

# Saproxylic community, guild and species responses to varying pheromone components of a pine bark beetle

Iñaki Etxebeste<sup>1\*</sup>, José L. Lencina<sup>2</sup> and Juan Pajares<sup>1</sup>

<sup>1</sup>Sustainable Forest Management Research Institute, University of Valladolid -CIFOR-INIA, Avd. Valladolid 44, 34004 Palencia, Spain: <sup>2</sup>Department of Zoology and Physical Anthropology, University of Murcia, Apdo. 4021. 30071 Murcia, Spain

## Abstract

Some bark beetle species (Coleoptera: Scolytinae) produce aggregation pheromones that allow coordinated attack on their conifer hosts. As a new saproxylic habitat is founded, an assemblage of associated beetles kairomonally respond to bark beetle infochemicals. *Ips sexdentatus* is one of the major damaging insects of *Pinus* spp. in Southern Europe. Its response to varying ipsenol (Ie) percentages in relation to ipsdienol (Id) was studied in northwestern Spain, along with the entire saproxylic beetle assemblage captured at multiple-funnel traps. Response profile modeling was undertaken for *I. sexdentatus* sexes and sex-ratios, associated species and for selected trophic groups using a reference Gaussian model. In addition, the effects on the saproxylic assemblages were analyzed. *I. sexdentatus* response curve peaked at 22.7% Ie content, while remaining taxa that could be modeled, peaked above ca. 40% Ie. Predator guilds showed a linear relationship with Ie proportion, while competitors showed a delayed response peak. Consequently, species assemblages differed markedly between varying pheromone component mixtures. Given that the evaluated pheromonal proportions mimicked that of logs being colonized by *I. sexdentatus*, results suggested that the registered differential responses at different levels might provide *I. sexdentatus* with a temporal window that maximizes conspecific attraction while reducing interference with competitor and predatory guilds. Described responses might help improve the monitoring of the population status of target bark beetles and their associates, but also point toward the by-catch of many natural enemies, as well as rare saproxylic beetle species, interfering with the aims of sustainable forest management.

**Keywords:** *Ips sexdentatus*, scolytinae, *Pinus*, saproxylics, integrated pest management, infochemicals

(Accepted 6 December 2012; First published online 1 March 2013)

## Introduction

Although most of the bark beetle species (Coleoptera: Curculionidae: Scolytinae) found in temperate and boreal forests are innocuous, the activity of aggressive bark beetles results in the destruction of millions of cubic meters of conifer trees per year in production forests around the world (Lieutier *et al.*, 2004; Blomquist *et al.*, 2010; Bussler *et al.*, 2011). Given the

---

\*Author for correspondence: Phone: +34 679 00 22 88  
Fax: +34 979 10 84 19  
E-mail: inaki@goisolutions.net

right conditions, such as increased mature host stands, favorable climate (Carroll *et al.*, 2004), or increased host availability through thinning operations, forest fires (Santolamazza-Carbone *et al.*, 2011; Etxebeste *et al.*, 2012) natural windthrows (Wermelinger, 2004), populations boost to epidemic levels and attack is shifted toward healthy trees. Alternatively, those human-competitor bark beetles constitute keystone species in natural ecosystems as they initiate break-down of trees contributing significantly to the foundation of saproxylic habitats (Grove, 2002; Foit, 2010; Santolamazza-Carbone *et al.*, 2011).

Aggregation pheromones are used by some bark beetles to coordinate mass attack on host conifers (Wood, 1982; Seybold *et al.*, 2006). Many of these pheromones are composed of monoterpenoids that are synthesized *de novo* once colonizing beetles initiate boring through host bark (Blomquist *et al.*, 2010). In most monogamous bark beetle species, the female begins releasing long-range aggregation pheromones (e.g., *Dendroctonus* spp.), whereas it is the male that does so in polygamous species (e.g., *Ips* spp.; Wood, 1982). Once nuptial chambers are carved out under the bark, eggs are laid along maternal galleries. Larvae mine the phloem after hatching, and pupate in oval chambers. New adults then chew through the bark and disperse in search of new hosts. Aggregation pheromones seem to benefit bark beetles in a number of ways, e.g. overcoming tree defenses by increasing the number of attacking beetles (Raffa & Berryman, 1983), interspecific resource partitioning (Poland & Borden, 1994) or diluting predation (Aukema & Raffa, 2004a), but as fellow colonists are attracted, competition increases, especially among non-aggressive bark beetles (Latty, *et al.*, 2009). Besides communication with conspecifics, location of foraging grounds by a diverse guild of eavesdroppers occurs through kairomonal attraction: habitat-specialist predators, host-specific parasitoids and competing subcortical herbivores follow bark beetle infochemicals, endangering their reproductive success (Wood, 1982; Poland & Borden, 1994; Ross & Daterman, 1995; Raffa, 2001). In fact, accumulated evidence suggests that competitors, predators and parasites strongly influence the population and behavioral ecology of bark beetles (Schroeder & Weslien, 1994a, b; Weslien, 1994; Herard & Mercadier, 1996; Reeve, 1997; Boone *et al.*, 2008). Even if other sensorial cues might be involved (e.g. visual cues; Strom *et al.*, 1999), subtle nuances of the pheromonal blend composition may allow bark beetles to avoid eavesdropping. For example, different populations of a bark beetle may display geographical or seasonal variations in chiral production and response, as is the case of *Ips pini* (Vité *et al.*, 1978; Teale & Lanier, 1991), but more remarkably, this species has been found to escape predators by having divergent chiral preferences (Raffa & Klepzig, 1989). Exploitation of these differences through selective pest removal, enemy augmentation strategies and improved monitoring has been proposed (Aukema *et al.*, 2000a, b; Aukema & Raffa, 2005). Minor components of bark beetle aggregation pheromones have also been shown to modify the response of associated species (Seybold *et al.*, 1992; Allison *et al.*, 2001; Pajares *et al.*, 2004; Etxebeste *et al.*, 2012), but responses of associate species assemblages to common chemical signals, as well as the interactions among those insects, have rarely been studied (Aukema & Raffa, 2004a). Although the importance of the variation of pheromone component ratios has long been appreciated (Roelofs, 1978; Teale *et al.*, 1994), only a few studies have tried to model coleopteran responses to these gradients. Besides, although

many of the infochemicals involved in the communication of several bark beetles have been described, information on the natural proportions among components of the infochemical blends is missing or could be strongly biased by the methodology, which could result in misleading conclusions on behavioral responses to synthetic infochemicals (Pureswaran & Sullivan, 2012). A Gaussian curve model has been proposed as a method for measuring the peak width of the response window, and hence describing the stability and variation of the pheromone signal (Schlyter *et al.*, 2001). This methodology could provide information for the refinement of pheromone formulations, facilitating useful information for enhanced control programs, while allowing the comparison and description of the response profiles of associated species and trophic guilds.

The six-toothed pine bark beetle (*Ips sexdentatus* Boern.), is a widely distributed species through the Eurasian continent, where it commonly behaves as a secondary pest (Gil & Pajares, 1986). Nevertheless, outbreaks may occur if suitable conditions are given, as happened after Klaus, an extratropical cyclone that struck south-western France in 2009. A large amount of felled trees prompted an increase in *I. sexdentatus* population resulting in about additional 3.9 million cubic meters of *Pinus pinaster* Aiton lost by the end of 2010 caused by the activity of this bark beetle (EFI, 2010). Pioneering works within *Ips* genus established ipsdienol (Id; 2-methyl-6-methylene-2, 7-octadien-4-ol) as the main pheromonal component regulating *I. sexdentatus* aggregation (Vité *et al.*, 1972, 1974), thereafter its attractiveness has been confirmed in several field experiments (Vité *et al.*, 1974; Klimetzek & Vité, 1986; Etxebeste *et al.*, 2012). Even if racemic ipsenol (Ie; 2-methyl-6-methylene-7-octen-4-ol) has been detected in hindgut and frass extracts of this species (Vité *et al.*, 1972; Francke *et al.*, 1986; Kohnle, 1991), its synergic effect on the aggregation power of Id has not been shown until recently (Etxebeste *et al.*, 2012). Absolute figures for *I. sexdentatus* sex-ratios responding to aggregation pheromones have been provided (e.g. Klimetzek & Vité, 1986), but detailed studies are missing. Even if the response to certain infochemicals of some predators and other species associated with *I. sexdentatus* has been described (Pajares *et al.*, 2004; Ibeas *et al.*, 2007; Etxebeste & Pajares, 2011; Etxebeste *et al.*, 2012), information of additional associated saproxylic beetles is also missing.

Thus, an experiment evaluating multiple funnel traps catches of *I. sexdentatus* and associated beetles was conducted in order to study the response to experimental lures with varying components of the *I. sexdentatus* pheromone complex at community, guild and species levels. In other words, the objectives of this research were to (i) characterize the intra-specific response of *I. sexdentatus*, describing response maxima and variations in the sex ratio; (ii) characterize the inter-specific response of saproxylic beetle species; (iii) determine changes in the guild structure; and (iv) study the species assemblage change through the evaluated compound gradient.

## Materials and methods

### Study area and experimental design

The experiment took place between 6 August and 20 September 2008, and was carried out at a site in northwestern Spain, approximately enclosed within the square defined by the 29T 7390 4729 coordinates of the Universal Transverse

Mercator system, ranging in elevation from 1050 to 1130 m. a.s. l. The area was mainly composed of reforested stands of about 30-year-old *Pinus nigra salzmannii* J. F. Arnold, although a few ca. 50-year-old *P. pinaster* stands could also be found among patches of *Quercus pyrenaica* Willd. A large fire burned across the area 2 years before, providing large amount of breeding material for *I. sexdentatus*, and hence its population level was still high at the onset of the experiment, reflected on a few bark beetle infestation foci present in the area. The mean day temperature through the experimental period averaged 17.5°C, while minimum and maximum temperatures averaged 10.5 and 24.5°C, respectively (Leon Airport, Castile and Leon, Spain).

A total of seven experimental blocks were located along firebreaks and dirt roads that held uniform conditions across the experimental sites. Within each block, seven 12-unit multiple funnel traps (Former Phero Tech Inc., now Contech Enterprises Inc., British Columbia, Canada; Lindgren, 1983), suspended 2m above ground from metal poles and spaced >75m apart, comprised the sampling units of the study. In order to test for the effects of the temporal variation in pheromone composition, lures containing increasing percentages of racemic Ie in relation to the total blend of Ie and racemic Id were designed together with chemists at SEDQ LLC (Barcelona, Spain). Selected percentage levels were defined after the natural evolution of the emission of these compounds from logs colonized by *I. sexdentatus* (Kohnle, 1991; Etxebeste *et al.*, unpublished data), and previous results that defined a pheromonal lure of *I. sexdentatus* (Etxebeste *et al.*, 2012). In order to avoid a confounding effect on the response of *I. sexdentatus*, the main pheromonal compound (Id; Vité *et al.*, 1974) was kept at a constant dosage through tested blend percentages (95 mg per lure sachet, resulting in ca. 1.1 mg day<sup>-1</sup> release rate). The first 0% level carried no Ie, while increasing Ie amounts released from separate devices helped obtain the selected remaining 1, 5, 10, 50, 90 and 95% Ie levels. For the 1, 5 and 10% Ie levels, closed 250 µl polyethylene (PE) vials were loaded with 1, 5 and 10.5 mg of Ie. Remaining levels were prepared loading Ie into aluminum sachets with PE windows varying in size: for the 50% level 95 mg of Ie were loaded in a sachet with the same window size as in the design used for Id; for the 90 and 95% Ie levels 440 mg of Ie were loaded into two and four sachets with larger PE windows, respectively. The performance of the release devices had been tested during previous works carried in the same experimental area (Etxebeste *et al.*, 2012.). All infochemical purities were reported to be above 95% (SEDQ LLC.). The resulting seven treatment levels were then randomly assigned to each sampling unit. To provide a blank control, traps were not baited until the second week of the experimental period. To reduce positional effects, re-randomizations of the assigned sampling units to treatments were implemented every week, and both, traps and lures, were moved to the resulting new positions within experimental blocks. Sample collection was conducted on a weekly basis, and these were preserved in 70% ethanol until identification and counting.

Taxonomic classification was undertaken by specialists, and beetles were identified to species level according to the nomenclature of the Fauna Europaea Web Service (2010). In addition, all *I. sexdentatus* individuals were sexed under the stereo microscope by checking the elytral spine structure (Gil & Pajares, 1986). Saproxyl and non-saproxyl beetles were distinguished using different resources (e.g., Dajoz, 2000; Kenis & Hilszczanski, 2004; Nieto & Alexander, 2010). Each

species was assigned to a trophic group (guild) according to those proposed by Bouget *et al.* (2005), and thus falling into one of four main guilds: saproxylphages, xylofungivores, xylophages, and predators. Xylophages in turn, were further classified as intraguild competitors or intraguild predators, based on the potential ability of certain xylophage larvae to act as facultative predators of other phloem inhabiting species (e.g., Dodds *et al.*, 2001). As many species might switch between guilds depending on their life stage, categorization was made regarding the closest linkage to *I. sexdentatus* (e.g., if the adult of a certain Buprestidae species is known to feed on pollen, but larvae grow on woody cambium, the species was assigned to the xylophage guild). However, because both larval and adult ecology for many captured species are largely unknown, the description of anatomical structures of collected taxa and the ecology of closely related species were also used in guild classification. Voucher specimens have been deposited at the Entomology Collection of the Department of Plant Production and Forest Resources of the University of Valladolid.

### Statistical analysis

Data from the multiple funnel traps were pooled for the experimental period and insect count sums per treatment and experimental block were used as the response variable in a series analysis aimed at studying the effect of varying Ie percentages in the pheromonal blend at the species, trophic guild and species assemblage level. All calculations and analyses were carried out under the R statistical environment and language (The R Development Core Team, 2011).

### Species response

Statistical analyses at the species level were only conducted on those taxa with a minimum of 20 specimens caught over the experimental period. Differences in mean number of accumulated individuals between Ie percentage levels were tested with multiple comparison of means (Tukey HSD), applying the Bonferroni correction to the value of  $\alpha$  for the confidence intervals (Reeve & Strom, 2004) in a generalized linear model (GLM) using the log-link function to account for the Poisson error structure. In the case of *I. sexdentatus* sex-ratio analysis, data were fitted in a GLM using logit-link function to account for a binomial error structure (Crawley, 2007). Species (and *I. sexdentatus* sex-ratio) data were additionally fitted to the Gaussian model described by Schlyter *et al.* (2001) through non-linear regression. Such a model allows for the location of peaks ( $\mu_r$ ) and widths ( $2\sigma_r$ ) of the response window of analyzed species. In order to allow fitted models to be comparable across different species, raw data were scaled dividing the response variable by the maximum empirical value within each experimental block. Modeling of *I. sexdentatus* sex-ratio was performed with the raw male-to-female catch ratio. As the explanatory variable, proportion ( $p$ ) of Ie in the lure, did not meet with the assumption of normality, it was transformed taking the arcsin of the square root of  $p$ . Modeling was performed following the non-linear least squares (NLS) method using the Gauss-Newton algorithm. Prior to the analysis, dot-plots aided at setting the starting parameters for the non-linear regression and discarding species that did not show a Gaussian response, which in turn were analyzed through linear regression with the aim of describing dose-response relationships.

Table 1. Effect of increasing Ie proportion in lures on mean accumulated catches ± SEM (n = 7) of *I. sexdentatus* (total, female, male and sex ratio) and other known saproxylic beetles. Shown species had accumulated catches of 20 or more individuals. See Appendix Table A1 for full scientific names and classifications. F and P(>F) values of the treatment effects at the analysis of variance (ANOVA) of fitted GLM are presented for each species. Asterisks after P(>F) values highlight: \*, <0.05; \*\*, <0.01 and \*\*\*, <0.001 significances. Shared letters within the same species indicate that means are not significantly different (Tukey's HSD test, Bonferroni's adjustment, P < 0.05).

| Species                     | F <sub>6,36</sub> | P(>F)     | Total | Ipsenol percentage in pheromonal blend |                    |                  |                   |                    |                  |                  |
|-----------------------------|-------------------|-----------|-------|--|--------------------|------------------|-------------------|--------------------|------------------|------------------|
|                             |                   |           |       | 0                                      | 1                  | 5                | 10                | 50                 | 90               | 95               |
| <i>I. sexdentatus</i>       | 11.15             | <0.001*** | 6706  | 115 ± 18.18 ab                         | 138.86 ± 31.52 ab  | 236.57 ± 45.22 c | 189.71 ± 48.18 ac | 155.14 ± 26.37 ac  | 58.86 ± 8.7 b    | 63.86 ± 21.3 b   |
| Females                     | 9.33              | <0.001*** | 4298  | 74.14 ± 10.55 abc                      | 87.86 ± 19.34 abcd | 141.29 ± 27.66 d | 120.86 ± 29.73 ad | 101.43 ± 17.55 abd | 41.71 ± 5.19 c   | 46.71 ± 14.84 bc |
| Males                       | 13.09             | <0.001*** | 2397  | 40.86 ± 8.17 ab                        | 51 ± 12.46 ab      | 95.14 ± 18.08 c  | 69.57 ± 18.38 ac  | 52.14 ± 8.62 ab    | 17.14 ± 3.85 b   | 16.57 ± 6.89 b   |
| Sex ratio                   | 5.8               | <0.001*** | 0.56  | 0.55 ± 0.07 ab                         | 0.59 ± 0.07 ab     | 0.68 ± 0.06 a    | 0.56 ± 0.03 ab    | 0.53 ± 0.03 abc    | 0.39 ± 0.05 bc   | 0.31 ± 0.08 c    |
| <i>A. ferus</i>             | 0.99              | 0.448     | 139   | 1.14 ± 0.99                            | 1 ± 0.72           | 2.29 ± 1.64      | 7 ± 6.67          | 5.57 ± 5.41        | 0.57 ± 0.43      | 2.29 ± 1.96      |
| <i>A. griseus</i>           | 1.68              | 0.154     | 36    | 0.29 ± 0.18                            | 0                  | 0.29 ± 0.18      | 0                 | 1 ± 0.38           | 1.29 ± 0.84      | 2.29 ± 1.52      |
| <i>B. novemmaculata</i>     | 1.34              | 0.266     | 57    | 0.57 ± 0.43                            | 1.14 ± 0.67        | 1.14 ± 0.67      | 1 ± 0.58          | 1 ± 0.53           | 1.43 ± 0.43      | 2.29 ± 0.64      |
| <i>H. ligniperda</i>        | 4.66              | 0.001***  | 481   | 3 ± 1.23 a                             | 2.86 ± 1.64 a      | 10.29 ± 5.51 ab  | 6.14 ± 3.05 a     | 7.86 ± 3.84 a      | 25 ± 10.1 b      | 13.57 ± 3.56 ab  |
| <i>H. pini</i>              | 2.35              | 0.051     | 50    | 0                                      | 0.14 ± 0.14        | 0.29 ± 0.18      | 0.29 ± 0.18       | 0                  | 1.57 ± 0.48      | 4.86 ± 3.09      |
| <i>C. erosus</i>            | 1.65              | 0.162     | 758   | 12 ± 3.03                              | 14 ± 3.02          | 19.43 ± 5.42     | 18.43 ± 4.83      | 17.86 ± 4          | 11.29 ± 2.07     | 15.29 ± 4.29     |
| <i>P. parallelolippidus</i> | 0.95              | 0.469     | 68    | 2.29 ± 1.13                            | 0.57 ± 0.2         | 2.29 ± 1.81      | 2.29 ± 1.21       | 0.29 ± 0.18        | 0.71 ± 0.29      | 1.29 ± 0.75      |
| <i>Q. abietum</i>           | 0.66              | 0.684     | 22    | 0.29 ± 0.18                            | 0.29 ± 0.18        | 0.29 ± 0.29      | 0.71 ± 0.18       | 0.71 ± 0.29        | 0.43 ± 0.43      | 0.43 ± 0.3       |
| <i>Q.uedius</i> sp.         | 2.73              | 0.027*    | 62    | 1.86 ± 0.34 a                          | 0.71 ± 0.29 a      | 0.71 ± 0.36 a    | 0.71 ± 0.36 a     | 0.57 ± 0.3 a       | 2.86 ± 1.2 a     | 1.43 ± 0.53 a    |
| <i>R. ferrugineus</i>       | 8.07              | <0.001*** | 1321  | 7.71 ± 5.29 a                          | 5.71 ± 1.66 a      | 18.43 ± 4.85 ab  | 23.71 ± 12.26 ab  | 23 ± 7.19 ab       | 47.57 ± 11.37 bc | 62.57 ± 19.75 C  |
| <i>S. royi</i>              | 0.89              | 0.511     | 22    | 0                                      | 0.14 ± 0.14        | 1 ± 1            | 0                 | 0                  | 0.43 ± 0.2       | 1.57 ± 1.41      |
| <i>T. caerulea</i>          | 1.37              | 0.254     | 267   | 2 ± 1.84                               | 1.29 ± 0.68        | 4.86 ± 4.53      | 4.29 ± 3.8        | 5.29 ± 4.2         | 4 ± 1.59         | 16.43 ± 13.44    |
| <i>T. dermestoides</i>      | 1.25              | 0.303     | 20    | 0.86 ± 0.46                            | 0.29 ± 0.18        | 0.43 ± 0.2       | 0.29 ± 0.18       | 0.43 ± 0.43        | 0.14 ± 0.14      | 0.43 ± 0.3       |
| <i>T. formicarius</i>       | 8.15              | <0.001*** | 53    | 0.14 ± 0.14 ab                         | 0 a                | 0.43 ± 0.2 ab    | 0.43 ± 0.43 ab    | 1.86 ± 0.63 bc     | 2.29 ± 0.47 c    | 2.43 ± 0.69 C    |

Guild structure

Accumulated catches per defined guild were analyzed following the same methodology described for species. Equivalently, their response toward increasing percentages of Ie, was modeled according to the described procedure for intraguild competitors, intraguild predators and predator guilds. In order to allow comparison with the described response for *I. sexdentatus*, it has to be noted that the intraguild competitors group did not include this species. Detritivore, saproxylophage, and xylofungivore guild response modeling have not been included because of low accumulated catch data, and lack of apparent differentiated response to the tested Ie gradient. In addition, the effect of varying Ie percentage in lures on diversity within guilds was estimated using the Shannon index, evaluated through GLM and Tukey HSD, as in previous analyses.

Species assemblages

Singletons and doubletons (species captured only once or twice) were removed from the set of tallied saproxylic beetle species prior to the analysis. Treatment effect on assemblage composition was analyzed using Adonis. This type of analysis, analogous to MANOVA multivariate analysis of variance, allows for a multivariate permutational analysis of the assemblage variation attributed to experimental treatments (Oksanen et al., 2008). In order to create the required dissimilarity matrix, Arrhenius z beta-diversity was calculated for all pairwise comparisons of the treatment level, which was further analyzed using Adonis set to 999 permutations. Beta-diversity was chosen as the measure for dissimilarity, as it accounts for the differentiation in composition among habitats. An ordination plot using no-metric multidimensional scaling (NMDS) of the Bray-Curtis dissimilarity index of Wisconsin transformed data was also produced (Oksanen et al., 2008). To this ordination, ellipses representing factor class standard error areas at 95% confidence intervals were added. Ipsenol percentage in the lure was also fitted to distance matrix using the envfit function of the vegan package (Oksanen et al., 2008).

Results

Species response

Traps caught virtually no beetles during the initial week of the experiment, when no lures were attached to traps. During the remaining experimental period, 101 beetle species (10,533 specimens) were captured, 10,232 of which (97%) were classified as saproxylics, pooled into 65 distinct taxa (table A.1). Although for 24 differentiated taxa the species could not be established, at least one new elaterid species was identified among trapped beetles (*Athous (Orthatous)* n. sp.; Sáez Bolaño J. A., personal communication). As could be expected from using its major pheromonal compounds, *I. sexdentatus* comprised 63% of all captured beetles, whereas those species with capture levels above 20 individuals made up 98% of the total of trapped specimens.

Significant treatment effects could be detected for *I. sexdentatus* catches (table 1). Total male and female catches were found to be highest when 5% Ie was present in the lure, although these figures were not significantly different from close blends (table 1). Even if differences among treatments were similar for male and female *I. sexdentatus*, sex-ratio varied significantly, achieving its lowest value at the 95% Ie level,



Table 2. Parameter estimates with their SE and goodness of fit of non-linear regression of Gaussian curve to response of *I. sexdentatus* (total, male and females) and other known saproxyl beetles. Cumulative response of intraguild competitors is also shown. In all cases, three and four regression and residual degrees of freedom, respectively.

| Taxon                  | Goodness of fit | Location ( $\mu_r$ ) | Width/2 ( $1\sigma_r$ ) | Form factor ( $f$ ) | Location ( $\mu_r$ ) <sup>1</sup> | Width ( $1\sigma_r$ ) <sup>1</sup> |
|------------------------|-----------------|----------------------|-------------------------|---------------------|-----------------------------------|------------------------------------|
| <i>I. sexdentatus</i>  | 0.861           | 0.479 ± 0.061        | 0.486 ± 0.076           | 0.402 ± 0.043       | 0.227                             | 0.100                              |
| Males                  | 0.837           | 0.450 ± 0.068        | 0.426 ± 0.084           | 0.344 ± 0.049       | 0.201                             | 0.106                              |
| Females                | 0.882           | 0.496 ± 0.054        | 0.515 ± 0.068           | 0.428 ± 0.039       | 0.244                             | 0.090                              |
| Sex-ratio              | 0.901           | 0.334 ± 0.135        | 0.902 ± 0.173           | 0.548 ± 0.090       | 0.111                             | 0.165                              |
| <i>A. griseus</i>      | 0.878           | 1.062 ± 0.091        | -0.418 ± 0.102          | -0.220 ± 0.037      | 0.903                             | 0.154                              |
| <i>O. erosus</i>       | 0.766           | 0.641 ± 0.049        | 0.727 ± 0.102           | 0.553 ± 0.051       | 0.399                             | 0.094                              |
| <i>T. formicarius</i>  | 0.992           | 1.170 ± 0.044        | 0.446 ± 0.040           | 0.373 ± 0.026       | 0.980                             | 0.063                              |
| Intraguild competitors | 695             | 1.026 ± 0.269        | 0.949 ± 0.376           | 0.711 ± 0.238       | 0.869                             | 0.454                              |

<sup>1</sup> Values retransformed from the arcsin of the square root of  $p$ .

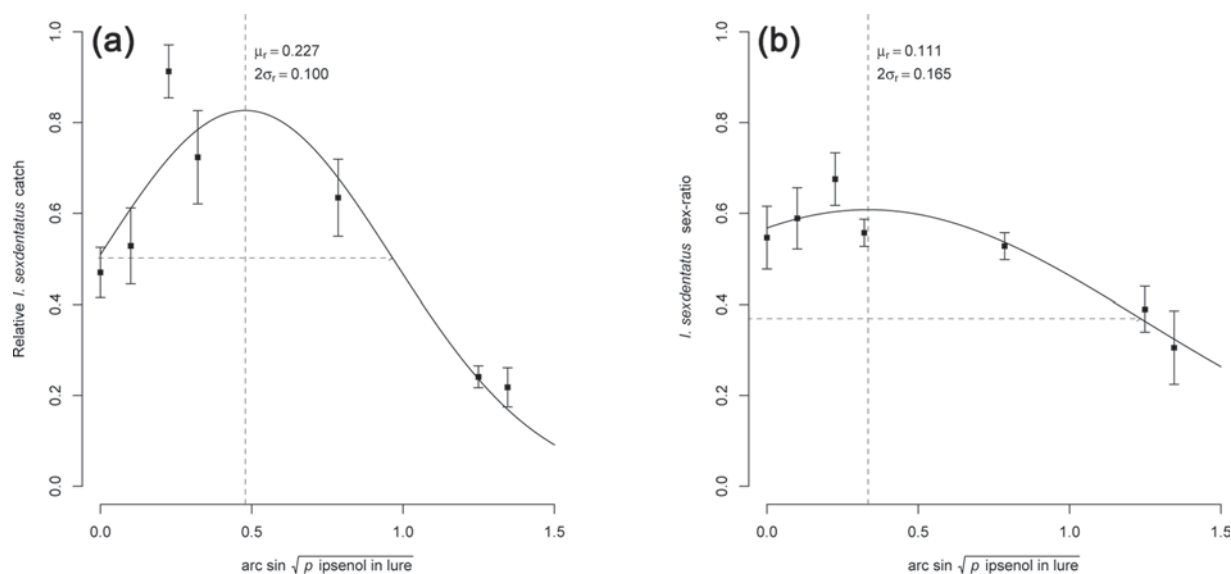


Fig. 1. Accumulated trap-catch data and the sex-ratio of *I. sexdentatus* in response to increasing proportions of Ie in the pheromonal blend. Back-transformed location of the response peaks ( $\mu_r$ ) and width ( $2\sigma_r$ ) of the response window are shown in the plot. (a) Mean relative *I. sexdentatus* catch  $\pm$  SEM ( $n=7$ ) and fitted Gaussian response curve. (b) Registered and predicted sex-ratio for *I. sexdentatus* after fitting the Gaussian curve. Catch data ( $y$ -axis) were rescaled so that the treatment with highest catch = 1 for each data set. See table 2 for parameter estimates.

with ca. 3 females captured per male (table 1). From the remaining taxa, only *Hylurgus ligniperda*, *Quedius* sp., *Rhizophagus ferrugineus* and *Thanasimus formicarius* were found to be significantly affected by the Ie proportion in the pheromonal blend. Means could not be separated for *Quedius* sp. (table 1).

Profiling of the response of species with total capture levels over 20 specimens showed that modeling of their response would not be possible in all cases. Only *I. sexdentatus*, *Acanthocinus griseus*, *Orthotomicus erosus* and *T. formicarius* responses could be fitted to the Gaussian curve (table 2). The *I. sexdentatus* response profile showed the characteristic bell-shaped profile (fig. 1a), and the response peak ( $\mu_r$ ) could be set at 0.227 Ie/Id + Ie, not far from its sex-ratio peak ( $\mu_r=0.111$ ; fig. 1b). The dose-response relationship of the remaining taxa was studied using linear regression (table 3). The accumulated catches of *Buprestis novemmaculata*, *H. ligniperda*, *R. ferrugineus*, and *Temnochila caerulea* were found to be positively correlated with Ie proportion in the lure (table 3).

### Guild structure

Xylophage and predator guilds followed catch trends seen for *I. sexdentatus* and *R. ferrugineus* (table 1), the most abundant species within each guild, respectively (table 4 and table A.1). Remaining guilds occurred in low numbers with no apparent variation in their response (table 4). Alternatively, Shannon index analysis revealed a significant increase in xylophage diversity with increased Ie percentage (table 5). Beyond the initial classification of captured taxa within the four main saproxyl guilds, those captured beetle species that have been shown to interact with conifer bark beetles were further tabulated into the three trophic groups related to *I. sexdentatus* and prone to detect its pheromones (table A.1; Herard & Mercadier, 1996; Kenis *et al.*, 2004). As shown in tables 2 and 3, modeling of the response of intraguild competitor and predators, and of the selected species of the predatory guild showed that the Gaussian curve could be fitted for competitors peaking close to that of *I. sexdentatus*, whereas both groups of

Table 3. Parameter estimates with their SE and goodness of fit of linear regression of the response of trapped saproxylic beetles and trophic guilds that did not show a Gaussian response. In all the cases, one and five regression and residual degrees of freedom, respectively. Asterisks after  $P(>F)$  values highlight: \*, <0.05; \*\*, <0.01 and \*\*\*, <0.001 significances.

| Taxon                    | Slope $\pm$ SE     | Intercept $\pm$ SE | Adj. $R^2$ | $F_{1,5}$ | $P(>F)$   |
|--------------------------|--------------------|--------------------|------------|-----------|-----------|
| <i>A. ferus</i>          | -0.006 $\pm$ 0.095 | 0.252 $\pm$ 0.073  | -0.199     | 0.003     | 0.955     |
| <i>B. novemmaculata</i>  | 0.269 $\pm$ 0.061  | 0.197 $\pm$ 0.047  | 0.755      | 19.47     | 0.007**   |
| <i>H. ligniperda</i>     | 0.437 $\pm$ 0.103  | 0.122 $\pm$ 0.079  | 0.738      | 17.93     | 0.008**   |
| <i>H. pini</i>           | 0.412 $\pm$ 0.124  | -0.008 $\pm$ 0.095 | 0.627      | 11.1      | 0.020*    |
| <i>P. parallelopedus</i> | -0.077 $\pm$ 0.097 | 0.282 $\pm$ 0.075  | -0.067     | 0.622     | 0.466     |
| <i>Q. abietum</i>        | -0.026 $\pm$ 0.137 | 0.297 $\pm$ 0.105  | -0.191     | 0.036     | 0.857     |
| <i>Quedum</i> sp.        | 0.142 $\pm$ 0.190  | 0.260 $\pm$ 0.146  | -0.079     | 0.559     | 0.488     |
| <i>R. ferrugineus</i>    | 0.519 $\pm$ 0.064  | 0.080 $\pm$ 0.049  | 0.915      | 66.25     | <0.001*** |
| <i>S. reyi</i>           | 0.236 $\pm$ 0.119  | 0.032 $\pm$ 0.092  | 0.328      | 3.93      | 0.104     |
| <i>T. caerulea</i>       | 0.512 $\pm$ 0.082  | 0.019 $\pm$ 0.063  | 0.865      | 39.39     | 0.001***  |
| <i>T. dermestoides</i>   | -0.298 $\pm$ 0.188 | 0.632 $\pm$ 0.144  | 0.202      | 2.522     | 0.173     |
| Intraguild predators     | 0.344 $\pm$ 0.08   | 0.123 $\pm$ 0.062  | 0.741      | 18.17     | 0.008**   |
| Predators                | 0.501 $\pm$ 0.087  | 0.01 $\pm$ 0.067   | 0.841      | 32.73     | 0.002**   |
| Saproxylophages          | 0.161 $\pm$ 0.104  | 0.315 $\pm$ 0.080  | 0.186      | 2.373     | 0.184     |
| Xylofungivores           | 0.238 $\pm$ 0.184  | 0.149 $\pm$ 0.142  | 0.100      | 1.667     | 0.253     |

potential *I. sexdentatus* predators showed a positive linear relationship with Ie percentage (fig. 2). Although a significant effect on the capture level could not be detected (table 4), the number of saproxylophages showed a positive significant linear relationship with the Ie percentage (table 3).

#### Species assemblages

The composition of beetles differed between levels of tested Ie percentages, and three clear groups could be established after Adonis (table 6). Low levels of Ie (0–10%) harbored compositions that could not be distinguished. Alternatively, the saproxylic beetle assemblages responding to 50, 90 and 95% Ie levels differed from this group and showed their own composition. The ordination of the data sets for the 49 experimental units and the subsequent fitting of the Ie percentage level as an explanatory factor confirmed results from Adonis (fig. 3).

#### Discussion

Our results provide new evidence of the role of *I. sexdentatus* infochemicals, could have in the attraction of several species lead to the establishment of its associated saproxylic beetle community, while assessing the response windows of the targeted bark beetle and that of a diverse assemblage of species belonging to different trophic guilds. Even if some of the captured beetles could have been randomly intercepted by multi-funnel traps, or through the visual attraction exhorted by the trunk-like silhouette of the trap (Strom *et al.*, 1999), the almost complete lack of captures in unbaited traps during the first experimental week, and their overlapping life histories, suggests an underlying kairomonal attraction of many of the tallied beetles. Many bark beetles have a well-known role in founding the saproxylic habitat, and hence it would not be surprising that a cohort of saproxylic beetles could use their infochemicals, in addition to those of their plant hosts, to locate appropriate foraging grounds (Wood, 1982; Grove, 2002; Seybold *et al.*, 2006; Foit, 2010). In any case, significant response changes to tested infochemical proportion range could only be proven for a few species. The vast majority of registered specimens corresponded to saproxylic taxa (table A.1), from which many have been listed in earlier studies and reviews describing bark beetle associated

entomofauna (Herard & Mercadier, 1996; Kenis *et al.*, 2004; Foit, 2010; Santolamazza-Carbone *et al.*, 2011). The discovery of a new elaterid species (*Athous (Orthatous)* n. sp.), together with the capture of the rare *Lathropus sepicola* (Baena *et al.*, 2011), *Chrysanthia reitteri* (Oedemeridae) and *Pachybrachis (Pachybrachis) suffriani* (Chrysomelidae), endemisms to the Iberian Peninsula (Lencina *et al.*, 2008), among ca. 100 identified species in a single experiment covering just part of *I. sexdentatus*' flight period, and using just two of the infochemicals involved in its communication (Francke *et al.*, 1986), somewhat illustrates the relatively little sampling effort received historically by the saproxylic beetle group in this region. Furthermore, even if the experimental period covered the peak of *I. sexdentatus* flight period, catches of some of its most important natural enemies were probably lower than what they could have been if sampling had been performed earlier or the experiment had lasted longer, e.g. very few *T. formicarius* were caught (grand total of 53 individuals) in comparison with trials performed in the same experimental area during spring and early summer (Etxebeste & Pajares, 2011; Etxebeste *et al.*, 2012).

The results obtained are in agreement with previous findings for the response of *I. sexdentatus* to the combined Ie and Id release, which pointed that highest catches were obtained when Ie:Id ratio values were close to natural, although the response peak could not be established (Vité *et al.*, 1972; Kohnle *et al.*, 1992; Etxebeste *et al.*, 2012). Figure 4, describing the evolution of the Ie proportion in the blend released by *I. sexdentatus* males boring into two different hosts (Kohnle, 1991; Etxebeste *et al.*, unpublished data), could provide the rationale behind the registered response maxima as well as for the parameter estimates of fitted Gaussian curve models. The response peak for both female and male *I. sexdentatus* was predicted to lie around the 20% Ie value (fig. 1), which would correspond to the registered blend released between days 5 and 10 of colonization, which in turn corresponds to the observed aggregation phase of the beetle (Wood, 1982; Francke *et al.*, 1986; Kohnle, 1991). In addition, the release rate of Id has been found to apparently peak 3–7 d after male settlement (Kohnle, 1991). Volatiles released during the first week after pioneer arrival have been studied in detail for *Ips typographus* (Birgersson *et al.*, 1984; Birgersson & Bergstrom, 1989), showing too that highest pheromone release rates occurred 3–4 d after the carving of nuptial chambers.

Table 4. Effect of increasing *Ie* proportion in lures on mean accumulated catches  $\pm$  SEM ( $n=7$ ) of the five main trophic groups. Asterisks after  $P(>F)$  values highlight: \*, <0.05; \*\*, <0.01 and \*\*\*, <0.001 significances of treatment effects. Shared letters within the same guild indicate that means are not significantly different (Tukey's HSD test, Bonferroni's adjustment,  $P<0.05$ ).

|                 | $F_{6,36}$ | $P(>F)$   | Ipsenol percentage in pheromonal blend |                       |                      |                       |                        |                     |                      |
|-----------------|------------|-----------|--|-----------------------|----------------------|-----------------------|------------------------|---------------------|----------------------|
|                 |            |           | 0                                      | 1                     | 5                    | 10                    | 50                     | 90                  | 95                   |
| Predators       | 13.15      | <0.001*** | 15.71 $\pm$ 5.58 a                     | 10.29 $\pm$ 2.52 a    | 29.43 $\pm$ 6.56 ab  | 33.57 $\pm$ 13.17 ab  | 32 $\pm$ 7.42 ab       | 61 $\pm$ 10.67 bc   | 94.14 $\pm$ 19.43 c  |
| Saproxylophages | 1.08       | 0.39      | 1.43 $\pm$ 0.97                        | 1.00 $\pm$ 0.53       | 0.71 $\pm$ 0.29      | 2.14 $\pm$ 1.18       | 1.29 $\pm$ 0.71        | 2.14 $\pm$ 1.18     | 2.00 $\pm$ 1.23      |
| Xylofungivores  | 2.36       | 0.050     | 0.57 $\pm$ 0.43                        | 0.43 $\pm$ 0.2        | 0.57 $\pm$ 0.43      | 0 $\pm$ 0             | 0.43 $\pm$ 0.2         | 0.29 $\pm$ 0.29     | 1.29 $\pm$ 0.18      |
| Xylophages      | 8.37       | <0.001*** | 132.14 $\pm$ 21.15 ab                  | 157.86 $\pm$ 32.13 ab | 270.71 $\pm$ 50.29 C | 222.86 $\pm$ 56.59 ac | 188.57 $\pm$ 34.00 abc | 98.57 $\pm$ 12.63 b | 100.57 $\pm$ 25.38 b |

Table 5. Effect of increasing *Ie* proportion in lures on mean Shannon diversity index  $\pm$  SEM ( $n=7$ ) within each trophic group Asterisks after  $P(>F)$  values highlight: \*, <0.05; \*\*, <0.01 and \*\*\*, <0.001 significances of treatment effects. Shared letters within the same guild indicate that means are not significantly different (Tukey's HSD test, Bonferroni's adjustment,  $P<0.05$ ).

|                 | $F_{6,36}$ | $P(>F)$   | Ipsenol percentage in pheromonal blend |                     |                     |                     |                     |                     |                    |
|-----------------|------------|-----------|--|---------------------|---------------------|---------------------|---------------------|---------------------|--------------------|
|                 |            |           | 0                                      | 1                   | 5                   | 10                  | 50                  | 90                  | 95                 |
| Predators       | 1.68       | 0.155     | 1.151 $\pm$ 0.16                       | 1.069 $\pm$ 0.215   | 0.679 $\pm$ 0.229   | 0.828 $\pm$ 0.136   | 0.621 $\pm$ 0.082   | 0.892 $\pm$ 0.15    | 0.797 $\pm$ 0.097  |
| Saproxylophages | 0.46       | 0.833     | 0.211 $\pm$ 0.211                      | 0.248 $\pm$ 0.168   | 0.099 $\pm$ 0.099   | 0.372 $\pm$ 0.187   | 0.235 $\pm$ 0.154   | 0.325 $\pm$ 0.231   | 0.317 $\pm$ 0.229  |
| Xylofungivores  | 0.77       | 0.595     | 0.091 $\pm$ 0.091                      | 0 $\pm$ 0           | 0.157 $\pm$ 0.157   | 0 $\pm$ 0           | 0 $\pm$ 0           | 0.099 $\pm$ 0.099   | 0.198 $\pm$ 0.128  |
| Xylophages      | 18.35      | <0.001*** | 0.457 $\pm$ 0.06 a                     | 0.459 $\pm$ 0.111 a | 0.468 $\pm$ 0.052 a | 0.494 $\pm$ 0.085 a | 0.564 $\pm$ 0.072 a | 0.877 $\pm$ 0.065 b | 1.141 $\pm$ 0.10 b |

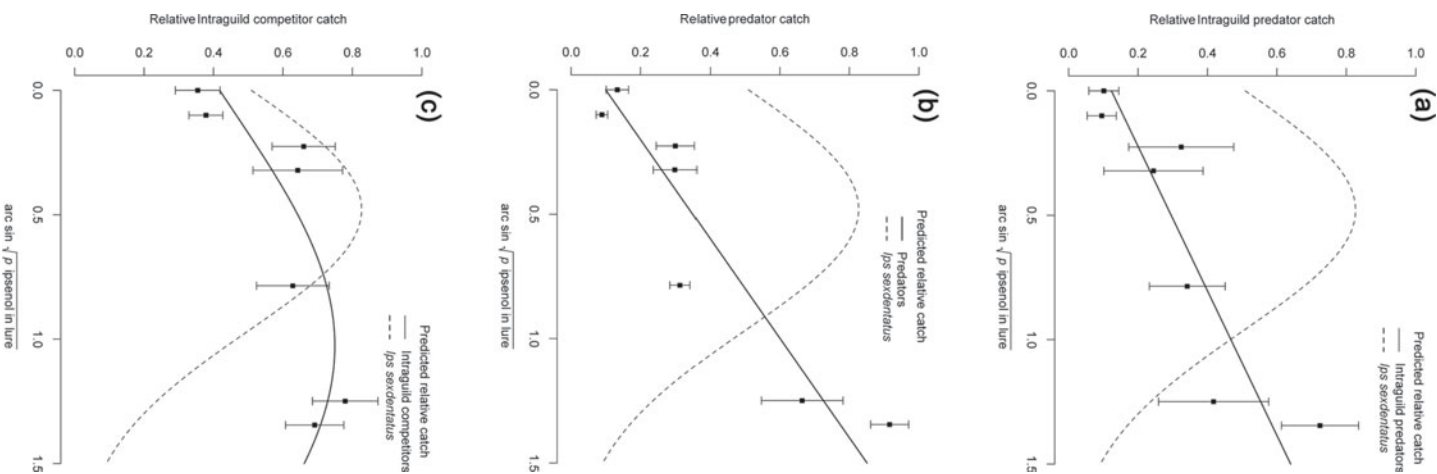


Fig. 2. Accumulated trap catches of competitor and predator guild associates and their modeled linear and Gaussian fitting in response to increasing proportions of Ipsenol in the pheromonal blend. Predicted *I. scaberrithus* response is also shown for comparison. (a) Intraguild (xylophage) predator response. (b) Predator response. (c) Intraguild (xylophage) competitor response. See Materials and methods section for guild descriptions. Catch data ( $y$ -axis) were rescaled so that the treatment with highest catch = 1 for each data set. See tables 2 and 3 for parameter estimates.

Table 6. *P* values of pairwise comparison tests (ADONIS, 1000 permutations) of saproxylic beetle beta diversity between treatment levels.

|    | Ipsenol percentage in pheromonal blend |       |       |        |        |        |
|----|--|-------|-------|--------|--------|--------|
|    | 1                                      | 5     | 10    | 50     | 90     | 95     |
| 0  | 0.466                                  | 0.063 | 0.058 | 0.028* | 0.014* | 0.016* |
| 1  |  | 0.309 | 0.637 | 0.015* | 0.012* | 0.019* |
| 5  |  |       | 0.086 | 0.049* | 0.018* | 0.014* |
| 10 |  |       |       | 0.017* | 0.015* | 0.040* |
| 50 |  |       |       |        | 0.014* | 0.017* |
| 90 |  |       |       |        |        | 0.306  |

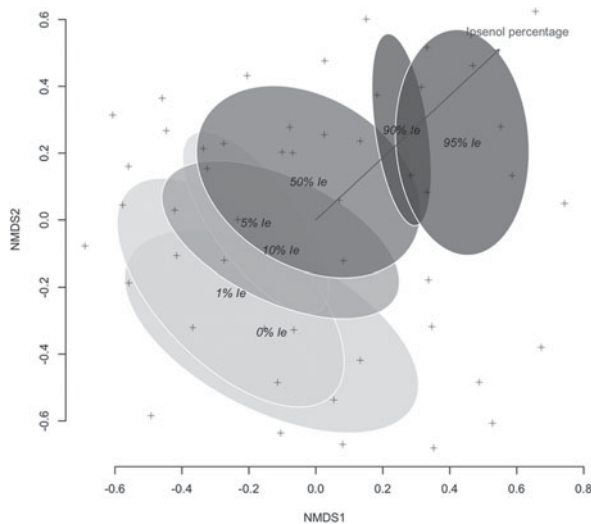


Fig. 3. Ordination of Bray–Curtis similarities (stress 0.268) of species and specimen abundance at multiple funnel traps using NMDS. Each cross represents each of the 49 sampling units. Ellipses represent factor class standard error area at 95% confidence interval, after fitting the Ipsenol percentage in the lure as a factor onto the ordination (shown vector,  $R^2=0.58$ ,  $P<0.001$ , 1000 permutations).

Both quantitative and qualitative change in the pheromone signal during initial colonizing days could provide bark beetles with the information on the substrate status. The sequence and mode in which pioneer beetles settle strongly modifies their reproductive success and survival (e.g., Aukema & Raffa, 2004a, b; Latty *et al.*, 2009; Latty & Reid, 2009). Yet, it is not clear whether pioneering confers any net advantage in reproductive success, as on the one hand pioneering *Dendroctonus ponderosae* were found to have reduced broods in comparison with early responders (Latty & Reid, 2009), while on the other hand, an increased risk for predation with time of arrival in responding males of *I. pini* has been reported (Aukema & Raffa, 2004b). Furthermore, as colonization proceeds, and the density of attackers increases, the number of eggs may decrease exponentially (Jactel & Lieutier, 1987). Registered response for *I. sexdentatus* reveals that the largest proportion of males got trapped at the pheromonal blend corresponding to the early stages of substrate colonization (fig. 1b), but also during the lowest predatory guild response (fig. 2). Catch maxima are reached

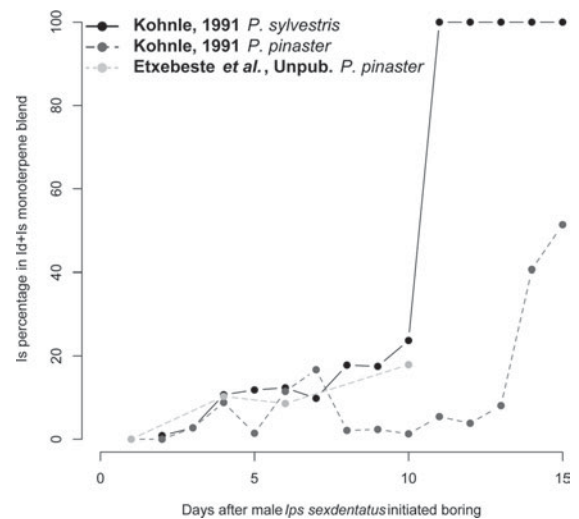


Fig. 4. Temporal evolution of the percentage of Ie in relation to Id in frass extracts of boring *I. sexdentatus* males. Data from Kohnle (1991) obtained through closed-loop stripping analysis, followed by filter extraction by MeCl and CS<sub>2</sub> in *P. pinaster* and *P. sylvestris*, respectively, and GC-MS. Unpublished data from Etxebeste *et al.* obtained through solid-phase microextraction of frass samples of two series of six *I. sexdentatus* males boring into a *P. pinaster* 50 cm long bole each.

'later' at the cost of suffering higher rates of predation, but specially competition (fig. 2; Schroeder & Weslien, 1994a; Dodds *et al.*, 2001; Raffa, 2001; Aukema & Raffa, 2004b). In other words, both modeled responses and the Ie percentage evolution in fig. 4 suggest that released pheromone blend is low in Ie while pioneering *I. sexdentatus* males initiate boring, then male *I. sexdentatus* responders gradually join the pioneers, facing the risk posed by host defenses but avoiding excessive competition. As colonization proceeds and Ie percentage increases in the pheromonal blend, proportionally more female *I. sexdentatus* arrive to a defenseless colonization spot, but facing higher competition and predation. Similar reflection of the steps in the behavioral sequence linked to changes on the proportions of the pheromone components has been reported for *Dendroctonus frontalis* (Pureswaran & Sullivan, 2012).

The responses registered for associated species further support the scenario described for the aggregation of *I. sexdentatus* and for the role of the changing pheromonal component proportion may have on it. Even if significant changes in response were detected only on nine of the captured species, in addition to signaling for the colonization phase, *I. sexdentatus* seems to avoid natural enemy and competing taxa also through a tuned pheromonal signal. Correspondingly, potential intraguild predators, such as *A. griseus* (Fabricius) or *B. novemmaculata* L., showed highly differentiated response profiles to that of *I. sexdentatus*, as their modeled response window were found to be positively correlated with or peaked at higher Ie proportion values (tables 2 and 3). Although a few studies have evaluated the impact on bark beetle larval survival of associated phloem feeder larvae, very high reductions on brood survival have been reported (Schroeder & Weslien, 1994a; Dodds *et al.*, 2001). Predatory species too followed the same pattern. While predator diversity does not change through the Ie gradient



(table 5), both accumulated predator guild and main predator (i.e. *R. ferrugineus*, *T. formicarius* and *T. caerulea*) responses increased with Ie percentage. Furthermore, 40% of registered saproxyl beetles belonged to this guild (table A.1), although only a few had accumulated catches above the arbitrarily set 20 individual threshold. *T. formicarius* and *T. caerulea* have been previously described to follow a similar pattern (Ettebeste *et al.*, 2012). Although attraction of *R. ferrugineus* to host volatiles had been reported before (e.g., Schroeder & Weslien, 1994b), the presented results provide the first evidence of this species being kairomonally cued to bark beetle pheromone components, which captured an especially large number of individuals (table 1). A similar conclusion can be reached when the results for *Hypophloeus pini* are considered. The genus is known to prey on bark beetles (Kenis *et al.*, 2004), but to our knowledge, kairomonal attraction has not been reported earlier. In addition, and although results have not been included in order to restrict the scope to coleopteran species, *Scoloposcelis pulchella* (Heteroptera, Anthocoridae), known for preying on bark beetle larvae (Herard & Mercadier, 1996; Kenis *et al.*, 2004), was captured in large numbers, and showed a positive linear relationship with Ie too.

The diversity of xylophages increased significantly along the Ie gradient (table 5), while competitor species response peaked at intermediate values (fig. 2c). *H. ligniperda* and *O. erosus* were the two main competitor species caught in the study. Kairomonal attraction to *I. sexdentatus* pheromonal components has been reported earlier for *H. ligniperda* (Ettebeste *et al.*, 2012), and according to the results presented in this work, this secondary bark beetle shows a positively correlated response with Ie proportion. In previous trials, Ie alone was found to attract less individuals than when released along with Id (Ettebeste *et al.*, 2012). Taken together, these results suggest that *H. ligniperda* may eavesdrop on *I. sexdentatus* and arrive at colonization spots once its settlement has finished, and taking advantage of a weakened host. As for *O. erosus*' response, it was not strongly affected by the change in the pheromone blend (table 1), confirming previous reports (Ettebeste *et al.*, 2012). This bark beetle requires another infochemical, 2-methyl-3-buten-2-ol, in addition to Id for its pheromone (Klimetzek & Vité, 1986; Seybold *et al.*, 2006). Even so, its response could be modeled to a response peak close to the 40% Ie level, which was thus differentiated from the response peak for *I. sexdentatus*.

In addition, all these functional changes in the responses of species and trophic guilds were reflected in the results of assemblage analysis. Three main groups could be differentiated (table 6; fig. 3), which corresponded to (i) the area of *I. sexdentatus* maximal response (0–10% Ie), (ii) the competitor area (50% Ie), and (iii) the predator group area (90 and 95% Ie). Thus, beyond the described responses of saproxyl species and guilds, well-differentiated species assemblages were caught along the Ie gradient, highlighting the role of *I. sexdentatus* infochemicals in assisting resource partitioning.

Although Ie has been associated with the pheromone of *Ips* and other bark beetle genera (e.g., Francke *et al.*, 1986; Seybold *et al.*, 2006) is not but one of the several volatile compounds with behavioral effects detected for *I. sexdentatus* (Kohnle, 1991). A far more complex signal scenario arises when other infochemicals are considered. On the one hand, host volatiles are used by several guilds to locate foraging grounds (e.g. Schroeder & Weslien, 1994b), while on the other hand, derivative volatiles emitted by bark beetles, as for example,

*cis-verbenol*, do enhance bark beetle response (*I. sexdentatus*) but are also used by natural enemies to locate their prey (*T. caerulea*; Ettebeste *et al.*, 2012). Moreover, many of the infochemicals involved have stereoisomers, to which bark beetles can respond in a specific manner aiding them escape from predators or competitors (Raffa & Klepzig, 1989; Raffa, 2001). Furthermore, a recent report has shown how secondary species that join the pioneering bark beetles may exert negative effects in addition to mere competition, as their pheromone components may attract third-party predators that influence reproductive success of pioneers (Boone *et al.*, 2008). In summary, even if presented results do reflect the steps seen in bark beetle settlement in terms of pheromone blend change, the complete representation of this process would involve the description of the responses of each of the associated species in terms of enantiomeric composition, synergists, kairomones, concentrations, and variations in space and time (Raffa, 2001).

The implications of detailed characterization of the response to pheromone blends in bark beetle management have been previously recognized (Raffa & Klepzig, 1989; Grégoire *et al.*, 1992). On one side, figures derived from monitoring programs aimed to estimate both scolytid and associated beetle populations based on pheromone-baited traps are probably inaccurate, and need to be adjusted to response disparities and, on the other, the use of pheromones as the control method by mass-trapping and related tactics is frequently hindered by the negative impact that these programs have on natural enemy populations (Raffa & Klepzig, 1989; Ettebeste *et al.*, 2012). Furthermore, the capture of rare species that might be associated with *I. sexdentatus* founded habitats in a rather 'small' experiment raises the question of what consequences these programs could have on the conservation of saproxyl species. Excessive forest hygiene or salvage logging has been pointed to as causes of loss of mature timber, which hosts many of those species (Grove, 2002; Foit, 2010). Appropriate hosts in managed stands occur highly dispersed temporarily and geographically, especially for secondary bark beetles such as *I. sexdentatus*, which normally require a weakened host to settle. Thus, artificial bark beetle 'foundations', i.e. pheromone-baited traps, might produce an unwanted impact on the saproxyl community, as is the case with some of the known predators (Ettebeste *et al.*, 2012). Results also demonstrate that pheromone-baited traps, although highly specific, may be used in addition to traps baited with host volatiles or photoelectors (e.g., Wermelinger, 2002) to sample for saproxyl species that eavesdrop on bark beetle infochemicals.

Additional research exploring interactions among bark beetles and associated insects in the phloem of host trees appears important if we are to increase our understanding of bark beetle population dynamics (e.g. Billings, 1988; Aukema *et al.*, 2000b; Dodds *et al.*, 2001). At the specific *I. sexdentatus* case, characterization of the response at the stereochemistry level of the pheromone components may further clarify the response patterns of this bark beetle, as well as that of its associated saproxyl beetles as a basis for a sustainable pest management.

### Acknowledgements

We acknowledge the aid given by Dr Volker Assing (Hanover, Germany) and José A. Sáez Bolaño (Badajoz, Spain)

in the identification of rove beetles (Staphylinidae) and specimens of the Elateridae and Trosidae families, respectively. We also acknowledge the aid provided by a number of people, especially Gonzalo Álvarez and Estela Sánchez from the Sustainable Forest Management Research Institute (University of Valladolid-INIA), Ana B. Martín, Gema Pérez and Luis Miguel from the Regional Forest Health Centre at Calabazanos (Palencia, Castile and Leon) and Dionisio Pozo from the Department of Environment of the Regional Castile and Leon Government. This work has been financed by the Spanish Science and Education Ministry, within the AGL 2004-07507-C04-04 and AGL 2007-61152 research projects. The first author was supported by a scholarship within a fellowship between the University of Valladolid and the Department of Environment of the Castile and Leon Autonomous Government.

### References

- Allison, J.D., Borden, J.H., McIntosh, R.L., de Groot, P. & Gries, R. (2001) Kairomonal response by four *Monochamus* species (Coleoptera: Cerambycidae) to bark beetle pheromones. *Journal of Chemical Ecology* **27**, 633–646.
- Aukema, B.H. & Raffa, K.F. (2004a) Does aggregation benefit bark beetles by diluting predation? Links between a group-colonisation strategy and the absence of emergent multiple predator effects. *Ecological Entomology* **29**, 129–138.
- Aukema, B.H. & Raffa, K.F. (2004b) Gender- and sequence-dependent predation within group colonizers of defended plants: a constraint on cheating among bark beetles? *Oecologia* **138**, 253–258.
- Aukema, B.H. & Raffa, K.F. (2005) Selective manipulation of predators using pheromones: responses to frontalin and ipsdienol pheromone components of bark beetles in the Great Lakes region. *Agricultural and Forest Entomology* **7**, 193–200.
- Aukema, B.H., Dahlsten, D.L. & Raffa, K.F. (2000a) Exploiting behavioral disparities among predators and prey to selectively remove pests: maximizing the ratio of bark beetles to predators removed during semiochemically based trap-out. *Environmental Entomology* **29**, 651–660.
- Aukema, B.H., Dahlsten, D.L. & Raffa, K.F. (2000b) Improved population monitoring of bark beetles and predators by incorporating disparate behavioral responses to semi-chemicals. *Environmental Entomology* **29**, 618–629.
- Baena, M., Lencina, J.L. & Andújar, C. (2011) Presencia de *Lathropus sepicola* (Müller, 1821) (Coleoptera: Laemphloeidae) en Sierra Madrona, Ciudad Real (España). *Boletín de la Sociedad Entomológica Aragonesa* **49**, 332.
- Billings, R.F. (1988). Forecasting southern pine beetle infestation trends with pheromone traps. In *Proceedings of the Symposium: Integrated Control of Scolytid Bark Beetles; 4 July, 1988. IUFRO and 17th International Congress of Entomology*, Vancouver, BC. Virginia Polytechnic Institute and State University, Blacksburg, VA. 295–306.
- Birgersson, G. & Bergstrom, G. (1989) Volatiles released from individual spruce bark beetle entrance holes – quantitative variations during the first week of attack. *Journal of Chemical Ecology* **15**, 2465–2483.
- Birgersson, G., Schlyter, F., Lofqvist, J. & Bergstrom, G. (1984) Quantitative variation of pheromone components in the spruce bark beetle *Ips typographus* (Coleoptera, Scolytidae) from different attack phases. *Journal of Chemical Ecology* **10**, 1029–1055.
- Blomquist, G.J., Figueroa-Teran, R., Aw, M., Song, M.M., Gorzalski, A., Abbott, N.L., Chang, E. & Tittiger, C. (2010) Pheromone production in bark beetles. *Insect Biochemistry and Molecular Biology* **40**, 699–712.
- Boone, C.K., Six, D.L. & Raffa, K.F. (2008) The enemy of my enemy is still my enemy: competitors add to predator load of a tree-killing bark beetle. *Agricultural and Forest Entomology* **10**, 411–421.
- Bouget, C., Brustel, H. & Nageleisen, L.M. (2005) Nomenclature of wood-inhabiting groups in forest entomology: synthesis and semantic adjustments. *Comptes Rendus Biologies* **328**, 936–948.
- Bussler, H., Bouget, C., Brustel, H., Brandle, M., Riedinger, V., Brandl, R. & Muller, J. (2011) Abundance and pest classification of scolytid species (Coleoptera: Curculionidae, Scolytinae) follow different patterns. *Forest Ecology and Management* **262**, 1887–1894.
- Carroll, A.L., Taylor, S., Régnière, J. & Safranyik, L. (2004). Effects of climate change on range expansion by the mountain pine beetle in British Columbia. In *Mountain Pine Beetle Symposium: Challenges and Solutions. 30–31 October 2003*, Kelowna, British Columbia.
- Crawley, M.J. (2007). *The R Book*. Chichester, Wiley.
- Dajoz, R. (2000). *Insects and Forests: The Role and Diversity of Insects in the Forest Environment*. Londres, Intercept.
- Dodds, K.J., Graber, C. & Stephen, F.M. (2001) Facultative intraguild predation by larval Cerambycidae (Coleoptera) on bark beetle larvae (Coleoptera: Scolytidae). *Environmental Entomology* **30**, 17–22.
- EFI (2010) Destructive Storms in European Forests: Past and Forthcoming Impacts. European Forest Institute, Atlantic European Regional Office – EFIATLANTIC, Cestas, France. p. 138.
- Etxebeste, I. & Pajares, J.A. (2011) Verbenone protects pine trees from colonization by the six-toothed pine bark beetle, *Ips sexdentatus* Boern. (Col.: Scolytinae). *Journal of Applied Entomology* **135**, 258–268.
- Etxebeste, I., Álvarez, G., Pérez, G. & Pajares, J.A. (2012) Field response of the six-toothed pine bark beetle, *Ips sexdentatus* (Col.: Curculionidae, Scolytinae), to pheromonal blend candidates. *Journal of Applied Entomology* **136**, 431–444.
- Fauna Europaea. (2010) Fauna Europaea version 2.4. Available online at <http://www.faunaeur.org> (accessed 2011).
- Foit, J.É. (2010) Distribution of early-arriving saproxylic beetles on standing dead Scots pine trees. *Agricultural and Forest Entomology* **12**, 133–141.
- Francke, W., Pan, M.L., Bartels, J., Konig, W.A., Vité, J.P., Krawielitzki, S. & Kohnle, U. (1986) The odor bouquet of three pine engraver beetles (*Ips* spp.). *Journal of Applied Entomology-Zeitschrift Fur Angewandte Entomologie* **101**, 453–461.
- Gil, L.A. & Pajares, J.A. (1986) *Los Escoltídeos de las Coníferas en la Península Ibérica*. Madrid, Instituto Nacional de Investigaciones Agrarias.
- Grégoire, J.C., Couillien, D., Drumont, A., Meyer, H. & Francke, W. (1992) Semiochemicals and the management of *Rhizophagus grandis* Gyll (Col., Rhizophagidae) for the bio-control of *Dendroctonus micans* Kug (Col., Scolytidae). *Journal of Applied Entomology-Zeitschrift Fur Angewandte Entomologie* **114**, 110–112.
- Grove, S.J. (2002) Saproxylic insect ecology and the sustainable management of forests. *Annual Review of Ecology and Systematics* **33**, 1–23.
- Herard, F. & Mercadier, G. (1996) Natural enemies of *Tomicus piniperda* and *Ips acuminatus* (Col., Scolytidae) on *Pinus*

- sylvestris* near Orleans, France: temporal occurrence and relative abundance, and notes on eight predatory species. *Entomophaga* **41**, 183–210.
- Ibeas, F., Gallego, D., Diez, J.J. & Pajares, J.A.** (2007) An operative kairomonal lure for managing pine sawyer beetle *Monochamus galloprovincialis* (Coleoptera: Cerymbycidae). *Journal of Applied Entomology* **131**, 13–20.
- Jactel, H. & Lieutier, F.** (1987) Effects of attack density on fecundity of the Scots pine-beetle *Ips sexdentatus* Boern (Col., Scolytidae). *Journal of Applied Entomology-Zeitschrift Fur Angewandte Entomologie* **104**, 190–204.
- Kenis, M. & Hilszczanski, J.** (2004) Natural enemies of cerambycidae and buprestidae infesting living trees. pp. 475–498 in Lieutier, F., Day, K.R., Battisti, A., Grégoire, J.-C. & Evans, H.F. (Eds) *Bark and Wood Boring Insects in Living Trees in Europe, a Synthesis*. Dordrecht, Kluwer Academic Publishers.
- Kenis, M., Wermelinger, B. & Grégoire, J.-C.** (2004) Research on Parasitoids and predators of Scolytidae – a review. pp. 237–290 in Lieutier, F., Day, K.R., Battisti, A., Grégoire, J.-C. & Evans, H.F. (Eds) *Bark and Wood Boring Insects in Living Trees in Europe, a Synthesis*. Dordrecht, Kluwer Academic Publishers.
- Klimetzek, D. & Vité, J.P.** (1986) The role of insect produced attractants on the aggregation behavior of the Mediterranean pine engraver beetle *Orthotomicus erosus*. *Journal of Applied Entomology-Zeitschrift Fur Angewandte Entomologie* **101**, 239–243.
- Kohnle, U.** (1991) Verhaltensmodifizierende Duftstoffe in der Aggregation von Borkenkäfern der Gattung *Ips* DeGeer (Col., Scolytidae). *Freiburger Waldschutz-Abhandlungen, Herausgegeben vom Forstzoologischen Institut*. Freiburg im Breisgau, Albert-Ludwigs-Universität Freiburg i. Br. p. 156.
- Kohnle, U., Meyer, M. & Kluber, J.** (1992) Formulation of population attractant for the pine bark beetle, *Ips sexdentatus* (Col., Scolytidae). *Allgemeine Forst Und Jagdzeitung* **163**, 81–87.
- Latty, T.M. & Reid, M.L.** (2009) First in line or first in time? Effects of settlement order and arrival date on reproduction in a group-living beetle *Dendroctonus ponderosae*. *Journal of Animal Ecology* **78**, 549–555.
- Latty, T.M., Magrath, M.J.L. & Symonds, M.R.E.** (2009) Harem size and oviposition behaviour in a polygynous bark beetle. *Ecological Entomology* **34**, 562–568.
- Lencina, J.L., Gallego, D. & Andújar, C.** (2008) Nuevos datos de Oedemeridae Latreille, 1810 de la Península Ibérica (Coleoptera). *Heteropterus Revista de Entomología* **8**, 95–107.
- Lieutier, F., Day, K.R., Battisti, A., Grégoire, J.-C. & Evans, H.F.** (Eds.) (2004) *Bark and Wood Boring Insects in Living Trees in Europe, a Synthesis*. Dordrecht, Kluwer Academic Publishers.
- Lindgren, B.S.** (1983) A multiple funnel trap for scolytid beetles (Coleoptera). *Canadian Entomologist* **115**, 299–302.
- Nieto, A. & Alexander, K.N.A.** (2010) *European Red List of Saproxyl community, guild and species responses of a pine bark beetle*. Luxembourg, Publications Office of the European Union.
- Oksanen, J., Roeland, K., Legendre, P., O'Hara, B., Simpson, G. L., Solymos, P., Stevens, H.H. & Wagner, H.** (2008) *Vegan: Community Ecology Package*. R package version 1.15-1.
- Pajares, J.A., Ibeas, F., Diez, J.J. & Gallego, D.** (2004) Attractive responses by *Monochamus galloprovincialis* (Col., Cerambycidae) to host and bark beetle semiochemicals. *Journal of Applied Entomology* **128**, 633–638.
- Poland, T.M. & Borden, J.H.** (1994) Semiochemical-based communication in interspecific interactions between *Ips pini* (Say) and *Pityogenes knechteli* (Swaine) (Coleoptera, Scolytidae) in lodgepole pine. *Canadian Entomologist* **126**, 269–276.
- Pureswaran, D.S. & Sullivan, B.T.** (2012) Semiochemical emission from individual galleries of the Southern Pine Beetle, (Coleoptera: Curculionidae: Scolytinae), attacking standing trees. *Journal of Economic Entomology* **105**, 140–148.
- Raffa, K.F.** (2001) Mixed messages across multiple trophic levels: the ecology of bark beetle chemical communication systems. *Chemoecology* **11**, 49–65.
- Raffa, K.F. & Berryman, A.A.** (1983) The role of host plant-resistance in the colonization behavior and ecology of bark beetles (Coleoptera: Scolytidae). *Ecological Monographs* **53**, 27–49.
- Raffa, K.F. & Klepzig, K.D.** (1989) Chiral escape of bark beetles from predators responding to a bark beetle pheromone. *Oecologia* **80**, 566–569.
- Reeve, J.D.** (1997) Predation and bark beetle dynamics. *Oecologia* **112**, 48–54.
- Reeve, J.D. & Strom, B.L.** (2004) Statistical problems encountered in trapping studies of scolytids and associated insects. *Journal of Chemical Ecology* **30**, 1575–1590.
- Roelofs, W.L.** (1978) Threshold hypothesis for pheromone perception. *Journal of Chemical Ecology* **4**, 685–699.
- Ross, D.W. & Daterman, G.E.** (1995) Efficacy of an antiaggregation pheromone for reducing Douglas-fir beetle, *Dendroctonus pseudotsugae* Hopkins (Coleoptera: Scolytidae), infestation in high risk stands. *Canadian Entomologist* **127**, 805–811.
- Santolamazza-Carbone, S., Pestaña, M. & Vega, J.A.** (2011) Post-fire attractiveness of maritime pines (*Pinus pinaster* Ait.) to xylophagous insects. *Journal of Pest Science* **84**, 343–353.
- Schlyter, F., Svensson, M., Zhang, Q.H., Knizek, M., Krokene, P., Ivarsson, P. & Birgersson, G.** (2001) A model for peak and width of signaling windows: *Ips duplicatus* and *Chilo partellus* pheromone component proportions – does response have a wider window than production? *Journal of Chemical Ecology* **27**, 1481–1511.
- Schroeder, L.M. & Weslien, J.** (1994a) Interactions between the phloem-feeding species *Tomicus piniperda* (Col., Scolytidae) and *Acanthocinus aedilis* (Col., Cerambycidae), and the predator *Thanasimus formicarius* (Col., Cleridae) with special reference to brood production. *Entomophaga* **39**, 149–157.
- Schroeder, L.M. & Weslien, J.** (1994b) Reduced offspring production in bark beetle *Tomicus piniperda* in pine bolts baited with ethanol and alpha-pinene, which attract antagonistic insects. *Journal of Chemical Ecology* **20**, 1429–1444.
- Seybold, S., Huber, D., Lee, J., Graves, A. & Bohlmann, J.** (2006) Pine monoterpenes and pine bark beetles: a marriage of convenience for defense and chemical communication. *Phytochemistry Reviews* **5**, 143–178.
- Seybold, S.J., Teale, S.A., Wood, D.L., Zhang, A.J., Webster, F.X., Lindahl, K.Q. & Kubo, I.** (1992) The role of lanierone in the chemical ecology of *Ips pini* (Coleoptera, Scolytidae) in California. *Journal of Chemical Ecology* **18**, 2305–2329.
- Strom, B.L., Roton, L.M., Goyer, R.A. & Meeker, J.R.** (1999) Visual and semiochemical disruption of host finding in the southern pine beetle. *Ecological Applications* **9**, 1028–1038.
- Teale, S.A. & Lanier, G.N.** (1991) Seasonal variability in response of *Ips pini* (Coleoptera, Scolytidae) to Ipsdienol in New York. *Journal of Chemical Ecology* **17**, 1145–1158.
- Teale, S.A., Hager, B.J. & Webster, F.X.** (1994) Pheromone-based assortative mating in a bark beetle. *Animal Behaviour* **48**, 569–578.

- The R Development Core Team** (2011). R: A Language and Environment for Statistical Computing. Vienna, Austria, R Foundation for Statistical Computing.
- Vité, J.P., Bakke, A. & Renwick, J.A.A.** (1972) Pheromones in *Ips* (Col.: Scolytidae) – occurrence and production. *Canadian Entomologist* **104**, 1967–1975.
- Vité, J.P., Bakke, A. & Hughes, P.R.** (1974) sex attractant of bark beetles, *Ips sexdentatus*. *Naturwissenschaften* **61**, 365–366.
- Vité, J.P., Ohloff, G. & Billings, R.F.** (1978) Pheromonal chirality and integrity of aggregation response in southern species of bark beetle *Ips* sp. *Nature* **272**, 817–818.
- Wermelinger, B.** (2002) Development and distribution of predators and parasitoids during two consecutive years of an *Ips typographus* (Col., Scolytidae) infestation. *Journal of Applied Entomology* **126**, 521–527.
- Wermelinger, B.** (2004) Ecology and management of the spruce bark beetle *Ips typographus* – a review of recent research. *Forest Ecology and Management* **202**, 67–82.
- Weslien, J.** (1994) Interactions within and between species at different densities of the bark beetle *Ips typographus* and its predator *Thanasimus formicarius*. *Entomologia Experimentalis et Applicata* **71**, 133–143.
- Wood, D.L.** (1982) The role of pheromones, kairomones, and allomones in the host selection and colonization behavior of bark beetles. *Annual Review of Entomology* **27**, 411–446.



Appendix: Table A1. List of saproxylic beetles captured at multiple funnel traps baited with *I. sexdentatus* pheromone blends at a site in Northwest Spain (UTM 29T 7390 4729), arranged by trophic guilds. Species richness (S) and total catches per species and trophic guild are provided too.

| Family                 | Subfamily       | Species   | Total catch |
|------------------------|-----------------|---|-------------|
| Xylofungivores         |                 | S = 11  | 25          |
| ANOBIIDAE              | Dorcatominae    | <i>Stagetus elongatus</i> (Mulsant & Rey, 1861)                             | 1           |
| CIIDAE                 |                 | Ciidae 001  | 1           |
|                        |                 | Ciidae 002  | 3           |
| CRYPTOPHAGIDAE         |                 | Cryptophagidae 001  | 1           |
| EUCINETIDAE            |                 | <i>Nycteus</i> sp.  | 3           |
| LATRIDIIDAE            | Latridiinae     | <i>Cartodere (Aridius) nodifer</i> (Westwood, 1839)                         | 1           |
| LEIODIDAE              | Leiodinae       | <i>Agathidium</i> sp.   | 6           |
|                        |                 | <i>Leiodes</i> sp.  | 4           |
|                        |                 | Leiodidae 001   | 2           |
|                        |                 | Leiodidae 002   | 2           |
| STAPHYLINIDAE          | Tachyporinae    | <i>Mycetoporus</i> sp.  | 1           |
| <b>Saproxylophages</b> |                 | <b>S = 12</b>   | <b>75</b>   |
| ANOBIIDAE              | Ernobiinae      | <i>Ernobius gigas</i> (Mulsant & Rey, 1863)                                 | 11          |
| ANTHRIBIDAE            | Anthribinae     | <i>Allandrus undulatus</i> (Panzer, 1795)                                   | 7           |
| BUPRESTIDAE            | Buprestinae     | <i>Anthaxia (Melanthaxia) morio</i> (Fabricius, 1792)                       | 12          |
|                        |                 | <i>Anthaxia (Melanthaxia)</i> sp.   | 2           |
|                        |                 | <i>Phaenops cyanea cyanea</i> (Fabricius, 1775)                             | 6           |
|                        | Chrysobothrinae | <i>Chrysobothris (Chrysobothris) solieri</i> (Laporte & Gory, 1839)         | 1           |
| CERAMBYCIDAE           | Lepturinae      | <i>Stictoleptura rubra</i> (Linnaeus, 1758)                                 | 1           |
| ELATERIDAE             |                 | <i>Athous (Orthatous)</i> n. sp.  | 10          |
|                        |                 | <i>Athous (Orthatous)</i> sp.   | 1           |
|                        |                 | <i>Cardiophorus (Cardiophorus) signatus</i> (Olivier, 1790)                 | 2           |
|                        |                 | <i>Elathous rufus</i> (Candeze, 1860)                                       | 2           |
| TRHOSCIDAE             |                 | <i>Trixagus dermestoides</i> (Linnaeus, 1766)                               | 20          |
| <b>Predators</b>       |                 | <b>S = 26</b>   | <b>1933</b> |
| CARABIDAE              | Lebiinae        | <i>Calodromius spilotus</i> (Illiger, 1798) <sup>1</sup>                    | 7           |
|                        |                 | <i>Dromius (Dromius) agilis</i> (Fabricius, 1787) <sup>1</sup>              | 1           |
| CLERIDAE               | Clerinae        | <i>Allonyx quadrimaculatus</i> (Schaller, 1783) <sup>1</sup>                | 7           |
|                        |                 | <i>T. formicarius</i> (Linnaeus, 1758) <sup>1</sup>                         | 53          |
| CRYPTOPHAGIDAE         |                 | Cryptophagidae 002  | 2           |
|                        |                 | Cryptophagidae 003  | 4           |
| HISTERIDAE             | Abraeinae       | <i>Plegaderus (Plegaderus) saucius</i> (Erichson, 1834) <sup>1</sup>        | 9           |
|                        |                 | <i>Teretrius (Neotepetrius) parasita</i> (Marseul, 1862)                    | 1           |
|                        | Dendrophilinae  | <i>Paromalus (Paromalus) parallelopipedus</i> (Herbst, 1792) <sup>1</sup>   | 68          |
|                        | Histerinae      | <i>Cylister elongatus</i> (Thunberg, 1787) <sup>1</sup>                     | 1           |
| LAEMOPHLOEIDAE         |                 | <i>L. sepicola</i> (Muller, 1821)   | 1           |
| MALACHIIDAE            | Malachiinae     | <i>Axinotarsus (Axinotarsus) marginalis</i> (Laporte de Castelnau, 1840)    | 1           |
| MONOTOMIDAE            |                 | <i>Rhizophagus (Rhizophagus) ferrugineus</i> (Paykull, 1800) <sup>1</sup>   | 1321        |
| MYCETOPHAGIDAE         | Mycetophaginae  | <i>Litargus (Litargus) connexus</i> (Geoffroy, 1785) <sup>1</sup>           | 4           |
| NITIDULIDAE            | Eपुरaeinae      | <i>Eपुरaea</i> sp. <sup>1</sup>   | 3           |
| SALPINGIDAE            | Salpinginae     | <i>Sphaeriestes (Sphaeriestes) reyi</i> (Abeille de Perrin, 1874)           | 22          |
| STAPHYLINIDAE          | Aleocharinae    | <i>Leptusa pulchella</i> (Mannerheim, 1831)                                 | 3           |
|                        | Staphylininae   | <i>Platydracus (Platydracus) chalconcephalus</i> (Fabricius, 1801)          | 2           |
|                        |                 | <i>Quedius</i> sp. <sup>1</sup>   | 62          |
|                        |                 | <i>Quedius (Microsaurus) abietum</i> (Kiesenwetter, 1858) <sup>1</sup>      | 22          |
|                        | Tachyporinae    | <i>Sepedophilus</i> sp.   | 2           |
|                        |                 | <i>Sepedophilus testaceus</i> (Fabricius, 1793) <sup>1</sup>                | 8           |
| TENEBRIONIDAE          | Diaperinae      | <i>Hypophloeus linearis</i> (Fabricius, 1790) <sup>1</sup>                  | 1           |
|                        |                 | <i>H. pini</i> (Panzer, 1799) <sup>1</sup>                                  | 50          |
| TROGOSITIDAE           | Trogositinae    | <i>T. caerulea</i> (Olivier, 1790) <sup>1</sup>                             | 267         |
| ZOPHERIDAE             | Colydiinae      | <i>Aulonium ruficorne</i> (Olivier, 1790) <sup>1</sup>                      | 11          |
| <b>Xylophages</b>      |                 | <b>S = 16</b>   | <b>8199</b> |
| BUPRESTIDAE            | Buprestinae     | <i>Buprestis (Buprestis) novemmaculata</i> (Linnaeus, 1758) <sup>2</sup>    | 57          |
| CERAMBYCIDAE           | Cerambycinae    | <i>Xylotrechus arvicola</i> (Olivier, 1795)                                 | 2           |
|                        | Lamiinae        | <i>A. griseus</i> (Fabricius, 1792) <sup>2</sup>                            | 36          |
|                        |                 | <i>Monochamus galloprovincialis</i> (Olivier, 1795) <sup>2</sup>            | 2           |
|                        | Spondylidinae   | <i>Arhopalus ferus</i> (Mulsant, 1839) <sup>2</sup>                         | 139         |
| CURCULIONIDAE          | Entiminae       | <i>Brachyderes (Brachyderes) incanus</i> (Linnaeus, 1758) <sup>2</sup>      | 1           |
|                        |                 | <i>Brachyderes (Brachyderes) lusitanicus</i> (Fabricius, 1781) <sup>2</sup> | 1           |
|                        | Mesoptiliinae   | <i>Magadalis (Magdalis) rufa</i> (Germar, 1824)                             | 2           |
|                        | Platypodinae    | <i>Platypus cylindrus</i> (Fabricius, 1792) <sup>3</sup>                    | 2           |
|                        | Scolytinae      | <i>Crypturgus cinereus</i> (Herbst, 1793) <sup>3</sup>                      | 1           |

Appendix: Table A1. (Cont.)

| Family         | Subfamily    | Species<br>$S = 11$   | Total catch<br>25 |
|----------------|--------------|---|-------------------|
| Xylofungivores |              | <i>H. ligniperda</i> (Fabricius, 1787) <sup>3</sup>   | 481               |
|                |              | <i>I. sexdentatus</i> (Borner, 1776)  | 6706              |
|                |              | <i>O. erosus</i> (Wollaston, 1857) <sup>3</sup>   | 758               |
|                |              | <i>Pityogenes bidentatus</i> (Herbst, 1784) <sup>3</sup>                                    | 1                 |
|                |              | <i>Pityogenes quadridens</i> (Hartig, 1834) <sup>3</sup>                                    | 9                 |
|                |              | <i>Lasioryhynchites</i> ( <i>Stenorhynchites</i> ) <i>coeruleocephalus</i> (Schaller, 1783) | 1                 |
| RHYNCHITIDAE   | Rhynchitinae |   |                   |

<sup>1</sup> Species with known linkage to bark beetles and used for the predator guild response modeling.

<sup>2</sup> Xylophage species with larvae sharing the same host substrate and consuming bark beetle larvae on conifers, and used for the intraguild predator response modeling.

<sup>3</sup> Xylophage species potentially competing with *I. sexdentatus* for feeding and breeding substrate, and used for the intraguild competitor response.