

Molecular targets for the development of new acaricides against *Rhipicephalus microplus*: a review

Review

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Abstract

The cattle tick *Rhipicephalus microplus* is an ectoparasite with high economic importance to bovine culture, mainly in tropical and subtropical regions. The resistance of the tick from the commercial acaricides has hindered its control, thus motivating the search for new strategies. The purpose of this study was to perform a critical review about the main molecular targets of *R. microplus* that are useful for the discovery of new acaricides. Bibliographic search was conducted in the databases PubMed, ScienceDirect and CAB Direct, using the following descriptors: ‘*Rhipicephalus microplus*’, ‘*Boophilus microplus*’, ‘molecular targets’ and ‘action’, published between 2010 and 2021. Out of the 212 publications identified, 17 articles were selected for study inclusion. This review described 14 molecular targets and among these 4 are targets from commercial acaricides. Most of them are enzymes to catalyse important reactions to tick survival, related to energetic metabolism, mechanisms of biotransformation and neurotransmission. The data will be helpful in the development of new more effective and selective acaricides.

Introduction

The infestation by the ectoparasite *Rhipicephalus microplus* is one of the main parasitic problems associated with losses in cattle production, mainly in tropical and subtropical regions of the world. The infestation by this tick leads to weight loss, reduced milk production, decreased leather quality and higher production costs. Also, it is one of the vectors of potentially fatal tick-borne diseases, such as anaplasmosis and babesiosis (Lew-Tabor and Rodriguez Valle, 2016; Braz *et al.*, 2019).

The main method of tick control has been generally based on the continuous and intensive use of synthetic acaricides on infested cattle. Currently, there are six main classes of acaricides for tick control: organophosphates, formamidines (amitraz), pyrethroids, macrocyclic lactones, phenylpyrazoles (fipronil) and benzoylphenyl ureas (fluazuron). Most of these products act on the tick’s nervous system (neurotoxic action), interacting with ion channels, neurotransmitter receptors or enzymes, except for benzoylphenyl ureas, which act by inhibiting chitin synthesis (inhibitors of development) (Agwunobi *et al.*, 2021).

The systematic and indiscriminate use of these pesticides has favoured the selection of resistant tick strains to different chemical groups available on the market (Silva *et al.*, 2021). The high fecundity and the short life cycle in a single host of *R. microplus* associated with the high exposure to acaricides have contributed to the acceleration of the resistance process. A recent study reported five annual peaks of *R. microplus* infestations on animals and pastures that were correlated with climatic conditions, mainly an increase in the environmental temperature (Nicaretta *et al.*, 2021). *Rhipicephalus microplus* has been ranked 6th among the resistant arthropods and resistance to almost every chemical has been reported from different parts of the world (Ghosh *et al.*, 2017; Kumar *et al.*, 2019).

Considering that one of the main mechanisms of resistance is associated with genetic alterations that cause changes in the site of action, there is growing interest in the identification of new molecular targets for the research and development of antiparasitic drugs. It is worth mentioning that a synthetic acaricide with a new action mechanism, used in the control of the cattle tick, has not been released since the introduction of fipronil, in the mid-1990s, which was registered in the USA in 1996 (Agwunobi *et al.*, 2021). After this period, other new classes of ectoparasiticides were discovered, such as spinosad and isoxazolines, but they are currently not registered for use in cattle. Spinosad is a fermentation metabolite of the actinomycete, *Saccharopolyspora spinosa*, and its mechanism of action is the disruption in the binding of acetylcholine (ACh) in nicotinic ACh receptors at the postsynaptic cell. The isoxazolines (fluralaner, afoxolaner, lotilaner and sarolaner) and the bispyrazole (tigolaner) act by the inhibition of γ -aminobutyric acid (GABA) chloride channels and glutamate-gated chloride channels (GABA_ACl and GluCl_s, respectively) of insects and acarids (Rufener *et al.*, 2017; Selzer and Epe, 2021).

A promising strategy for the research of new active compounds is the application of *in silico* methods, such as screening by fragment, pharmacophore models and molecular docking (Leveridge *et al.*, 2018). These analyses allow one to evaluate the prediction of potential ligand–target interactions, requiring the characterization of molecular targets (protein, receptors) (Davis, 2020). Among the acaricide targets, macromolecules related to essential metabolic pathways or specific to parasites are promising to identify antiparasitic candidates with lower toxicity.

In this context, considering the resistance mechanisms developed by *R. microplus* and the need to discover new molecular targets for pest control, we have investigated previous studies on molecular targets of *R. microplus* with potential use for the research of new acaricide compounds.

Materials and methods

Literature searches were conducted to identify studies describing molecular targets of *R. microplus* in electronic databases: PubMed, Science Direct and CAB Direct. The following descriptors were used: ‘*Rhipicephalus microplus*’, ‘*Boophilus microplus*’, ‘molecular targets’ and ‘action’, using the Boolean operator ‘AND’. The inclusion criteria were: (1) peer-reviewed articles (experimental research and review manuscripts) without geographical limitation; (2) articles written in the English language and published between January 2010 and June 2021 and (3) studies related to the molecular target of the *R. microplus* species for acaricide product. Articles not found through databases and those that did not meet the selection criteria, comprising editorials, letters, book chapters, studies about molecular targets to vaccines and research in other tick species were excluded.

Literature was initially screened by the title and abstract, and citations judged as potentially eligible were obtained as full text. The full-text publications were evaluated considering inclusion and exclusion criteria previously described. The selection was performed by two independent evaluators, and, in case of disagreement, the study was discussed between reviewers so that a consensus could be reached. Figure 1 shows a flowchart of the methodology employed to perform this work and the number of articles evaluated.

Results and discussion

Firstly, 212 studies were identified, of which 17 met selection criteria (15 articles were searched in ScienceDirect, 1 article on PubMed and CAB Direct was found in both platforms). It can

see a greater number of publications in 2013 (4), while no publications about the theme were found in 2010, 2012 and 2016. The studies were performed in 6 different countries, and Brazil and USA had the larger number of publications (6 articles in each). Brazil plays an important role in global livestock production and tick infestation in cattle causes considerable economic losses in this country (Calvano *et al.*, 2021).

Molecular targets of *R. microplus*

This review identified 14 molecular targets of *R. microplus* associated with anti-tick activity. Among these, there are targets related to enzymes that participate in the energetic metabolism [glycogen synthase kinase 3 β (GSK-3 β), hexokinase, triosephosphate isomerase (TIM), NADH cytochrome c reductase, succinate cytochrome c reductase and glutamate dehydrogenase], in the detoxification mechanism [glutathione S-transferase (GST)], in neurotransmission [glutamate decarboxylase (GAD)] and in neuropeptide receptors [leucokinin-like peptide receptor (LKR) and CAP2b receptor (periviscerokinin)]. Four known targets of commercial acaricides, with neurotoxicity action, were found: acetylcholinesterase (AChE; organophosphates and carbamates), GABACls and GluClIs (macrocytic lactones and isoxazolines) and octopamine/tyramine receptor (amitraz) (Table 1).

The characteristics of each target identified in this work, as well as their potential use in the development of acaricides, are discussed below.

Glutathione S-transferase

GSTs (EC: 2.5.1.18) are present in a great variety of tissues in almost all of the eukaryotes. They are an enzyme family involved in the biotransformation of several xenobiotics, in the presence of glutathione (GSH) (Parizi *et al.*, 2011).

The increase in GST activity in resistant strains of tick species, including *R. microplus*, was reported in literature (Nandi *et al.*, 2015; Fular *et al.*, 2018). This increase is probably related to the GST participation in the metabolic detoxification of xenobiotics in arthropods because it catalyses the conjugation of glutathione (GSH) with molecules containing an electrophilic centre (thioether bond is formed between the sulphur of GSH and the substrate). The conjugated products are more water soluble, and consequently can be quickly eliminated (Bandara and Karunaratne, 2017; Le Gall *et al.*, 2018).

Besides the function of detoxification of xenobiotics, the GSTs also perform other important physiological roles, such as

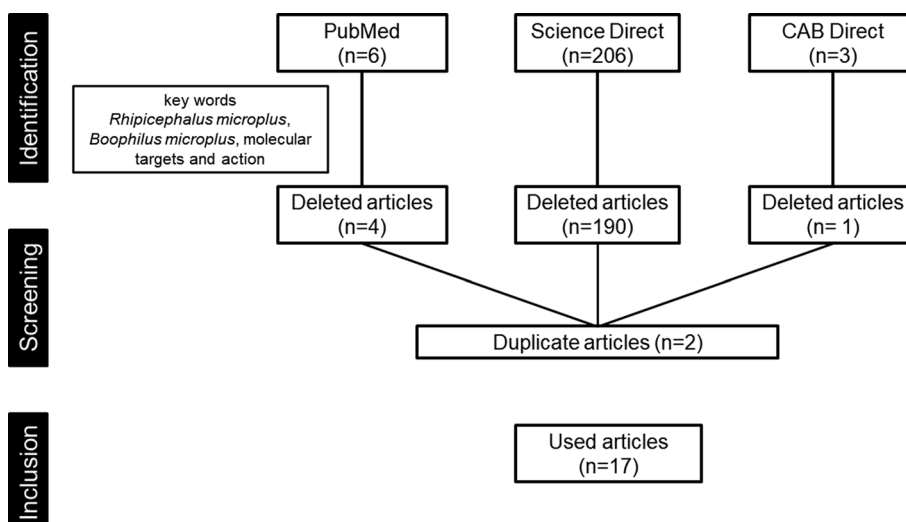


Fig. 1. Flowchart of the process of articles included to review the molecular targets of *Rhipicephalus microplus* for the development of new tick control compounds.

Table 1. Molecular targets and their physiological functions in *R. microplus*

Molecular targets	Localization	Physiological function	Sequence length (residues)	Molecular weight (kDa)	Reference
Acetylcholinesterase (AChE)	Nervous system	Acetylcholine (ACh) hydrolyses	595	65.7	Swale <i>et al.</i> (2013), Temeyer <i>et al.</i> (2013), Prado-Ochoa <i>et al.</i> (2014), Santos <i>et al.</i> (2018), Silva <i>et al.</i> (2021)
Glutathione S-transferase (GST)	Cytoplasm	Detoxification of xenobiotics	223	25.6	Jia <i>et al.</i> (2018), Sabadin <i>et al.</i> (2021)
Glycogen synthase kinase 3 β (GSK-3 β)	Cytoplasm, mitochondria and nucleus	Glucose metabolism (gluconeogenesis)	410	45.8	Waltero <i>et al.</i> (2020)
Hexokinase	Cytoplasm and mitochondria	Glucose metabolism (glycolysis)	492	55.2	Braz <i>et al.</i> (2019)
Triosephosphate isomerase	Cytoplasm	Glucose metabolism (glycolysis)	249	27.0	Moraes <i>et al.</i> (2011), Braz <i>et al.</i> (2019)
NADH cytochrome c reductase	Mitochondria	Cellular respiration (oxidative phosphorylation)	494	55.9	Braz <i>et al.</i> (2019)
Succinate cytochrome c reductase	Mitochondria	Cellular respiration (oxidative phosphorylation)	285	31.8	Braz <i>et al.</i> (2019)
Glutamate dehydrogenase	Mitochondria	Cellular respiration (oxidative phosphorylation)	555	61.9	Braz <i>et al.</i> (2019)
Glutamate decarboxylase	Nervous system	GABA synthesis	503	57.2	Ilg <i>et al.</i> (2013)
Leucokinin-like peptide receptor (LKR)	Nervous system	Diverse functions (myotropic and diuretic activity)	397	44.9	Brock <i>et al.</i> (2019)
Octopamine and tyramine receptors	Nervous system	Diverse functions (process of egg-laying, feeding rates)	80	8,5	Gross <i>et al.</i> (2015), Gross <i>et al.</i> (2017)
γ -Aminobutyric acid-gated chloride channels	Nervous system	Neurotransmitters receptors mediate the inhibitory synaptic transmission	537	59.3	Gassel <i>et al.</i> (2014), Rufener <i>et al.</i> (2017)
L-Glutamate-gated chloride channels	Nervous system	Neurotransmitters receptors mediate the inhibitory synaptic transmission	450	52.3	Gassel <i>et al.</i> (2014), Rufener <i>et al.</i> (2017)
CAP2b receptor (periviscerokinin)	Nervous system	Regulation of the processes of diuresis and/or antidiuresis	446	49.5	Yang <i>et al.</i> (2013)

sequestration and transport of endogenous hydrophobic compounds, such as hormones, steroids, haem, bilirubin and bile acids. This enzyme showed a strong positive correlation with lipid peroxidation, an indication that it plays a role in oxidant protection in *R. microplus* eggs (Freitas *et al.*, 2007).

Sabadin *et al.* (2021) analysed expression profiles of three genes that codify GST (SG2, GST4 and GST3) in engorged and partially engorged females of *R. microplus* and demonstrated that the GST3 gene was transcribed in all tested organs (salivary glands, ovaries, midguts and fat bodies), while GST4 genes were transcribed in the fat body. This ubiquitous transcription pattern suggests that the GST3 gene has substrates present in several organs, probably related to a common cellular function such as protection against oxidative stress, acting in S-glutathionylation.

Considering these physiological functions, GST represents a potential molecular target of acaricides since the inhibition of GST activity can cause tick death. The association of commercial acaricides with GST inhibitors may decrease the resistance of ticks to antiparasitic products. Le Gall *et al.* (2018) demonstrated that a combined treatment of ivermectin and diethyl maleate, a GST-specific inhibitor, increased the mortality by 30% in an ivermectin-resistant strain compared with the mortality of the

same strain treated only with ivermectin. The *in vitro* effect of terpenes isolated from *Arisaema anurans* on *R. microplus* was associated with multiple mechanisms of action, including the inhibition of GST activity (Jia *et al.*, 2018).

Enzymes related to energetic metabolism

The carbohydrate metabolism represents an essential role in the physiology and development for most living organisms, such as in arthropods' embryogenesis. A number of biosynthetic pathways are activated to support the process of cellular growth and differentiation, as well as embryonic growth. The high energy demanded by the developing embryo is supplied by the catabolism of biomolecules such as carbohydrates and lipids, which participate in other biosynthetic pathways (Moraes *et al.*, 2007; Silva *et al.*, 2019).

Variations in glucose and glycogen levels during *R. microplus* embryogenesis are related to glycolysis and gluconeogenesis (Moraes *et al.*, 2007). In the first stage from embryogenesis (from oviposition to the cellular blastoderm), an increase in the consumption of glycogen occurs, while the second stage (from the blastoderm to larval hatching) is characterized by intense gluconeogenesis (Braz *et al.*, 2019).

Hexokinase (EC 2.7.1.1), the first enzyme of glycolytic pathways, phosphorylates glucose to glucose-6-phosphate, the essential substrate to cellular biosynthesis (Moraes *et al.*, 2007). In *R. microplus*, this enzyme is found on cytosol or in the outer membrane of mitochondria. The inhibition of hexokinase can cause deleterious effects on the tick, such as the production of reactive oxygen species (ROS) and reduction of ATP levels (Fraga *et al.*, 2013; Braz *et al.*, 2019). Then, the enzymes involved in energetic metabolism during embryogenesis represent potential molecular targets for parasitic control, since they may interfere with embryonic development and consequently interrupt the life cycle of the tick (Logullo *et al.*, 2009).

Another important enzyme present in the glycolytic and gluconeogenic pathways is TIM (EC 5.3.1.1), responsible for the catalysis of the reversible interconversion of dihydroxyacetone phosphate and glyceraldehyde-D-phosphate. TIM is the only enzyme of *R. microplus* whose crystallographic structure was obtained from X-ray diffraction, and it was deposited in the Protein Data Bank (PDB ID: 3TH6) (Moraes *et al.*, 2011). This enzyme belongs to the ($\beta\alpha$)8-barrel class of proteins in which 8 parallel β -strands form the central core of the barrel, surrounded by 8 α -helices (Yang *et al.*, 2017; He *et al.*, 2018).

TIM from various parasites has been extensively studied, and their three-dimensional (3D) structure and catalytic properties from different species are very similar. This enzyme plays an essential role in the production of ATP in glycolysis and represents a promising target for drug design against pathogens, which depends on glycolytic metabolism (Kumar *et al.*, 2012; Romero-Romero *et al.*, 2018). Despite a great structural similarity among the species, one can use non-conserved amino acid residues to get selective inhibitors to homologue enzymes (Tungtur *et al.*, 2010).

Investigations about the effects of dimerization on the activity and the role of loop motions in the catalysis of TIM indicate that dimerization is necessary for their stability and proper conformation of the active site loops. Thermodynamic studies indicate that the energy required to dissociate the homodimer in TIM is greater than the energy required to unfold the monomers. Furthermore, several studies proposed that a cysteine (Cys) residue located at the interface stands out as a promising target because molecules that interact with exposed cysteines, such as methyl methane thiosulphonate (MMTS), inhibit TIMs from various species (Kumar *et al.*, 2012; Lopez-Zavala *et al.*, 2016; Chang *et al.*, 2018).

Moraes *et al.* (2011) investigated TIM from *R. microplus* embryos (BmTIM) and observed that BmTIM is a homodimer with amino acid sequence, secondary structure and kinetic parameters like other TIMs already described. However, BmTIM is richer in Cys moieties, which are significantly accessible to interact with molecule inhibitors. The authors also reported that BmTIM shows high sensitivity to the action of the thiol reagents dithionitrobenzoic acid and mMMTS. TIM inhibition can promote an increase in the formation of methylglyoxal (intermediary metabolite of the reaction catalysed by TIM), a harmful molecule for ticks (Braz *et al.*, 2019).

Another important enzyme in arthropod metabolism is GSK-3 (EC: 2.7.11.26), which acts mainly in the glycogen metabolism regulation, through the inhibition of glycogen synthase enzyme during embryonic development. This enzyme is present in the cytoplasm, mitochondria, nucleus and other subcellular compartments. GSK-3 is considered as a multifunctional enzyme since it is involved in various cellular processes such as glucose metabolism, apoptosis, cell proliferation, transcription, cell migration and immune response (Saraswati *et al.*, 2018). These multiple actions can be explained by its capacity to interact with some different substrates. Studies reported that about 100 proteins are

phosphorylated by GSK-3 (Sutherland, 2011; Waltero *et al.*, 2020). In the tick *R. microplus*, only an isoform from the enzyme was identified, GSK-3 β , and the ovaries represent the main transcription site. GSK-3 β inhibition may lead to a reduction of oviposition and egg viability (Waltero *et al.*, 2020).

Enzymes related to mitochondrial respiratory chain have also been studied for the identification of new potential molecular targets for tick control. The respiratory chain consists of four multi-enzymatic protein complex and electron carriers, such as Q co-enzyme and cytochrome c. Among these enzymes, the complex NADH cytochrome c reductase and succinate cytochrome c reductase stands out, which participates in important stages of ATP production. Mitochondrial respiratory chain changes are related to the increase in intracellular levels of ROS, which can compromise the development of ticks at different stages (Braz *et al.*, 2019).

Braz *et al.* (2019) investigated the *in vitro* activity of 3-bromopyruvate (3-BrPA) against different molecular targets from *R. microplus* and demonstrated that 3-BrPA inhibited the activity from hexokinase, TIM, NADH cytochrome c reductase, succinate cytochrome c reductase and glutamate dehydrogenase. These effects against different metabolic pathways were related to the death of embryo-derived cells and the reduction from cellular respiration in developing embryos.

Octopamine and tyramine receptors

Octopamine (OA) and tyramine (TA), biogenic amines found in several invertebrates, are characterized as neurotransmitters, neurohormones and neuromodulators. In arthropods, the tyraminer-gic/octopaminergic system bears a relation with the adrenergic system in vertebrates (Cossío-Bayúgar *et al.*, 2012).

These neurotransmitters act in different physiological functions and behaviours from invertebrates, such as egg-laying, aggressive behaviours, feeding rates and defensive jumping. Firstly, TA was considered only as a precursor for the synthesis of OA. However, literature suggests that it is an independent neurotransmitter because it has a different localization within the insect nervous system, different receptors and transporters; in some cases, it can behave as an OA antagonist (Cossío-Bayúgar *et al.*, 2015).

The octopaminergic/tyraminer-gic receptors have been classified in different groups, namely α -adrenergic-like (α OCT), β -adrenergic-like (β OCT) and octopamine/tyramine (OCT/Tyr) or tyraminer-gic type 1 (TA/AO or TAR1), tyraminer-gic type 2 (TAR2) and 3 (TAR3), although TAR3 is only identified in *Drosophila melanogaster* (Wu *et al.*, 2014; Baron *et al.*, 2015). The activation of these receptors induces transient alterations in the intracellular concentration of cyclic-AMP and calcium. The OA receptors have been extensively studied as acaricide targets, and an example of OA agonist is amitraz, which causes neuronal hyperexcitability and tick death (Ohta and Ozoe, 2014; Baron *et al.*, 2015).

The effect of OA, TA and 83 adrenergic compounds was screened for their ability to block oviposition in *R. microplus* by Cossío-Bayúgar *et al.* (2012). They identified inhibition of oviposition in this tick at pharmacological concentrations of different molecules: α -agonists (10), α -antagonists (3), β -adrenergic agonists (5), β -antagonists (7) and norepinephrine. The most potent molecules are TA and other α -adrenergic, while the OA was not active, suggesting that oviposition is primarily controlled by the tyraminer-gic pathway. In ticks, oviposition occurs on a massive and nearly explosive basis, probably requiring extensive smooth muscle contraction, which is mediated by TA (Cossío-Bayúgar *et al.*, 2012).

The TA receptor type 1 from *R. microplus* was expressed to heterologous form in ovary cells from Chinese hamster and it

showed 39 times more power to TA compared to OA. Furthermore, the expressed receptor was strongly antagonized by yohimbine and cyproheptadine, and mildly antagonized by mianserin and phentolamine. Agonistic or modulatory activity against the expressed receptor was observed by tolazoline, naphazoline and BTS-27271 (amitraz metabolite). However, BTS-27271 activity was only observed in the presence of TA (Gross *et al.*, 2015). The activity of terpenoids against the TA receptor from *R. microplus* was related by Gross *et al.* (2017), who observed agonist action from pulegone and a modulator activity by carvacrol, isoeugenol and 1,4-cineole. These authors indicate that the TA receptor can represent a possible target for the action of new compounds for tick control.

Neuropeptide receptors: leucokinin-like peptide receptor and periviscerokinin receptor

Ticks, particularly haematophagous ectoparasite, consume large amounts of blood in a single feeding and need to remove excessive water after a blood meal. Like haematophagous insects, this complex regulation is mediated by different neuropeptides (Neupert *et al.*, 2005).

Leucokinin-like neuropeptides, also known as leucokinin or kinins, were found in invertebrates and they are associated with different physiological effects, such as myotropic and diuretic activities. The receptors of this family of peptide (LKR, leucokinin receptor and kinins receptor) are coupled to the G protein (GPRS) and are present in all life stages of a tick in both sexes (Holmes *et al.*, 2000; Pietrantonio *et al.*, 2018; Brock *et al.*, 2019).

Leucokinin-like neuropeptides play important functions in insects, including digestive enzyme release, regulation of water balance and pre-ecdysis control (Kersch and Pietrantonio, 2011; Pietrantonio *et al.*, 2018). However, knowledge about physiological functions from this family of neuropeptides in ticks is still limited (Brock *et al.*, 2019). In an analogy with insects, tissues expressing leucokinin receptors in ticks may be involved in water balance or neuromodulation such as the Malpighian tubules and hindgut, or the central nervous system (synganglion) (Holmes *et al.*, 2000).

Brock *et al.* (2019) identified, by immunolocalization, the presence of LKR in the midgut of *R. microplus* female. These authors carried out the process of silencing the LKR genes and observed alterations in reproductive parameters of females, such as decreases in weights of egg masses and in the percentages of eggs hatched per egg mass, as well as delays in time of oviposition and egg hatching. Considering that LKR is a selective G protein-coupled receptor (GPCR) only found in invertebrates, it can be considered a promising target for *R. microplus* control.

Another important neuropeptide found in arthropods is the cardioacceleratory peptide 2b (CAP2b)/periviscerokinin (PVK), also associated with the regulation of the processes of diuresis and/or antidiuresis in a variety of insects (Nachman *et al.*, 2013). Yang *et al.* (2013) cloned and functionally characterized the first Rhimi-CAP2b-R receptor from *R. microplus*. The analyses of the predicted protein sequence, phylogenetics and functional expression indicate that this receptor is the orthologue of the insect CAP2b/PVK. The expression of Rhimi-CAP2b-R was detected in synganglion, salivary gland, Malpighian tubule and in the ovary of female ticks.

γ -Aminobutyric acid- and L-glutamate-gated chloride channels

GABA_ACl and GluCl₁ are members of the Cys-loop receptor channel family characterized by a large N-terminal extracellular domain, four transmembrane domains (M1–M4) and a long, variable intracellular loop connecting the M3 and M4 segments

as the hallmarks of every subunit (Ozoe, 2013). Both channels are widely distributed in the nervous system and neuromuscular junctions from invertebrates, and act as neurotransmitter receptors to mediate inhibitory synaptic transmission.

The GABA_ACl and GluCl₁ channels are molecular targets from acaricides used in the control of ectoparasites, as phenylpyrazoles, macrocyclic lactones and isoxazolines. The compounds of phenylpyrazole classes (fipronil) act, especially, to block the GABA_ACl promoting parasite death by hyperexcitability. Macrocyclic lactones (ivermectins and milbemycins) activate GABA_ACl and GluCl₁ in invertebrates (Ozoe, 2013; Rufener *et al.*, 2017; Bae and Kwon, 2020). These actions result in a block of nervous stimulus transmission, causing immobilization and paralysis of the parasite. Compounds of the isoxazoline class (fluralaner, lotilaner) act as non-competitive antagonists from GABA_ACl channels, and to a lesser extent, they also show inhibitory action against GluCl₁ (Gassel *et al.*, 2014; Rufener *et al.*, 2017).

Glutamate decarboxylase

Besides GABA_ACl channels, other components from the GABAergic system can represent molecular targets, but that is still too little scientifically explored (Ilg *et al.*, 2013). This system is composed of four primary parts: GABA synthesis by GAD action, GABA catabolism by GABA-transaminase, GABA transporters and GABA receptors (Dionisio *et al.*, 2011; Ilg *et al.*, 2013; Tang *et al.*, 2020).

GAD (EC 4.1.1.15) is an enzyme that catalyses the irreversible α -decarboxylation of L-glutamate to GABA. GAD is a key enzyme in the dynamic regulation of neural network excitability because both its substrate and product are neurotransmitters and exhibit opposite actions (L-glutamate – excitatory effect and GABA – inhibitory action). GAD is considered as an initiator on the GABAergic system and is frequently used as a molecular marker from GABAergic neurons (Li *et al.*, 2016; Tang *et al.*, 2020).

The GAD activity was reported in various arthropods, including insects (cockroach, grasshopper, moth, bee and fly) and ectoparasites (*Ctenocephalides felis* and *R. microplus*) and the functional properties of arthropod GADs are analogous to mammals (Ueno, 2000).

Considering that the block of chloride channels controlled by GABA is lethal to ticks, it is expected that the negative regulation of the production of GABA, through GAD inhibition, has the potential to produce the same phenotype. Ilg *et al.* (2013) evaluated the effect of 109 591 compounds on the GAD of *C. felis* and *R. microplus* using high-throughput inhibitor screening methods. Barbituric acid and phenothiazine derivative show a more potent effect on arthropod GADs ($IC_{50} = 14.9 \mu M$ and $28.4 \mu M$, respectively) in comparison with mouse GAD forms ($IC_{50} = 59.3 \mu M$ and $>100 \mu M$, respectively). These two compounds appear to be promising molecules for the design of parasitocidal products.

Acetylcholinesterase

AChE, target carbamate and organophosphate acaricide, is a central enzyme from the eukaryotic nervous system and it acts on Ach hydrolyses. When AChE is inactivated, there is the accumulation of Ach and the excitation neural time is consequently extended, causing paralysis and the death of ticks (Temeyer *et al.*, 2013).

AChE transcription is different among the species: a single gene is responsible for the transcription in vertebrates (Massoulié *et al.*, 1998), while in invertebrates, it is associated with different genes (Fournier *et al.*, 1992; Ilg *et al.*, 2010). In *R. microplus*, three genes were identified that codify AChEs: RmAChE1 (Baxter and Barker, 1998), RmAChE2 (Hernandez

et al., 1999; Baxter and Barker, 2002) and RmAChE3 (Temeyer *et al.*, 2004).

RmAChE1, RmAChE2 and RmAChE3 were biochemically characterized from the baculovirus expression by Temeyer *et al.* (2010), who attested that the three forms had properties to degrade Ach. RmAChE 1 shows a higher affinity to Ach and is considered as the most important isoform for tick survival. The post-transcriptional gene silencing by RNA interference was used by Temeyer *et al.* (2013) to investigate the functional role from three RmAChEs, and they suggest that there is a functional complementation by RmAChEs genes *in vivo*, but their specific functional role is still not elucidated.

The different classes of synthetic and natural compounds have been studied against *R. microplus* cholinesterase. Swale *et al.* (2013) characterized the inhibitor profiles of AChE from *R. microplus* (RmAChE1) compared to human and bovine AChEs, and showed that RmAChE had low sensitivity to tacrine, and a serial of dimer bis(*n*)-tacrine compounds. The crystallographic structure of *R. microplus* cholinesterases is not still available, thus 3D models were developed by molecular homology to predict the *in silico* potential of AChE inhibitors. Cerqueira *et al.* (2021) built a 3D model of AChE1 and conducted docking studies with flavonoids. Among the evaluated flavonoids, quercetin presented high affinity to RmAChE1 and exhibited *in vitro* larvicidal effect.

Prado-Ochoa *et al.* (2014) evaluated the *in vitro* effects produced by the new synthetic carbamates [ethyl-(4-bromophenyl) carbamate and ethyl-(4-chlorophenyl) carbamate] on two *R. microplus* strains. The anti-tick effects of these compounds were independent of AChE inhibition and the action of carbamates probably occurred due to morphological alterations in the reproductive organs (vitellogenesis and viability of the ovarian cells). The *in vitro* larvicidal effect of alkaloids (berberine and piperine) on *R. microplus* was correlated with AChE inhibition by Silva *et al.* (2021). These authors characterized the molecular inhibition profile through *in silico* studies (docking and molecular dynamic) and reported that berberine interacts with RmAChE1 by π -stacking, cation- π and hydrophobic bonds with peripheral anionic site residues.

Among the targets described in this review, only four are associated with mechanisms of action of acaricides available on the market. However, the *R. microplus* presented several enzymes and receptors that are of pharmaceutical interest and could be used in the search for new acaricides.

Concluding remarks

The characterization of molecular targets from a tick is essential for the search for novel, more effective and more selective anti-tick compounds, with less risk to human and animal health. In this review, we were able to describe 14 molecular targets of *R. microplus*: four known targets of commercial products with action in the nervous system (AChE, GABA- and L-glutamate-gated chloride channels and OA and TA receptors) and 10 new molecular targets (enzymes and neuropeptide receptors) that can cause changes to the homeostasis, reproductive parameters and neurotoxic effects. The enzymes are related to energetic metabolism (hexokinase, triosephosphate isomerase, GSK-3 β), respiratory process (NADH cytochrome c reductase, succinate cytochrome c reductase and glutamate dehydrogenase), neurotransmitter activation (GAD) and detoxification mechanism (GST). The neuropeptide receptors [LKR and CAP2b (periviscerokinin) receptor] are important for hydric equilibrium and neuromodulation. These targets can be useful to identify compounds able to act in resistant strains through new action mechanisms or different molecular interactions in known targets.

This review gives an overview of molecular targets of *R. microplus* that are essential to parasite life, over the past 10 years. Knowledge about the molecular target enables the search for molecules with greater selectivity, improving their action and reducing toxicity. The new approaches and targets described in this study provide the basis for the research of novel antiparasitic agents, especially in target-based drug design, which represents a valuable route for drug design.

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References

- Agwunobi DO, Yu Z and Liu J (2021) A retrospective review on ixodid tick resistance against synthetic acaricides: implications and perspectives for future resistance prevention and mitigation. *Pesticide Biochemistry and Physiology* **173**, 104776.
- Bae J-W and Kwon W-S (2020) Investigating the effects of fipronil on male fertility: insight into the mechanism of capacitation. *Reproductive Toxicology* **94**, 1–7.
- Bandara KMJ and Karunaratne SHPP (2017) Mechanisms of acaricide resistance in the cattle tick *Rhipicephalus (Boophilus) microplus* in Sri Lanka. *Pesticide Biochemistry and Physiology* **139**, 68–72. doi: 10.1016/j.pestbp.2017.05.002
- Baron S, Van Der Merwe NA, Madder M and Maritz-Olivier C (2015) SNP analysis infers that recombination is involved in the evolution of amitraz resistance in *Rhipicephalus microplus*. *PLoS ONE* **10**. doi: 10.1371/journal.pone.0131341.
- Baxter GD and Barker SC (1998) Acetylcholinesterase cDNA of the cattle tick, *Boophilus microplus*: characterisation and role in organophosphate resistance. *Insect Biochemistry and Molecular Biology* **28**, 581–589.
- Baxter GD and Barker SC (2002) Analysis of the sequence and expression of a second putative acetylcholinesterase cDNA from organophosphate susceptible and organophosphate-resistant cattle ticks. *Insect Biochemistry and Molecular Biology* **32**, 815–820.
- Braz V, Gomes H, Galina A, Saramago L, Braz G, Vaz I Jr., Logullo C, Fonseca RN, Campos E and Moraes J (2019) Inhibition of energy metabolism by 3-bromopyruvate in the hard tick *Rhipicephalus microplus*. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology* **218**, 55–61.
- Brock CM, Temeyer KB, Tidwell J, Yang, Y, Blandon MA, Carreón-Camacho D, Longnecker MT, Almazán C, Pérez De León AA and Pietrantonio PV (2019) The leucokinin-like peptide receptor from the cattle fever tick, *Rhipicephalus microplus*, is localized in the midgut periphery and receptor silencing with validated double-stranded RNAs causes a reproductive fitness cost. *International Journal for Parasitology* **49**, 287–299.
- Calvano MPCA, Brumatti RC, Barros JC, Garcia MV, Martins KR and Andreotti R (2021) Bioeconomic simulation of *Rhipicephalus microplus* infestation in different beef cattle production systems in the Brazilian Cerrado. *Agricultural Systems* **194**, 103247.
- Cerqueira APM, Santana IB, Araújo, JSC, Lima HG, Batatinha MJM, Branco A, Santos Junior MCD and Botura MB (2021) Homology modeling, docking, molecular dynamics and *in vitro* studies to identify *Rhipicephalus microplus* acetylcholinesterase inhibitors. *Journal of Biomolecular Structure & Dynamics* **1**, 1–11. doi: 10.1080/07391102.2021.1889666

- Chang TC, Park JH, Colquhoun AN, Khoury CB, Seangmany NA and Schwans JP (2018) Evaluating the catalytic importance of a conserved Glu97 residue in triosephosphate isomerase. *Biochemical and Biophysical Research Communications* **505**, 492–497.
- Cossio-Bayúgar R, Miranda-Miranda E, Narváez Padilla V, Olvera-Valencia F and Reynaud E (2012) Perturbation of tyraminerpic/octopaminergic function inhibits oviposition in the cattle tick *Rhipicephalus (Boophilus) microplus*. *Journal of Insect Physiology* **58**, 628–633.
- Cossio-Bayúgar R, Miranda-Miranda E, Fernández-Rubalcaba M, Padilla V and Reynaud E (2015) Adrenergic ligands that block oviposition in the cattle tick *Rhipicephalus microplus* affect ovary contraction. *Scientific Reports* **5**, 15109.
- Davis RL (2020) Mechanism of action and target identification: a matter of timing in drug discovery. *iScience* **23**. doi: 10.1016/j.isci.2020.101487.
- Dionisio L, Rosa MJR, Bouzat C and Esandi MDC (2011) An intrinsic GABAergic system in human lymphocytes. *Neuropharmacology* **60**, 513–519.
- Fournier D, Mutero A and Rungger D (1992) Drosophila acetylcholinesterase – expression of a functional precursor in *Xenopus* oocytes. *European Journal of Biochemistry* **203**, 513–519.
- Fraga A, Ribeiro L, Lobato M, Santos V and Silva JR (2013) Glycogen and glucose metabolism are essential for early embryonic development of the red flour beetle *Tribolium castaneum*. *PLoS ONE* **8**. doi: 10.1371/journal.pone.0065125.
- Freitas DRJ, Rosa RM, Moraes J, Campos E, Logullo C, Vaz IS Jr. and Masuda A (2007) Relationship between glutathione S-transferase, catalase, oxygen consumption, lipid peroxidation and oxidative stress in eggs and larvae of *Boophilus microplus* (Acarina: Ixodidae). *Comparative Biochemistry and Physiology – Part A Molecular & Integrative Physiology* **146**, 688–694.
- Fular A, Sharma AK, Kumar S, Nagar G, Chigure G, Ray D and Ghosh S (2018) Establishment of a multi-acaricide resistant reference tick strain (IVRI-V) of *Rhipicephalus microplus*. *Ticks and Tick-borne Diseases* **9**, 1184–1191. doi: 10.1016/j.ttbdis.2018.04.014
- Gassel M, Wolf C, Noack S, Williams H and Ilg T (2014) The novel isoxazoline ectoparasiticide fluralaner: selective inhibition of arthropod γ -aminobutyric acid- and L-glutamate-gated chloride channels and insecticidal/acaricidal activity. *Insect Biochemistry and Molecular Biology* **45**, 111–124.
- Ghosh S, Gupta S, Ajith Kumar KG, Sharma AK, Kumar S, Nagar G, Kumar R, Paul S, Fular A, Chigure G, Nandi A, Manjunathachar HV, Mohammad A, Verma MR, Saravanan BC and Ray D (2017) Characterization and establishment of a reference deltamethrin and cypermethrin resistant tick line (IVRI-IV) of *Rhipicephalus (Boophilus) microplus*. *Pesticide Biochemistry and Physiology* **138**, 66–70.
- Gross A, Temeyer K, Day T, Pérez De León A, Kimber M and Coats J (2015) Pharmacological characterization of a tyramine receptor from the southern cattle tick, *Rhipicephalus (Boophilus) microplus*. *Insect Biochemistry and Molecular Biology* **63**, 47–53.
- Gross AD, Temeyer KB, Day TA, Pérez De León AA, Kimber MJ and Coats JR (2017) Interaction of plant essential oil terpenoids with the southern cattle tick tyramine receptor: a potential biopesticide target. *Chemico-Biological Interactions* **263**, 1–6.
- He R, Zhang H, Shen N, Guo C, Ren Y, Xie Y, Gu X, Lai W, Peng X and Yang G (2018) Molecular characterization and allergenicity potential of triosephosphate isomerase from *Sarcoptes scabiei*. *Veterinary Parasitology* **257**, 40–47.
- Hernandez RH, He H, Chen AC, Ivie GW, George JE and Wagner GG (1999) Cloning and sequencing of a putative acetylcholinesterase cDNA from *Boophilus microplus* (Acarina: Ixodidae). *Journal of Medical Entomology* **36**, 764–770.
- Holmes SP, He H, Chen AC, Ivie GW and Pietrantonio PV (2000) Cloning and transcriptional expression of a leucokinin-like peptide receptor from the Southern cattle tick, *Boophilus microplus* (Acarina: Ixodidae). *Insect Molecular Biology* **9**, 457–465.
- Ilg T, Schmalz S, Werr M and Cramer J (2010) Acetylcholinesterases of the cat flea *Ctenocephalides felis*: identification of two distinct genes and biochemical characterization of recombinant and in vivo enzyme activities. *Insect Biochemistry and Molecular Biology* **2**, 153–164.
- Ilg T, Berger M, Noack S, Rohwer A and Gabel M (2013) Glutamate decarboxylase of the parasitic arthropods *Ctenocephalides felis* and *Rhipicephalus microplus*: gene identification, cloning, expression, assay development, identification of inhibitors by high throughput screening and comparison with the orthologs from *Drosophila melanogaster* and mouse. *Insect Biochemistry and Molecular Biology* **43**, 162–177.
- Jia M, He Q, Wang W, Dai J and Zhu L (2018) Chemical composition and acaricidal activity of *Arisaema anurans* essential oil and its major constituents against *Rhipicephalus microplus* (Acarina: Ixodidae). *Veterinary Parasitology* **261**, 59–66.
- Kersch CN and Pietrantonio PV (2011) Mosquito *Aedes aegypti* (L.) leucokinin receptor is critical for *in vivo* fluid excretion post blood feeding. *FEBS Letters* **585**, 3507–3512.
- Kumar K, Bhargava P and Roy U (2012) Cloning, overexpression and characterization of *Leishmania donovani* triosephosphate isomerase. *Experimental Parasitology* **130**, 430–436.
- Kumar R, Kumar Sharma A and Ghosh S (2019) Menace of acaricide resistance in cattle tick, *Rhipicephalus microplus* in India: status and possible mitigation strategies. *Veterinary Parasitology* **278**, 108993.
- Le Gall VL, Klafke GM and Torres TT (2018) Detoxification mechanisms involved in ivermectin resistance in the cattle tick, *Rhipicephalus (Boophilus) microplus*. *Scientific Reports* **8**. doi:10.1038/s41598-018-30907-7.
- Leveridge M, Chung CW, Gross JW, Phelps CB and Green D (2018) Integration of lead discovery tactics and the evolution of the lead discovery toolbox. *SLAS Discovery: Advances* **23**, 881–897.
- Lew-Tabor AE and Rodriguez Valle M (2016) A review of reverse vaccinology approaches for the development of vaccines against ticks and tick borne diseases. *Ticks and Tick-borne Diseases* **7**, 573–585.
- Li M, Wang L, Qiu L, Wang W, Xin L, Xu J, Wang H and Song L (2016) A glutamic acid decarboxylase (GAD) highly expressed in hemocytes of Pacific oyster *Crassostrea gigas*. *Developmental and Comparative Immunology* **63**, 56–65.
- Logullo C, Witola WH, Andrade C, Abreu L, Gomes J, Vaz IS, Imamura S, Konnai S, Ohashi K and Onuma M (2009) Expression and activity of glycogen synthase kinase during vitellogenesis and embryogenesis of *Rhipicephalus (Boophilus) microplus*. *Veterinary Parasitology* **161**, 261–269.
- Lopez-Zavala AA, Carrasco-Miranda JS, Ramirez-Aguirre CD, López-Hidalgo M, Benitez-Cardoza CG, Ochoa-Leyva A, Cardona-Felix CS, Diaz-Quezada C, Rudiño-Piñera E, Sotelo-Mundo RR and Bribea LG (2016) Structural insights from a novel invertebrate triosephosphate isomerase from *Litopenaeus vannamei*. *Biochimica et Biophysica Acta (BBA) – Proteins and Proteomics* **1864**, 696–1706.
- Massoulié J, Anselmet A, Bon S, Krejci E, Legay C, Morel N and Simon S (1998) Acetylcholinesterase: C-terminal domains, molecular forms and functional localization. *Journal of Physiology* **92**, 183–190.
- Moraes J, Galina A, Alvarenga PH, Rezende GL, Masuda A, Vaz IS Jr. and Logullo C (2007) Glucose metabolism during embryogenesis of the hard tick *Boophilus microplus*. *Comparative Biochemistry and Physiology. B, Biochemistry & Molecular Biology* **146**, 528–533.
- Moraes J, Arreola R, Cabrera N, Saramago L, Freitas D, Masuda A, Vaz IS Jr., Gomez-Puyou MT, Perez-Montfort R, Gomez-Puyou A and Logullo C (2011) Structural and biochemical characterization of a recombinant triosephosphate isomerase from *Rhipicephalus (Boophilus) microplus*. *Insect Biochemistry and Molecular Biology* **41**, 400–409.
- Nachman RJ, Wang XJ, Etkorn FA, Kaczmarek K, Zabrocki J, Lopez J and Coast GM (2013) Evaluation of insect CAP2B analogs with either an (E)-alkene, trans- or a (Z)-alkene, cis-Pro isostere identifies the Pro orientation for antidiuretic activity in the stink bug. *Peptides* **41**, 101–106.
- Nandi A, Singh H and Singh NK (2015) Esterase and glutathione S-transferase levels associated with synthetic pyrethroid resistance in *Hyalomma anatolicum* and *Rhipicephalus microplus* ticks from Punjab, India. *Experimental & Applied Acarology* **66**, 141–157.
- Neupert S, Predel R, Russell WK, Davies R, Pietrantonio PV and Nachman RJ (2005) Identification of tick periviscerokinin, the first neurohormone of Ixodidae: single cell analysis by means of MALDI-TOF/TOF mass spectrometry. *Biochemical and Biophysical Research Communications* **338**, 1860–1864.
- Nicaretta JE, Zapa DMB, Couto LFM, Heller LM, Cavalcante ASA, Cruvinel LB, Melo Júnior RD, Ferreira LL, Nascimento RMD, Soares VE, Borges LMF, Monteiro CMO and Lopes WDZ (2021) *Rhipicephalus microplus* seasonal dynamic in a Cerrado biome, Brazil: an update data considering the global warming. *Veterinary Parasitology* **296**, 109506.
- Ohta H and Ozoe Y (2014) Molecular signalling, pharmacology, and physiology of octopamine and tyramine receptors as potential insect pest control targets. *Advances in Insect Physiology* **46**, 73–166.
- Ozoe Y (2013) γ -Aminobutyrate- and glutamate-gated chloride channels as targets of insecticides. *Advances in Insect Physiology* **44**, 211–286.

- Parizi LF, Utiumi KU, Imamura S, Onuma M, Ohashi K, Masuda A and Vaz IS Jr (2011) Cross immunity with *Haemaphysalis longicornis* glutathione S-transferase reduces an experimental *Rhipicephalus* (*Boophilus*) *microplus* infestation. *Experimental Parasitology* **127**, 113–118.
- Pietrantonio PV, Xiong C, Nachman RJ and Shen Y (2018) G protein-coupled receptors in arthropod vectors: omics and pharmacological approaches to elucidate ligand-receptor interactions and novel organismal functions. *Current Opinion in Insect Science* **29**, 12–20.
- Prado-Ochoa MG, Ramírez-Noguera P, Díaz-Torres R, Garrido-Fariña GI, Vázquez-Valadez VH, Velázquez-Sánchez AM, Muñoz-Guzmán MA, Angeles E and Alba-Hurtado F (2014) The action of two ethyl carbamates on acetylcholinesterase and reproductive organs of *Rhipicephalus microplus*. *Veterinary Parasitology* **199**, 215–224.
- Romero-Romero S, Becerril-Sesín LA, Costas M, Rodríguez-Romero A and Fernández-Velasco DA (2018) Structure and conformational stability of the triosephosphate isomerase from *Zea mays*. Comparison with the chemical unfolding pathways of other eukaryotic TIMs. *Archives of Biochemistry and Biophysics* **658**, 66–76.
- Rufener L, Danelli V, Bertrand D and Sager H (2017) The novel isoxazoline ectoparasiticide lotilaner (Credelio™): a non-competitive antagonist specific to invertebrates γ -aminobutyric acid-gated chloride channels (GABA_ACl_s). *Parasites & Vectors* **10**. doi: 10.1186/s13071-017-2470-4
- Sabadin GA, Salomon TB, Leite MS, Benfato MS, Oliveira PL and Silva Vaz I Jr. (2021) An insight into the functional role of antioxidant and detoxification enzymes in adult *Rhipicephalus microplus* female ticks. *Parasitology International* **81**, 102274.
- Santos FO, Lima HG, de Souza Santa Rosa S, das Mercês NB, Serra TM, Uzeda RS, Reis IMA, Botura MB, Branco A and Batatinha MJM (2018) In vitro acaricide and anticholinesterase activities of digitaria insularis (Poaceae) against *Rhipicephalus* (*Boophilus*) *microplus*. *Veterinary Parasitology* **255**, 102–106.
- Saraswati AP, Ali Hussaini SM, Krishna NH, Babu BN and Kamal A (2018) Glycogen synthase kinase-3 and its inhibitors: potential target for various therapeutic conditions. *European Journal of Medicinal Chemistry* **144**, 843–858.
- Selzer PM and Epe C (2021). Antiparasitics in animal health: quo vadis? *Trends in Parasitology* **37**, 77–89.
- Silva RM, Vital WO, Fonseca RN, Martins YPM, Lemos FJA, Vaz IS Jr. and Logullo C (2019) Hypometabolic strategy and glucose metabolism maintenance of *Aedes aegypti* egg desiccation. *Comparative Biochemistry and Physiology* **227**, 56–63.
- Silva GD, Lima HG, Freitas HF, Rocha Pita SS, Santos Luz Y, Figueiredo MP, Uzeda RS, Branco A, Costa SL, Batatinha MJM and Botura MB (2021) In vitro and in silico studies of the larvicidal and anticholinesterase activities of berberine and piperine alkaloids on *Rhipicephalus microplus*. *Ticks and Tick Borne Diseases* **12**. doi: 10.1016/j.ttbdis.2020.101643
- Sutherland C (2011) What are the bona fide GSK3 substrates? *International Journal of Alzheimer's Disease* 505607. doi: 10.4061/2011/505607.
- Swale DR, Tong F, Temeyer KB, Li A, Lam PC, Totrov MM, Carlier PR, Pérez De León AA and Bloomquist JR (2013) Inhibitor profile of bis (*n*)-tacirines and *N*-methylcarbamates on acetylcholinesterase from *Rhipicephalus* (*Boophilus*) *microplus* and *Phlebotomus papatasi*. *Pesticide Biochemistry and Physiology* **106**. <https://doi.org/10.1016/j.pestbp.2013.03.005>.
- Tang CD, Li X, Shi HL, Jia YY, Dong ZX, Jiao ZJ, Wang LF, Xu JH, Yao LG and Kan YC (2020) Efficient expression of novel glutamate decarboxylases and high level production of γ -aminobutyric acid catalyzed by engineered *Escherichia coli*. *International Journal of Biological Macromolecules* **160**, 372–379.
- Temeyer KB, Darvey RB and Chen AC (2004) Identification of a third *Boophilus microplus* (Acari: Ixodidae) cDNA presumptively encoding an acetylcholinesterase. *Journal of Medicine and Entomology* **41**, 259–268.
- Temeyer KB, Pruett J and Olafson P (2010) Baculovirus expression, biochemical characterization and organophosphate sensitivity of rBmAChE1, rBmAChE2, and rBmAChE3 of *Rhipicephalus* (*Boophilus*) *microplus*. *Veterinary Parasitology* **172**, 114–121.
- Temeyer KB, Olafson PU, Brake DK, Tuckow AP, Li AY and Perez de Leon AA (2013) Acetylcholinesterase of *Rhipicephalus* (*Boophilus*) *microplus* and *Phlebotomus papatasi*: gene identification, expression, and biochemical properties of recombinant proteins. *Pesticide Biochemistry and Physiology* **106**, 118–123.
- Tungtur S, Meinhardt S and Swint-Kruse L (2010) Comparing the functional roles of nonconserved sequence positions in homologous transcription repressors: implications for sequence/function analyses. *Journal of Molecular Biology* **395**, 785–802.
- Ueno H (2000) Enzymatic and structural aspects on glutamate decarboxylase. *Journal of Molecular Catalysis B: Enzymatic* **10**, 67–79.
- Waltero C, Martins R, Calixto C, Fonseca RN, Abreu LA, Silva Vaz I Jr. and Logullo C (2020) The hallmarks of GSK-3 in morphogenesis and embryonic development metabolism in arthropods. *Insect Biochemistry and Molecular Biology* **118**, 103307.
- Wu SF, Xu G, Qi YX, Xia RY, Huang J and Ye GY (2014) Two splicing variants of a novel family of octopamine receptors with different signalling properties. *Journal of Neurochemistry* **129**, 37–47.
- Yang Y, Bajracharya P, Castillo P, Nachman RJ and Pietrantonio PV (2013) Molecular and functional characterization of the first tick CAP2b (periviscerokinin) receptor from *Rhipicephalus* (*Boophilus*) *microplus* (Acari: Ixodidae). *General and Comparative Endocrinology* **194**, 142–151.
- Yang Y, Chen Z-W, Hurlburt B, Li G-L, Zhang Y-X, Fei D-X, Shen H-W, Cao M-J and Liu G-M (2017) Identification of triosephosphate isomerase as a novel allergen in *Octopus fangsiao*. *Molecular Immunology* **85**, 35–46.