

NATURALLY DYED WOOL AND SILK AND THEIR ATOMIC C:N RATIO FOR QUALITY CONTROL OF ^{14}C SAMPLE TREATMENT

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ABSTRACT. Quality control of sample material (e.g. charcoal, collagen) is receiving considerable attention in the effort to obtain more reliable ^{14}C dates. The atomic carbon to nitrogen (C:N) ratio is a useful indicator of contamination and/or degradation of bone collagen. Wool and silk are also composed of proteinaceous material such as bone collagen, and the C:N ratio may also be a useful quality indicator for archaeological wool and silk. Analyses of modern undyed, mordanted, non-mordanted, and naturally dyed silk and wool were done in order to determine a C:N range that indicates the sample quality. The C:N range can be different for every material as the amino acid composition of wool, silk, and bone collagen are distinct. The measured minimum and maximum C:N values were used to set up a C:N range of uncontaminated and undegraded wool and silk. Then, the C:N ratio and ^{14}C were analyzed of archaeological wool and silk samples. The applicability of the C:N ratio as a quality indicator for archaeological silk and wool was shown by the good agreement of the ^{14}C dates with the presumed historical dates for the uncontaminated samples and the disagreement of the ^{14}C dates with the presumed historical dates for contaminated samples.

KEYWORDS: C:N ratio, wool, silk, quality control.

INTRODUCTION

Wool and silk, and textiles in general, are gaining more attention as suitable radiocarbon dating material due to their short lifespan, potentially presenting the true age of an object made of these materials (Rageth 2004; Van Strydonck et al. 2004; van der Plicht et al. 2004; Kim et al. 2008; Mannering et al. 2010; Kuzmin et al. 2012; Van Strydonck and Grömer 2013; Vedeler and Bender Jørgensen 2013; Hajdas et al. 2014).

The quality control of sample material (e.g. charcoal, collagen) is receiving considerable attention in order to obtain more reliable ^{14}C dates (DeNiro 1985; Alon et al. 2002; Van Strydonck et al. 2005; Boudin et al. 2011). Boudin et al. (2011) demonstrated that humic substances are a major contaminant in archaeological wool and silk samples by nondestructive fluorescence spectroscopy analyses. However, degraded wool and silk may contain other contaminants such as mold, fungus, dirt, or other carbon-containing materials (Kim et al. 2008). Therefore, the atomic carbon to nitrogen (C:N) ratio could serve as a quality control indicator of archaeological wool and silk as it does for bone collagen to indicate contamination and/or degradation (DeNiro 1985; Schoeninger et al. 1989; Ambrose 1990). The C:N range can be different for every protein-containing material as the amino acid composition of wool, silk, and collagen are distinct. Wool consists mainly of the protein keratin, which is a α -helix structure with cystine, leucine, glutamic acid, arginine, and serine as the most abundant amino acids (Sibley and Jakes 1984). However, wool proteins are composed mainly of 21 amino acids, with variations in amino acid content due to breed of sheep, diet, climate, and other external influences (Gillespie et al. 1969; Maclaren and Milligan 1981; Leeder and Marshall 1982; Jones et al. 1998). Silk protein (fibroin) is a polypeptide polymer comprising 15 amino acids; the largest percentage (80%) in the fiber consists of the amino acids glycine, alanine, and serine in an approximate 3:2:1 ratio. These substances are small in size, with no large side groups. Therefore, the polypeptide chains can pack together closely as β -sheets (Sibley and Jakes 1984; Becker et al. 1997). For archaeological human and animal hair isotopic analysis, the C:N ratio can be used to indicate if the hair keratin is contaminated or not. Analysis of modern human hair defines a C:N range for uncontaminated archaeological hair between 2.9 and 3.8 (O'Connell

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and Hedges 1999a, 1999b; O'Connell et al. 2001). Taylor et al. (1995) conducted ^{14}C , C:N, and amino acid composition analyses on hair. The glycine/glutamine and glycine/aspartic acid ratios indicate well-preserved chemical structures of the hair. The C:N ratios of the archaeological hair fall within the range of 2.9 and 3.8, determined by modern human hair analyses. The ^{14}C dates of the hair were also in agreement with the archaeological expectations. This suggests that the C:N ratio may be a useful indicator for quality ^{14}C control of human hair and undyed wool (animal hair). However, mordanting and/or dyeing may alter the C:N ratio. Boudin et al. (2013) proposed a C:N range for modern undyed, mordanted, non-mordanted, and naturally dyed silk (all *Bombyx mori*). Based on their analyses of a small data set, the value was found to be between 2.9 and 3.4.

Dyeing is an ancient art, which predates written records. It was practiced during the Bronze Age in Europe. Primitive dyeing techniques included sticking plants to fabric or rubbing crushed pigments into cloth. The methods became more sophisticated with time and techniques were developed using natural dyes from crushed fruits, berries, and other plants, which were boiled into the fabric and gave light and water fastness (resistance). Until the mid-19th century all dyestuffs were made from natural material, mainly vegetable and animal matter. Synthetic organic dyes were introduced in the mid-19th century, with mauveine as the first synthetic organic dye produced in AD 1856 (Herbst and Hunger 1997).

Few natural dyes are color-fast with fibers. Mordants are substances used to fix a dye to the fibers (IUPAC 2006). Different types of mordants are available, including metals in the form of salts (chrome, copper, tin, iron, aluminium), tannins, cream of tartar, baking soda, and vinegar. Different mordants will give different color with the same dye (Aspland 1997).

The classification of natural dyestuffs can be determined on the basis of the following:

- Chemical constitution of the dyestuff molecule. This can be based on the chromophore structure, as with anthraquinones (e.g. madder) or flavonoids (e.g. weld) or the derivatives of one dye (Dedhia 1998; Hofenk de Graaff 2004).
- The application method of the dyestuff. In the case of natural dyestuffs, only three groups are of importance (Gulrajani and Gupta 1992; Hofenk de Graaff 2004):
 1. Dyestuffs that are soluble in water but form metal complexes on the fiber. They can only be applied with the help of water-soluble metal salts (mordants) and are therefore called *mordant dyes*.
 2. Dyestuffs that are insoluble in water and therefore have to be first converted to their water-soluble form before application to the fibers. These are *vat dyestuffs*.
 3. Dyestuffs that dissolve in water and can be applied as such. These are *direct dyestuffs*.

Mordanting and dyeing of wool and silk may alter the C:N ratio because most dyes and some mordants contain carbon and/or nitrogen atoms in their molecular structure, which are added to the textile fiber or fabric. Therefore, in this study, the C:N ratio of modern undyed, mordanted, non-mordanted, and naturally dyed silk (*Bombyx mori*) and wool were measured to determine a C:N range of uncontaminated and undegraded wool and silk. The viability of the established C:N range as a quality indicator for archaeological silk and wool was tested by comparing ^{14}C analysis of archaeological wool and silk with the presumed historical dates.

MATERIALS AND METHODS

Sample Selection

Different modern wool and silk fibers were dyed in the Textile Laboratory of the Royal Institute of Cultural Heritage (RICH), Brussels, Belgium. Some of the most frequently applied dyestuffs as listed in Table 1 (kermes, cochineal, madder, brazilwood, red sandalwood, Persian berries, barberry root, safflower, turmeric, dyer's broom, woad, indigo, orchil, galls) for wool and silk were used, but also less common dyestuffs were applied in this study as listed in Table 2.

Archaeological silk and wool samples from different geographical origins and with different color were selected to study (see Tables 4 and 5).

Design

First, C:N analyses of modern undyed, mordanted, non-mordanted, and naturally dyed silk and wool were conducted in order to determine their C:N range, by using the minimum and maximum value, for uncontaminated and undegraded wool and silk. This study design is based on the papers of DeNiro (1985) and O'Connell and Hedges (1999a) wherein modern bone collagen and hair were analyzed to define a C:N range for uncontaminated bone collagen and hair.

Secondly, the C:N ratio, dye, and ^{14}C analyses were conducted of archaeological wool and silk samples. The viability of the C:N ratio as a quality indicator for archaeological silk and wool was tested by comparing the ^{14}C dates with the presumed historical dates. If the C:N ratio of the bulk did not fall within the proposed C:N range in this study and indicated contamination, cross-flow nanofiltration on the bulk samples was performed to remove contaminants and improve the sample quality by observing a C:N decrease (bulk samples that underwent cross-flow nanofiltration are referred to as permeate). The efficiency of cross-flow nanofiltration to obtain better sample quality and the used protocol are described in Boudin et al. (2013, 2014).

METHODS

Sample Pretreatment

Preliminary tests with the standard pretreatment protocol for archaeological wool and silk were performed on modern wool and silk. No C:N alteration of modern wool and silk was observed. Therefore, the modern undyed and dyed silk and wool samples were not pretreated prior to analysis. The archaeological wool and silk samples (also referred to as bulk samples) were pretreated with hexane, acetone, ethanol, Milli-Q™ water (Merck Millipore, Belgium), 1% NaOH, and 1% HCl for bulk ^{14}C and C:N analyses as described in detail in Boudin et al. (2011).

Atomic C:N Ratio

Analyses were performed in duplicate on a ThermoFlash EA/HT elemental analyzer. The standard used was acetanilide (0.5–1 mg was used).

^{14}C Dating

^{14}C dates were measured on the AMS at the Leibniz Labor für Altersbestimmung und Isotopenforschung in Kiel, Germany (lab code KIA) (Nadeau et al. 1998) or at the Royal

Table 1 The most frequently used natural dyestuffs for wool and silk, the timescale of practice, and its practice location (Hofenk de Graaff 2004).

| | Timescale of dye practice | Dye practice location |
|------------------------------|--|--|
| Red dyestuffs | | |
| <i>Alkanna</i> | Antiquity | Egypt, Mesopotamia, Romans |
| <i>Henna</i> | Antiquity | Palestine, Syria, Egypt |
| <i>Coccid dyestuffs</i> | | |
| Kermes | Since antiquity | Europe, Near East, Asia |
| Polish cochineal | Since antiquity | Europe and Mediterranean countries |
| Armenian cochineal | Since antiquity | Europe, Near East, Far East |
| Cochineal | Since AD 1518 in Europe Since ancient times in Mexico Since antiquity in India and Japan Since AD 1790 in Europe | |
| <i>Madder-type dyestuffs</i> | | |
| Madder | Since antiquity | Worldwide |
| Wild madder | Since antiquity | Mediterranean countries |
| Munjeet | Since antiquity in Far East Since 13th century in Europe | |
| Chay root | Since antiquity | India |
| Galium species | Since ancient times | Worldwide |
| Relbunium species | Pre-Columbian | South and Middle America |
| Indian mulberry | Since antiquity | India, Indonesia, and Polynesian Islands |
| <i>Redwoods</i> | | |
| Brazilwood | Since ancient times in Orient and Far East Since Early Middle Ages in Europe | |
| Red sandalwood | Since ancient times in India Since middle of 16th century AD in Europe | |
| <i>Safflower</i> | Since antiquity in the Orient and southern Asia 2nd century BC in China and Japan 6th century BC in Persia Since 18th century in Europe | |
| Yellow dyestuffs | | |
| <i>Annatto</i> | Pre-Colombian in Peru Since 16th century in Europe | |
| <i>Barberry root</i> | Before AD 1200 | China North and South America Tibet |
| <i>Yellowwoods</i> | | |
| Young fustic | Since antiquity | |
| Fustic | Since 16th century | Europe, Brazil, Argentina, Middle America |
| Quercitron | Since 16th century AD | Europe |
| <i>Persian berries</i> | Since antiquity | Europe, Near East |
| <i>Saffron</i> | Since antiquity | Far and Near East, and on a lesser scale in Europe |
| <i>Turmeric (curcuma)</i> | Since classical times | Polynesia, Micronesia, Melanesia, East Indies, Mesopotamia, Rome, and Greece |
| <i>Weld</i> | Before AD 1492 | Europe |
| <i>Dyer's broom</i> | Since prehistoric times and early Middle Ages | Europe |
| <i>Sawwort</i> | Medieval times | Europe |

| | | |
|------------------------------|---|---|
| Blue dyestuffs | | |
| <i>Logwood</i> | Since 16th century AD | Europe |
| <i>Indigoid dyestuffs</i> | | |
| Woad | Since antiquity | Europe, Egypt, Mesopotamia, Syria and Palestine |
| Indigo | Since antiquity in India Since early 17th century AD in Europe | |
| Indigo carmine | Since AD 1740 | Europe |
| Purple dyestuff | | |
| <i>Tyrian purple</i> | Since antiquity, but production declined after AD 700 in Greece, Persia, and ancient Rome Dyeing continued until AD 1200 in Byzantium and Sicily Produced from <i>Murex</i> shells: provokes reservoir effect | |
| <i>Orchil</i> | Antiquity but its use seems to have been lost in Middle Ages. In AD 1300, orchil was again imported into Europe by a Florentine merchant and more often used in Europe in the 16th century | |
| Black dyestuffs | | |
| <i>Gallotannin dyestuffs</i> | | |
| Galls | Ancient times: worldwide for silk Medieval times western Europe for wool | |
| Alder bark | Since antiquity | worldwide |
| Sumac | Since antiquity | worldwide |
| Catechu | Since antiquity | Far East |
| <i>Black walnut</i> | Since ancient times in the Near East Medieval times in Europe and America | |

Institute for Cultural Heritage, Brussels (lab code RICH) (Boudin et al. 2015). CO₂ was obtained by sample combustion in the presence of CuO and Ag. Graphitization was done with H₂ over a Fe catalyst. Targets were prepared at the Royal Institute for Cultural Heritage in Brussels (Van Strydonck and Van der Borg 1990–1991). ¹⁴C calibrations were performed using OxCal 3 (Bronk Ramsey 1995, 2001) and IntCal13 calibration curve data (Reimer et al. 2013).

Dye Analysis

Each dye analysis requires a sample between 0.2 and 0.5 mg. After pre-examination with a binocular microscope to avoid visible contamination, the dyes were extracted from the yarn using the following extraction method. To the sample of yarn, 250 μL water/methanol/37% hydrochloric acid (1/1/2, v/v/v) is added. The mixture is heated for 10 min at 105°C in open Pyrex™ tubes in a heating block. After cooling, it is filtered through a porous polyethylene frit. The filtrate is dried in an evacuated desiccator over NaOH pellets. The dry residues are taken up in 50 mL of methanol/water (1/1, v/v) and 20 mL of this solution is injected for analysis. The high-performance liquid chromatography (HPLC) equipment consists of a high-pressure pump (Model M615, Waters, USA), a photodiode array detector (Model 996, Waters, USA), and a system for data storage, manipulation, and retrieval (Empower, Waters, USA).

Table 2 Name (species) of dye source and its dye color of uncommon natural dyes used in this study.

| Name (species) of dye source | Dye color |
|--|---|
| Grass (<i>Miscanthus tinctorius</i>) | yellow to light brown |
| Pagoda tree (<i>Sophora japonica</i>) | yellow to olive green |
| Red cedar (<i>Juniperus</i> berries) | brown |
| Pot marygold (<i>Calendula officinalis</i>) | yellow |
| Yellow chamomile (<i>Anthemis tinctoria</i>) | yellow |
| Birch (<i>Betula</i>) | yellow (leaves)-green (leaves)-reddish brown (bark)- yellowish brown (lichen) |
| Chamomile (<i>Chamaemelum nobile</i>) | yellow |
| Elder (<i>Sambucus nigra</i>) | green |
| Black mangrove (<i>Bruguiera gymnorhiza</i>) | red-brown |

Chromatographic conditions were applied using a temperature-controlled (20–22°C) Lichrosorb RP-18 column (4.0 × 125 mm, 5 mm particle size, VWR, Belgium). Three solvents are used: water, methanol, and 5% (w/v) phosphoric acid in water. The elution program is 60A/30B/10C for 3 min, followed by a linear gradient to 10A/80B/10C for 26 min. A flow rate of 1.2 mL/min was used.

Identification of the dye components was done by comparison of the spectral data with the reference spectra in the RICH database (130 references) at the maximum absorbance wavelength of each peak (Wouters 1985; Van den Berghe et al. 2009).

RESULTS AND DISCUSSION

C:N Analyses of Modern Wool and Silk

Comparing the C:N ratios of untreated textiles with mordanted and/or dyed textiles indicates that mordanting and/or dyeing may slightly change the C:N ratio (see Table 3). However, almost all mordanted and/or dyed textiles fall into the C:N ranges (see Table 3):

1. For hair/wool, proposed by O'Connell and colleagues (O'Connell and Hedges 1999a, 1999b; O'Connell et al. 2001): between 2.9 and 3.8;
2. For silk, proposed by Boudin et al. (2013): between 2.9 and 3.4.

This means that the C:N ratio may be a good indicator to control the sample quality of naturally dyed (until AD 1856) archaeological wool and silk for ¹⁴C dating. The minimum value for wool in this study is 3.4 and is in perfect agreement with the theoretical C:N value of keratine. This allows us to refine the C:N range proposed by O'Connell and colleagues (O'Connell and Hedges 1999a, 1999b; O'Connell et al. 2001). The C:N range for uncontaminated wool should be between 3.4 and 3.8. The C:N range proposed by Boudin et al. (2013) for silk is valid for dyed and/or mordanted wool or silk, based on this data set, and can thus serve as an indication if a sample is contaminated or not.

Table 3 Atomic C:N ratio of undyed/dyed/mordanted modern wool and silk. The typical values are 3.0–3.2 for silk and 3.4–3.7 for wool.

| Sample material | Dye and/or mordant | Atomic C:N |
|--------------------------------|---|------------|
| SILK | | |
| <i>Silk yarn I</i> | | |
| | Barberry root: mordanted and unmordanted | 3.2 |
| | Indigo: different dyeing duration | 3.2 |
| | Grass (<i>Miscanthus tinctorius</i>): alum mordant | 3.8 |
| | Pagoda tree (<i>Sophora japonica</i>): alum mordant | 3.3 |
| | Madder | 3.1 |
| <i>Silk yarn J</i> | | |
| | Untreated | 3.1 |
| | Indigo | 3.0 |
| | Brazilwood: alum mordant | 3.0 |
| <i>Silk yarn K</i> | | |
| | Cochineal: alum mordant | 3.0 |
| | Indigo: unmordanted | 3.0 |
| | Mordanted (alum + cream of tartar) | 3.0 |
| | Dyer's broom: alum mordant (with potash) | 3.1 |
| WOOL | | |
| <i>Rowan purelife</i> | | |
| <i>British sheep breeds dk</i> | | |
| | Untreated | 3.4 |
| | Mordanted, undyed | 3.5 |
| | Indigo | 3.5 |
| | Persian berrie (<i>Rhamnus catharticus</i>): mordant: 8% alum + 7% cream of tartar | 3.6 |
| | Kermes: alum mordant | 3.5 |
| | Woad | 3.5 |
| | Red cedar (<i>Juniperus</i> berries): mordant: 8% alum + 7% cream of tartar | 3.6 |
| | Pot marygold (<i>Calendula officinalis</i>): mordant: 8% alum + 7% cream of tartar | 3.6 |
| | Safflower: mordant: 8% alum + 7% cream of tartar | 3.6 |
| | Red sandalwood: mordant: 8% alum + 7% cream of tartar | 3.6 |
| | Indian lac: mordant: 8% alum + 7% cream of tartar | 3.6 |
| | Turmeric: mordant: 8% alum + 7% cream of tartar | 3.6 |
| | Anatto: mordant: 8% alum + 7% cream of tartar | 3.6 |
| | Yellow chamomile (<i>Anthemis tinctoria</i>): mordant: 8% alum + 7% cream of tartar | 3.7 |
| | Birch (<i>Betula</i> sp.): mordant: 8% alum + 7% cream of tartar | 3.7 |
| | Chamomile (<i>Chamaemelum nobile</i>): mordant: 8% alum + 7% cream of tartar | 3.7 |
| | Dyer's broom: mordant: 8% alum + 7% cream of tartar | 3.6 |
| | Elder (<i>Sambucus nigra</i>): mordant: 8% alum + 7% cream of tartar | 3.8 |
| | Galls | 3.7 |
| | Madder | 3.6 |

Table 3 (Continued)

| Sample material | Dye and/or mordant | Atomic C:N | |
|---|--|---|-----|
| | Brazilwood | 3.6 | |
| | Orchil | 3.6 | |
| | Alum mordant medieval dyeing | 3.6 | |
| | <i>Caesalpinia</i> ; mordant: 25% alum, method extraction vinager | 3.6 | |
| | <i>Rubia tinctoria</i> ; mordant: 25% alum or 100% clubmoss | 3.6 | |
| | Cudbear powder, unmordanted | 3.5 | |
| | <i>Bruguiera gymnorhiza</i> , mordanted alum or unmordanted | 3.5 | |
| | <i>Miscanthus tinctorius</i>, mordanted alum or unmordanted | 4.2 | |
| | <i>Sophora japonica</i>, mordanted alum or unmordanted | 4.3 | |
| | Wool A | Untreated | 3.4 |
| | | Washed | 3.4 |
| | | Mordanted | 3.5 |
| | | Green weed no potassy (alum mordanted wool) | 3.5 |
| Dyer's green weed with potassium | | 3.5 | |
| Indigo on wool | | 3.5 | |
| Indigo on wool (recipe I) (no mordanted wool) | | 3.4 | |
| Safflower yellow | | 3.5 | |
| Safflower (red) on wool | | 3.6 | |
| Sappanwood on wool (mordanted) | | 3.5 | |
| Cochineal, alum mordanted | | 3.4 | |
| Gallic acid, mordanted with FeSO₄ and/or CuSO₄ | | 4.1 | |
| <i>Rubia peregrina</i> , dyes extracted from different plants | | 3.6 | |
| <i>Cocheneal</i> , unmordanted, and mordanted with CuSO ₄ or ZnCl ₂ | | 3.6 | |
| Wool B | | <i>Ochrolechia tartarea</i> (lichen, moss) | 3.6 |
| | <i>Umbilicaria pustulata</i> (lichen, moss) | 3.6 | |
| | <i>Parmelia omphalodes</i> (lichen, moss) | 3.7 | |
| | <i>Parmelia saxatilis</i> (lichen, moss) | 3.6 | |
| | <i>Xanthoria parietina</i> (lichen, moss) | 3.7 | |

The only exceptions (marked bold in Table 3) are the textiles dyed with *Miscanthus tinctorius*, *Sophora japonica*, and gallic acid. *Miscanthus tinctorius* and *Sophora japonica* were only used in Eastern Asian textiles (Bechtold and Mussak 2009). *Miscanthus tinctorius* ranges in color from yellow to light brown depending on the mordant, while *Sophora japonica* is yellow to olive green depending on the mordant. Gallic acid was used globally. Although gallic acid was often applied as a mordant, it can also be used as a dye, but very rarely (like in this study). When applied as a dye, it is more concentrated and consequently increases the C:N more than when used as a mordant. The obtained color of the dye gallic acid varies from yellowish brown to dark brown depending on the mordant.

Table 4 Archaeological site, sample color (used dye shown in parentheses if dye analysis was conducted), laboratory code, ¹⁴C ages (BP), calibrated ages (2σ), presumed historical date, and atomic C:N ratio of analyzed archaeological uncontaminated wool samples.

| Sample name | Archaeological site | Sample color | Atomic C:N | Lab code | ¹⁴ C age (BP) | Calibrated age (2σ) | Presumed historical date |
|--------------------|---------------------|------------------|------------|------------|--------------------------|----------------------|--------------------------|
| WOOL | | | | | | | |
| E29479 | Egypt | ochre yellow | 3.7 | KIA-42363 | 1555 ± 30 | AD 420 (95.4%) 580 | AD 400–700 |
| E31963 | Egypt | red | 3.7 | KIA-42370 | 1560 ± 35 | AD 410 (95.4%) 580 | AD 400–700 |
| N1027 | Egypt | yellow-beige | 3.7 | KIA-42369 | 890 ± 30 | AD 1040 (95.4%) 1220 | ? |
| E29477 | Egypt | yellow-beige | 3.7 | KIA-42372 | 1530 ± 30 | AD 430 (95.4%) 600 | AD 400–700 |
| KTN 2298 | Egypt | beige | 3.7 | KIA-47945 | 1510 ± 25 | AD 430 (10.3%) 490 | AD 500–700 |
| | | | | | | AD 530 (85.1%) 620 | |
| Colutus | Egypt | blue (indigoid) | 3.8 | RICH-20058 | 1663 ± 31 | AD 250 (7.8%) 300 | AD 400–500 |
| | | | | | | AD 310 (85.2%) 440 | |
| | | | | | | AD 490 (2.4%) 530 | ? |
| Bouirelet | Egypt | red | 3.7 | KIA-39433 | 1675 ± 30 | AD 250 (11.8%) 300 | ? |
| | | | | | | AD 320 (83.6%) 430 | |
| E32155 | Egypt | yellow | 3.8 | RICH-20309 | 1520 ± 30 | AD 430 (95.4%) 610 | 1st millennium AD |
| E31971 | Egypt | orange | 3.8 | RICH-20305 | 1318 ± 30 | AD 650 (95.4%) 780 | 1st millennium AD |
| E31972 | Egypt | red | 3.8 | RICH-20308 | 1515 ± 30 | AD 430 (95.4%) 620 | 1st millennium AD |
| E29477 bouirelet | Egypt | yellow | 3.7 | KIA-42372 | 1530 ± 30 | AD 420 (95.4%) 580 | ? |
| E31963 bouirelet | Egypt | red | 3.7 | KIA-42370 | 1560 ± 30 | AD 410 (95.4%) 580 | ? |
| KTN 2187 | Egypt | yellow-brown | 3.7 | RICH-20382 | 1562 ± 32 | AD 430 (95.4%) 610 | AD 600–800 |
| KTN 857-03 | Egypt | red-orange | 3.7 | RICH-20377 | 1315 ± 28 | AD 650 (95.4%) 780 | AD 400–700 |
| KTN 614 | Egypt | green-red-yellow | 3.7 | RICH-20380 | 1321 ± 29 | AD 650 (95.4%) 780 | AD 600–800 |
| Hallstatt 79-448 | Austria | yellow-beige | 3.6 | KIA-42753 | 2485 ± 30 | 780 (92.1%) 490 BC | 800–400 BC |
| | | | | | | 470 (3.3%) 410 BC | |
| Hallstatt 07-065 C | Austria | red | 3.8 | KIA-42749 | 2980 ± 30 | 1370 (2.1%) 1340 BC | 1350 BC |
| | | | | | | 1320 (93.3%) 1110 BC | |
| Hallstatt 78-551 | Austria | red-brown | 3.8 | KIA-42754 | 2445 ± 30 | 760 (23.7%) 680 BC | 800–400 BC |
| | | | | | | 670 (10.5%) 610 BC | |
| Hallstatt 79-429 | Austria | yellow-brown | 3.6 | KIA-42752 | 2525 ± 30 | 600 (61.2%) 400 BC | 800–400 BC |
| | | | | | | 800 (29.5%) 720 BC | |
| Hallstatt 79-442 | Austria | blue-black | 3.6 | KIA-42750 | 2555 ± 30 | 700 (65.9%) 530 BC | 800–400 BC |
| | | | | | | 810 (53.5%) 740 BC | |
| | | | | | | 690 (16.4%) 660 BC | |
| | | | | | | 650 (25.5%) 550 BC | |
| Hallstatt 78-552 | Austria | light brown | 3.5 | KIA-42755 | 2380 ± 30 | 720 (2.8%) 690 BC | 800–400 BC |
| | | | | | | 540 (92.6%) 390 BC | |

Table 4 (Continued)

| Sample name | Archaeological site | Sample color | Atomic C:N | Lab code | ¹⁴ C age (BP) | Calibrated age (2σ) | Presumed historical date |
|-----------------------|------------------------------|---|------------|-------------------------|--------------------------|--|---|
| SILK | | | | | | | |
| Zandaniji Maaseik | South Iran | yellow-green (<i>Rexeda luteola</i> + Morus type) | 3.1 | KIA-40826 | 1325 ± 25 | AD 650 (77.5%) 720 | AD 700–900 |
| Davidzijde Tx 606 | ? Iraq | red (Indian lac) red-orange-green | 3.2 3.1 | KIA-40827 KIA-48819 | 1285 ± 25 1245 ± 35 | AD 660 (95.4%) 780 AD 670 (95.4%) 880 | AD 700–800 AD 744–750 (Marwan II) AD 700–900 |
| Tx 372 | East Iran or Central Asia | yellowish brown (ellagic acid) - dark blue (red wood) - beige (indigoid) | 3.1 | KIA-48816 | 1190 ± 35 | AD 710 (5.1%) 750 AD 760 (83.2%) 900 | AD 700–900 |
| Tx 373 Tx 609 | Byzantium Byzantium | brown dark brown-dark green-dark red (mainly tannic acids, kermes, madder, indigoid) | 3.3 3.1 | KIA-48815 KIA-48818 | 1255 ± 30 1085 ± 30 | AD 670 (95.4%) 870 AD 890 (95.4%) 1020 | AD 700–900 AD 700–900 |
| Tx 371 | Byzantium | orange (kermes, madder, traces of tannic acids)- green-white | 3.0 | KIA-48817 | 1195 ± 30 | AD 710 (4.3%) 750 AD 760 (88.8%) 900 AD 920 (2.2%) 940 | AD 700–900 |
| KTN 2400 | Central Asia | brown | 3.2 | RICH-20059 | 1338 ± 31 | AD 640 (80.9%) 720 AD 740 (14.5%) 770 | AD 600–800 |
| KTN 2480 DM1923-01 | Central Asia Egypt | brown beige | 3.1 3.1 | RICH-20059 KIA-42113 | 1288 ± 28 1220 ± 35 | AD 660 (95.4%) 780 AD 680 (95.4%) 890 | AD 600–800 AD 600–900 |

Table 5 Archaeological site, sample color, laboratory code, ¹⁴C ages (BP), calibrated ages (2σ), presumed historical date, and atomic C:N ratio of analyzed archaeological contaminated wool and silk samples before (bulk) and after (permeate) cross-flow nanofiltration.

| Sample name | Archaeological site | Sample color | Atomic | | Lab code | ¹⁴ C age (BP) | Calibrated age (2σ) | Presumed historical date |
|-------------|---------------------|--------------|-----------|-----------|-----------|--------------------------|---------------------|--------------------------|
| | | | C:N ratio | C:N ratio | | | | |
| WOOL | | | | | | | | |
| Beerlegem | Belgium | brown | 3.9 | | KIA-42365 | 1705 ± 30 | AD 250 (95.4%) 410 | AD 587 |
| bulk | | | 3.4 | | KIA-46083 | 1510 ± 25 | AD 430 (10.3%) 490 | |
| permeate | | | | | | | AD 530 (85.1%) 620 | |
| Mainz 2 | Germany | black | 4.0 | | KIA-41535 | 2075 ± 25 | 180 (95.4%) 30 BC | 5 BC |
| bulk | | | 3.5 | | KIA-46193 | 2005 ± 20 | 50 BC (95.4%) AD 55 | |
| permeate | | | | | | | | |
| Mainz 5 | Germany | black | 4.0 | | KIA-37616 | 2080 ± 30 | 200 (94.3%) 20 BC | 5 BC |
| bulk | | | 3.5 | | KIA-47417 | 1915 ± 35 | 10 BC (1.1%) AD | |
| permeate | | | | | | | AD (92.8%) 180 | |
| | | | | | | | AD 190 (2.6%) 220 | |
| SILK | | | | | | | | |
| Baldwin 46 | Belgium | dark brown | 3.6 | | KIA-47415 | 1260 ± 30 | AD 660 (91.9%) 830 | AD 1068 |
| bulk | | | 2.9 | | KIA-47418 | 1175 ± 30 | AD 840 (3.5%) 870 | |
| permeate | | | | | | | AD 770 (83.1%) 900 | |
| | | | | | | | AD 910 (12.3%) 970 | |
| Baldwin 43 | Belgium | dark brown | 3.7 | | KIA-47414 | 1210 ± 30 | AD 690 (12.2%) 750 | AD 1068 |
| bulk | | | 2.9 | | KIA-47811 | 1180 ± 30 | AD 760 (83.2%) 900 | |
| permeate | | | | | | | AD 770 (86.0%) 900 | |
| | | | | | | | AD 910 (9.4%) 970 | |

C:N Ratio, Dye, and ¹⁴C Analyses of Archaeological Wool and Silk

Table 4 lists all the archaeological wool and silk samples from which the C:N ratio falls within the proposed C:N range of 3.4–3.8 for wool and 2.9–3.4 for silk, indicating uncontaminated samples. The ¹⁴C dates of all the samples are in perfect agreement with the presumed historical date, suggesting the samples are not contaminated.

The sample color was visually determined prior to analysis and dye analyses were conducted on some samples. The detected dyes are shown in parentheses next to the determined sample color in Table 4. The analyzed archaeological yellowish and brown silk and wool samples from Egypt, Austria, south Iran, Byzantium, and Central Asia have a C:N ratio lower than the upper boundary, respectively, 3.8 for wool and 3.4 for silk. This excludes exogenous contamination or the use of gallic acid as a dye (see Table 4).

The C:N ratios of the bulk samples (wool and silk) fall outside the proposed C:N ranges, indicating contamination (see Table 5). Moreover, the ¹⁴C ages of samples with untypical C:N ratios do not show a good agreement with the presumed historical date. Cross-flow nanofiltration decreased the C:N ratio and thus improved the sample quality. The obtained C:N ratio of the permeates falls within the proposed C:N ranges of uncontaminated wool and silk, indicating the absence of contaminants in the permeates. The ¹⁴C dates of the cross-flow nanofiltrated wool samples (permeate) are in perfect agreement with the presumed historical dates, suggesting uncontaminated wool samples after cross-flow nanofiltration. The ¹⁴C dates of the permeate of the silk samples (Baldwin 46 and Baldwin 43) are not in agreement with the presumed historical date, but this disagreement is probably due to the reuse of older (parts of) textiles as suggested by Van Strydonck and Bénézet (2014).

The brown color of the contaminated samples Beerlegem, Baldwin 46, and Baldwin 43 are likely due to the presence of humic substances, as shown by nondestructive spectrofluorescence analyses in Boudin et al. (2011, 2014), and also demonstrated by the decrease of C:N ratio of the permeate after cross-flow nanofiltration as well as the good agreement of the ¹⁴C date with the presumed historical date.

CONCLUSION

The C:N ratio is a good indicator for sample quality (contamination) of naturally dyed (until AD 1856) archaeological wool and silk for ¹⁴C dating. The C:N range indicating good (uncontaminated) sample quality is between 2.9 and 3.4 for silk and between 3.4 and 3.8 for wool. If the C:N ratio of the archaeological sample is greater than the upper boundary of the C:N range of wool and silk, respectively, 3.8 and 3.4, then the following may be true:

1. The sample may be dyed with *Miscanthus tinctorius* or with *Sophora japonica* in the case of Eastern Asian textiles or with gallic acid (used worldwide). A dye analysis can reveal if one of these three dyes was applied on the textile. If the dye analysis is positive, the sample is not contaminated and can undergo ¹⁴C analysis.
2. However, if the dye analysis is negative (none of the aforementioned dyes detected), the sample is contaminated and should be rejected for ¹⁴C dating, with the too-high C:N ratio indicating contamination. An option to avoid dye analysis is taking samples for ¹⁴C dating with a different color than the color obtained by applying these three dyes.

REFERENCES

- Alon D, Mintz G, Cohen I, Weiner S, Boaretto E. 2002. The use of Raman spectroscopy to monitor the removal of humic substances from charcoal: quality control for ^{14}C dating of charcoal. *Radiocarbon* 44(1):1–11.
- Ambrose SH. 1990. Preparation and characterization of bone and tooth collagen for isotopic analysis. *Journal of Archaeological Science* 17(4):431–51.
- Aspland JR. 1997. *Textile Dyeing and Coloration*. Durham: American Association of Textile Chemists and Colorists. p 244–5.
- Bechtold T, Mussak R. 2009. *Handbook of Natural Colorants*. Chichester: Wiley & Sons.
- Becker MA, Magoshi Y, Sakai T, Tuross NC. 1997. Chemical and physical properties of old silk fabrics + biochemical analysis of 17 Japanese silk kimono lining fabrics. *Studies in Conservation* 42(1):27–37.
- Boudin M, Boeckx P, Vandenaabeele P, Mitschke S, Van Strydonck M. 2011. Monitoring the presence of humic substances in wool and silk by the use of nondestructive fluorescence spectroscopy: quality control for ^{14}C dating of wool and silk. *Radiocarbon* 53(3):429–42.
- Boudin M, Boeckx P, Vandenaabeele P, Van Strydonck M. 2013. Improved radiocarbon dating for contaminated archaeological bone collagen, silk, wool and hair samples via cross-flow nanofiltrated amino acids. *Rapid Communications for Mass Spectrometry* 27(18):2039–50.
- Boudin M, Boeckx P, Vandenaabeele P, Van Strydonck M. 2014. An archaeological mystery revealed by radiocarbon dating of cross-flow nanofiltrated amino acids derived from bone collagen, silk, and hair: case study of the bishops Baldwin I and Radbod II from Noyon-Tournai. *Radiocarbon* 56(2):603–17.
- Boudin M, Van Strydonck M, van den Brande T, Synal H-A, Wacker L. 2015. A new AMS facility at the Royal Institute for Cultural Heritage, Brussels, Belgium. *Nuclear Instruments and Methods in Physics Research B* 361:120–3.
- Bronk Ramsey C. 1995. Radiocarbon calibration and analysis of stratigraphy: the OxCal program. *Radiocarbon* 37(2):425–30.
- Bronk Ramsey C. 2001. Development of the radiocarbon calibration program. *Radiocarbon* 43(2A):355–63.
- Dedhia EM. 1998. Natural dyes. *Colourage* 45(3):45–9.
- DeNiro MJ. 1985. Postmortem preservation and alteration of *in vivo* bone collagen isotope ratios in relation to palaeodietary reconstruction. *Nature* 317(6040):806–9.
- Gillespie JM, Broad A, Reis PJ. 1969. Further study on the dietary-regulated biosynthesis of high-sulphur wool proteins. *Biochemical Journal* 112(1):41–9.
- Gulrajani ML, Gupta D. 1992. *Natural Dyes and Application to Textiles*. New Delhi: Department of Textile Technology, Indian Institute of Technology.
- Hajdas I, Cristi C, Bonani G, Maurer M. 2014. Textiles and radiocarbon dating. *Radiocarbon* 56(2):637–43.
- Herbst W, Hunger K. 1997. Industrial organic pigments. Production, properties, applications. *Journal of American Institute of Conservation* 45:107–25.
- Hofenk de Graaff J. 2004. *The Colourful Past: Origins, Chemistry and Identification of Natural Dyestuffs*. London: Archetype Publications.
- Holme I. 2006. Sir William Henry Perkin: a review of his life, work and legacy. *Coloration Technology* 122(5):235–51.
- IUPAC. 2006. *Compendium of Chemical Terminology*, 2nd edition (the “Gold Book”). Compiled by A D McNaught and A Wilkinson. Oxford: Blackwell Scientific Publications (1997). XML on-line corrected version: <http://goldbook.iupac.org> (2006-) created by M Nic, J Jirat, B Kosata; updates compiled by A Jenkins. ISBN 0-9678550-9-8. doi:10.1351/goldbook
- Jones LN, Rivett DE, Tucker DJ. 1998. Wool and related mammalian fibres. In: Lewin M, Pearce EM, editors. *Handbook of Fibre Chemistry*. New York: Marcel Dekker. p 356–414.
- Kim K, Southon J, Imamura M, Sparks R. 2008. Development of sample pretreatment of silk for radiocarbon dating. *Radiocarbon* 50(1):131–8.
- Kuzmin Y, Keally C, Jull A, Burr G, Klyuev N. 2012. The earliest surviving textiles in East Asia from Chertovy Vorota Cave, Primorye Province, Russian Far East. *Antiquity* 86(332): 325–37.
- Leeder JD, Marshall RC. 1982. Readily-extracted proteins from merino wool. *Textile Research Journal* 52(4):245–9.
- Ling HT. 2009. Natural dyes in Eastern Asia, Vietnam and neighbouring countries. In: Bechtold T, Mussak R, editors. *Handbook of Natural Colorants*. Chichester: Wiley & Sons. p 65–72.
- Maclaren JA, Milligan B. 1981. *Wool Science - The Chemical Reactivity of the Wool Fibre*. Marrickville: Science Press. p 1–16.
- Mannering U, Possnert G, Heinemeier J, Gleba M. 2010. Dating Danish textiles and skins from bog finds by means of ^{14}C AMS. *Journal of Archaeological Science* 37(2):261–8.
- Nadeau M-J, Grootes PM, Schliecher M, Hasselberg P, Rieck A, Bitterling M. 1998. Sample throughput and data quality at the Leibniz-Labor AMS facility. *Radiocarbon* 40(1):239–45.
- O’Connell TC, Hedges REM. 1999a. Investigations into the effect of diet on modern human hair isotopic values. *American Journal of Physical Anthropology* 108(4):409–25.
- O’Connell TC, Hedges REM. 1999b. Isotopic comparison of hair and bone: archaeological analyses. *Journal of Archaeological Science* 26(6):661–5.

- O'Connell TC, Hedges REM, Healey MA, Simpson AHRW. 2001. Isotopic comparison of hair, nail and bone: modern analyses. *Journal of Archaeological Science* 28(11):1247–55.
- Rageth J. 2004. Radiocarbon dating of textiles. *Orientations* 35(4):57–62.
- Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PG, Bronk Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Hafflidason H, Hajdas I, Hatté C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 and Marine13 radiocarbon age calibration curves 0–50,000 years cal BP. *Radiocarbon* 55(4):1869–87.
- Schoeninger MJ, Moore KM, Murray ML, Kingston JD. 1989. Detection of bone preservation in archaeological and fossil samples. *Applied Geochemistry* 4:281–92.
- Sibley LR, Jakes KA. 1984. Survival of protein fibers in archaeological contexts. *Science and Archaeology* 26:17–27.
- Taylor RE, Hare E, Prior CA, Kirner DL, Wan L, Burky RR. 1995. Radiocarbon dating of biochemically characterized hair. *Radiocarbon* 37(2):319–30.
- Van den Berghe I, Gleba M, Mannering U. 2009. Towards the identification of dyestuffs in Early Iron Age Scandinavian peat bog textiles. *Journal of Archaeological Science* 36(9):1910–21.
- van der Plicht J, van der Sanden WAB, Aerts AT, Streurma HJ. 2004. Dating bog bodies by means of ¹⁴C-AMS. *Journal of Archaeological Science* 31:471–91.
- Van Strydonck Bénazeth D. 2014. Four Coptic textiles from the Louvre collection ¹⁴C redated after 55 years. *Radiocarbon* 56(1):1–5.
- Van Strydonck M, Grömer K. 2013. Analysis reports – ¹⁴C-dating of textiles from the Hallstatt salt mine. In: Grömer K, Kern A, Reschreiter H, Rösler-Mautendorfer H, editors. *Textiles from Hallstatt. Weaving Culture in Bronze and Iron Age Salt Mines. Textilien aus Hallstatt. Gewebte Kultur aus dem bronze- und eisenzeitlichen Salzbergwerk*. Budapest: Archaeolingua 29. p 189–92.
- Van Strydonck M, Van der Borg K. 1990–1991. The construction of a preparation line for AMS-targets at the Royal Institute for Cultural Heritage, Brussels. *Bulletin Koninklijk Instituut voor Kunstpatrimonium* 23:228–34.
- Van Strydonck M, De Moor A, Bénazeth D. 2004. ¹⁴C dating compared to art historical dating of Roman and Coptic textiles from Egypt. *Radiocarbon* 46(1):231–44.
- Van Strydonck M, Boudin M, Eryvnc A. 2005. Humans and myotragus: the issue of sample integrity in radiocarbon dating. In: Alcover JA, Bover P, editors. *Proceedings of the International Symposium “Insular Vertebrate Evolution: The Palaeontological Approach.”* Palma. *Onografies de la Societat d'Història Natural de les Balears* 12:369–76.
- Vedeler M, Bender Jørgensen L. 2013. Out of the Norwegian glaciers: Lendbreen—a tunic from the early first millennium AD. *Antiquity* 87(337): 788–801.
- Wouters J. 1985. High performance liquid chromatography of anthraquinones analysis of plant and insect extracts and dyed textiles. *Studies in Conservation* 30:119–28.