Original Article



Statewide surveillance of carbapenemase-producing carbapenem-resistant *Escherichia coli* and *Klebsiella* species in Washington state, October 2012–December 2017

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Abstract

Background: Carbapenem-resistant Enterobacterales (CRE) are common causes of healthcare-associated infections and are often multidrug resistant with limited therapeutic options. Additionally, CRE can spread within and between healthcare facilities, amplifying potential harms.

Objective: To better understand the burden, risk factors, and source of acquisition of carbapenemase genes in clinical *Escherichia coli* and *Klebsiella* spp isolates from patients in Washington to guide prevention efforts.

Design: Multicenter prospective surveillance study.

Methods: *Escherichia coli* and *Klebsiella* spp isolates meeting the Washington state CRE surveillance case definition were solicited from clinical laboratories and tested at Washington Public Health Laboratories using polymerase chain reaction (PCR) for the 5 most common carbapenemase genes: bla_{NDA} , bla_{NDM} , bla_{IMP} , bla_{OXA-48} . Case patients positive by PCR were investigated by the public health department.

Results: From October 2012 through December 2017, 363 carbapenem-resistant *E. coli* and *Klebsiella* spp isolates were tested. Overall, 45 of 115 carbapenem-resistant *K. pneumoniae* (39%), 1 of 8 *K. oxytoca* (12.5%), and 28 of 239 carbapenem-resistant *E. coli* (11.7%) were carbapenemase positive. Of 74 carbapenemase-positive isolates, bla_{KPC} was most common (47%), followed by bla_{NDM} (30%), bla_{OXA-48} (22%), and bla_{IMP} (1%). Although all cases had healthcare exposure, bla_{KPC} acquisition was associated with US health care, whereas non- bla_{KPC} acquisition was associated with international health care or travel.

Conclusions: We report that bla_{KPC} , the most prevalent carbapenemase in the United States, accounts for nearly half of carbapenemase cases in Washington state and that most KPC-cases are likely acquired through in-state health care.

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Antimicrobial resistance is one of the most serious global health threats of the 21st century. Currently carbapenems, β -lactam antibiotics that target and arrest bacterial cell wall synthesis serve as "last resort" therapeutic agents for multidrug-resistant infections caused by bacteria in the Enterobacterales family (formerly called Enterobacteriaceae).^{1,2} Infections with carbapenem-resistant Enterobacterales (CRE), and particularly carbapenemase-producing Enterobacterales (CPE), increased during the first decade of this century with a coincident increase in patient morbidity and mortality due in part to limited treatment options.^{3,4} In response, the

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Centers for Disease Control and Prevention (CDC) has categorized CRE as an urgent public health threat and recommends aggressive coordinated action to minimize emergence and spread.¹

Various mechanisms of carbapenem-resistance in Enterobacterales have been described; however, resistance mediated by production of carbapenemases (enzymes that hydrolyze and inactivate carbapenem antibiotics) are of particular concern.⁵ Carbapenemase genes are generally found on mobile genetic elements that can increase exponentially in a population through the combination of horizontal gene transfer and clonal expansion.⁵ The most commonly reported carbapenemases in Enterobacterales worldwide are Klebsiella pneumoniae carbapenemase (KPC), New Delhi metallo-β-lactamase (NDM), oxacillin-hydrolyzing β -lactamase-48 (OXA-48), Verona integron-encoded metallo- β lactamase (VIM), and imipenem-hydrolyzing β -lactamase (IMP).¹ KPC was first identified in the United States in 1996 and is considered endemic in many areas of the world, including parts of the United States.^{1,6,7} NDM and OXA-48 have primarily been associated with health care on the Indian subcontinent and in Europe, respectively.⁸ NDM cases reported in the United States have mostly been identified in patients with either international travel or health care.^{1,9} IMP and VIM are predominately identified in Asia with sporadic cases identified in the United States.¹⁰ The epidemiology of CRE indicates that CPE have spread, which in turn has prompted active surveillance to direct targeted actions to prevent transmission.¹¹

CPE are increasingly common causes of healthcare-associated infections and are typically seen in patients with prior healthcare exposure.¹²⁻¹⁴ The US Centers for Disease Control and Prevention (CDC) has published tool kits for monitoring and preventing spread of CPE.¹⁵ These guidance documents stress the importance of understanding the epidemiology of CRE and CPE locally, regionally, and nationally to develop appropriate prevention strategies. Therefore, systematic surveillance of circulating carbapenemresistance mechanisms is necessary to track prevalence and coordinate public health response.

In October 2012, Washington State Public Health Laboratories (WAPHL) began prospective, statewide surveillance and carbapenemase testing for CRE isolates. The objectives of this work were (1) to quantify prevalence and assess molecular characteristics of carbapenem-resistant *Escherichia coli* and *Klebsiella* spp circulating in Washington state, (2) to determine broad sources of carbapenemase acquisition including Washington healthcare facility or foreign healthcare exposure/travel, and (3) to compare Washington CPE surveillance to data from other states to guide prevention efforts. We present patient and microbiological characteristics from 5 years of CRE surveillance in Washington.

Methods

Statewide submission of CRE bacterial isolates

The Washington State Department of Health (DOH) solicited CRE clinical isolates from clinical laboratories across Washington state from October 2012 through December 2017 for submission to WAPHL. The CRE surveillance case definition changed twice during the surveillance period. From October 2012 through December 2013, submission criteria included all genera of Enterobacterales resistant to all third-generation cephalosporins tested and nonsusceptible to 1 or more carbapenems. Criteria for submission from January 2014 through April 2015 were E. coli and Klebsiella spp resistant to all third-generation cephalosporins tested and nonsusceptible to 1 or more carbapenems. Starting May 2015 through December 2017, surveillance criteria were all E. coli, Klebsiella spp, and Enterobacter spp resistant to any carbapenem. Various submission criteria are summarized in Supplementary Table 1 (online). Due to variability in submission criteria, only carbapenem-resistant E. coli and Klebsiella spp were included in this study and CR-Enterobacter isolates were excluded.

Information accompanying isolates included date of collection, specimen source, organism identification, and patient demographic data including gender and age.

Microbiology and molecular testing methods

The identification of all CR-*E. coli* and CR-*Klebsiella* isolates received from laboratories was confirmed using traditional biochemicals as previously described.¹⁶ Additionally, isolates underwent confirmatory antibiotic susceptibility testing using Kirby-Bauer disk diffusion and/or Etest methods as described by the Clinical Laboratory Standards Institute (CLSI). Current, available cephalosporin and carbapenem CLSI break points were used based on year tested.¹⁷⁻²² Any isolates that failed identification or antimicrobial susceptibility confirmatory testing were excluded

from the study. Depending on the testing protocol in place at the time of submission to WAPHL, some CR-*E. coli* and CR-*Klebsiella* isolates underwent phenotypic assay for carbapenemase production in addition to PCR for carbapenemase genes. The modified Hodge test (MHT) was performed from October 2012 through April 2015, and PCR alone without MHT was performed from May 2015 until September 22, 2017, when testing including both the modified carbapenem inactivation test (mCIM) test and PCR began. The MHT and mCIM were performed as previously described.^{23,24}

The identification of carbapenemase-encoding genes was determined using single-plex laboratory developed traditional polymerase chain reactions (PCR) validated to detect the 5 most common carbapenemase genes: bla_{KPC} , bla_{NDM} , bla_{OXA-48} , bla_{VIM} , and bla_{IMP} . The PCR amplification primer sequences were adapted from previous published reports^{10,25,26} and are listed in Supplementary Table 2 (online). Genomic DNA was extracted from CR-Klebsiella and CR-E. coli isolates using a rapid-boil method. For each bacterial isolate, a 1-µL loopful of colonies was taken from extended-spectrum β-lactamase (ESBL) HardyCHROM (Hardy Diagnostics, Santa Maria, CA) or blood agar, suspended in 1× TE buffer, boiled at 100°C for 5 minutes. Extracted DNA was amplified using the HotStarTaq Master Mix Kit (Qiagen, Hilden, Germany). Reactions were carried out under the following conditions: initial denaturation (94°C for 2 minutes), followed by 30 cycles of denaturation (94°C for 30 seconds), annealing (at specified T_{ann} per Supplementary Table 2 for 15 seconds) and extension (72°C for 1 minute). Amplicons and products were analyzed on electrophoresis gels or the TapeStation 2200 genetic analyzer (Agilent Technologies, Santa Clara, CA) and visualized for appropriate band size compared to positive control strains and control DNA ladder.

Surveillance data analysis

To prevent analysis of duplicate isolates, each unique genus/ species/carbapenemase combination was counted only once during the surveillance period for each case patient. Additional isolates from case patients were counted if a different carbapenemase or the same carbapenemase in a different genus or species was identified.

To determine risk factors for carbapenemase infection in Washington state, local DOH staff investigated all carbapenemase-positive case patients. Medical record review was completed for all case patients, and an interview with hospital infection prevention staff and/or interview with the patient or proxy was completed when possible. Summary data collected through these investigations were reported to the state using a standardized case report form and included information such as underlying health conditions, healthcare exposures, and international travel in the prior year. Healthcare exposure questions were developed based on healthcare exposure definitions outlined in previous reports,^{14,27,28} and they focused on hospitalizations, admissions to a long-term care facility, surgeries, dialysis, and indwelling medical devices in the year prior to date of collection.

We used an online CDC CRE tracking database to compare the number and nature of carbapenemase case reports in Washington to those in other states.²⁹

Statistical analysis

Statistical summaries were performed using Excel version 16.27 software (Microsoft, Redmond, WA) to describe frequency and proportion of carbapenem-resistant *E. coli* and *Klebsiella* spp

 Table 1. Clinical Characteristics and Demographics for Carbapenem-Resistant

 Enterobacterales (CRE)-Positive Patients

Characteristic	Total Patients (n=357), No. (%) ^a			
Demographics				
Male	140 (39.2)			
Female	217 (60.8)			
Median age, y (range)	66 (0–101)			
	Total Isolates (n=363), No. (%)			
Location at time of culture ^b				
Outpatient	57 (15.7)			
Acute-care hospital	74 (20.4)			
Long-term care facility	18 (5.0)			
Long-term care hospital	11 (3.0)			
Unknown	203 (55.9)			
Specimen type				
Urine	187 (51.5)			
Blood	24 (6.6)			
Respiratory	15 (4.1)			
Wound	15 (4.1)			
Stool	13 (3.6)			
Other site	46 (12.7)			
Unknown	63 (17.4)			

^a6 patients had 2 isolates.

^bPatient location was not collected for all isolates without carbapenemase.

tested, proportion of carbapenemase-positive isolates of the total number of isolates tested, and proportion of each carbapenemase gene detected in respective genera.

Results

Patient demographics and source of carbapenem-resistant E. coli and Klebsiella spp isolates tested

From October 2012 through December 2017, 363 unique CRE isolates from 357 patients were tested at WAPHL. Isolates were submitted by 61 unique clinical laboratories from patients receiving medical care across 21 total counties spanning the geographical area of Washington state. Of these, 239 (66%) isolates were *E. coli* and 124 (34%) were *Klebsiella* spp. Patient demographics and source of isolates are summarized in Table 1. Just more than half (51.5%) of the isolates were recovered from urine specimens.

The number of carbapenem resistant isolates submitted and tested at WAPHL per year increased steadily from 2012 to 2017, with *E. coli* remaining the predominant organism throughout (Fig. 1). *Klebsiella pneumoniae* (n = 115) was the primary *Klebsiella* spp tested, followed by *K. oxytoca* (n = 8) and *K. variicola* (n = 1).

Carbapenemase genes detected

Of the 363 carbapenem-resistant *E. coli* and *Klebsiella* spp isolates tested, 74 isolates (20%) from 70 patients were carbapenemase positive (Fig. 2). These isolates originated in 51 unique healthcare facilities across Washington, most of which (n = 24, 47.1%) were acute-care hospitals (Table 2). Among the Washington isolates,

Table 2.	Clinical	Characteristics	and	Demographics	of	Case	Patients	With
Carbapenemase-Positive Escherichia coli or Klebsiella spp								

Clinical Characteristics of Case Patients (n=70)	No. (%)
Patient age, y	
0–4	3 (4)
5–17	1 (<1)
18-64	40 (57)
>64	26 (37)
Sex, male	38 (54)
Travel and healthcare exposure	
Patients with prior US healthcare exposure ^a	42 (60)
Patients with prior international exposure ^a	28 (40)
Travel and health care ^b	18
Travel	10
Location at time of culture ^d	
Acute-care hospital	35 (50)
Long-term care hospital	9 (12.8)
Skilled nursing facility	10 (14.3)
Outpatient	16 (22.9)
Specimen type (n=74 E. coli or Klebsiella spp isolates)	
Urine	45 (61)
Bronchial wash/Sputum	7 (9)
Wound or nodule	6 (8)
Blood	3 (4)
Stool/Rectal	5 (7)
Other ^c	7 (8)

^aExposure included any health care 12 months prior to isolate collection.^bHealth care is defined as hospitalization, long-term care admission, surgery, dialysis, indwelling devices. Does not include outpatient care.¹Includes isolates from bone, abdominal fluid, abscess, cerebral spinal fluid, and pleural fluid.^dIncluded 51 unique healthcare facilities; 24 acute-care hospitals, 2 long-term acute-care hospitals, 9 skilled nursing facilities, and 16 outpatient locations.

K. pneumoniae were most likely to encode a carbapenemase, with 45 of 115 isolates (39%) testing carbapenemase positive compared to only 28 of 239 E. coli isolates (12%) (Fig. 2). Two-thirds of 45 carbapenemase-positive K. pneumoniae isolates carried the bla_{KPC} gene, and the 28 carbapenemase-positive E. coli isolates were predominately positive for bla_{NDM} (57%) (Fig. 3). Of 8 K. oxytoca isolates, 1 (12%) was positive for the bla_{KPC} gene (Fig. 3) and the single K. variicola isolate submitted was carbapenemase negative. Overall, *bla_{KPC}* was the most common carbapenemase gene detected (n = 35, 47%), followed by bla_{NDM} (n = 22, 30%), bla_{OXA-48} (n = 16, 22%), and bla_{IMP} (n = 1, 1%). The bla_{VIM} gene was not detected in any E. coli or Klebsiella spp tested and analyzed during this study. All carbapenemase-positive CR-E. coli and CR-Klebsiella isolates were positive by either MHT or mCIM, depending on the method used at the time, and no isolates positive by MHT or mCIM tested negative for a carbapenemase gene by PCR.

Demographics of carbapenemase-positive case patients and molecular epidemiology of carbapenemase genes

In total, 70 case patients had a carbapenemase-positive *E. coli* or *Klebsiella* spp isolate. Four patients had 2 carbapenemase-positive

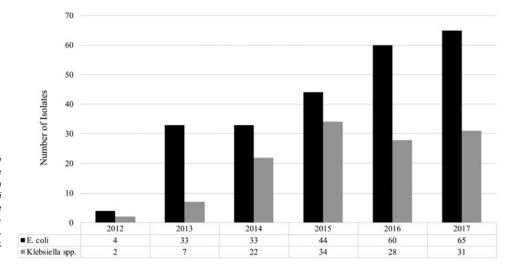
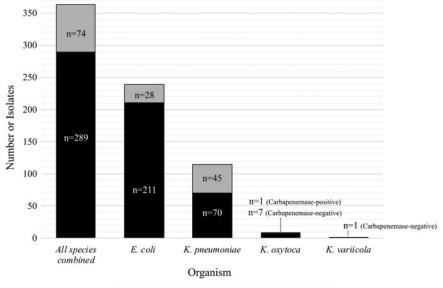


Fig. 1. Number of carbapenem-resistant *Escherichia coli* and *Klebsiella* spp isolates submitted to the Washington State Public Health Laboratories each year from 2012 to 2017. Number of total *E. coli* and *Klebsiella* spp isolates tested is shown on the y-axis and the numbers are captured in the table provided below the corresponding year on the x-axis. *E. coli* and *Klebsiella* spp. are shown by the black and grey bars respectively.



Carbapenemase-positive Carbapenemase-negative

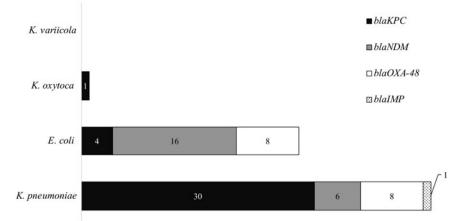


Fig. 2. Portion of carbapenemase-producing, carbapenemresistant *Escherichia coli* and *Klebsiella* spp. isolates submitted to the Washington State Public Health Laboratories from 2012 to 2017. The number of isolates is shown on the y-axis and bacterial species including *E. coli, K. pneumonia, K. oxytoca,* and *K. variicola* on the x-axis. The numbers of carbapenemase-producing and carbapenemase-negative isolates from each species are shown by grey bars and black bars respectively; "n" indicates the total number.

Fig. 3. Distribution and number of carbapenemase genes among *Escherichia coli* and *Klebsiella* spp tested at the Washington State Public Health Laboratories from 2012 to 2017. The different species are shown on the y-axis, and the horizontal bars are sectioned by carbapenemase gene identified. Each gene, including bla_{KPC} (black bars), bla_{NDM} (grey bars), bla_{IMP} (dotted bar), and bla_{OXA-dB} (white bars), are shown for each species if detected. Abbreviations: bla, β -lactamase.

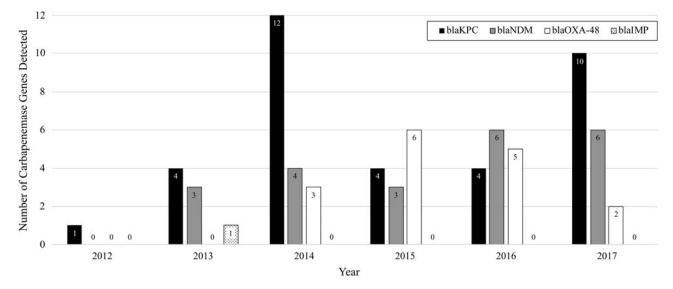


Fig. 4. Molecular epidemiology of carbapenemase genes detected each year from 2012 to 2017. The number of carbapenemase genes is depicted on the y-axis and year on the x-axis; *bla_{KPC}* (black bars), *bla_{NDM}* (grey bars), *bla_{OXA-48}* (white bars), and *bla_{IMP}* (dotted bar) are shown. Abbreviations: *bla*, β-lactamase.

isolates, each with a unique genus-species-carbapenemase combination (Supplementary Table 3 online). Most patients (58%) were between the ages of 18 and 64 years old and were male, and most carbapenemase-positive isolates (45 of 74, 61%) were cultured from urine (Table 2). From 2012 to 2014 and in 2017, bla_{KPC} was the most common carbapenemase gene detected; however other years varied, with bla_{OXA-48} and bla_{NDM} being the most common detected in 2015 and 2016 respectively (Fig. 4).

Characteristics associated with carbapenemase positivity and suspected source of carbapenemase acquisition

As determined by epidemiologic investigations, of 70 case patients with carbapenemase-producing E. coli or Klebsiella spp, 42 (60%) had healthcare exposure in Washington, 18 (26%) had international healthcare or travel exposure, and 10 (14%) had only international travel but no health care in the 12 months prior to isolate collection (Table 2). Of the 28 case patients that had international health care or travel, country of exposure was identified for 27 (97%). Most had traveled to India (n = 14) or the surrounding area: Bangladesh (n = 1), Sri Lanka (n = 1), and the United Arab Emirates (n = 1). Other locations of travel were China and Southeast Asia (n = 5), Ghana (n = 1), Ethiopia (n = 1), Cameroon (n = 1), Italy (n = 1), and Russia (n = 1). Washington healthcare exposures were assessed as the likely sources of acquisition of bla_{KPC} , whereas international health care or travel were probable sources of non- bla_{KPC} carbapenemases (Fig. 5). Furthermore, patient data for specific dates and length of stay acquired through epidemiological investigations indicated that patients with international exposure positive for non-bla_{KPC} carbapenemases did not receive overlapping care from any of our submitting facilities.

Comparison to other state and national CRE data

Based on publicly available CDC surveillance CRE data (as of December 2017), all 50 states have reported bla_{KPC} .²⁹ Washington is among the 34 states that have reported bla_{NDM} , 27 states that have reported bla_{OXA-48} , and 13 states that have reported bla_{IMP} . Based on these national surveillance data, Washington has the third-highest number of case reports for

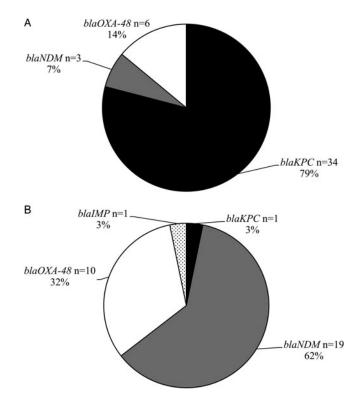


Fig. 5. Total number and percentage of carbapenemase genes from patients with either exposure to US health care or international travel or health care. (A) Proportion and percentages of carbapenemase genes linked to US healthcare exposure. (B) Proportion and percentages of carbapenemase genes linked to international travel or healthcare exposure: bla_{KPC} (black), bla_{NDM} (grey), bla_{IMP} (dotted), and bla_{out-a} (white). Abbreviations: bla, β-lactamase.

 bla_{NDM} and the second-highest number of case reports for bla_{OXA-48} in the United States.

Discussion

The CRE are of significant concern to clinicians and public health officials. Carbapenemases in CPE are typically found on plasmids

or transposons, which can be shared and therefore spread rapidly among different species of bacteria.^{5,30} These infections have few treatment options, and they are associated with poorer clinical outcomes.^{4,31} Estimating prevalence and surveillance of CRE within an institution, state, and/or region is an important step in understanding the burden and distribution of carbapenem resistance caused by carbapenemases, leading to implementation of appropriate prevention measures.

Prior to 1996, CPE was not reported in the United States, but recent studies have shown that CPE has been reported in all US states.^{1,29} Our Washington surveillance results align with national data showing that bla_{KPC} is the most prevalent carbapenemase. KPC is considered endemic in some parts of the United States and is so prevalent that the CDC and some states do not track individual case reports.^{29,32} A prospective multicenter study of hospitals in Ohio, Pennsylvania, and Michigan completed by Duin et al³³ highlighted the endemicity of KPC-producing CPE in these states with primary focus on K. pneumoniae; they reported only a single NDM positive isolate. Furthermore, a large populationand laboratory-based active surveillance study including data from Colorado, Georgia, Maryland, Minnesota, New Mexico, New York, and Oregon by Guh et al¹⁴ showed that CP-CRE account for a small subset of infections when compared to the total number of infections caused by Enterobacterales. Across these 7 states, they only identified KPC and no other carbapenemases. Interestingly, NDM-producing CPE outbreaks have been identified in both Colorado and Illinois,³⁴ and a previous multistate study characterizing NDM-positive Enterobacterales encompassed only 8 isolates from Massachusetts, California, Illinois, Virginia, and Maryland.³⁵ Washington state is among the small subset of states that have reported the presence of other carbapenemase types, including IMP. Compared to other antibiotic-resistant pathogens, such as methicillin-resistant Staphylococcus aureus (MRSA) or Clostridium difficile, the CRE incidence is lower.³⁶ However, limited treatment options and poor clinical outcomes associated with CRE infections demand interventions to prevent emergence and for controlling further spread of CPE.³⁶ The epidemiologic investigations for CPE cases in Washington state suggest that most non-KPC carbapenemases, such as NDM and OXA-48, were potentially acquired outside of the United States via international exposure. These findings support previous published literature that NDM is often associated with travel in highly endemic countries.37,38 Furthermore, non-KPC-positive patients who had international exposure did not receive overlapping care from any submitting facilities, indicating that the NDM and/or OXA-48 detected in this group were likely not transmitted within Washington state healthcare facilities. Notably, Washington state began CRE surveillance and performed PCR testing of CR-isolates for 5 carbapenemases earlier than many other states, which may in part explain the larger diversity and higher number of carbapenemase cases found relative to other states. Taken together, these data show that Washington state has a higher number of NDM and OXA-48 carbapenemases than most other states, but it is not clear whether this is due to more widely available detection or to increased prevalence.

During this surveillance period, some patients showed evidence of persistent CPE colonization, based on recurrent positive cultures for the same species/carbapenemase combination on different days. In total, 15 of the 70 carbapenemase-positive patients (21.4%) tested positive more than once with a carbapenemasepositive *E. coli* or *Klebsiella* isolate. We documented persistent colonization for longer than 3 months in 7 (47%) of these cases (6 KPC and 1 NDM), with 2 (13%) having recurrent infections multiple times >2 years after first identification (1 KPC and 1 NDM). These data suggest that persistent colonization may occur and that special screening and infection prevention interventions are required to prevent transmission.

Our study has several limitations. CRE surveillance definitions and isolate submission criteria changed between October 2012 and December 2017. Our results did not capture other carbapenemresistant pathogens that could have potentially harbored carbapenemases, such as other genera of Enterobacterales, Pseudomonas spp, and Acinetobacter spp. From October 2012 through April 2015, antibiotic resistance criteria for isolate submission were more exclusive (Supplementary Table 1 online) and required resistance to all third-generation cephalosporins tested, which may have resulted in nonsubmission of carbapenem-resistant isolates that would have been accepted starting in May 2015. Due to the variability in submission criteria, we only included carbapenemresistant E. coli and Klebsiella spp in this report as the 2 genera collected throughout the entire surveillance period. The CRE definition has evolved over time in an effort to maximize detection and reporting of carbapenemase genes. Notably, K. variicola has a history of being misidentified by microbiology laboratories, and this could explain why we identified only 1 K. variicola in our isolate cohort.³⁹ Another important factor to consider is that interviews with patient, proxy, and/or infection preventionists were only completed when possible; therefore, epidemiological investigations may have missed some health care and/or travel exposures in our carbapenemase-positive case patients. However, an extensive medical record review was conducted for each case patient, which informed the vast majority of the analysis presented here. Lastly, our surveillance system design only detected infections confirmed by the submitting laboratories; therefore, infections for which cultures were not obtained were missed.

This study highlights the prevalence of carbapenem-resistant *E. coli* and *Klebsiella* isolates with the overarching goal to determine the distribution of carbapenem-resistance resulting from the presence of carbapenemase genes circulating in patients in Washington state. CRE and CPE prevalence data are critical components of more comprehensive national CRE and CPE data that ultimately help to elucidate the burden, prevalence, and type of carbapenemase genes and in response provide a framework for potential infection and outbreak response measures.

Supplementary material. To view supplementary material for this article, please visit https://doi.org/10.1017/ice.2020.26

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Conflicts of interest. All authors report no conflicts of interest relevant to this article.

References

- 1. Centers for Disease Control and Prevention. *Antibiotic Resistance Threats in the United States*. Atlanta: CDC; 2013.
- Sievert DM, Ricks P, Edwards JR, et al. Antimicrobial-resistant pathogens associated with healthcare-associated infections: summary of data reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2009–2010. Infect Control Hosp Epidemiol 2013;34:1–14.

- 3. Centers for Disease Control and Prevention. Vital signs: carbapenemresistant Enterobacteriaceae. *Morb Mortal Wkly Rep* 2013;62:165–170.
- Patel G, Huprikar S, Factor SH, Jenkins SG, Calfee DP. Outcomes of carbapenem-resistant *Klebsiella pneumoniae* infection and the impact of antimicrobial and adjunctive therapies. *Infect Control Hosp Epidemiol.* 2008;29:1099–1106.
- Queenan AM, Bush K. Carbapenemases: the versatile lactamases. Clin Microbiol Rev 2007;20:440–458.
- 6. Molton JS, Tambyah PA, P Ang BS, Lin Ling M, Fisher DA. The global spread of healthcare-associated multidrug-resistant bacteria: a perspective from Asia. *Clin Infect Dis* 2013;56:1310–1318.
- 7. Yigit H, Queenan AM, Anderson GJ, et al. Novel carbapenemhydrolyzing-lactamase, KPC-1, from a carbapenem-resistant strain of *Klebsiella pneumoniae. Antimicrob Agents Chemother* 2001;45:1151–1161.
- Lascols C, Hackel M, Marshall SH, *et al.* Increasing prevalence and dissemination of NDM-1 metallo-β-lactamase in India: data from the SMART study (2009). *J Antimicrob Chemother* 2011;66:1992–1997.
- 9. Codjoe F, Donkor E. Carbapenem resistance: a review. Med Sci 2017;6(1):1.
- Nordmann P, Naas T, Poirel L. Global spread of carbapenemase-producing Enterobacteriaceae. *Emerg Infect Dis* 2011;17:1791–1798.
- 11. Schwaber MJ, Lev B, Israeli A, *et al.* Containment of a country-wide outbreak of carbapenem-resistant *Klebsiella pneumoniae* in Israeli hospitals via a nationally implemented intervention. *Clin Infect Dis* 2011;52:848–855.
- Tängdén T, Giske CG. Global dissemination of extensively drug-resistant carbapenemase-producing Enterobacteriaceae: clinical perspectives on detection, treatment and infection control. J Intern Med 2015;277:501–512.
- Ben-David D, Maor Y, Keller N, et al. Potential role of active surveillance in the control of a hospital-wide outbreak of carbapenem-resistant Klebsiella pneumoniae infection. Infect Control Hosp Epidemiol 2010;31:620–626.
- Guh AY, Bulens SN, Mu Y, *et al.* Epidemiology of carbapenem-resistant Enterobacteriaceae in 7 US communities, 2012–2013. *JAMA* 2015;314: 1479–1487.
- Centers for Disease Control and Prevention. Facility Guidance for Control of Carbapenem-Resistant Enterobacteriaceae (CRE) Update-CRE Toolkit. Atlanta: CDC; 2015.
- Traub WH, Raymond EA, Linehan J. Identification of Enterobacteriaceae in the clinical microbiology laboratory. *Appl Microbiol* 1970;20:303–308.
- Performance Standards for Antimicrobial Susceptibility Testing. 22nd Ed. CLSI Supplement M100. Vol 32. Wayne, PA: Clinical Laboratory Standards Institute; 2012.
- Performance Standards for Antimicrobial Susceptibility Testing. 23rd Ed. CLSI Supplement M100. Wayne, PA: Clinical Laboratory Standards Institute; 2013.
- Performance Standards for Antimicrobial Susceptibility Testing. 24th Ed. CLSI Supplement M100. Wayne, PA: Clinical Laboratory Standards Institute; 2014.
- Performance Standards for Antimicrobial Susceptibility Testing. 25th Ed. CLSI Supplement M100. Wayne, PA: Clinical Laboratory Standards Institute; 2015.
- Performance Standards for Antimicrobial Susceptibility Testing. 26th Ed. CLSI Supplement M100. Wayne, PA: Clinical Laboratory Standards Institute; 2016.

- Performance Standards for Antimicrobial Susceptibility Testing. 27th Ed. CLSI Supplement M100. Wayne, PA: Clinical Laboratory Standards Institute; 2017.
- Pierce VM, Simner PJ, Lonsway DR, *et al.* Modified carbapenem inactivation method for phenotypic detection of carbapenemase production among Enterobacteriaceae. *J Clin Microbiol* 2017;55:2321–2333.
- 24. Ramana K, Rao R, Sharada C V, Kareem M, Reddy R, Ratna Mani M. Modified Hodge test: a useful and the low-cost phenotypic method for detection of carbapenemase producers in Enterobacteriaceae members. *J Nat Sci Biol Med.* 2013;4:346–348.
- 25. Mataseje LF, Boyd DA, Willey BM, *et al.* Plasmid comparison and molecular analysis of *Klebsiella pneumoniae* harbouring blakpc from New York City and Toronto. *J Antimicrob Chemother* 2011;66:1273–1277.
- Doyle D, Peirano G, Lascols C, Lloyd T, Church DL, Pitouta JDD. Laboratory detection of Enterobacteriaceae that produce carbapenemases. *J Clin Microbiol* 2012;50:3877–3880.
- Munoz-Price LS, Poirel L, Bonomo RA, et al. Clinical epidemiology of the global expansion of *Klebsiella pneumoniae* carbapenemases. *Lancet Infect Dis* 2013;13:785–796.
- Guh AY, Limbago BM, Kallen AJ. Epidemiology and prevention of carbapenem-resistant Enterobacteriaceae in the United States. *Expert Rev Anti Infect Ther* 2014;12:565–580.
- Tracking CRE. Centers for Disease Control and Prevention website. https://www.cdc.gov/hai/organisms/cre/trackingcre.html. Published 2017. Accessed February 10, 2020.
- 30. Frieden TR, Harold Jaffe DW, Cardo DM, et al. Morbidity and Mortality Weekly Report Centers for Disease Control and Prevention MMWR Editorial and Production Staff MMWR Editorial Board.
- 31. Patel TS, Nagel JL Clinical outcomes of enterobacteriaceae infections stratified by carbapenem MICs. J Clin Microbiol 2015;53:201–205.
- 32. Lazarovitch T, Amity K, Coyle JR, *et al.* The complex epidemiology of carbapenem-resistant *Enterobacter* infections: a multicenter descriptive analysis. *Infect Control Hosp Epidemiol.* 2015;36:1283–1291.
- 33. Van Duin D, Perez F, Rudin SD, et al. Surveillance of carbapenem-resistant Klebsiella pneumoniae: tracking molecular epidemiology and outcomes through a regional network. Antimicrob Agents Chemother 2014;58: 4035–4041.
- 34. van Duin D, Doi Y. The global epidemiology of carbapenemase-producing Enterobacteriaceae. *Virulence* 2017;8:460–469.
- Rasheed JK, Kitchel B, Zhu W, *et al.* New Delhi metallo-β-lactamase– producing enterobacteriaceae, United States. *Emerg Infect Dis* 2013;19: 870–878.
- 36. Livorsi DJ, Chorazy ML, Schweizer ML, et al. A systematic review of the epidemiology of carbapenem-resistant Enterobacteriaceae in the United States. Antimicrob Resist Infect Control 2018;7:55.
- Van der Bij AK, Pitout JDD. The role of international travel in the worldwide spread of multiresistant enterobacteriaceae. J Antimicrob Chemother 2012;67:2090–2100.
- Bush K. Proliferation and significance of clinically relevant β-lactamases. *Ann N Y Acad Sci* 2013;1277:84–90.
- Fontana L, Bonura E, Lyski Z, Messer W. The brief case: Klebsiella variicola identifying the misidentified. J Clin Microbiol 2019;57(1):e00825–18.