Growth and home range size of the gracile mouse opossum *Gracilinanus microtarsus* (Marsupialia: Didelphidae) in Brazilian cerrado

Fernanda Rodrigues Fernandes^{*,1}, Leonardo Dominici Cruz⁺, Eduardo Guimarães Martins^{+,2} and Sérgio Furtado dos Reis⁺

Abstract: Differences in growth patterns between the sexes of the gracile mouse opossum *Gracilinanus microtarsus* and the consequences for home range size were investigated in a savanna habitat (cerrado) of south-eastern Brazil. A total of 51 juvenile individuals of *Gracilinanus microtarsus* was monitored using capture–mark–recapture from November 2005 to August 2006. The increase in body mass of gracile mouse opossums was described using the Gompertz growth model. Male gracile mouse opossums grew faster than females (dimorphic ratio of 1.5). Home range size, estimated with the minimum convex polygon method, was positively related to body mass. Model selection using Akaike's Information Criterion (AIC_c) and incorporating body mass, sex and season as independent variables showed that the best-supported model describing variance in home range sizes included only body mass. Our data suggest that a greater body mass gain in juvenile males is probably the proximate cause of sexual dimorphism in adult gracile mouse opossums and that energetic needs required for growth have a greater influence in home range size.

Key Words: body mass, cerrado, growth, home range, marsupial, population ecology

INTRODUCTION

Sexual size dimorphism (SSD) is widespread and variable among animals and the ultimate causes of SSD are associated with three major processes: sexual selection, fecundity selection and ecological causation (Blanckenhorn 2005, Shine 1989). From a proximate perspective, the SSD of a given species is caused by sexbiased maternal investment (before birth or during food provisioning/suckling) or differences between the sexes in development time and growth rate (the larger sex has to develop faster or for a longer period of time) (Foster & Taggart 2008, Koskela *et al.* 2004).

Mammals are generally dimorphic in size with a bias toward males, with males being at least 10% larger than

females in over 45% of species (Lindenfors *et al.* 2007). Male-biased SSD may evolve through sexual selection on male body size: larger males can be more successful at acquiring mating opportunities through male-male combat, leading to the evolution of larger male body size (Cox *et al.* 2003, Weckerly 1998).

SSD is common in didelphids (Cáceres *et al.* 2007, Isaac 2006, Loretto & Vieira 2008) with males being larger than females in most species. Didelphids are considered polygynous, and in this promiscuous mating system males compete for access to breeding females, which favours phenotypic adaptations that enhance the ability of a male to prevail in male-male contests; e.g. a rapid growth rate and large body mass (Moore 1990, Schulte-Hostedde *et al.* 2001).

Dimorphism in body mass can contribute to differences in the behaviour, demography, life history, physiology, ecology and evolution of males and females within a population (Cox *et al.* 2003). Variability in growth parameters between the sexes may have consequences in

^{*} Programa de Pós-graduação em Ecologia, Universidade Estadual de Campinas, Instituto de Biologia, Departamento de Parasitologia, Caixa Postal 6109, Campinas, São Paulo, 13083-970, Brazil

[†] Programa de Pós-graduação em Ciências Biológicas (Zoologia), Instituto de Biociências, Universidade Estadual Paulista Júlio de Mesquita Filho, Rio Claro, São Paulo, Brazil

[‡] Departamento de Parasitologia, Instituto de Biologia, Universidade Estadual de Campinas, Campinas, São Paulo, Brazil (Accepted 8 October 2009)

¹ Corresponding author. Email: nandafernandes@gmail.com

² Current address: Department of Forest Sciences, The University of British Columbia, Vancouver, BC V6T 1Z4, Canada.

space-use patterns (Dahle *et al.* 2006, Kelt & Van Vuren 1999). In polygynous mating systems females tend to have more stable home ranges whereas males tend to be more vagile, resulting in differences in use of space between the sexes (Loretto & Vieira 2005).

Home range is the area traversed by the individual in its normal activities of food gathering, mating and caring for young (Burt 1943). McNab (1963) suggested that mammal home range area reflects metabolic needs. Many studies have employed allometric relationships to demonstrate the relationship between home range size and body mass (Kelt & Van Vuren 2001, Linstedt *et al.* 1986). Besides body mass, others factors like sex, diet, population density, resource seasonality and habitat heterogeneity can cause variation in home range size (Dahle & Swenson 2003, Harestad & Bunnel 1979).

Home range size can differ between the sexes because males and females are dimorphic in size and have differences in metabolic need (Cederlund & Source 1994), use resources differently (Safi *et al.* 2007) or have different reproductive strategies (Dahle & Swenson 2003).

Seasonal differences in home range size can result from differences in the availability of resources at different times of the year, so at times when food is scarce or when there is the need to find mates (during the breeding season), individuals travel greater distances compared to periods in which food is abundant or during times outside the breeding season (Getz & McGuire 2008, Lesmeister *et al.* 2009).

In this study, we gathered data on a cohort of *Gracilinanus microtarsus*, a species of south-eastern Brazil with SSD to test the following hypotheses: (1) that the proximate cause of sexual size dimorphism in *G. microtarsus* is the difference in growth velocity (growth constant) between the sexes, and (2) that the home range size is positively related to body mass and consequently the home range size is larger in males than in females.

METHODS

Study species

Gracilinanus microtarsus Wagner 1842 is a small, solitary, arboreal and nocturnal didelphid marsupial that inhabits cerrado (savanna) and the Atlantic forest of south-eastern Brazil (Costa *et al.* 2003, Martins *et al.* 2006a). This species shows sexual size dimorphism, with adult males (30–45 g) being larger than adult females (20–30 g) (Martins *et al.* 2006a) and occurs at high densities in the cerrado (Martins *et al.* 2006a). Mating in this species is concentrated in a short period between August and September (the end of the cool-dry season) and individuals reproduce for the first time when they are approximately 1 y old, so females rear their offspring during the first half

of the warm-wet season (October–December) (Martins *et al.* 2006a).

Study area

Our study was carried out in savanna habitat at the Reserva Biológica de Mogi Guaçu (RBMG) located in the district of Martinho Prado, Mogi Guaçu, São Paulo $(22^{\circ}15'-22^{\circ}18'S; 47^{\circ}08'-47^{\circ}13'W)$. Vegetation at the RBMG consists of cerrado, which is a neotropical savanna formation comprising different vegetation physiognomies that differ in the density and composition of woody and ground-layer plants, forming a continuum from open and dry grassland to dense forest (Goodland 1971). The RBMG is a remnant of the physiognomy known locally as 'cerrado sensu stricto' which is woodland with scattered trees 5-8 m tall and closed scrub (Oliveira-Filho & Ratter 2002). The climate of the region has two well-defined seasons: the warm-wet season occurs from October to March whereas the cool-dry season occurs from April to September. The mean annual rainfall and the mean annual temperature are, respectively, 1430 mm and $21 \degree C$ (data from the meteorological station of RBMG).

Data collection

Fieldwork was conducted from November 2005 to August 2006. In November 2005, a cohort of G. microtarsus was captured and then monitored until they reached the sub-adult stage. The marsupials were captured monthly within the 4 mo of the warm-wet season (November 2005, December 2005, January 2006 and March 2006) and within the 4 mo of the cool-dry season (May 2006, June 2006, July 2006 and August 2006). The data location points were obtained on 10 consecutive days in each mo. An 11×11 trapping grid (22 500 m²) with 121 trapping stations located 15 m apart was used. A single Sherman live-trap $(7.5 \times 9.0 \times 23.5 \text{ cm})$ was set on trees at each trapping-station c. 1.75 m above ground and baited with banana and peanut butter. The individuals captured were marked with a numbered leg-band and their sex and weight were recorded with a Pesola[®] balance (precision =1 g) (Costa et al. 2003). Location points of the captures was recorded and defined as x and y co-ordinates in the space of trapping-stations.

Live specimens are more prone to measurement error than are dead specimens (Blackwell *et al.* 2006), however it was necessary to monitor live individuals for a certain period of time to obtain growth and home range data. Weight record error was minimised because all individuals (males and females) were weighed by the same observer in all samples, using the same balance and procedure.

Data analysis

Body mass data from females and males were fitted using the non-linear Gompertz growth model $W = Ae^{-e^{-K(t-1)}}$ where W represents body mass at time t, A the asymptotic body mass, I the inflection point and K a growth constant (Begall 1997). The day of first capture of juveniles was designated as day 0 for modelling procedures and we fixed the asymptotic mass at mean values of adult females and males captured early in the study area ($A_{\circ} = 29 \pm 2$ g, n = 10 and $A_{\sigma} = 38 \pm 2$ g, n = 6). The parameters K and I were estimated by least squares using the Gauss-Newton algorithm, and the adequacy of the model was determined from a determination coefficient (R^2 of the correlation between observed and predicted body mass values) (Souza 1998). The growth constant K can be regarded as a measure of growth; i.e. the higher the K. the faster the growth; and because of its independence of adult weight, intraspecific comparisons are possible (Begall 1997). Thus, differences in model parameters between females and males were evaluated using a t-test with $\alpha = 0.05$. The analyses were conducted using SAS.

Data location points were used to estimate home range sizes of individuals of G. microtarsus through the minimum convex polygon (MCP) method (Mohr 1947) using the program CALHOME (Kie et al. 1996). In the MPC method the outer data points obtained for an individual animal are connected by a connection rule in which no internal angle of the polygon exceeds 180° (Mohr 1947). The area of the polygon is then calculated and taken as an estimate of the home range size. As minimum convex polygon estimates are dependent on the number of captures per individual, all estimates of home range sizes were based on individuals with at least five location points, for which estimates were more reliable (Cáceres & Monteiro-Filho 2001). In the CALHOME program we chose 90% MCPs, so that the program excluded 10% of the outer data points to generate home range areas. Therefore, a 90% convex polygon is the smallest area derived by connecting location points such that the resultant polygon encloses 90% of all observations. The 90% MCPs are less dependent on the sample size, because as new positions are added, more outlying positions are excluded (Dahle & Swenson 2003).

We applied a linear regression by plotting the distribution of the logarithm of home range size (m^2) against log_{10} body mass (g) to determine if there was a significant relationship between home range size and body mass. We only used data from individuals that had home range areas located totally inside the trapping grid away from the boundaries for the linear regression because sometimes the home range areas can be underestimated if they are located on the trapping-grid boundaries.

We used home range estimates obtained from the MPC method to model variation in home range size as function

of body mass, sex and season. Generalized linear models were used with the response variable (home range size) distributed according to a Gamma distribution and with the inverse link function (Dobson 2002, McCullagh & Nelder 1989). We fitted models including the following effects (notation): (1) mass (mass); (2) sex (sex); (3) season (season); (4) additive effect of mass and season (mass + season); (5) additive effect of mass and sex (mass + sex); (6) additive effect of sex and season (sex + season); (7) additive effect of mass, sex and season (mass + sex + season); (8) additive effect of sex and season and interaction between sex and season (sex + season + sex × season); (9) additive effect of mass, sex and season and interaction between sex and season (mass + sex + season + sex × season).

The adequacy of the general model to data was tested using a Pearson chi-squared test (Sokal & Rohlf 1995). The selection of most parsimonious model to describe the data was based on Akaike's Information Criterion, AIC, using the corrected version for small sample size, AIC_c (Burnham & Anderson 1998). AIC_c was calculated as AIC_c = $-2 \ln L(\hat{\theta}|m) + 2K \left(\frac{n}{n-K-1}\right)$, where $\ln L(\hat{\theta}|m)$ is the natural logarithm of the likelihood function evaluated at the maximum likelihood estimates of a given model (*m*), *K* is the number of parameters in the model and *n* is the sample size. According to this criterion, the model with the lowest AIC_c value is the most parsimonious model. Differences of AIC_c values between m model and the most parsimonious model, Δ_m , were used to compare the support of different models in the set of candidate models. As suggested by Burnham & Anderson (1998), models with $0 \le \Delta_m \le 2$ were considered high support models. Statistics analyses were conducted using the GLM procedure in SAS.

RESULTS

Body mass

A total of 51 juvenile individuals of *Gracilinanus microtarsus* were captured from November 2005 to August 2006. During the initial fieldwork in November, individuals captured had body masses that ranged from 6 g to 11 g. The mean body mass for both sexes in each month is shown in Table 1.

The Gompertz model described the body mass growth for both females and males ($R^2_{\varphi} = 0.646$ and $R^2_{\sigma'} = 0.789$) (Table 2, Figure 1). After the age of 180 d, body mass of males and females did not overlap (Figure 1). The *K* and *I* parameters differed between females and males in the Gompertz model with males showing the growth constant (*K*) larger than females (t = 3.15; df = 103; P = 0.003) and the inflection point (*I*) later than females (t = 2.12; df = 103; P = 0.04) (Table 2). The dimorphic

Table 1. Mean \pm SD body mass of males and females of *Gracilinanusmicrotarsus* captured in RBMG, Martinho Prado, São Paulo, south-
eastern Brazil.

		Body mass (g)		
Months	Season	Female (n)	Male (n)	
November	Warm-wet	$8 \pm 2 (7)$	11(1)	
December	Warm-wet	$10 \pm 2(15)$	$13 \pm 2 (10)$	
January	Warm-wet	$13 \pm 2 (15)$	$14 \pm 1 (7)$	
March	Warm-wet	$14 \pm 3 (9)$	$18 \pm 2 (9)$	
May	Cool-dry	15 ± 1 (6)	$18 \pm 2 (6)$	
June	Cool-dry	$16 \pm 1 (5)$	$23 \pm 3(5)$	
July	Cool-dry	$17 \pm 2 (4)$	$26 \pm 2(5)$	
August	Cool-dry	18 ± 1 (2)	$29 \pm 4 (7)$	

Table 2. Gompertz growth model parameters of *Gracilinanus microtarsus* ($_{\varphi} =$ female, $_{\sigma} =$ male) captured in RBMG, Martinho Prado, São Paulo, south-eastern Brazil, from November 2005 to August 2006. ($\hat{K} =$ the growth constant, $\hat{I} =$ the inflection point).

Parameter	Estimate	Asymptotic SE	Estimate Asymptotic SE	95% Confidence Interval
\widehat{K}_{Q}	0.00316	0.000325	9.72	0.0029-0.0046
\widehat{K}_{\circ}	0.00483	0.000403	11.98	0.0048-0.0068
$\widehat{I}_{\mathcal{Q}}$	26	10	2.6	6-47
\widehat{I}_{σ}	55	9	6.1	37-73



Figure 1. Body growth of the females (observed values = open circles; Gompertz curve = dotted line) and males (observed values = closed circles; Gompertz curve = solid line) of *Gracilinanus microtarsus* captured in RBMG, Martinho Prado, São Paulo, south-eastern Brazil, from November 2005 to August 2006. $R^2_{\phi} = 0.646$ and $R^2_{\sigma} = 0.789$.

ratio $(\hat{K}_{\sigma}/\hat{K}_{\varphi})$ was 1.5, indicating that males grew faster than females.

Home range

We used data location points from 26 individuals (12 females and 14 males) to estimate home range size due to requirements of the MPC method. These data were used

Table 3. Home range area of Gracil	linanus microtarsus ($_{\circ} =$ female, $_{\circ} =$
male) captured in both seasons	in RBMG, Martinho Prado, São
Paulo, south-eastern Brazil, from	November 2005 to August 2006,
as estimated by the MPC method.	

Individuals	Sex	Body mass (g)	Home range size (m^2)	Season
B89	Ŷ	14	1463	warm-wet
		17	1688	cool-dry
B92	Q	11	2587	warm-wet
		15	900	cool-dry
B120	Q	12	2363	warm-wet
		16	2700	cool-dry
B123	Q	14	563	warm-wet
		15	675	cool-dry
B131	ď	16	1800	warm-wet
		22	6300	cool-dry
B152	ď	21	1350	warm-wet
		21	1350	cool-dry

for the analysis of models that best explained the variance in home range size.

Analysing only the individuals for which a home range area was estimated in both seasons, we noted that home range size increased proportionately to the body mass gain. Table 3 shows that females B89 and B120 increased 3 g and 4 g, respectively, in their body mass and increased 225 m² and 337 m² in their home range, respectively, from the warm-wet season to the cool-dry season. The female B123 increased 1 g in its body mass and 112 m² in its home range from the warm-wet season to the cooldry season. Whereas male B131 increased 6 g in its body mass and 4500 m² in its home range from the warm-wet season to the cool-dry season, male B152 remained with the same weight and the same home range size from the warm-wet season to the cool-dry season.

Although female B92 had a body mass gain of 4 g, there was a reduction in the size of her home range from the warm-wet season to the cool-dry season.

Plotting home range size against body mass of individuals with home range areas located totally inside the trapping grid, we observed that the linear regression has a significantly positive slope (y = 0.88 + 1.84x; R² = 0.39; F = 5.84; df = 1.9; P = 0.04; n = 10; Figure 2).

The general model (mass + sex + season + sex × season) fit the data well ($\chi^2 = 23.1$; df = 31; P = 0.85). The model selection using Akaike's Information Criterion (AIC_c) showed that the most parsimonious model to describe variance in home range sizes included only body mass as the independent variable. The summary of the statistics of model selection and parameter estimates of the best model is shown in Tables 4 and 5, respectively.

DISCUSSION

The juveniles of *Gracilinanus microtarsus* grew throughout the study to reach adult body size. No female individual

Table 4. Selection of the most parsimonious model based on Akaike's information criterion (AIC_c) for the description of home range data of *Gracilinanus microtarsus* captured in RBMG, Martinho Prado, São Paulo, south-eastern Brazil, from November 2005 to August 2006. AIC_c = Akaike's Information Criterion corrected for small samples, Δ_m = differences of AIC_c values between a given model *m* and the most parsimonious model, w_i = the proportional likelihood of the models, *K* = the number of parameters in the model.

Model	AIC _c	Δ_m	w _i	K
mass*	0.94^{*}	0.00*	0.49*	2*
mass + season	3.21	2.27	0.16	3
mass + sex	3.32	2.38	0.15	3
season	4.86	3.92	0.07	2
mass + sex + season	5.74	4.80	0.04	4
sex + season	6.49	5.55	0.03	3
sex	6.62	5.67	0.03	2
$sex + season + sex \times season$	8.34	7.39	0.01	4
$mass + sex + season + sex \times season$	8.40	7.45	0.01	5

Table 5. Parameter estimates of the best model for the description of homerange data of *Gracilinanus microtarsus* captured in RBMG, MartinhoPrado, São Paulo, south-eastern Brazil, from November 2005 to August2006.

Model	Estimates	SE	Wald statistic	Р
Intercept	0.00133	0.000249	28.3	< 0.00001
Mass	-0.000029	0.000010	8.61	0.003336

had reached adult body mass by the end of the fieldwork and only a few male individuals had already reached adult body mass in July and August. Adult males of the gracile mouse opossum weigh 30–45 g and adult females weigh 20–30 g (Martins *et al.* 2006a).

Differences in growth patterns between the sexes, with males having a faster body mass gain, resulted in sexual size dimorphism in this marsupial. This proximate



Figure 2. Plot of the distribution of home range size against body mass of juveniles of *Gracilinanus microtarsus* captured in RBMG, Martinho Prado, São Paulo, south-eastern Brazil, from November 2005 to August 2006. $R^2 = 0.39$; F = 5.84; df = 1.9; P = 0.04; regression line = dotted line.

mechanism probably occurs after birth (during suckling or after they begin to forage by themselves), because marsupial offspring born after a very short gestation period at an early stage of development weigh less than 1 g at birth (Isaac 2006). Many marsupials and other mammals present differences in post-weaning growth rates and juvenile males often grow faster than juvenile females (Badyaev 2002, Isaac 2006, Lee & Cockburn 1985).

The majority of mammals have a predominantly polygynous mating system, with males competing for access to breeding females (Krebs & Davies 1981) and it can therefore be predicted that selection will favour phenotypic adaptations that enhance the ability of a male to grow rapidly and have a large body size. However, the mating success of females is commonly less dependent on body size and therefore females are expected to invest resources into reproduction rather than body growth (Isaac 2006, Schulte-Hostedde *et al.* 2001).

Gracilinanus microtarsus has a polygynous mating system and mating is concentrated in a short period between August and September (the end of cool-dry season) allowing females to rear their offspring during the first half of the warm-wet season (October to December) (Martins *et al.* 2006a) when insects, the main food resource for this species (Martins & Bonato 2004, Martins *et al.* 2006b), are highly abundant in the cerrado (Pinheiro *et al.* 2002). Therefore, weaning probably occurs at the beginning of the warm-wet season.

Martins et al. (2006c) demonstrated that males of G. microtarsus are partially semelparous (a condition in which mortality after the first mating is high but not complete, with a small fraction of males surviving to a second breeding season). One of the forces that drives semelparity (a single reproductive episode followed by death) in males is inter-male competition (Hollelev et al. 2006). Intense male-male competition during a short mating period and low probability of male survival to the next breeding season due to severe physiological stress results in selection for males to expend maximum effort in a single breeding season (Boonstra 2005, Oakwood et al. 2001). A larger body size can intensify reproductive effort since it increases mating access and maintains intromission despite aggression from other males (Holleley et al. 2006, Schulte-Hostedde et al. 2001, Weckerly 1998), but the stress leads to reduced survival (Martins et al. 2006c, Oakwood et al. 2001).

Home range size was positively related to body mass, so heavier individuals had larger home ranges. Larger species have higher energy demands and necessitate larger areas for food gathering (Harestad & Bunnel 1979, Kelt & Van Vuren 1999, 2001; McNab 1963), unless food exists in superabundance. At the intraspecific level, larger individuals have larger home ranges because they probably have larger energy demands and need to cover larger distances to find the amount of food that supplies their requirements. However, home range sizes can be underestimated because the MPC method describes only a 2-dimensional area, and, since *G. microtarsus* is an arboreal species, individuals can utilise both the vertical and horizontal dimensions of the habitat, perhaps using one dimension to a greater extent than the other.

Since *G. microtarsus* is sexually dimorphic in size, sex should be an important factor influencing home range size. Differences between male and female home range sizes as a consequence of size dimorphism are observed in some marsupials, like *Burramys parvus* (Broome 2001), *Dasyurus maculatus* (Belcher & Darrant 2004), *Didelphis aurita* (Cáceres & Monteiro-Filho 2001), *Micoureus demerarae* (Moraes-Júnior & Chiarello 2005) and *Phascogale tapoatafa* (Soderquist 1995). However, because almost no individual had reached adult body mass at the end of data collection, sexual dimorphism was not as pronounced and was unlikely to cause a great effect on home range size.

Reproductive season is another factor that probably causes variation in home range size. However, this factor could not be investigated because we did not observe a reproductive season during the data sampling period. In some species that have a promiscuous mating system, reproductive season can cause great variation in male home range size. In this season, males increase their home range sizes while searching for breeding females to mate (Loretto & Vieira 2005, Moraes-Júnior & Chiarello 2005).

Habitat quality and structure can also influence home range size. In habitats with abundant food resources, the resource quality alone might dictate smaller home range sizes than in habitats where these factors are scarce (Ims 1987, Stradiotto *et al.* 2009). Therefore, stability in habitat quality may be reflected in stability of home range sizes. In addition to habitat quality, habitat structure may influence the size and spatial distribution of the home range (Lambert *et al.* 2008, Lucherini & Lovari 1996). The female B92 may have found a microhabitat of better quality and therefore not expanded the size of her home range despite having increased her body mass from the warm-wet season to the cold-dry season.

In the present study, body mass was the most important factor influencing home range size, probably because energetic needs required for growth can be an important factor in juveniles that are growing to reach adult body size.

ACKNOWLEDGEMENTS

We are very grateful to João Del Giudice Neto and Marcos Mecca Pinto of Mogi Guaçu Biological Reserve for logistical support. Fernanda Rodrigues Fernandes and Leonardo Dominici Cruz were supported by a scholarship from Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq, Brazil: 135028/2005-1 and 131806/2006-8). This research was supported by grants from Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP, Brazil).

LITERATURE CITED

- BADYAEV, A. V. 2002. Growing apart: an ontogenetic perspective on the evolution of sexual size dimorphism. *Trends in Ecology and Evolution* 17:369–378.
- BEGALL, S. 1997. The application of the Gompertz model to describe body growth. *Growth, Development and Aging* 61:61–67.
- BELCHER, C. A. & DARRANT, J. P. 2004. Home range and spatial organization of the marsupial carnivore, *Dasyurus maculatus maculatus* (Marsupialia: Dasyuridae) in south-eastern Australia. *Journal of Zoology* 262:271–280.
- BLACKWELL, G. L., BASSETT, S. M. & DICKMAN, C. R. 2006. Measurement error associated with external measurements commonly used in small-mammal studies. *Journal of Mammalogy* 87:216–223.
- BLANCKENHORN, W. U. 2005. Behavioral causes and consequences of sexual size dimorphism. *Ethology* 111:977–1016.
- BOONSTRA, R. 2005. Equipped for life: the adaptive role of the stress axis in male mammals. *Journal of Mammalogy* 86:236–247.
- BROOME, L. S. 2001. Density, home range, seasonal movements and habitat use of the mountain pygmy-possum *Burramys parvus* (Marsupialia: Burramyidae) at Mount Blue Cow, Kosciusko National Park. *Austral Ecology* 26:275–292.
- BURNHAM, K. P. & ANDERSON, D. R. 1998. Model selection and multimodel inference: a practical information-theoretic approach. (Second edition). Springer, New York. 488 pp.
- BURT, W. H. 1943. Territoriality and home ranges concepts as applied to mammals. *Journal of Mammalogy* 24:346–352.
- CÁCERES, N. C. & MONTEIRO-FILHO, E. L. A. 2001. Food habits, home range and activity of *Didelphis aurita* (Mammalia, Marsupialia) in a forest fragment of Southern Brazil. *Studies on Neotropical Fauna and Environment* 36:85–92.
- CÁCERES, N. C., NAPOLI, R. P., LOPES, W. H., CASELLA, J. & GAZETA, G. S. 2007. Natural history of the marsupial *Thylamys macrurus* (Mammalia, Didelphidae) in fragments of savannah in southwestern Brazil. *Journal of Natural History* 41:1979–1988.
- CEDERLUND, G. & SOURCE, H. S. 1994. Home-range size in relation to age and sex in moose. *Journal of Mammalogy* 75:1005–1012.
- COSTA, L. P., LEITE, Y. L. R. & PATTON, J. L. 2003. Phylogeography and systematic notes on two species of gracile mouse opossums, genus *Gracilinanus* (Marsupialia: Didelphidae) from Brazil. *Proceedings of the Biological Society of Washington* 116:275–292.
- COX, R. M., SKELLY, S. L. & JOHN-ALDER, H. B. 2003. A comparative test of adaptive hypotheses for sexual size dimorphism in lizards. *Evolution* 57:1653–1669.
- DAHLE, B. & SWENSON, J. E. 2003. Home ranges in adult Scandinavian brown bears (*Ursus arctos*): effects of mass, sex, reproductive category, population density and habitat type. *Journal of Zoology* 260:329–335.
- DAHLE, B., OLE-GUNNAR, S. & SWENSON, J. E. 2006. Factors influencing home-range size in subadult brown bears. *Journal of Mammalogy* 87:859–865.

- DOBSON, A. J. 2002. An introduction to generalized linear models. (Second edition). Chapman & Hall, Boca Raton. 225 pp.
- FOSTER, W.K. & TAGGART, D.A. 2008. Gender and parental influences on the growth of a sexually dimorphic carnivorous marsupial. *Journal* of Zoology 275:221–228.
- GETZ, L. L. & MCGUIRE, B. 2008. Factors influencing movement distances and home ranges of the Short-tailed Shrew (*Blarina brevicauda*). Northeastern Naturalist 15:293–302.
- GOODLAND, R. 1971. A physiognomic analysis of the 'cerrado' vegetation of Central Brasil. *Journal of Ecology* 59:411–419.
- HARESTAD, A. S. & BUNNEL, F. L. 1979. Home range and body weight a reevaluation. *Ecology* 60:389–402.
- HOLLELEY, C. E., DICKMAN, C. R., CROWTHER, M. S. & OLDROYD, B. P. 2006. Size breeds success: multiple paternity, multivariate selection and male semelparity in a small marsupial, *Antechinus* stuartii. Molecular Ecology 15:3439–3448.
- IMS, R. A. 1987. Responses in spatial organization and behaviour to manipulations of the food resource in the vole *Clethrionomys rufocanus*. *Journal of Animal Ecology* 56:585–596.
- ISAAC, J. L. 2006. Sexual dimorphism in a marsupial: seasonal and lifetime differences in sex-specific mass. *Australian Journal of Zoology* 54:45–50.
- KELT, D. A. & VAN VUREN, D. 1999. Energetic constraints and the relationship between body size and home range area in mammals. *Ecology* 80:337–340.
- KELT, D. A. & VAN VUREN, D. 2001. The ecology and macroecology of mammalian home range area. *American Naturalist* 157:637–645.
- KIE, J. G., BALDWIN, J. A. & EVANS, C. J. 1996. CALHOME: a program for estimating animal home ranges. Wildlife Society Bulletin 24:342– 344.
- KOSKELA, E., HUITU, O., KOIVULA, M., KORPIMÄKI, E. & MAPPES, T. 2004. Sex-biased maternal investment in voles: importance of environment conditions. *Proceedings of the Royal Society of London, B. Biological Sciences* 271:1385–1391.
- KREBS, J. R. & DAVIES, N. B. 1981. An introduction to behavioural ecology. Blackwell Scientific Press, Oxford. 292 pp.
- LAMBERT, M. S., QUY, R. J., SMITH, R. H. & COWAN, D. P. 2008. The effect of habitat management on home-range size and survival of rural Norway rat populations. *Journal of Applied Ecology* 45:1753– 1761.
- LEE, A. K. & COCKBURN, A. 1985. Evolutionary ecology of marsupials. Cambridge University Press, New York. 274 pp.
- LESMEISTER, D. B., GOMPPER, M. E. & MILLSPAUGH, J. J. 2009. Habitat selection and home range dynamics of Eastern Spotted Skunks in the Ouachita Mountains, Arkansas, USA. *Journal of Wildlife Management* 73:18–25.
- LINDENFORS, P., GITTLEMAN, J. L. & JONES, K. E. 2007. Sexual size dimorphism in mammals. Pp. 16–26 in Fairbairn, D. J., Blanckenhorn, W. U. & Székely, T. (eds.). Sex, size, and gender roles: evolutionary studies of sexual size dimorphism. Oxford University Press, New York.
- LINSTEDT, S. L., MILLER, B. J. & BUSKIRK, S. W. 1986. Home range, time, and body size in mammals. *Ecology* 67:413–418.
- LORETTO, D. & VIEIRA, M. V. 2005. The effects of reproductive and climatic seasons on movements in the Black-eared opossum (*Didelphis aurita* Wied-Neuwied, 1826). *Journal of Mammalogy* 86:287–293.

- LORETTO, D. & VIEIRA, M V. 2008. Use of space by the marsupial Marmosops incanus (Didelphimorphia, Didelphidae) in the Atlantic Forest, Brazil. Mammalian Biology 73:255–261.
- LUCHERINI, M. & LOVARI, S. 1996. Habitat richness affects home range size in the red fox *Vulpes vulpes*. *Behavioural Processes* 36:103– 106.
- MARTINS, E. G. & BONATO, V. 2004. On the diet of *Gracilinanus* microtarusus (Marsupialia, Didelphidae) in an Atlantic Rainforest fragment in southeastern Brazil. *Mammalian Biology* 69:58– 60.
- MARTINS, E. G., BONATO, V., DA-SILVA, C. Q. & REIS, S. F. 2006a. Seasonality in reproduction, age structure and density of the gracile mouse opossum *Gracilinanus microtarsus* (Marsupialia: Didelphidae) in a Brazilian cerrado. *Journal of Tropical Ecology* 22:461– 468.
- MARTINS, E. G., BONATO, V., PINHEIRO, H. P. & REIS, S. F. 2006b. Diet of the gracile mouse opossum (*Gracilinanus microtarsus*) (Didelphimorphia: Didelphidae) in a Brazilian cerrado: patterns of food consumption and intrapopulation variation. *Journal of Zoology* 269:21–28.
- MARTINS, E. G., BONATO, V., DA-SILVA, C. Q. & REIS, S. F. 2006c. Partial semelparity in the neotropical didelphid marsupial *Gracilinanus microtarsus. Journal of Mammalogy* 87:915–920.
- McCULLAGH, P. & NELDER, J. A. 1989. Generalized linear models. (Second edition). Chapman & Hall, New York. 532 pp.
- McNAB, B. K. 1963. Bioenergetics and the determination of home range size. American Naturalist 97:133–140.
- MOHR, C. O. 1947. Table of equivalent populations of North American small mammals. *American Midland Naturalist* 37:223–249.
- MOORE, A. J. 1990. The evolution of sexual dimorphism by sexual selection: the separate effects of intrasexual selection and intersexual selection. *Evolution* 44:315–331.
- MORAES-JÚNIOR, E. A. & CHIARELLO, A. G. 2005. A radio tracking study of home range and movements of the marsupial *Micoureus demerarae* (Thomas) (Mammalia, Didelphidade) in the Atlantic forest of south-eastern Brazil. *Revista Brasileira de Zoologia* 22:85– 91.
- OAKWOOD, M., BRADLEY, A. J. & COCKBURN, A. 2001. Semelparity in a large marsupial. *Proceedings of the Royal Society of London, B. Biological Sciences* 268:407–411.
- OLIVEIRA-FILHO, A. T. & RATTER, J. A. 2002. Vegetation physiognomies and woody flora of the Cerrado biome. Pp. 91–120 in Oliveira, P. S. & Marques, R. J. (eds). *The cerrados of Brazil: ecology and natural history of a Neotropical savanna*. Columbia University Press, New York. 424 pp.
- PINHEIRO, F., DINIZ, I. R., COELHO, D. & BANDEIRA, M. P. S. 2002. Seasonal pattern of insect abundance in the Brazilian cerrado. *Austral Ecology* 27:132–136.
- SAFI, K., KÖNIG, B. & KERTH, G. 2007. Sex differences in population genetics, home range size and habitat use of the parti-colored bat (*Vespertilio murinus*, Linnaeus 1758) in Switzerland and their consequences for conservation. *Biological Conservation* 137:28– 36.
- SCHULTE-HOSTEDDE, A. E., MILLAR, J. S. & HICKLING, G. J. 2001. Sexual dimorphism in body composition of small mammals. *Canadian Journal of Zoology* 79:1016–1020.

- SHINE, R. 1989. Ecological causes for the evolution of sexual dimorphism: a review of the evidence. *Quarterly Review of Biology* 64:419–461.
- SODERQUIST, T. L. 1995. Spatial organization of the arboreal carnivorous marsupial *Phascogale tapoatafa*. *Journal of Zoology* 237:385–398.
- SOKAL, R. R. & ROHLF, F. J. 1995. Biometry: the principles and practice of statistics in biological research. (Third edition). W. H. Freeman & Co, New York. 887 pp.
- SOUZA, G. S. 1998. Introdução aos modelos de regressão linear e não linear. EMBRAPA/SPI, Brasília. 505 pp.
- STRADIOTTO, A., CAGNACCI, F., DELAHAY, R., TIOLI, S., NIEDER, L. & RIZZOLI, A. 2009. Spatial organization of the yellow-necked mouse: effects of density and resource availability. *Journal of Mammalogy* 90:704–714.
- WECKERLY, F. 1998. Sexual-dimorphism: influence of body mass and mating systems in the most dimorphic mammals. *Journal of Mammalogy* 79:33–52.