, especially on smallsized bats. The second pair of bat fly species showing a significant negative density correlation occurs on different body regions of the host species Artibeus jamaicensis, namely wing membranes and on the fur. We did not expect this strong negative interaction between these two ecomorphological groups, especially not on large-sized host bats. However, a peculiarity of the host bat species A. jamaicensis is its flexibility in roost sites, using caves, hollow trees or the foliage (Handley and Gardner, 1991). As bat flies are obligated to leave their host bat to deposit the pupae in the roost (Overal, 1980; Dick and Patterson, 2006), the bats' roost is highly important for maintaining the bat fly life cycle. Every roost type offers a different habitat longevity as well as a characteristic microclimate and substrate (Patterson et al. 2007; Dittmar et al. 2009), which may favour the abundance of one bat fly species over the other. Depending on the roost characteristics, the manner of pupiposition may also differ among bat fly species. Pupiposition can occur at different distances to the host bat, with winged species depositing farther away than species with reduced wings (Ross, 1961; Overal, 1980; Dittmar et al. 2009). Pupae of e.g. Trichobius-, Speiseria- or Megistopoda-species are deposited to the substrate of the roost walls (Overal, 1980; Fritz, 1983; Dittmar et al. 2009), whereas in contrast, pupae of flies of the genus Strebla and Nycterophilia do not possess pronounced attachment processes and therefore might be dropped freely in the roost (Wenzel in Marshall, 1981; Fritz, 1983). Fritz (1983) recovered in experiments with caged host bats pupae of St. guajiro and St. wiedemanni on the ground, whereas pupae of T. joblingi and Sp. ambigua were directly attached to the cage. This indicates that not only competition but also a wide range of other factors such as roosting habits may influence the occurrence of streblid bat fly species on their host bats (Wenzel et al. 1966;

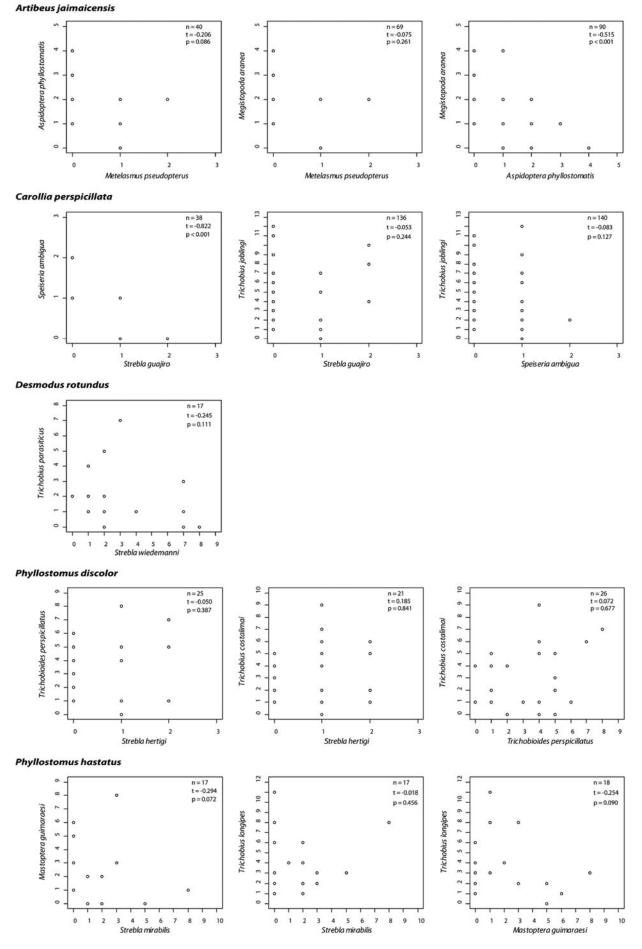


Fig. 4. Scatterplots of densities for all bat fly species pairs occurring on their respective host bat species.

ter Hofstede and Fenton, 2005; Patterson *et al.* 2007). Roost characteristics may, therefore, play a major role in the bat fly species composition and may even mask competition between congeneric bat fly species.

In sum, we confirmed three distinct ecomorphologies in streblid bat flies, each preferring a different habitat on their host. We found significant differences in leg morphology associated with these distinct ecomorphological groups. As to date, no detailed phylogeny of bat flies is available, it remains uncertain whether these ecomorphological groups reflect phylogeny or whether adaptations to different habitat use evolved several times independently within the New World Streblidae. The division of the bat host into three structurally distinct habitats contributes to avoiding competition among the ectoparasites. Additionally, we found evidence for density-dependent competition between species belonging to the same ecomorphological group. Finally, we found evidence that bat fly species composition might differ depending on the ecological background, with quality and availability of roosting sites favouring the abundance of one bat fly species over the other. Further studies are needed focusing on co-occurrences and community ecology of bat flies, integrating roosting information of host bats in the field.

Supplementary material. The supplementary material for this article can be found at https://doi.org/10.1017/S0031182018000318

Acknowledgements. We thank the Smithsonian Tropical Research Institute for the use of facilities and for logistical support, especially Oris Acevedo and Belkys Jimenez. Special thanks go to Stefan Brändel and his field crew for extensive bat fly sampling. We also thank Carl Dick for valuable input and comments on this manuscript. Animal capture and handling were approved by the Panamanian Government (MiAmbiente SE/A-75-13, SE/A-69-14) and the Institutional Animal Care and Use Committee (IACUC, 2013-0401-2016).

Financial support. This study was funded by the German Research Foundation (DFG) SPP 1596 Grant TS 81/6-1 to M.T., which had no influence in study design or interpretation of the results.

References

- Beuttell K and Losos JB (1999) Ecological morphology of Caribbean anoles. Herpetological Monographs 13, 1–28.
- Bush AO, Lafferty KD, Lotz JM, Shostak AW (1997) Parasitology meets ecology on its own terms: Margolis *et al.* revisited. *Journal of Parasitology* 83, 575–583.
- Cirranello A, Simmons NB, Solari S and Baker RJ (2016) Morphological diagnoses of higher-level phyllostomid taxa (Chiroptera: Phyllostomidae). *Acta Chiropterologica* **18**, 39–71.
- Dick C (2005) Ecology and host specificity of Neotropical bat flies (Diptera: Streblidae) and their chiropteran hosts. Lubbock, Texas, USA: Texas Tech University, p. 265.
- Dick CW and Dick SC (2006) Effects of prior infestation on host choice of bat flies (Diptera: Streblidae). *Journal of Medical Entomology* **43**, 433–436.
- Dick CW and Gettinger D (2005) A faunal survey of streblid flies (Diptera: Streblidae) associated with bats in Paraguay. *Journal of Parasitology* **91**, 1015–1024.
- Dick CW and Miller J (2010) Streblidae. In Brown B, Borkent A, Cumming J, Wood D, Woodley N and Zumbado M (eds)). *Manual of Central American Diptera*. Costa Rica: Los Angeles County Museum of Natural History and Instituto Nacional de Biodiversidad, vol. 2, pp. 1249–1260.
- Dick CW and Patterson BD (2006) Bat flies: obligate ectoparasites of bats. In Morand S, Krasnov BR and Poulin R (eds)). *Micromammals and Macroparasites: From Evolutionary Ecology to Management*. Tokyo: Springer, pp. 179–191.
- Dick CW and Patterson BD (2007) Against all odds: explaining high host specificity in dispersal-prone parasites. *International Journal for Parasitology* 37, 871–876.
- Dittmar K, Porter ML, Murray S and Whiting MF (2006) Molecular phylogenetic analysis of nycteribiid and streblid bat flies (Diptera: Brachycera,

Dittmar K, Dick CW, Patterson BD, Whiting MF and Gruwell ME (2009) Pupal deposition and ecology of bat flies (Diptera: Streblidae): *Trichobius* sp. (Caecus group) in a Mexican cave habitat. *Journal of Parasitology* **95**, 308–314.

gins. Molecular Phylogenetics and Evolution 38, 155-170.

- Dittmar K, Morse SF, Dick CW and Patterson BD (2015) Bat fly evolution from the Eocene to the present (Hippoboscoidea, Streblidae and Nycteribiidae). In Morand S, Krasnov BR and Littlewood DTJ (eds). *Parasite Diversity and Diversification: Evolutionary Ecology Meets Phylogenetics*. Cambridge: Cambridge University Press, pp. 246–264.
- Fritz GN (1983) Biology and ecology of bat flies (Diptera: Streblidae) on bats in the genus *Carollia*. *Journal of Medical Entomology* **20**, 1–10.
- **Full RJ and Tu MS** (1991) Mechanics of a rapid running insect: two-, fourand six-legged locomotion. *Journal of Experimental Biology* **156**, 215–231.
- **Giraudoux P** (2016) pgirmess: Data analysis in ecology. R package version 1.6.5. Available at https://CRAN.R-project.org/package=pgirmess.
- Gotelli NJ and Rohde K (2002) Co-occurrence of ectoparasites of marine fishes: a null model analysis. *Ecology Letters* 5, 86–94.
- Graciolli G (2010) Nycteribiidae. In Brown B, Borkent A, Cumming J, Wood D, Woodley N and Zumbado M (eds). *Manual of Central American Diptera*. Costa Rica: Los Angeles County Museum of Natural History and Instituto Nacional de Biodiversidad, vol. 2, pp. 1261–1266.
- Graham D (1985) Pattern and control of walking in insects. Advances in Insect Physiology 18, 31–140.
- **Guerrero R** (1993) Catalogo de los Streblidae (Diptera: Pupipara) parasitos de murcielagos (Mammalia: Chiroptera) del Nuevo Mundo I. Clave para los géneros y Nycterophiliinae. *Acta Biologica Venezuelica* **14**, 61–75.
- **Guerrero R** (1994a) Catalogo de los Streblidae (Diptera: Pupipara) parasitos de murcielagos (Mammalia: Chiroptera) del Nuevo Mundo II. Los grupos: pallidus, caecus, major, uniformis y longipes del género *Trichobius* (Gervais, 1844). *Acta Biologica Venezuelica* **15**, 1–18.
- **Guerrero R** (1994b) Catalogo de los Streblidae (Diptera: Pupipara) parasitos de murcielagos (Mammalia: Chiroptera) del Nuevo Mundo IV. Trichobiinae con alas desarrolladas. *Boletín Entomologica Venezolana* **9**, 161–192.
- **Guerrero R** (1995a) Catalogo de los Streblidae (Diptera: Pupipara) parasitos de murcielagos (Mammalia: Chiroptera) del Nuevo Mundo III. Los grupos: dugesii, dunni y phyllostomae del género *Trichobius* (Gervais, 1844). *Acta Biologica Venezuelica* **15**, 1–27.
- **Guerrero R** (1995b) Catalogo de los Streblidae (Diptera: Pupipara) parasitos de murcielagos (Mammalia: Chiroptera) del Nuevo Mundo V. Trichobiinae con alas reducidas o ausentes y miscelaneos. *Boletin de Entomologia Venezolana* **10**, 135–160.
- **Guerrero R** (1996) Catálogo de los Streblidae (Diptera: Pupipara) parásitos de murciélagos (Mammalia: Chiroptera) del Nuevo Mundo VI. Streblinae. *Acta Biologica Venezuelica* **16**, 1–26.
- **Guerrero R** (1997) Catálogo de los Streblidae (Diptera: Pupipara) parásitos de murciélagos (Mammalia: Chiroptera) del Nuevo Mundo VII. Lista de especies, hospedadores y países. *Acta Biologica Venezuelica* **17**, 9–24.
- **Guerrero R** (1998) Notes on Neotropical batflies (Diptera, Streblidae). I. The genus *Trichobius*, with description of two new species and one new subspecies from Venezuela. *Acta Parasitologica* **43**, 86–93.
- Handley COW and Gardner AL (1991) Demography and natural history of the common fruit bat, *Artibeus jamaicensis*, on Barro Colorado Island, Panamá. *Smithsonian Contributions to Zoology* **155**, 1–173.
- Hsu M-H, Hsu T-C and Wu W-J (2002) Distribution of cat fleas (Siphonaptera: Pulicidae) on the cat. *Journal of Medical Entomology* **39**, 685–688.
- Iturralde-Vinent MA and MacPhee R (1996) Age and paleogeographical origin of Dominican amber. Science 273, 1850.
- Lapinski W, Walther P and Tschapka M (2015) Morphology reflects microhabitat preferences in an assemblage of neotropical wandering spiders. *Zoomorphology* 134, 219–236.
- Linhares AX and Komeno CA (2000) Trichobius joblingi, Aspidoptera falcata, and Megistopoda proxima (Diptera: Streblidae) parasitic on Carollia perspicillata and Sturnira lillium (Chiroptera: Phyllostomidae) in southeastern Brazil: sex ratios, seasonality, host site preference, and effect of parasitism on the host. Journal of Parasitology **86**, 167–170.
- Losos JB and Sinervo B (1989) The effects of morphology and perch diameter on sprint performance of *Anolis* lizards. *Journal of Experimental Biology* 145, 23–30.
- Marshall AG (1981) The Ecology of Ectoparasitic Insects. Academic Press Inc Ltd, London, UK, p. 446.

- Morand S, Krasnov BR and Poulin R (2006) Micromammals and Macroparasites: From Evolutionary Ecology to Management. Tokyo: Springer, p. 640.
- Mouillot D, George-Nascimento M and Poulin R (2003) How parasites divide resources: a test of the niche apportionment hypothesis. *Journal of Animal Ecology* 72, 757–764.
- Oksanen J, Blanchet F, Friendly M, Kindt R, Legendre P, McGlinn D, Minchin PR, O'Hara R, Simpson G, Solymos P, Stevens M, Szoecs E and Wagner H (2016). vegan: Community ecology package. R package version 2.4–1. Available at https://CRAN.R-project.org/package=vegan.
- **Overal WL** (1980) Host-relations of the bat fly *Megistopoda aranea* (Diptera: Streblidae) in Panama. *University of Kansas Science Bulletin* **52**, 1–20.
- Patterson BD, Dick CW and Dittmar K (2007) Roosting habits of bats affect their parasitism by bat flies (Diptera: Streblidae). *Journal of Tropical Ecology* 23, 177–189.
- Patterson BD, Dick CW and Dittmar K (2009) Nested distributions of bat flies (Diptera: Streblidae) on Neotropical bats: artifact and specificity in host-parasite studies. *Ecography* 32, 481–487.
- Poinar G and Brown A (2012) The first fossil streblid bat fly, Enischnomyia stegosoma n. gen., n. sp.(Diptera: Hippoboscoidea: Streblidae). Systematic Parasitology 81, 79–86.
- Poulin R and Morand S (2000) The diversity of parasites. Quarterly Review of Biology 75, 277–293.
- Presley SJ (2007) Streblid bat fly assemblage structure on Paraguayan Noctilio leporinus (Chiroptera: Noctilionidae): nestedness and species co-occurrence. Journal of Tropical Ecology 23, 409–417.
- Presley SJ (2010) Interspecific aggregation of ectoparasites on bats: importance of hosts as habitats supersedes interspecific interactions. Oikos 120, 832–841.
- R Development Core Team (2016) R: A Language and Environment for Statistical Computing. Vienna, Austria: R Foundation for Statistical Computing.

- Rasband W (1997–2016). *ImageJ*. Bethesda, Maryland, USA: US National Institutes of Health. Available at http://imagej.nih.gov/ij/.
- Rojas D, Warsi OM and Dávalos LM (2016) Bats (Chiroptera: Noctilionoidea) challenge a recent origin of extant neotropical diversity. *Systematic Biology* 65, 432–448.
- **Ross A** (1961) Biological studies on bat ectoparasites of the genus *Trichobius* (Diptera: Streblidae) in North America, north of Mexico. *Wasmann Journal of Biology* **19**, 229–246.
- Schlee D (1990) Das Bernstein-Kabinett. Stuttgarter Beitraege zur Naturkunde, Serie C 28, 1–100.
- Schoener TW (1974) Resource partitioning in ecological communities. Science 185, 27–39.
- Spielberg S (1997) The Lost World: Jurassic Park. USA: Universal Pictures.
- Tello SJ, Stevens RD and Dick CW (2008) Patterns of species co-occurrence and density compensation: a test for interspecific competition in bat ectoparasite infracommunities. *Oikos* 117, 693–702.
- ter Hofstede HM and Fenton MB (2005) Relationships between roost preferences, ectoparasite density, and grooming behaviour of neotropical bats. *Journal of Zoology* **266**, 333–340.
- ter Hofstede HM, Fenton MB and Whitaker Jr JO (2004). Host and host-site specificity of bat flies (Diptera: Streblidae and Nycteribiidae) on Neotropical bats (Chiroptera). *Canadian Journal of Zoology* **82**, 616–626.
- Wenzel RL (1976) The streblid batflies of Venezuela (Diptera: Streblidae). Brigham Young University Science Bulletin 20, 1–177.
- Wenzel RL, Tipton VJ and Kiewlicz A (1966). The streblid bat flies of Panama (Diptera: Calyptera: Streblidae). In Wenzel RL and Tipton VJ (eds). *Ectoparasites of Panama*, Vol. Fieldiana. Zoology. Special Publications. Chicago: Field Museum of Natural History, pp. 405–676.
- Windsor DA (1998) Controversies in parasitology, most of the species on earth are parasites. *International Journal for Parasitology* 28, 1939–1941.