

Review Article

Normal and abnormal pulmonary arteriovenous shunting: occurrence and mechanisms

Julien I.E. Hoffman

Department of Pediatrics, University of California, San Francisco, CA, United States of America

Abstract Severe cyanosis due to pulmonary arteriovenous fistulas occurs often after a bidirectional superior cavopulmonary anastomosis (Glenn operation) and also in some congenital anomalies in which hepatic venous blood bypasses the lungs in the first passage. Relocation of hepatic flow into the lungs usually causes these fistulas to disappear. Similar pulmonary arteriovenous fistulas are observed in hereditary haemorrhagic telangiectasia, and in liver disease (hepatopulmonary syndrome). There is no convincing identification yet of a responsible hepatic factor that produces these lesions. Candidates for such a factor are reviewed, and the possibility of angiotensin or bradykinin contributing to the fistulas is discussed.

Keywords: Cavopulmonary anastomosis; hepatopulmonary syndrome; angiogenesis; hepatic venous flow; pulmonary arteriovenous fistulas

Received: 30 August 2012; Accepted: 4 January 2013; First published online: 5 March 2013

IN SEVERAL DISEASES, A LARGE INCREASE IN VENOUS admixture in the pulmonary venous return causes cyanosis that may be very severe. These diseases include:

1. Congenital heart lesions after partial or complete cavopulmonary anastomosis from Glenn, Kawashima, and Fontan–Kreutzer operations.
2. Rarely in congenital heart disease without previous surgery.
3. Pulmonary arteriovenous fistula without other congenital heart diseases, often associated with hereditary haemorrhagic telangiectasia (Osler–Rendu–Weber syndrome).
4. Certain types of liver disease – the hepatopulmonary syndrome.

An important question is whether these disorders have any causal mechanisms in common.

In this review, the normal venous admixture is described, and then the pathological entities are discussed.

Normal subjects

Intrapulmonary shunting

The oxygen tension is always higher in the alveoli than in the systemic arteries, a difference termed the alveolar–arterial difference. There are several mechanisms for the lower arterial oxygen tension:

1. If any blood perfuses inadequately ventilated alveoli, there will be a ventilation–perfusion mismatch and some blood will not be fully oxygenated by those alveoli. Breathing 100% oxygen abolishes this component of the alveolar–arterial difference.^{1,2}
2. Diffusion limitation causes incomplete oxygenation of the perfused blood. There is no evidence that this can occur normally at rest,^{3,4} but it may occur with extreme exercise⁵ or hepatic cirrhosis.^{6,7}
3. Anatomic connections allow venous blood to bypass the alveoli. These are termed shunts, and they result in venous admixture that is not affected by breathing 100% oxygen.
 - a. Some of these are post-pulmonary capillary shunts of deoxygenated blood from the Thebesian veins that enter the left atrium or ventricle and deep bronchial veins that drain

Correspondence to: Professor J.I.E. Hoffman, BS Hons (Wits), MD (Wits), 925 Tiburon Boulevard, Tiburon, CA 94920-1525. Tel: 415-435-6941; Fax: 415-435-6941; E-mail: julien.hoffman@ucsf.edu

- into pulmonary veins. These shunts are always present, but contribute very little blood flow.
- b. There may be direct pulmonary artery-to-vein connections that bypass pulmonary capillaries, but these are negligible in the normal subject (see below).

Detecting anatomic intrapulmonary shunts

Anatomic shunts may be detected in four ways:

1. By intravenous or pulmonary arterial injection of particles of radio-labelled macroaggregated albumin ($^{99m}\text{TcMAA}$) that are too large to pass through normal lung capillaries. Any radioactive counts in peripheral organs such as the kidney or brain indicate bypass of the alveoli – a patent foramen ovale must be excluded – and shunt proportion can be estimated by comparing the counts in the lungs to the counts in the whole body. The method is very sensitive but depends on the size of the particles.
2. Contrast echocardiography involves injecting intravenously agitated saline that contains small air bubbles and determining whether any are detected in the left atrium, making sure that they do not pass there via a patent foramen ovale. This method is sensitive but does not give absolute shunt quantity.
3. Pulmonary angiography shows either a blush due to excessive numbers of small pulmonary vessels that cannot be distinguished individually or else may show larger tortuous or even aneurysmal connections between the artery and vein. This method is far less sensitive than the two previous methods.^{8–10}
4. By measuring the alveolar–arterial difference while breathing 100% oxygen to eliminate the effects of diffusion and ventilation–perfusion mismatch. This method does not exclude intracardiac right-to-left shunts.

These methods reveal only pulmonary artery-to-pulmonary vein shunts, and do not detect bronchial or Thebesian venous drainage.

Normal intrapulmonary shunts

Normal subjects have a small alveolar–arterial difference of 5–15 mmHg, which is equivalent to

<5% of the cardiac output except for neonates in whom it may be more.¹¹ Most of this venous admixture is due to the ventilation–perfusion mismatch.

Contrast bubble echocardiography: Pulmonary arteriovenous shunts are rare at rest. In one study, small shunts were found in 0/8 adults while erect but in 2/8 of them while supine,¹² and another study from the same laboratory found no shunts at rest in 23 normal subjects.¹³ In both studies, the amount of shunting increased with exercise in parallel with an increased alveolar–arterial difference. All shunting disappeared within 3 minutes of stopping exercise. Another study of 30 young patients after corrective surgery for tetralogy of Fallot showed no shunts at rest.¹⁴ Larsson et al⁹ found no shunting in five normal subjects.

Shunting was observed in 13/13 late-term foetal lambs, 4/4 lambs under 3 days of age, but in 0/7 lambs over 4 weeks of age.¹⁵

In normal dogs, microbubbles did not pass through the lungs unless a vasodilator was used or more than 20 ml of saline was injected.¹⁶

Radiolabelled microspheres: Normal results are given in Table 1.

In dogs, <5% of microspheres 8 μm in diameter passed through the lungs, but occasionally after several circulations even microspheres as large as 50 μm in diameter might pass.²¹ Studies of excised ventilated and perfused lungs of baboons and humans with 25- and 50- μm -diameter microspheres showed <0.2% shunting.²²

Respiratory gas techniques: Vogiatzis et al³ using Fick techniques calculated that normal subjects during heavy exercise while breathing room air had a maximum of 3.5% (standard deviation (SD) 1.2%) venous admixture that decreased to 0.5% (SD 0.5%) when breathing 100% oxygen.

Anatomy of intrapulmonary shunts

Normal neonates may have small pulmonary arteriovenous connections²³ that may be the remnants of the primitive microvascular plexus in the embryo. These connections exist in adult human lungs as well. In 1965, Elliot and Reid²⁴ injected lung vessels and described supernumerary branches of the pulmonary

Table 1. Normal shunting measured by radioactive microspheres.

Publication	Microsphere diameter (μm)	% shunting	Subjects
Whyte et al ¹⁷	7–25	<5.1	Normal
Wolfe et al ¹⁸	20–50	3–6% of cardiac output	Normal
Cloutier et al ¹⁹	10–50	0.3–0.5% of cardiac output	Normal
Kim et al ¹⁴	Unspecified	<11%; mean 6.9%	Tetralogy of Fallot: post-operative
Vettukattil et al ²⁰	8–28	5.4% (SD 2.3%) of cardiac output	Normal or surgically corrected

artery that did not accompany airways, branched at right angles from the parent artery, often had a sphincter-like muscle at the origin, and might not communicate with pulmonary capillaries. Similar results were described by others,^{25,26} and in one of those studies microspheres >50 μm in diameter were found in the pulmonary veins.²⁶

Some studies showed arteriovenous connections subpleurally where alveoli are sparse. It is possible for smaller air bubbles or albumin macroaggregates to pass through the larger capillaries. In rats and dogs, Short et al²⁷ observed capillary diameters ranging up to 13 μm , with more large capillaries subpleurally, and these figures fit well with an average diameter of 10.2 μm in humans found by scanning electron microscopy.²⁸ Such connections might explain some of the microparticle passage through normal lungs, but would certainly not explain the alveolar–arterial difference.

Abnormal venous admixture

Cavopulmonary anastomoses

Glenn anastomosis: The Glenn anastomosis is usually the second stage of a planned Fontan–Kreutzer single-ventricle repair. Initially, the right pulmonary artery was transected and anastomosed end to end to the superior caval vein so that blood returning from the upper body was oxygenated in the right lung; this is the classical or unilateral Glenn operation. Return of partly deoxygenated venous blood from the lower body enters the right atrium. It then either bypasses the lungs to enter the systemic circulation through an atrial or ventricular septal defect – tricuspid or pulmonary atresia – or else divides into two streams, one entering the lungs and another, usually larger, passing directly to the systemic circulation. For many years, however, the superior caval vein has been anastomosed end to side to the top of the pulmonary artery so that flows from the upper body are oxygenated in both lungs; this is the bidirectional Glenn procedure, also known as the bidirectional cavopulmonary anastomosis. Depending on the circumstances, the main pulmonary artery may or may not contribute to pulmonary blood flow. There is, however, a variable contribution to superior caval vein flow from the azygos vein that in normal supine adults averages 86–174 ml/min.^{29–31} In most cavopulmonary anastomoses, the azygos vein is ligated at the time of surgery.

Mathur and Glenn³² described 5/63 patients who developed pulmonary arteriovenous fistulas after a classical unidirectional Glenn procedure, and McFaul et al³³ described four similar patients.

All of these patients had been followed up for many years, and the pulmonary arteriovenous fistulas were confined to the right lung. Trusler et al³⁴ estimated that 11–21% of patients develop pulmonary arteriovenous fistulas after a classical Glenn operation. With the subsequent use of a connection between the superior caval vein and both pulmonary arteries – bidirectional Glenn anastomosis – the fistulas were found in both lungs.

Kopf et al³⁵ in a long-term follow-up of 91 patients aged 2 days to 46 years after a Glenn anastomosis observed pulmonary arteriovenous fistulas in 18 (19.7%) of them. A similar study by Cloutier et al,¹⁹ with an average follow-up of 8.8 years, showed that all 20 patients had increased shunting of radioactive macroaggregates, although only five of these had shunting observed by contrast echocardiography. Feinstein et al⁸ noted angiographically diagnosed fistulas in 66% of lungs sampled. Kim et al¹⁴ found after an average of 22 months that 19/27 patients developed shunts after the bidirectional Glenn procedure, but only 4/27 showed pulmonary arteriovenous fistulas with angiography. In 14 patients studied 0.1–8.6 years after a bidirectional Glenn shunt for univentricular hearts, 10 (71%) had intrapulmonary shunting by contrast echocardiography but only 3 (21%) showed shunting by angiography.¹⁰

Another study observed that 45% of patients had developed shunts after an average of 11.2 years after the operation.⁹ There is an increase in the proportion of patients who develop shunts as the follow-up time increases,^{34,36} although occasionally shunts develop very soon after surgery.

Kawashima operation: In some patients, the intrahepatic portion of the inferior caval vein is absent, because of the failure of the right subcardinal vein to join the hepatic venous system.^{37,38} The venous blood returning from the lower body therefore cannot pass directly to the right atrium, but is diverted through the azygos and hemiazygos veins that develop from the supracardinal veins. These pathways drain into the superior caval vein, so that only the hepatic venous and coronary venous returns pass directly into the right atrium. This venous anomaly is particularly common in patients who have left atrial isomerism/polysplenia syndrome, but it can occur with other congenital cardiac anomalies or even in their absence.^{39–41}

In patients with this anomaly, Kawashima and colleagues connected the superior caval vein to the pulmonary artery^{42,43} and called it the “total cavopulmonary shunt operation”. After this procedure, all systemic venous return except that from the hepatic and coronary veins passes through the superior caval vein to the lungs. Pulmonary

arteriovenous fistulas after the Kawashima operation occur in >50% of patients.^{44–48}

Congenital heart disease without surgery

Rarely, patients with heterotaxy develop pulmonary arteriovenous fistulas spontaneously.^{49,50} There was one patient who had facial telangiectasia and a cerebral arteriovenous malformation but no family history of hereditary haemorrhagic telangiectasia.⁵⁰ Srivastava et al⁴⁶ observed such malformations in only 1/56 patients without surgery, and that patient had biliary atresia. Kawata et al⁵¹ observed pulmonary arteriovenous fistulas in 3/16 patients with left atrial isomerism but in 0/50 with right atrial isomerism.

Miscellaneous forms of congenital heart disease with direct drainage of hepatic veins into the left atrium have also been associated with pulmonary arteriovenous fistulas.^{40,52–57} There are also patients with isolated drainage of the inferior caval vein into the left atrium^{58–64} Some of these patients had pulmonary arteriovenous fistulas,^{40,54,57,59} some did not,^{56,62,63} and in the others fistulas were not assessed.

Pathology of pulmonary arteriovenous fistulas with congenital heart diseases: One patient developed pulmonary arteriovenous fistulas after a Kawashima operation and had thin-walled pulmonary arterioles and dilated small pulmonary veins, as well as some abnormal thin-walled vessels in the interstitium.⁶⁵ Two others had dilated terminal bronchial arteries and thickened veins, as well as clusters of thin-walled subpleural vessels.⁴⁶ One patient with polysplenia but no operation had multiple irregularly shaped vascular channels consistent with pulmonary arteriovenous fistulas.⁴⁹

Another two children had excessive small vessels that were thin walled, often dilated, with disordered collagen fibrils and decreased elastic tissue in their walls.⁶⁶ Some of these vessels appeared to arise from small pulmonary arteries. These abnormal vessels were either dilated (lakes) or were in clusters of small vessels (chains). Some of them appeared to arise from small pulmonary arteries. Immunohistochemistry suggested that the rate of cell proliferation was not increased, and the authors concluded that mechanisms other than vascular proliferation were involved.

In this and a later study,⁶⁷ children after a cavopulmonary anastomosis had increased numbers of small vessels on lung biopsy, even in the absence of clinically or angiographically demonstrated pulmonary arteriovenous fistulas. Further investigation of these abnormal vessels⁶⁸ showed increased staining for vascular endothelial growth factor and

its receptor, and decreased staining for CD31, an indicator of intercellular junctions.

What are these abnormal vessels? We need to distinguish enlargement of pre-existing but inapparent vessels from the budding of new vessels from existing vessels (angiogenesis), and this in turn from vasculogenesis in which angioblasts produce new blood vessels where previously there were none. Vasculogenesis is unlikely because it occurs only with embryonic development and with tumours in tissues devoid of blood vessels. There is evidence for both the other possibilities, and not enough study has been done to determine which predominates.

Isolated pulmonary arteriovenous fistulas

Isolated pulmonary arteriovenous fistulas in the absence of congenital heart disease are uncommon, and are characterised by thin-walled channels lined by endothelium and supported by scanty connective tissue stroma. The fistulas may resemble a plexiform mass of dilated but still small vessels, a tortuous communication between an artery and a vein, or a similar communication with a dilated aneurysmal sac.⁶⁹ Rarely, some of these fistulas are present at birth, and are thus truly congenital, but others are detected later and might or might not have been present at birth. There is a strong but not universal association with hereditary haemorrhagic telangiectasia.

Hepatopulmonary syndrome

The hepatopulmonary syndrome is defined as “an arterial oxygenation defect induced by intrapulmonary vascular dilatations (IPVD) associated with hepatic disease”.^{70,71} It is most often due to liver cirrhosis of any cause, with about 15–30% of cirrhotics having the syndrome,^{70,72,73} but any form of acute or chronic liver disease has been associated with hepatopulmonary syndrome. There is a closer association with portal hypertension than with decreased liver function. The lungs contain dilated pre- and post-capillary vessels,⁷⁴ as well as markedly dilated capillaries,^{6,7} so that there is a substantial diffusion defect.⁷⁰ Pulmonary arteriovenous fistulas and ventilation–perfusion mismatch also contribute to the increased venous admixture.^{74,75} There are also porto-pulmonary anastomoses secondary to the portal hypertension that occurs with most patients with hepatopulmonary syndrome, but these contribute little to the venous admixture.⁷⁶ Recently, a role for CD68(+) macrophages in the lung as a source of vascular endothelial growth factor and platelet-derived growth factor has been described.^{77,78} These macrophages were found in a cirrhotic model produced by chronic bile duct ligation in rats, and macrophage

depletion was shown to prevent or regress the hepatopulmonary syndrome. Similar macrophages were found in autopsies of cirrhotic patients.

Hepatopulmonary syndrome occurs in Budd–Chiari syndrome in which hepatic vein outflow is blocked and the liver is congested; 17/29 patients in one series had some degree of hepatopulmonary syndrome.⁷⁹ The syndrome can occur also with the Abernethy syndrome, in which either the portal vein is absent as a result of which mesenteric blood drains directly into the inferior caval vein or else there is a connection between the portal vein and the inferior caval vein.^{80,81} The liver may be normal.

Mechanisms causing pulmonary arteriovenous fistulas: When fistulas were reported after Glenn procedures, a proposed mechanism was the decreased pulsatility in the vessels of the affected lung(s). This hypothesis became untenable when these fistulas did not develop as frequently or even regressed after the complete Fontan–Kreutzer procedure.^{46,82} Some investigators reported the development of new pulmonary arteriovenous fistulas after the Fontan–Kreutzer operation,^{9,83,84} but in many of these patients new fistula development was associated with maldistribution of hepatic venous drainage into the pulmonary arteries, often with disappearance of the fistulas when the flow maldistribution was corrected.^{85–89}

The hepatic venous drainage was further considered a factor in those patients with congenital drainage of the hepatic veins or inferior caval vein into the left atrium who developed pulmonary arteriovenous fistulas that regressed after restoration of hepatic venous return directly to the pulmonary circulation. Furthermore, in cavopulmonary anastomoses, the hepatic venous drainage reaches the lungs only after passing through the systemic circulation, and re-routing hepatic venous drainage to the pulmonary arteries may cause the venous admixture to decrease and even disappear.^{44,52,82,90–99} Similar changes have been seen after correction of isolated inferior caval vein or hepatic venous drainage into the left atrium.^{53,54,100}

Dilution or loss of substance?: Owing to the fact that in cavopulmonary anastomosis hepatic venous blood does not pass through the right heart into the lungs, there are two possible mechanisms as causes of the resulting pulmonary arteriovenous fistulas. Either the abnormal pathway for hepatic venous blood dilutes some agonist or else the agonist has a very short half-life. In the normal circulation, about 20–25% of the cardiac output returns to the heart through the hepatic veins,^{101,102} so that in the right heart hepatic venous blood joins the remainder of the venous return and is diluted four- to fivefold by the time it reaches the lungs. After a Kawashima

operation, the hepatic venous blood mixes with the remaining venous return in the left heart, and is diluted to the same extent. After a bilateral Glenn operation in which no blood flows through the main pulmonary artery, the hepatic venous blood mixes with the venous return through the inferior caval vein and enters the left heart where it mixes with the blood that has returned through the superior caval vein; the resulting dilution is exactly the same. With both of these shunts, some hepatic venous blood that enters the aorta returns to the lungs via the superior caval vein. The dilution is the same as normal, but only about half the amount of a hepatic factor reaches the lungs in its first passage. This raises the possibility that less of a potential anti-angiogenic factor reaches the lungs in the first passage. However, some first-pass mechanism must be involved, because any long-lasting hepatic factor will eventually reach the lungs with a normal degree of dilution. Therefore, either there is a labile, short-lived hepatic factor that prevents vasodilatation and angiogenesis or else in passage through the systemic circulation angiogenic or vasodilator factors develop and reach the lungs. Both mechanisms might act synergistically in some patients.

Any explanation for the fistulas needs to account for the apparently greater incidence after the Kawashima operation than the conventional bidirectional Glenn operation, and why some patients do not develop pulmonary arteriovenous fistulas with these operations.

Possible mechanisms: Many parts of the body – for example, red and white blood cells, intestinal epithelium, hair root cells, cellular organelles, bone spicules, and myosin – exist in dynamic equilibrium. After some typical time period, the entity breaks down and is replaced. Owing to the fact that the amount of each entity normally remains relatively constant, there must be some feedback system that regulates its quantity. Although such equilibrium has not been studied for pulmonary blood vessels, it is conceivable that the endothelial cells with their exposure to physical forces and chemical activity undergo similar changes. Another type of equilibrium occurs when agonist and antagonist systems maintain the status quo until some trigger mechanism creates an imbalance in the equilibrium; an example of this is the clotting cascade.

A clue to the dynamic factors involved in the pulmonary blood vessels may be found in hereditary haemorrhagic telangiectasia. Several genetic mutations have been associated with hereditary haemorrhagic telangiectasia, the two most prominent involving the endoglin gene^{103,104} (~60%) and the activin receptor-like kinase 1 gene (~30%)¹⁰⁵ A rarer mutation (~4%) occurs in Smad 4.

These genes mediate signalling by transforming growth factor- β ligands^{106,107} (Fig 1).

There are important interactions among one of the main components of the transforming growth factor- β superfamily, bone morphogenetic protein-9, a circulating protein produced mainly in the liver,¹⁰⁸ activin receptor-like kinase 1, and Smads. Smads are a large family of small messenger proteins that form heterodimers with co-Smad 4 and carry signals from members of the transforming growth factor- β superfamily to the nucleus where they interact with oncogenes, kinases, and other factors to promote or inhibit expression of genes. The affected genes

depend on the Smad, cell type, and on the rest of the cell environment.¹⁰⁹

In a schema described by Lebrin et al,¹⁰⁷ transforming growth factor β acts via transforming growth factor- β receptor II to activate both angiogenic and anti-angiogenic pathways. The angiogenic system includes the activin receptor-like kinase 5-Smad 2/3 pathway that stimulates endothelial cell proliferation and migration. The anti-angiogenic system includes the activin receptor-like kinase 1-Smad 1/5/8 pathway that inhibits endothelial cell proliferation and migration. Endoglin is an accessory transforming growth factor- β receptor that is essential for activin receptor-like kinase 1 signalling and indirectly inhibits activin receptor-like kinase 5 signalling.

The steps in angiogenesis are beginning to be elucidated (Fig 2).

The angiogenic process begins with vasodilatation in which nitrous oxide plays a role.^{112–114} Angiopoietin-2 and vascular endothelial growth factor produce fenestrations, increase vascular permeability, and allow plasma proteins to extravasate and, with the aid of cadherins and integrins, form a scaffold for endothelial cell migration. This process is aided by membrane breakdown by activated matrix metalloproteinases. The cords of cells acquire a lumen, and then angiopoietin-1 tightens cell-cell contacts and transforming growth factor- β inhibits endothelial cell proliferation and migration. Peri-endothelial cells are recruited by platelet-derived growth factor-BB to form smooth muscle cells, and a new basement membrane forms. These processes are held in check by anti-angiogenic factors such as some members of the vascular endothelial growth factor superfamily, thrombospondin-1, and

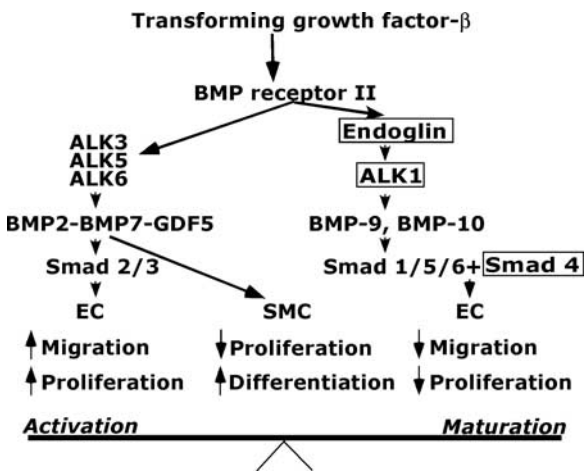


Figure 1. Basic angiogenesis balance. Concepts based on data published by Lebrin et al,¹⁰⁷ David et al,¹⁰⁸ Scharpfenecker et al,¹⁰⁹ and Carmeliet.¹¹² EC = endothelial cells; GDF5 = growth differentiation factor; SMC = smooth muscle cells; ALK = activin receptor-like kinase. The boxes indicate sites of known mutation.

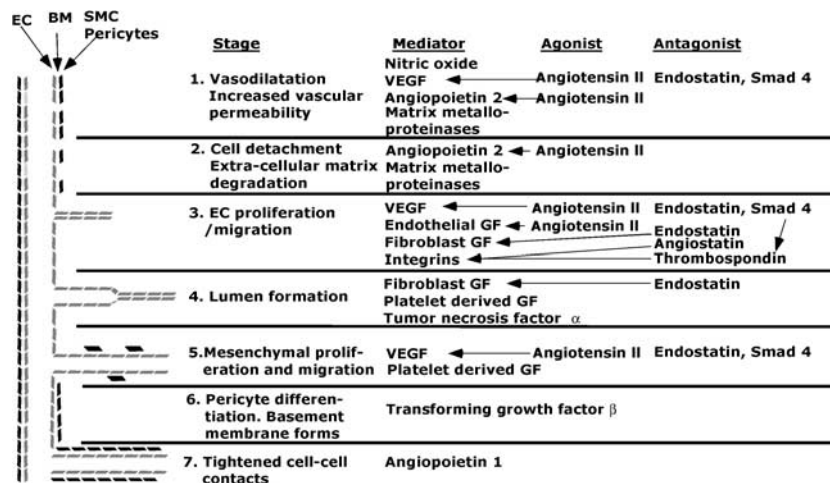


Figure 2. Diagram of stages and some major mediators, agonists and antagonists in angiogenesis. EC = endothelial cells; SMC = smooth muscle cells; BM = basement membrane; VEGF = vascular endothelial growth factor; GF = growth factor. The lines indicate which agonists or antagonists affect a give mediator. In stage 3, Smad 4 increases thrombospondin activity, and thereby increases inhibition of integrins.

endostatin and angiostatin. Undoubtedly, many other angiogenic and antiangiogenic factors exist and are deployed in various circumstances.^{110,111}

Given this array of factors, we should look for one or more factors with angiogenic abilities that reach the lungs in high concentrations only if first-pass hepatic venous drainage bypasses the lungs.

Endoglin per se is not likely to be responsible for the abnormal angiogenesis found in cavopulmonary anastomoses because it is produced widely by endothelial cells. The various isoforms of vascular endothelial growth factor are produced by many types of endothelial cells, and their production is increased by the hypoxia-inducible factor HIF. Their concentrations are increased in children with cyanotic heart disease,¹¹¹ and there is usually an inverse relation between vascular endothelial growth factor concentration and oxygen saturation.^{112–118} In one study of cyanotic children, not only did vascular endothelial growth factor and basic fibroblast growth factor have higher concentrations in cyanotic than acyanotic children, but in the cyanotic children vascular endothelial growth factor concentrations were considerably higher in the superior caval vein (mean 136 pg/ml) than the inferior caval vein (mean ~75 pg/ml).¹¹⁷ In all, 6/22 of these cyanotic patients had had a bidirectional Glenn operation, but unfortunately their results were not separated from the others. Nevertheless, these findings suggest an added layer of complexity in understanding the mechanisms underlying these pulmonary arteriovenous fistulas.

The angiopoietins, like the vascular endothelial growth factor family, are glycoproteins. Angiopoietin-1 is produced by mesenchymal cells, particularly smooth muscle cells. Angiopoietin-2 is produced by endothelial cells in many organs.

Liver factors that might affect angiogenesis include endostatin and angiostatin, both inhibitors of angiogenesis, and angiotensin that can stimulate angiogenesis. Therefore, a deficiency of the first two agents or a surplus of the third might play a role in the formation of pulmonary arteriovenous fistulas in liver disease or diversion of hepatic blood from first passage through the lung.

Endostatin is the C-terminal fragment of collagen XVIII, a key constituent of basement membrane.^{119,120} Collagen XVIII has two isoforms – a short form with 303 residues produced by stellate and endothelial cells in the liver that is attached to the basement membrane, and a long form with 493 residues that is produced by hepatocytes and circulates as a plasma protein.^{121,122} Endostatin is produced when the tumour suppressor p53 upregulates the activity of a (II) collagen prolyl-4-hydroxylase^{123,124} Although it meets some of the requirements

as a causative factor in pulmonary arteriovenous fistulas, endostatin seems to have its major effect on pathologically induced angiogenesis, as in cancer,¹²⁵ and has a half-life of about 1–2 hours.^{126,127} Therefore, unless there is some as yet unknown factor associated with the first pass of collagen XVIII through lung, endostatin may not be a primary factor in the formation of these fistulas. Recently, Field-Ridley et al¹²⁸ observed that after a Glenn procedure in patients endostatin levels decreased from 4.42 to 3.34 ng/ml, whereas collagen XVIII levels were doubled. The findings are of considerable interest, but a relationship to first-pass lung metabolism of collagen XVIII is yet to be demonstrated. p53 activity has been observed in human pulmonary macrovascular endothelial cells.¹²⁹ It is of interest that collagen XVIII gene expression is upregulated after cavopulmonary anastomosis in the rat.¹³⁰

Angiostatin is formed by proteolytic cleavage of plasminogen secreted by the liver.¹³¹ It is one possible missing liver factor, because it is present in hepatic venous blood and has a half-life of about 15 minutes.¹³² This, however, might be too long to involve a first-pass mechanism.

An agent that meets some of the requirements is angiotensin. Angiotensinogen, an α -2-globulin formed mainly in the liver, is cleaved by renin to form angiotensin I, a 10-amino-acid peptide with little biological activity. This in turn is cleaved to form the 8-amino-acid peptide angiotensin II by an exopeptidase – angiotensin-converting enzyme – secreted by pulmonary and renal vascular endothelial cells. Angiotensin II is known to induce angiopoietin and vascular endothelial growth factor,¹³³ and probably stimulates angiogenesis through the angiotensin-2 receptor.¹³⁴ Conversely, stimulating the angiotensin-1 receptor decreases angiogenesis. Bone marrow-derived endothelial cells might also be involved.¹³⁵ Angiotensin II is also known to stimulate the production of nitric oxide,¹³⁶ possibly via the angiotensin-2 receptors.¹³⁴ A similar peptidase termed angiotensin-converting enzyme-2 converts angiotensin II into a heptapeptide angiotensin III that acts via the *mas* receptor to antagonise almost all the effects of angiotensin II.^{137,138} Therefore, the resultant effect on angiogenesis in the lung will depend in part on the balance of these two angiotensins.

Angiotensin II has a short half-life in the circulation, about 16–30 seconds.¹³⁹ Therefore, one possible scenario is that with normal hepatic venous drainage into the lung, the angiotensin II that is produced has been partly metabolised to angiotensin III by the time it has passed through the systemic circulation and returned to the lung (Fig 3). Although the difference in time between

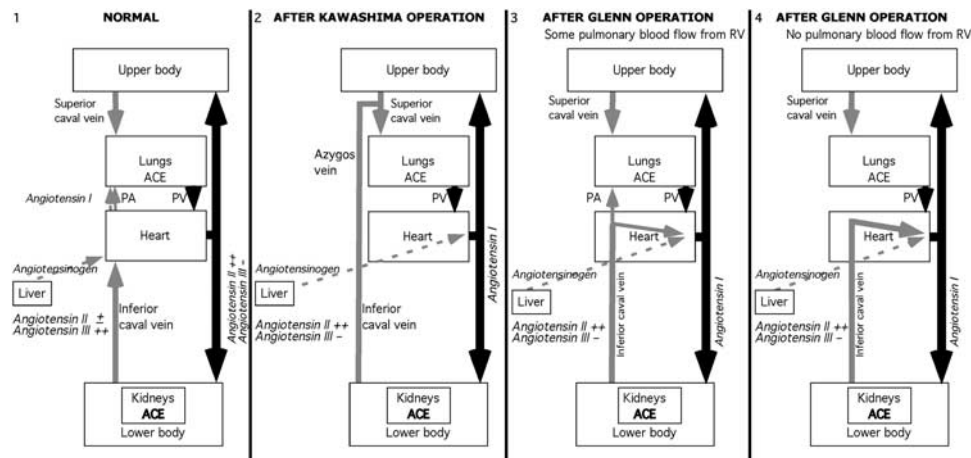


Figure 3.

Schema of possible role of angiotensin II in angiogenesis. Panel 1: Normal circulation. Hepatic and lower body venous blood join in the high inferior caval vein to reach the lung. Angiotensin II is produced as blood passes through the lung, but little of it reaches the lung. Panel 2: After a Kawashima operation, angiotensin II is produced in the kidneys and returns rapidly to the lungs via the inferior caval and azygos veins. Panel 3: After a Glenn operation, if some pulmonary blood flow reaches the lungs, then the lungs are exposed to a high concentration of angiotensin II, but in lesser amounts than after a Kawashima operation. Panel 4. If all pulmonary flow is from the superior caval vein, then the concentration of angiotensin II reaching the lung is much lower because it has to circulate first through the upper body. Black arrows = arterial blood. Gray solid arrows = caval venous blood. Gray dashed arrows = hepatic venous blood.

the appearance of angiotensin II in the aorta and in the renal vein is small, not only is there a small decrease in angiotensin II concentration by the time it reaches the kidney, but there is a substantial uptake of angiotensin II by the kidney that further reduces the concentration of lung-derived angiotensin II in the renal vein.^{140–142}

If hepatic venous blood is diverted to the systemic circulation, then angiotensin II is produced as it passes through the renal circulation (Fig 3). After a Kawashima operation, renal venous blood returns rapidly to the lung via the azygos and hemiazygos venous connections to the superior caval vein, so that angiotensin II is present in greater than normal concentrations and angiotensin III in lower concentrations when it reaches the lung. A similar short return pathway from the kidneys occurs after a Glenn operation in which some flow reaches the lung through the main pulmonary artery, as may occur with tricuspid atresia and miscellaneous anomalies including some double-inlet left ventricles and complex anomalies. On the other hand, the azygos vein is usually ligated during the Glenn operation. Therefore, if all pulmonary blood flow after the Glenn operation comes through the superior vena cava, as in pulmonary atresia, hypoplastic left heart syndrome, and many other complex anomalies, then most of the renal venous return reaches the lungs only after returning to the left ventricle, and circulating once more around the upper body before reaching the superior caval vein (Fig 3). In these latter patients, angiotensin has

been further metabolised and so is less important as an angiogenic stimulus. Therefore, although angiotensin II is a good candidate for angiogenesis after a Kawashima operation, it may be a poorer candidate after most traditional Glenn operations. However, because angiogenesis is a delicate balance between several angiogenic and anti-angiogenic factors, the balance might easily be upset in more than one way. No matter what the venous return pathway to the lungs is, there will be more angiotensin II and less angiotensin III when hepatic venous blood bypasses the lung in its first circulation.

In one study of children after a bidirectional Glenn operation, those with no other source of pulmonary blood flow had an intrapulmonary shunt averaging 34.9% of cardiac output (SD 15.8%), whereas those with additional flow through the main pulmonary artery had an average shunt of 12.0% (SD 2.6%).²⁰ One possible explanation of these findings is that the reduced amount of hepatic blood flow entering the pulmonary artery from the right ventricle contained a reduced amount of a short-lived anti-angiogenic factor, whereas hepatic blood reaching the pulmonary arteries only through the caval anastomosis had even less of this factor. To date, such a short-lived anti-angiogenic factor has not yet been identified. One candidate for this factor might be the incompletely defined anti-angiogenic protein isolated by Marshall et al¹⁴³ who showed that media from cultured rat hepatocytes inhibited the growth of bovine capillary endothelium but not epithelial or tumour cells. The half-life of this

protein is unknown. Another candidate is angiotensin III, which also has a short half-life.

The role of angiotensin needs further study. Angiotensin-1 receptors are in greater numbers than angiotensin-2 receptors, and angiotensin-1 receptor activation has variously been reported to promote and inhibit angiogenesis. For example, Amaral et al^{144,145} found that exercise induced angiogenesis in rat limb muscle, and that this action could be blocked by an angiotensin-1 receptor blocker. On the other hand, Walther et al¹³⁴ did experiments that indicated that the angiotensin-2 receptors but not the angiotensin-1 receptors were involved in angiogenesis. Of more importance is the fact that tissues have a local renin-angiotensin system that might act independently of any circulating agonists.^{146,147} Nevertheless, there seems enough reason to investigate whether angiotensin is one of the factors promoting angiogenesis in these diseases.

The fact that angiotensin is involved in some way was demonstrated by the finding that after creating a unilateral cavopulmonary anastomosis in sheep the shunted lung had decreases in angiotensin-converting enzyme and angiotensin II, and increases in angiotensin-2 receptors, but these levels returned to normal after 15 weeks. However, angiotensin-1 receptors were persistently upregulated.^{148,149}

Bradykinin is an angiogenic factor (among its many actions) that is produced by the action of plasma kallikrein on high molecular weight kininogen that is produced by the liver. It is normally metabolized completely as it passes through the lung into an inert component and a long-acting anti-angiogenic form of high molecular weight kininogen. If bradykinin does not pass through the lung it circulate with a half life of about 30 seconds, possibly long enough to exert an angiogenic effect in the lung.

The role of liver factors has been invoked not only because of the effects of diverting hepatic blood from or returning it to the pulmonary artery, but also because of similarities to the hepatopulmonary syndrome. The comparison should not be pushed too far because of substantial differences between the various disorders. In hepatopulmonary syndrome, for example, the predominant pathology in the lung consists of dilated capillaries that are not a feature of other forms of pulmonary arteriovenous fistulas. The role of macrophages has also not been described other than in the hepatopulmonary syndrome, although their discovery is recent enough that they have not been looked for in other forms of pulmonary arteriovenous fistulas. Then, although in the cardiac forms of pulmonary arteriovenous fistulas there is clear evidence of a first-pass mechanism, in liver disease there must be a constant

overproduction of an angiogenic factor or failure of production of an anti-angiogenic factor. There is no need to postulate a labile factor. Angiogenic and vasodilator factors abound in cirrhosis of the liver. There is evidence of overproduction of nitric oxide in cirrhosis. One suggested mechanism for the pathology is stimulation of macrophage production of inducible nitric oxide synthase by tumour necrosis factor- α , and inhibition of tumour necrosis factor- α by pentoxifylline^{150,151} decreases nitric oxide production and ameliorates the hepatopulmonary syndrome. Furthermore, patients with cirrhotic livers have increased circulating endotoxin¹⁵² that is also vasodilator and angiogenic.¹⁵³ Finally, other characteristics of cirrhosis of the liver such as spider naevi and systemic peripheral vasodilatation are not a feature of other forms of pulmonary arteriovenous fistulas.

Conclusion

The role of hepatic factors in stimulating or inhibiting angiogenesis in the lung appears to require involvement of first-pass metabolism of hepatic venous blood through the lungs. Diversion of hepatic blood from pulmonary capillaries during the first pass implies that there is a short-lived factor that is either anti-angiogenic that bypasses the lungs or else a short-lived pro-angiogenic factor that fails to break down before reaching the lungs in the second passage. Both theories are ripe for further exploration. The fact that in liver disease with the hepatopulmonary syndrome there are also pulmonary arteriovenous fistulas may not be helpful in indicating the cause of pulmonary arteriovenous fistulas after a cavopulmonary anastomosis.

The mechanisms described above do not explain the variability in the timing or extent of pulmonary arteriovenous fistulas in different patients. Whether this is due to variations in the baseline concentrations of various factors involved in angiogenesis, such as vascular endothelial growth factor, or underlying genetic mechanisms is unknown.

Acknowledgement

The author thanks Dr Jeffrey Fineman for allowing him to see his unpublished manuscript, and Dr David Teitel for suggestions about mechanisms.

References

1. Rahn H, Fenn WO. A Graphical Analysis of the Respiratory Gas Exchange. American Physiological Society, Washington, DC, 1955.
2. Riley RL, Cournand A. Ideal alveolar air and the analysis of ventilation-perfusion relationships in the lungs. *J Appl Physiol* 1949; 1: 825-847.

3. Vogiatzis I, Zakynthinos S, Boushel R, et al. The contribution of intrapulmonary shunts to the alveolar-to-arterial oxygen difference during exercise is very small. *J Physiol* 2008; 586: 2381–2391.
4. Wagner PD, Gale GE, Moon RE, Torre-Bueno JR, Stolp BW, Saltzman HA. Pulmonary gas exchange in humans exercising at sea level and simulated altitude. *J Appl Physiol* 1986; 61: 260–270.
5. Hammond MD, Gale GE, Kapitan KS, Ries A, Wagner PD. Pulmonary gas exchange in humans during exercise at sea level. *J Appl Physiol* 1986; 60: 1590–1598.
6. Davis HH II, Schwartz DJ, Lefrak SS, Susman N, Schainker BA. Alveolar-capillary oxygen disequilibrium in hepatic cirrhosis. *Chest* 1978; 73: 507–511.
7. Schraufnagel DE, Kay JM. Structural and pathologic changes in the lung vasculature in chronic liver disease. *Clin Chest Med* 1996; 17: 1–15.
8. Feinstein JA, Moore P, Rosenthal DN, Puchalski M, Brook MM. Comparison of contrast echocardiography versus cardiac catheterization for detection of pulmonary arteriovenous malformations. *Am J Cardiol* 2002; 89: 281–285.
9. Larsson ES, Solymar L, Eriksson BO, de Wahl Granelli A, Mellander M. Bubble contrast echocardiography in detecting pulmonary arteriovenous malformations after modified Fontan operations. *Cardiol Young* 2001; 11: 505–511.
10. Chang RK, Alejos JC, Atkinson D, et al. Bubble contrast echocardiography in detecting pulmonary arteriovenous shunting in children with univentricular heart after cavopulmonary anastomosis. *J Am Coll Cardiol* 1999; 33: 2052–2058.
11. Nelson NM, Prod'Hom LS, Cherry RB, Lipsitz PJ, Smith CA. Pulmonary function in the newborn infant: the alveolar-arterial oxygen gradient. *J Appl Physiol* 1963; 18: 534–538.
12. Stickland MK, Welsh RC, Haykowsky MJ, et al. Intrapulmonary shunt and pulmonary gas exchange during exercise in humans. *J Physiol* 2004; 561: 321–329.
13. Eldridge MW, Dempsey JA, Haverkamp HC, Lovering AT, Hokanson JS. Exercise-induced intrapulmonary arteriovenous shunting in healthy humans. *J Appl Physiol* 2004; 97: 797–805.
14. Kim SJ, Bae EJ, Cho DJ, et al. Development of pulmonary arteriovenous fistulas after bidirectional cavopulmonary shunt. *Ann Thorac Surg* 2000; 70: 1918–1922.
15. McMullan DM, Hanley FL, Cohen GA, Portman MA, Riemer RK. Pulmonary arteriovenous shunting in the normal fetal lung. *J Am Coll Cardiol* 2004; 44: 1497–1500.
16. Butler BD, Hills BA. The lung as a filter for microbubbles. *J Appl Physiol* 1979; 47: 537–543.
17. Whyte MK, Peters AM, Hughes JM, et al. Quantification of right to left shunt at rest and during exercise in patients with pulmonary arteriovenous malformations. *Thorax* 1992; 47: 790–796.
18. Wolfe JD, Tashkin DP, Holly FE, Brachman MB, Genovesi MG. Hypoxemia of cirrhosis: detection of abnormal small pulmonary vascular channels by a quantitative radionuclide method. *Am J Med* 1977; 63: 746–754.
19. Cloutier A, Ash JM, Smallhorn JF, et al. Abnormal distribution of pulmonary blood flow after the Glenn shunt or Fontan procedure: risk of development of arteriovenous fistulae. *Circulation* 1985; 72: 471–479.
20. Vettukattil JJ, Slavik Z, Lamb RK, et al. Intrapulmonary arteriovenous shunting may be a universal phenomenon in patients with the superior cavopulmonary anastomosis: a radionuclide study. *Heart* 2000; 83: 425–428.
21. Ring GC, Blum AS, Kurbatov T, Moss WG, Smith W. Size of microspheres passing through pulmonary circuit in the dog. *Am J Physiol* 1961; 200: 1191–1196.
22. Lovering AT, Stickland MK, Kelso AJ, Eldridge MW. Direct demonstration of 25- and 50-microm arteriovenous pathways in healthy human and baboon lungs. *Am J Physiol Heart Circ Physiol* 2007; 292: H1777–H1781.
23. Groniowski J. Morphological investigations on pulmonary circulation in the neonatal period. *Am J Dis Child* 1960; 99: 516–523.
24. Elliott FM, Reid L. Some new facts about the pulmonary artery and its branching pattern. *Clin Radiol* 1965; 16: 193–198.
25. Tobin CE. Arteriovenous shunts in the peripheral pulmonary circulation in the human lung. *Thorax* 1966; 21: 197–204.
26. Wilkinson MJ, Fagan DG. Postmortem demonstration of intrapulmonary arteriovenous shunting. *Arch Dis Child* 1990; 65: 435–437.
27. Short AC, Montoya ML, Gebb SA, Presson RG Jr, Wagner WW Jr, Capen RL. Pulmonary capillary diameters and recruitment characteristics in subpleural and interior networks. *J Appl Physiol* 1996; 80: 1568–1573.
28. Miura A, Nakamura K, Matsubara H, et al. Differences of diameter of pulmonary capillary vessels in patients with pulmonary hypertension using scanning electron microscope. *Circulation* 2011; 122: Abstract 21460.
29. Lomas DJ, Hayball MP, Jones DP, Sims C, Allison ME, Alexander GJ. Non-invasive measurement of azygos venous blood flow using magnetic resonance. *J Hepatol* 1995; 22: 399–403.
30. Debatin JF, Zahner B, Meyenberger C, et al. Azygos blood flow: phase contrast quantitation in volunteers and patients with portal hypertension pre- and postintrahepatic shunt placement. *Hepatology* 1996; 24: 1109–1115.
31. Bosch J, Mastai R, Kravetz D, Bruix J, Rigau J, Rodes J. Measurement of azygos venous blood flow in the evaluation of portal hypertension in patients with cirrhosis. Clinical and haemodynamic correlations in 100 patients. *J Hepatol* 1985; 1: 125–139.
32. Mathur M, Glenn WW. Long-term evaluation of cavopulmonary artery anastomosis. *Surgery* 1973; 74: 899–916.
33. McPaul RC, Tajik AJ, Mair DD, Danielson GK, Seward JB. Development of pulmonary arteriovenous shunt after superior vena cava-right pulmonary artery (Glenn) anastomosis. Report of four cases. *Circulation* 1977; 55: 212–216.
34. Trusler GA, Williams WG, Cohen AJ, et al. William Glenn lecture: the cavopulmonary shunt. Evolution of a concept. *Circulation* 1990; 82: IV131–IV138.
35. Kopf GS, Laks H, Stansel HC, Hellenbrand WE, Kleinman CS, Talner NS. Thirty-year follow-up of superior vena cavopulmonary artery (Glenn) shunts. *J Thorac Cardiovasc Surg* 1990; 100: 662–670.
36. Duncan BW, Desai S. Pulmonary arteriovenous malformations after cavopulmonary anastomosis. *Ann Thorac Surg* 2003; 76: 1759–1766.
37. Chuang VP, Mena CE, Hoskins PA. Congenital anomalies of the inferior vena cava. Review of embryogenesis and presentation of a simplified classification. *Br J Radiol* 1974; 47: 206–213.
38. Mayo J, Gray R, St Louis E, Grosman H, McLoughlin M, Wise D. Anomalies of the inferior vena cava. *Am J Roentgenol* 1983; 140: 339–345.
39. Debich DE, Devine WA, Anderson RH. Polysplenia with normally structured hearts. *Am J Cardiol* 1990; 65: 1274–1275.
40. Guardado FJ, Byrd TM, Petersen WG. Azygos continuation of the inferior vena cava with anomalous hepatic vein drainage. *Am J Med Sci* 2012; 343: 259–261.
41. Celentano C, Malinger G, Rotmensch S, Gerboni S, Wolman Y, Glezerman M. Prenatal diagnosis of interrupted inferior vena cava as an isolated finding: a benign vascular malformation. *Ultrasound Obstet Gynecol* 1999; 14: 215–218.
42. Kawashima Y, Kitamura S, Matsuda H, Shimazaki Y, Nakano S, Hirose H. Total cavopulmonary shunt operation in complex cardiac anomalies. A new operation. *J Thorac Cardiovasc Surg* 1984; 87: 74–81.
43. Kawashima Y, Matsuki O, Yagihara T, Matsuda H. Total cavopulmonary shunt operation. *Semin Thorac Cardiovasc Surg* 1994; 6: 17–20.

44. Brown JW, Ruzmetov M, Vijay P, Rodefeld MD, Turrentine MW. Pulmonary arteriovenous malformations in children after the Kawashima operation. *Ann Thorac Surg* 2005; 80: 1592–1596.
45. Kutty S, Frommelt MA, Danford DA, Tweddell JS. Medium-term outcomes of Kawashima and completion Fontan palliation in single-ventricle heart disease with heterotaxy and interrupted inferior vena cava. *Ann Thorac Surg* 2010; 90: 1609–1613.
46. Srivastava D, Preminger T, Lock JE, et al. Hepatic venous blood and the development of pulmonary arteriovenous malformations in congenital heart disease. *Circulation* 1995; 92: 1217–1222.
47. Vollebregt A, Pushparajah K, Rizvi M, et al. Outcomes following the Kawashima procedure for single-ventricle palliation in left atrial isomerism. *Eur J Cardiothorac Surg* 2012; 41: 574–579.
48. Setyapranata S, Brizard CP, Konstantinov IE, Iyengar A, Cheung M, d'Udekem Y. Should we always plan a Fontan completion after a Kawashima procedure? *Eur J Cardiothorac Surg* 2011; 40: 1011–1015.
49. Papagiannis J, Kanter RJ, Effman EL, et al. Polysplenia with pulmonary arteriovenous malformations. *Pediatr Cardiol* 1993; 14: 127–129.
50. Gurses D, Ulger Z, Levent E, Ozyurek AR. A very rare case of polysplenia syndrome with congenital diffuse pulmonary arteriovenous fistulas. *Turk J Pediatr* 2006; 48: 96–99.
51. Kawata H, Kishimoto H, Ikawa S, et al. Pulmonary and systemic arteriovenous fistulas in patients with left isomerism. *Cardiol Young* 1998; 8: 290–294.
52. Agnoletti G, Borghi A, Annecchino FP, Crupi G. Regression of pulmonary fistulas in congenital heart disease after redirection of hepatic venous flow to the lungs. *Ann Thorac Surg* 2001; 72: 909–911.
53. Lee J, Menkis AH, Rosenberg HC. Reversal of pulmonary arteriovenous malformation after diversion of anomalous hepatic drainage. *Ann Thorac Surg* 1998; 65: 848–849.
54. Brochard P, Lejonc JL, Loisanca DY, Nitenberg A. A rare cause of cyanosis and polycythemia: anomalous systemic venous connections without associated intracardiac malformations. The blue milkman story. *Eur Heart J* 1981; 2: 227–233.
55. Kirsch J, Araoz PA, Breen JF, Chareonthaitawee P. Isolated total anomalous connection of the hepatic veins to the left atrium. *J Cardiovasc Comput Tomogr* 2007; 1: 55–57.
56. Kloppenburg GT, Post MC, Mager HJ, Schepens MA. Rerouting anomalous hepatic venous connection to the left atrium. *Ann Thorac Surg* 2010; 90: 638–640.
57. Stoller JK, Hoffman RM, White RD, Mee RB. Anomalous hepatic venous drainage into the left atrium: an unusual cause of hypoxemia. *Respir Care* 2003; 48: 58–62.
58. Al-Ammouri I, Shomali W, Alsmady MM, et al. Anomalous inferior vena cava drainage to the left atrium with successful staged repair in a 32-year-old woman with arthritis. *Pediatr Cardiol* 2010; 31: 912–914.
59. Black H, Smith GT, Goodale WT. Anomalous inferior vena cava draining into the left atrium associated with intact interatrial septum and multiple pulmonary arteriovenous fistulae. *Circulation* 1964; 29: 258–267.
60. Gardner DL, Cole L. Long survival with inferior vena cava draining into left atrium. *Br Heart J* 1955; 17: 93–97.
61. Sierig G, Vondrys D, Daehnert I. Anomalous drainage of the inferior caval vein to the left atrium. *Cardiol Young* 2005; 15: 85–87.
62. Kogon BE, Fyfe D, Butler H, Kanter KR. Anomalous drainage of the inferior vena cava into the left atrium. *Pediatr Cardiol* 2006; 27: 183–185.
63. Meadows WR, Bergstrand I, Sharp JT. Isolated anomalous connection of a great vein to the left atrium. The syndrome of cyanosis and clubbing, “normal” heart, and left ventricular hypertrophy on electrocardiogram. *Circulation* 1961; 24: 669–676.
64. Venables AW. Isolated drainage of the inferior vena cava to the left atrium. *Br Heart J* 1963; 25: 545–548.
65. Bernstein HS, Ursell PC, Brook MM, Hanley FC, Silverman NH, Bristow J. Fulminant development of pulmonary arteriovenous fistulas in an infant after total cavopulmonary shunt. *Pediatr Cardiol* 1996; 17: 46–50.
66. Duncan BW, Kneebone JM, Chi EY, et al. A detailed histologic analysis of pulmonary arteriovenous malformations in children with cyanotic congenital heart disease. *J Thorac Cardiovasc Surg* 1999; 117: 931–938.
67. Starnes SL, Duncan BW, Kneebone JM, et al. Pulmonary microvessel density is a marker of angiogenesis in children after cavopulmonary anastomosis. *J Thorac Cardiovasc Surg* 2000; 120: 902–907.
68. Starnes SL, Duncan BW, Kneebone JM, et al. Angiogenic proteins in the lungs of children after cavopulmonary anastomosis. *J Thorac Cardiovasc Surg* 2001; 122: 518–523.
69. Sloan RD, Cooley RN. Congenital pulmonary arteriovenous aneurysm. *Am J Roentgenol Radium Ther Nucl Med* 1953; 70: 183–210.
70. Rodriguez-Roisin R, Krowka MJ, Herve P, Fallon MB. Pulmonary-hepatic vascular disorders (PHD). *Eur Respir J* 2004; 24: 861–880.
71. Rodriguez-Roisin R, Krowka MJ. Hepatopulmonary syndrome – a liver-induced lung vascular disorder. *New Engl J Med* 2008; 358: 2378–2387.
72. Gupta D, Vijaya DR, Gupta R, et al. Prevalence of hepatopulmonary syndrome in cirrhosis and extrahepatic portal venous obstruction. *Am J Gastroenterol* 2001; 96: 3395–3399.
73. Varghese J, Ilias-basha H, Dhanasekaran R, Singh S, Venkataraman J. Hepatopulmonary syndrome – past to present. *Ann Hepatol* 2007; 6: 135–142.
74. Berthelot P, Walker JG, Sherlock S, Reid L. Arterial changes in the lungs in cirrhosis of the liver – lung spider nevi. *N Engl J Med* 1966; 274: 291–298.
75. Agusti AG, Roca J, Rodriguez-Roisin R. Mechanisms of gas exchange impairment in patients with liver cirrhosis. *Clin Chest Med* 1996; 17: 49–66.
76. Herve P, Lebrec D, Brenot F, et al. Pulmonary vascular disorders in portal hypertension. *Eur Respir J* 1998; 11: 1153–1166.
77. Thenappan T, Goel A, Marsboom G, et al. A central role for CD68(+) macrophages in hepatopulmonary syndrome. Reversal by macrophage depletion. *Am J Respir Crit Care Med* 2011; 183: 1080–1091.
78. Zhang ZJ, Yang CQ. Progress in investigating the pathogenesis of hepatopulmonary syndrome. *Hepatobiliary Pancreat Dis Int* 2010; 9: 355–360.
79. De BK, Sen S, Biswas PK, et al. Occurrence of hepatopulmonary syndrome in Budd–Chiari syndrome and the role of venous decompression. *Gastroenterology* 2002; 122: 897–903.
80. Alvarez AE, Ribeiro AF, Hessel G, Baracat J, Ribeiro JD. Abernethy malformation: one of the etiologies of hepatopulmonary syndrome. *Pediatr Pulmonol* 2002; 34: 391–394.
81. Howard ER, Davenport M. Congenital extrahepatic portocaval shunts – the Abernethy malformation. *J Pediatr Surg* 1997; 32: 494–497.
82. Knight WB, Mee RB. A cure for pulmonary arteriovenous fistulas? *Ann Thorac Surg* 1995; 59: 999–1001.
83. Kwon BS, Bae EJ, Kim GB, Noh CI, Choi JY, Yun YS. Development of bilateral diffuse pulmonary arteriovenous fistula after Fontan procedure: is there nonhepatic factor? *Ann Thorac Surg* 2009; 88: 677–680.
84. Moore JW, Kirby WC, Madden WA, Gaither NS. Development of pulmonary arteriovenous malformations after modified Fontan operations. *J Thorac Cardiovasc Surg* 1989; 98: 1045–1050.
85. Justino H, Benson LN, Freedom RM. Development of unilateral pulmonary arteriovenous malformations due to

- unequal distribution of hepatic venous flow. *Circulation* 2001; 103: E39–E40.
86. McElhinney DB, Marx GR, Marshall AC, Mayer JE, Del Nido PJ. Cavopulmonary pathway modification in patients with heterotaxy and newly diagnosed or persistent pulmonary arteriovenous malformations after a modified Fontan operation. *J Thorac Cardiovasc Surg* 2011; 141: 1362–1370; e1361.
 87. Nakamura Y, Yagihara T, Kagisaki K, Hagino I, Kobayashi J. Pulmonary arteriovenous malformations after a Fontan operation in the left isomerism and absent inferior vena cava. *Eur J Cardiothorac Surg* 2009; 36: 69–76.
 88. Pike NA, Vricella LA, Feinstein JA, Black MD, Reitz BA. Regression of severe pulmonary arteriovenous malformations after Fontan revision and “hepatic factor” rerouting. *Ann Thorac Surg* 2004; 78: 697–699.
 89. Nakata T, Fujimoto Y, Hirose K, et al. Fontan completion in patients with atrial isomerism and separate hepatic venous drainage. *Eur J Cardiothorac Surg* 2010; 37: 1264–1270.
 90. Aidala E, Chiappa E, Cascarano MT, Valori A, Abbruzzese PA. Partial hepatic vein diversion in pulmonary arteriovenous malformations in congenital heart disease. *Ann Thorac Surg* 2004; 78: 1089–1090.
 91. Baskett RJ, Ross DB, Warren AE, Sharratt GP, Murphy DA. Hepatic vein to the azygous vein anastomosis for pulmonary arteriovenous fistulae. *Ann Thorac Surg* 1999; 68: 232–233.
 92. Ichikawa H, Fukushima N, Ono M, et al. Resolution of pulmonary arteriovenous fistula by redirection of hepatic venous blood. *Ann Thorac Surg* 2004; 77: 1825–1827.
 93. Kim SJ, Bae EJ, Lee JY, Lim HG, Lee C, Lee CH. Inclusion of hepatic venous drainage in patients with pulmonary arteriovenous fistulas. *Ann Thorac Surg* 2009; 87: 548–553.
 94. Lopez Enriquez E, Martinez Catinchi F, Johnson C. Pulmonary arteriovenous fistula: report of a case. *Bol Asoc Med P R* 1976; 68: 217–220.
 95. McElhinney DB, Kreutzer J, Lang P, Mayer JE Jr, del Nido PJ, Lock JE. Incorporation of the hepatic veins into the cavopulmonary circulation in patients with heterotaxy and pulmonary arteriovenous malformations after a Kawashima procedure. *Ann Thorac Surg* 2005; 80: 1597–1603.
 96. Steinberg J, Alfieri GM, Brandt III B, et al. New approach to the surgical management of pulmonary arteriovenous malformations after cavopulmonary anastomosis. *Ann Thorac Surg* 2003; 75: 1640–1642.
 97. Wu IH, Nguyen KH. Redirection of hepatic drainage for treatment of pulmonary arteriovenous malformations following the Fontan procedure. *Pediatr Cardiol* 2006; 27: 519–522.
 98. Shah MJ, Rychik J, Fogel MA, Murphy JD, Jacobs ML. Pulmonary AV malformations after superior cavopulmonary connection: resolution after inclusion of hepatic veins in the pulmonary circulation. *Ann Thorac Surg* 1997; 63: 960–963.
 99. Burstein DS, Mavroudis C, Puchalski MD, Stewart RD, Blanco CJ, Jacobs ML. Pulmonary arteriovenous malformations in heterotaxy syndrome: the case for early, direct hepatic vein-to-azygos vein connection. *World J Pediatr Cong Heart Surg* 2011; 2: 119–128.
 100. Georghiou GP, Erez E, Bruckheimer E, Dagan O, Vidne BA, Birk E. Anomalous hepatic venous drainage. *Ann Thorac Surg* 2005; 80: 1113–1115.
 101. Bradley SE, Ingelfinger FJ, et al. The estimation of hepatic blood flow in man. *J Clin Invest* 1945; 24: 890–897.
 102. Wiklund L. Human hepatic blood flow and its relation to systemic circulation during intravenous infusion of bupivacaine or etidocaine. *Acta Anaesthesiol Scand* 1977; 21: 189–199.
 103. McAllister KA, Grogg KM, Johnson DW, et al. Endoglin, a TGF-beta binding protein of endothelial cells, is the gene for hereditary haemorrhagic telangiectasia type 1. *Nat Genet* 1994; 8: 345–351.
 104. McAllister KA, Lennon F, Bowles-Biesecker B, et al. Genetic heterogeneity in hereditary haemorrhagic telangiectasia: possible correlation with clinical phenotype. *J Med Genet* 1994; 31: 927–932.
 105. Berg JN, Gallione CJ, Stenzel TT, et al. The activin receptor-like kinase 1 gene: genomic structure and mutations in hereditary hemorrhagic telangiectasia type 2. *Am J Hum Genet* 1997; 61: 60–67.
 106. van den Driesche S, Mummery CL, Westermann CJ. Hereditary hemorrhagic telangiectasia: an update on transforming growth factor beta signaling in vasculogenesis and angiogenesis. *Cardiovasc Res* 2003; 58: 20–31.
 107. Lebrin F, Goumans MJ, Jonker L, et al. Endoglin promotes endothelial cell proliferation and TGF-beta/ALK1 signal transduction. *Embo J* 2004; 23: 4018–4028.
 108. David L, Mallet C, Keramidis M, et al. Bone morphogenetic protein-9 is a circulating vascular quiescence factor. *Circ Res* 2008; 102: 914–922.
 109. Schwarte-Waldhoff I, Schmiegel W. Smad4 transcriptional pathways and angiogenesis. *Int J Gastrointest Cancer* 2002; 31: 47–59.
 110. Nyberg P, Xie L, Kalluri R. Endogenous inhibitors of angiogenesis. *Cancer Res* 2005; 65: 3967–3979.
 111. Ribatti D. Endogenous inhibitors of angiogenesis: a historical review. *Leuk Res* 2009; 33: 638–644.
 112. Baghdady Y, Hussein Y, Shehata M. Vascular endothelial growth factor in children with cyanotic and acyanotic and congenital heart disease. *Arch Med Sci* 2010; 6: 221–225.
 113. El-Melegy NT, Mohamed NA. Angiogenic biomarkers in children with congenital heart disease: possible implications. *It J Pediatr* 2010; 36: 32.
 114. Aydin HI, Yozgat Y, Demirkaya E, et al. Correlation between vascular endothelial growth factor and leptin in children with cyanotic congenital heart disease. *Turk J Pediatr* 2007; 49: 360–364.
 115. Himeno W, Akagi T, Furui J, et al. Increased angiogenic growth factor in cyanotic congenital heart disease. *Pediatr Cardiol* 2003; 24: 127–132.
 116. Ootaki Y, Yamaguchi M, Yoshimura N, Oka S, Yoshida M, Hasegawa T. Vascular endothelial growth factor in children with congenital heart disease. *Ann Thorac Surg* 2003; 75: 1523–1526.
 117. Starnes SL, Duncan BW, Kneebone JM, et al. Vascular endothelial growth factor and basic fibroblast growth factor in children with cyanotic congenital heart disease. *J Thorac Cardiovasc Surg* 2000; 119: 534–539.
 118. Suda K, Matsumura M, Miyaniishi S, Uehara K, Sugita T, Matsumoto M. Increased vascular endothelial growth factor in patients with cyanotic congenital heart diseases may not be normalized after a Fontan type operation. *Ann Thorac Surg* 2004; 78: 942–946; discussion 946–947.
 119. Seppinen L, Pihlajaniemi T. The multiple functions of collagen XVIII in development and disease. *Matrix Biol* 2011; 30: 83–92.
 120. Zheng MJ. Endostatin derivative angiogenesis inhibitors. *Chin Med J* 2009; 122: 1947–1951.
 121. Clement B, Musso O, Lietard J, Theret N. Homeostatic control of angiogenesis: a newly identified function of the liver? *Hepatology* 1999; 29: 621–623.
 122. Musso O, Theret N, Heljasvaara R, et al. Tumor hepatocytes and basement membrane-producing cells specifically express two different forms of the endostatin precursor, collagen XVIII, in human liver cancers. *Hepatology* 2001; 33: 868–876.
 123. Teodoro JG, Parker AE, Zhu X, Green MR. p53-Mediated inhibition of angiogenesis through up-regulation of a collagen prolyl hydroxylase. *Science* 2006; 313: 968–971.

124. Lee TY, Tjin Tham Sjin RM, Movahedi S, et al. Linking antibody Fc domain to endostatin significantly improves endostatin half-life and efficacy. *Clin Cancer Res* 2008; 14: 1487–1493.
125. Folkman J. Antiangiogenesis in cancer therapy – endostatin and its mechanisms of action. *Exp Cell Res* 2006; 312: 594–607.
126. Herbst RS, Hess KR, Tran HT, et al. Phase I study of recombinant human endostatin in patients with advanced solid tumors. *J Clin Oncol* 2002; 20: 3792–3803.
127. Thomas JP, Arzoumanian RZ, Alberti D, et al. Phase I pharmacokinetic and pharmacodynamic study of recombinant human endostatin in patients with advanced solid tumors. *J Clin Oncol* 2003; 21: 223–231.
128. Field-Ridley A, Heljasvara R, Pihlajaniemi T, et al. Endostatin, an inhibitor of angiogenesis, decreases after bidirectional superior vena cava anastomosis. *Pediatr Cardiol* 2012; doi:10.1007/S00246-012-0441-2
129. Damico R, Simms T, Kim BS, et al. p53 mediates cigarette smoke-induced apoptosis of pulmonary endothelial cells: inhibitory effects of macrophage migration inhibitor factor. *Am J Respir Cell Mol Biol* 2011; 44: 323–332.
130. Tipps RS, Mumtaz M, Leahy P, Duncan BW. Gene array analysis of a rat model of pulmonary arteriovenous malformations after superior cavopulmonary anastomosis. *J Thorac Cardiovasc Surg* 2008; 136: 283–289.
131. Brauer R, Beck IM, Roderfeld M, Roeb E, Sedlacek R. Matrix metalloproteinase-19 inhibits growth of endothelial cells by generating angiostatin-like fragments from plasminogen. *BMC Biochem* 2011; 12: 38.
132. Gonzalez-Gronow M, Grenett HE, Fuller GM, Pizzo SV. The role of carbohydrate in the function of human plasminogen: comparison of the protein obtained from molecular cloning and expression in *Escherichia coli* and COS cells. *Biochim Biophys Acta* 1990; 1039: 269–276.
133. Fujiyama S, Matsubara H, Nozawa Y, et al. Angiotensin AT(1) and AT(2) receptors differentially regulate angiopoietin-2 and vascular endothelial growth factor expression and angiogenesis by modulating heparin binding-epidermal growth factor (EGF)-mediated EGF receptor transactivation. *Circ Res* 2001; 88: 22–29.
134. Walther T, Menrad A, Orzechowski HD, Siemeister G, Paul M, Schirner M. Differential regulation of in vivo angiogenesis by angiotensin II receptors. *FASEB J* 2003; 17: 2061–2067.
135. de Resende MM, Stodola TJ, Greene AS. Role of the renin angiotensin system on bone marrow-derived stem cell function and its impact on skeletal muscle angiogenesis. *Physiol Genomics* 2010; 42: 437–444.
136. Yin T, Ma X, Zhao L, Cheng K, Wang H. Angiotensin II promotes NO production, inhibits apoptosis and enhances adhesion potential of bone marrow-derived endothelial progenitor cells. *Cell Res* 2008; 18: 792–799.
137. Grace JA, Herath CB, Mak KY, Burrell LM, Angus PW. Update on new aspects of the renin–angiotensin system in liver disease: clinical implications and new therapeutic options. *Clin Sci (Lond)* 2012; 123: 225–239.
138. Clapp C, Thebault S, Jeziorski MC, Martinez De La Escalera G. Peptide hormone regulation of angiogenesis. *Physiol Rev* 2009; 89: 1177–1215.
139. Al-Merani SA, Brooks DP, Chapman BJ, Munday KA. The half-lives of angiotensin II, angiotensin II-amide, angiotensin III, Sar1-Ala8-angiotensin II and renin in the circulatory system of the rat. *J Physiol* 1978; 278: 471–490.
140. Bailie MD, Rector FC Jr, Seldin DW. Angiotensin II in arterial and renal venous plasma and renal lymph in the dog. *J Clin Invest* 1971; 50: 119–126.
141. Semple PE, Cumming AM, Millar JA. Angiotensins I and II in renal vein blood. *Kidney Int* 1979; 15: 276–282.
142. Admiraal PJ, Danser AH, Jong MS, Pieterman H, Derckx FH, Schalekamp MA. Regional angiotensin II production in essential hypertension and renal artery stenosis. *Hypertension* 1993; 21: 173–184.
143. Marshall B, Duncan BW, Jonas RA. The role of angiogenesis in the development of pulmonary arteriovenous malformations in children after cavopulmonary anastomosis. *Cardiol Young* 1997; 7: 370–374.
144. Amaral SL, Linderman JR, Morse MM, Greene AS. Angiogenesis induced by electrical stimulation is mediated by angiotensin II and VEGF. *Microcirculation* 2001; 8: 57–67.
145. Amaral SL, Papanek PE, Greene AS. Angiotensin II and VEGF are involved in angiogenesis induced by short-term exercise training. *Am J Physiol Heart Circ Physiol* 2001; 281: H1163–H1169.
146. Gwathmey TM, Alzayadneh EM, Pendergrass KD, Chappell MC. Novel roles of nuclear angiotensin receptors and signaling mechanisms. *Am J Physiol Regul Integr Comp Physiol* 2012; 302: R518–R530.
147. Gwathmey TM, Westwood BM, Pirro NT, et al. Nuclear angiotensin-(1-7) receptor is functionally coupled to the formation of nitric oxide. *Am J Physiol Renal Physiol* 2010; 299: F983–F990.
148. Malhotra SP, Reddy VM, Thelitz S, et al. Cavopulmonary anastomosis induces pulmonary expression of the angiotensin II receptor family. *J Thorac Cardiovasc Surg* 2002; 123: 655–660.
149. Malhotra SP, Riemer RK, Thelitz S, He YP, Hanley FL, Reddy VM. Superior cavopulmonary anastomosis suppresses the activity and expression of pulmonary angiotensin-converting enzyme. *J Thorac Cardiovasc Surg* 2001; 122: 464–469.
150. Gupta LB, Kumar A, Jaiswal AK, et al. Pentoxifylline therapy for hepatopulmonary syndrome: a pilot study. *Arch Int Med* 2008; 168: 1820–1823.
151. Zhang J, Ling Y, Tang L, et al. Pentoxifylline attenuation of experimental hepatopulmonary syndrome. *J Appl Physiol* 2007; 102: 949–955.
152. Lumsden AB, Henderson JM, Kutner MH. Endotoxin levels measured by a chromogenic assay in portal, hepatic and peripheral venous blood in patients with cirrhosis. *Hepatology* 1988; 8: 232–236.
153. Pidgeon GP, Harmey JH, Kay E, Da Costa M, Redmond HP, Bouchier-Hayes DJ. The role of endotoxin/lipopolysaccharide in surgically induced tumour growth in a murine model of metastatic disease. *Br J Cancer* 1999; 81: 1311–1317.