

Adulthood asthma as a consequence of childhood adversity: a systematic review of epigenetically affected genes

Yasemin Saygideger^{1,2} , Hakan Özkan³, Oya Baydar¹ and Ozge Yilmaz⁴

Original Article

Cite this article: Saygideger Y, Özkan H, Baydar O, and Yilmaz O. (2022) Adulthood asthma as a consequence of childhood adversity: a systematic review of epigenetically affected genes. *Journal of Developmental Origins of Health and Disease* **13**: 674–682. doi: [10.1017/S2040174422000083](https://doi.org/10.1017/S2040174422000083)

Received: 13 February 2021
Revised: 29 January 2022
Accepted: 4 February 2022
First published online: 8 March 2022

Keywords:

Asthma; epigenetics; child abuse; childhood adversity; asthma-related genes; inflammation

Address for correspondence:

Yasemin Saygideger, Department of Pulmonary, School of Medicine, Cukurova University, Adana, Turkey.
Email: ysaygideger@cu.edu.tr

Part of this work has been presented at Turkish Thoracic Society's 22nd Annual Congress in April 2019.

¹Department of Pulmonary, School of Medicine, Cukurova University, Adana, Turkey; ²Department of Translational Medicine, Graduate School of Health Sciences, Cukurova University, Adana, Turkey; ³Department of Pediatrics, Division of Neonatal Intensive Care, Metro Hospital, Adana, Turkey and ⁴Department of Pediatric Allergy and Immunology, School of Medicine, Celal Bayar University, Manisa, Turkey

Abstract

There is an accumulating data that shows relation between childhood adversity and vulnerability to chronic diseases as well as epigenetic influences that in turn give rise to these diseases. Asthma is one of the chronic diseases that is influenced from genetic regulation of the inflammatory biomolecules and therefore the hypothesis in this research was childhood adversity might have caused epigenetic differentiation in the asthma-related genes in the population who had childhood trauma. To test this hypothesis, the literature was systematically reviewed to extract epigenetically modified gene data of the adults who had childhood adversity, and affected genes were further evaluated for their association with asthma. PRISMA guidelines were adopted and PubMed and Google Scholar were included in the searched databases, to evaluate epigenetic modifications in asthma-related genes of physically, emotionally or sexually abused children. After retrieving a total of 5245 articles, 36 of them were included in the study. Several genes and pathways that may contribute to pathogenesis of asthma development, increased inflammation, or response to asthma treatment were found epigenetically affected by childhood traumas. Childhood adversity, causing epigenetic changes in DNA, may lead to asthma development or influence the course of the disease and therefore should be taken into account for the prolonged health consequences.

Introduction and aim

Chronic respiratory diseases, particularly asthma, is known to be regulated by cellular and immunologic responses to environmental and biological factors to varying degrees in different individuals. Prenatal in utero stress, microbiota at birth or various postnatal factors such as nutrition, infections, and second-hand smoke contribute to the development of these responses.^{1,2} Cumulative data also indicate a relation between early life adversities and chronic airway diseases. These studies not only focus on asthma, but also other chronic disorders including chronic obstructive pulmonary disease, cancer, and immune system disorders.^{3–10} They have evaluated different types of adversities ranging from sexual and physical abuse to gun violence and emotional abuse in children, and the outcome revealed that there was a strong correlation between childhood adversities and presence of adulthood chronic respiratory diseases (Table S1).

Child adversities are known as risk factors for psychosocial or mental disorders in different levels and found associated with addiction to smoke, alcohol, or drugs.¹¹ In this context, search for the epigenetic changes in acute and chronic stress including childhood adversities revealed that the methylation status of various genes changes due to these childhood conditions.¹² The proposed pathways regarding epigenetic changes suggest that stress activates hypothalamic pituitary adrenal axis (HPA axis) and this induce cortisol increase leading to direct or indirect regulation of immune system.

The hypothesis of this research was, epigenetic changes might lead to asthma in abused children. Therefore, we systematically screened the studies that listed significantly affected genes from epigenetic changes associated with child abuse and then further evaluated for the role of these genes in asthma pathogenesis.

Methods

We followed PRISMA Statement¹³ for review of the literature for childhood adversity and asthma in population-based researches. We used Cukurova University Library help to construct keywords as ("child abuse" OR "child adversity" OR "child sexual abuse" OR "child physical abuse" OR "child neglect" OR "child trauma") AND ("genetics" OR "epigenetics" OR (DNA

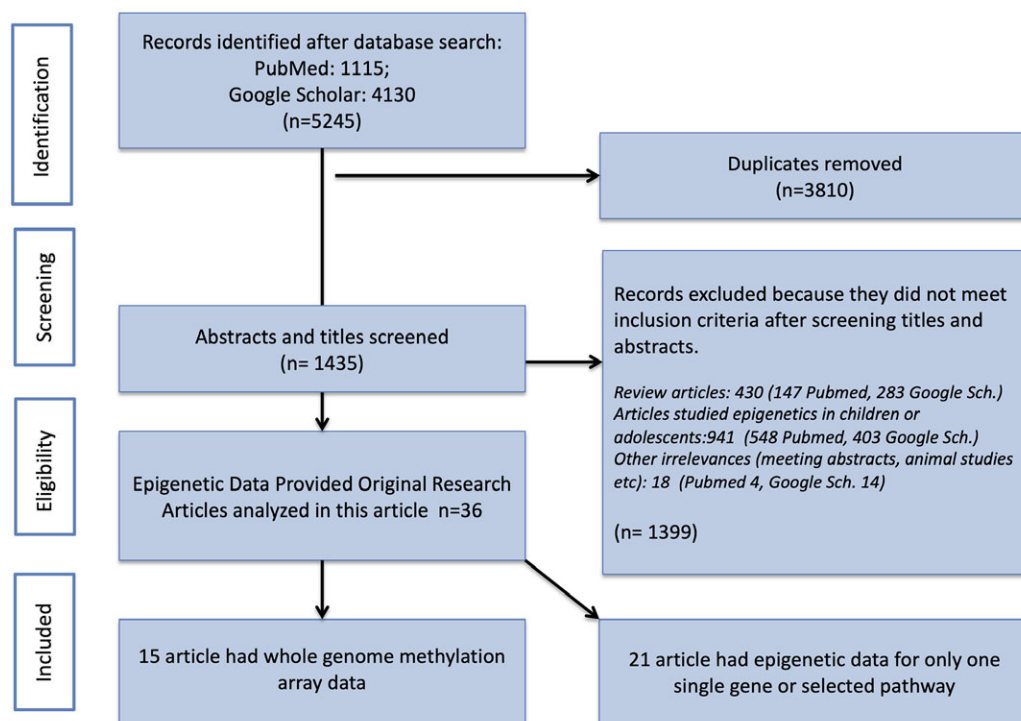


Fig. 1. Adopted PRISMA flowchart as of date 18 December 2019.

Methylation)) in NCBI-PubMed and Google Scholar to bring together the informative data that shows relation between childhood adversity and epigenetic changes. Only original research articles were included; review articles, preprint articles, articles involving adversities that occurred after age of 18 and articles that did not have the results of epigenetic analysis as well as the ones involving only economical adversities and pregnancy adversities were excluded. Initially, 5245 articles were retrieved, and majority of those articles were excluded after reviewing the abstracts and titles. Among the 36 articles that were identified after this first review, 15 had whole genome methylation arrays and 21 had methylation data for single genes or pathways, given in the adopted PRISMA flowchart (Fig. 1). Finally, we extracted significantly methylated genes from each study and evaluated the functions of these genes on *Gene Cards* and NCBI Gene web sites in order to identify the relations with asthma. We also searched PubMed to assess *in vivo*, *in vitro*, and clinical studies in relation with these differentially methylated genes and asthma. We then run a gene ontology (GO) analysis using an online tool¹⁴ and visualized the pathways related with the evaluated genes.

Results

After reviewing 36 full text articles, differentially methylated genes were recorded and analyzed for their role in asthma. The study sample sizes, methods, studied tissues, countries, and significantly differentially methylated genes and their possible contribution to asthma is listed in Table 1 for 15 whole genome analysis or microarray studies. Twenty-one single gene or pathway studies are excluded from the main list due to possible bias and listed in Table S2. All studies were performed in developed countries including USA, Canada, UK, Netherlands, and so forth, and they have extracted DNAs were from whole blood, saliva, T cells, and

monocytes. There was at least one affected gene related to asthma in each study while some of the studies showed multiple pathways involved in asthma pathogenesis (Table 1). Single gene or pathway studies mostly had data for post-traumatic stress disorder genes and focused on HPA axis and immune system-related genes, therefore, the genes that regulate HPA axis, *NR3C1* and *FKBP5*, were the most well-studied genes in the adults that had childhood adversities (Table S2). *NR3C1* and *FKBP5* are also involved in asthma pathogenesis by decreased suppression of inflammation and decrease response to corticosteroids, respectively. Other significantly deregulated genes that were affected by childhood adversities were interleukins, ADAM family, Wingless type-Integration family (WNT), STAT, and mitogene-activated protein kinase (MAPK) pathway and inflammatory-related genes (Table 1). Only one recent study had found no significant relation between childhood adversity and epigenetic regulation after applying multiple statistical corrections.²⁵ The total number of affected genes were

NR3C1 and asthma

The *NR3C1* gene encodes glucocorticoid receptor (GR), which has DNA binding, nuclear localization, ligand binding, and two activation function domains, and has the ability to inhibit the expression of asthma-related cytokines such as IL-5.²⁹ IL-5 induces the activity of eosinophils in asthma, after being released from the active macrophages and T lymphocytes. IL-5-induced leukotrienes, therefore, causes asthmatic reactions and symptoms. In a recent bioinformatics study that evaluated ignored genes that might have a function in allergic asthma, *NR3C1* was found to be downregulated.³⁰ It has also been shown that polymorphisms and mutations in specific sites of this gene has correlation with increase in transforming growth factor (TGF)- β expression in asthma patients,³¹ which is a

Table 1. Properties of 15 studies of childhood adversity and possible effects of differentially methylated genes in asthma

Article	Definition of Child Maltreatment	Sample size	Country	Age at the time of DNA sampling (Mean or range)	Tissue	Study design	Significantly differently methylated asthma related genes	Possible outcome for asthma
Yang et al. ¹²	Reported case records	96 cases 96 control	USA	10,2 (5–14)	Saliva	Whole genome methylation arrays	FANK-1 WNT pathway IL12B	Increased cell differentiation in the airway epithelium, increased inflammation
Mehta et al. ¹⁵	Childhood trauma questionnaire	32 cases 29 controls	USA	39,56 (cases) 43.69 (controls)	Blood	whole genome methylation arrays	IL5RA IL8 ADAM10 ADAM17	Increased inflammation, increased trans-endothelial leukocyte migration, epithelial permeability
Provençal et al. ¹⁶	Self-reported and criminal record	8 cases 12 controls	Canada	25,8 (cases) 25.2 (controls)	Blood-T cells and monocytes	Methylated DNA Immunoprecipitation + Microarray for selected cytokines and related transcription factors	transcription factors of cytokines: NF-KB NFAT5 STAT6	Increased inflammatory responses
Guiliemin et al. ¹⁷	Teacher assessment	5 female 12 male cases 19 female 25 male controls	Canada	25 (Cases) 24,19 (controls)	Blood-T cells	Methylated DNA Immunoprecipitation + Microarray (20,000 genes and 400 miRNAs)	Out of 448 differently methylated genes, Inflammatory response genes found to be significantly effected	Increased inflammation and increased risk for respiratory diseases
Provençal et al. ¹⁸	Self-reported and criminal record	8 cases 12 controls	Canada	25,8 (cases) 25.4 (controls)	Blood-T cells	Methylated DNA Immunoprecipitation + Microarray (20,000 genes and 400 miRNAs)	IL10 Signaling pathway genes Cytokine signaling between immune cells, Leukocyte extravasation signaling, MAPK signaling, etc.	Increased inflammation, increased trans-endothelial leukocyte migration, epithelial permeability
Suderman et al. ¹⁹	Confidential questionnaire (self-reported)	12 cases 28 controls	GBR	45 (all cases and controls)	Blood	Methylated DNA Immunoprecipitation + Microarray (20,533 genes and 489 miRNAs) and qPCR for selected genes	WNT signaling pathway	Progression of inflammation in asthma
Labonte et al. ²⁰	Childhood abuse interviews	25 cases 16 controls	Canada	37,3 (cases) 40,9 (controls)	Brain	Whole genome methylation arrays	RGS3 HCK	Abolished T-cell migration, inability of the cells to undergo the cytoskeletal reorganization required for mediator release
Schwaiger et al. ²¹	Childhood trauma questionnaire	30 cases 30 controls	Germany	52,57 (cases) 51,47 (controls)	Blood-Monocytes	Whole genome methylation arrays	NFKB STAT1 CREB	Impaired regulation of cytokine activity, abnormal allergic responses
Cicchetti et al. ²²	Child protective services	298 cases 250 controls	USA	9,4 (SD:0.88)	Saliva	SNP genotyping, and methylation assays for selected regions	ALDH2 NR3C1	Premature lung agigng, decreased suppression of inflammation
Cecil et al. ²³	Childhood trauma questionnaire	84 cases 40 controls	England	16–24 years	Buccal epithelial cells	Whole genome methylation arrays	RPTOR	Impaired inflammation and cell regulation
Kaufman et al. ²⁴	Recruited from a reported maltreated children cohort	122 cases 112 controls	USA	11,5 (SD: 1,9)	Saliva	Whole genome methylation arrays	<i>CXCL10</i>	Impaired eosinophil activation, increased asthma exacerbation

Table 1. (Continued)

	Childhood trauma questionnaire	1669 samples (from 2232 twins)	USA	18	Blood	Whole genome methylation arrays	No significance after corrections	n/a
Marzi et al. ²⁵	Childhood trauma questionnaire	1669 samples (from 2232 twins)	USA	18	Blood	Whole genome methylation arrays	No significance after corrections	n/a
Hautepen et al. ²⁶	Confidential questionnaire and self-reported	780/552 two cohorts	UK	47 (cases) 53 (controls)	Buccal cells Blood	Epigenome wide analysis (EWAS)	TOMSL (IKappabR) NPY	Regulation of the expression of inflammatory mediators Early onset asthma
Marinova et al. ²⁷	Childhood trauma questionnaire	30 cases 15 controls	Switzerland	75,9 (cases) 72,8 (controls)	Buccal	Whole genome methylation arrays	MTOR ATP2A3 ZC3H12D PLXNB1 ROBO1	Increased inflammation, increased risk for obesity related asthma, increased intracellular calcium Lymphocyte activation
Khulan et al. ²⁸	Childhood trauma questionnaire	83 cases 83 controls	Finland	64 (cases) 62,9 (controls)	Blood	Whole genome methylation arrays	FOXP4 CADM1	Airway hyperactivity Airway smooth muscle proliferation

well-known mediator that leads smooth muscle proliferation and airway remodeling. Regarding the epigenetic regulation of *NR3C1*, maternal stress during pregnancy, caused increased methylation in the umbilical cord blood mononuclear cells that cause decreased GR expression in the child, which may affect cytokine production and treatment response in the childhood asthma.³²

FKBP5 and asthma

FKBP5 or FK506-binding protein 51, is a member of immunophilin protein family that has peptidyl-prolyl isomerases to catalyze isomeric shape of proline amino acid and thus, act as co-chaperon to assist protein folding in various proteins. Increased levels of protein is found in central and peripheral airway brushings of severe asthma patients comparing to healthy subjects.³³ It also serves as a receptor for steroids and immune suppressive drugs and therefore, mostly studied for the effects of treatment response in asthma. FKBP5 express an inhibitory effect on GR function by binding to glucocorticoid-GR complex and delays nuclear translocation of the signal.³⁴ There are also other roles of this protein in different pathways related with asthma pathogenesis. Increased levels of FKBP5 induce T-cell proliferation and function by binding to calcineurin and activating nuclear factor of activated T-cells pathway.³⁴ Besides, stress induced decrease in *FKBP5* methylation, upregulated the protein levels in blood and immune cells and this increase in FKBP5 promotes NF- κ B (nuclear factor kappa-light-chain enhancer of activated B cells) mediated inflammation.³⁵ Thus, reduced methylation of FKBP5 in the abused children, might play role in the asthma development.

FANK1 and asthma

The relationship of FANK1 (Fibronectin type 3 and ankyrin repeat domains protein 1) with asthma is not well studied. There are studies that suggest the role of FANK1 in inhibition or activation of apoptosis. It is known to be expressed mostly by testis cells, but recently shown to be present also in T cells, alveolar, and epithelial cells of bronchi.³⁶ In a computational analysis of asthma-related genes, *FANK1* has come forward as one of the 10 genes found to be associated with asthma.³⁷ However, the functional relation currently remains unknown.

CHRNA5 and asthma

CHRNA5 (Neuronal acetylcholine receptor subunit alpha-5), is a cholinergic nicotinic receptor gene that acts in opening ion channels on the plasma membrane after acetylcholine binding. Expression of the gene is mostly found in airway myocytes and epithelial cells throughout the lungs.³⁸ In a meta-analysis that evaluate chromosome 15q25 region and airflow obstruction, Asp389Asn missense single nucleotide polymorphism in *CHRNA5* was associated with airflow obstruction in never smokers.³⁸ Silencing this gene in three-dimensional cell culture model, increased thickness of the epithelium, suggesting the lead to airway remodeling.³⁹ Studies show increased dependency to cigarette smoking in men^{40,41} but there is no data that shows relation with epigenetic regulation of *CHRNA5* and asthma but since genetically modification of the gene contributes airflow obstruction, and in vitro knock-down of the gene causes increased thickness in the airway models, it is possible that the increased methylation might contribute to asthma phenotype in the abused child.

ADAMs 10 and 17 and asthma

ADAM (A disintegrin and metalloproteinase domain containing protein) family proteins act as sheddase, with its adhesion and protease function, and shed various membrane proteins at the outer cell membrane that cause the maturation of those proteins and/or releasing them from the membrane. A single ADAM protein can cleave more than one protein as well as different ADAM members may cleave the same substrate. ADAM10 and ADAM17 have the similar active sites that both of them can cleave membrane bound TNF- α to its mature dissolvable state. ADAM10 has ability to shed ephrin/eph complexes between the cells, helps releasing soluble forms of IL6 and IL11 by mediating cleavage of IL6R and IL11RA, and also other surface proteins including heparin binding epidermal growth factor. ADAM10 also serves as a receptor for *S. aureus*, increasing the virulence of the bacteria. ADAM17 also cleaves IL6R, IL1RII, TGF- α , growth hormone receptor.⁴² Obviously, there are other many proteins and pathways effected by these two ADAMs that are not listed here. Thus, ADAM 10 and 17 playing role in endothelial permeability, smooth muscle transactivation, leukocyte recruitment, preventing resolution of inflammation and inducing inflammation, are important proteins in asthma pathogenesis and might be target for the treatment of the disease. Loss of function mutations of *ADAM10* and *ADAM17* are extremely rare and mice with double knockout of these genes do not survive.⁴² Overall, epigenetic regulation of these genes might affect asthma development in children.

SLC6A4 (SERT) and asthma

SLC6A4 gene, encodes sodium-dependent serotonin transporter and solute carrier family 6-member 4 (also SERT or 5HT transporter) protein that involves in recycling of serotonin (5-hydroxy tryptamine, 5HT) molecules by transporting them from the synaptic area to the presynaptic membrane. 5HT, besides its functions in the central nervous system, has ability to induce cytokine and chemokine synthesis, cell proliferation and tissue regeneration,⁴³ therefore, it is well studied in the pathogenesis of asthma. Increased levels of serotonin is found in symptomatic asthmatic patients and negatively correlated with FEV1 levels.⁴³ 5HT, stimulates PGE2 production by alveolar macrophages, activates eosinophils, and suppresses IL-12 expression.⁴³ 5HT also act as a platelet-stored vasoconstrictor. Termination of the serotonin signal depends on the levels of *SLC6A4* and activation of the receptor helps relief of asthma symptoms to release.⁴⁴ There are studies that evaluated the relationship between *SLC6A4* gene polymorphisms and asthma but have not found any correlations up to now with the studied ones.⁴⁵ *SLC6A4* knockout mice show anxiety-like behavior and changes in the methylation of the gene may contribute the levels of 5HT and therefore influence asthma genotype in the maltreated and affected children.

OXTR and asthma

OXTR gene codes for oxytocin receptor, which is a G protein coupled receptor that needs to activate phosphatidylinositol-calcium second messenger molecules. Studies have shown that *OXTR* is expressed both in human and mice airway smooth muscle cells and its expression is affected by cytokines such as IL-13 and TNF- α .⁴⁶ Oxytocine levels in BAL fluids were also found increased in asthmatic patients comparing to healthy control

subjects⁴⁶ suggesting that *OXTR* and its epigenetic regulation might have a role in the pathogenesis of asthma.

RPTOR, MTOR, and asthma

RPTOR or raptor (Regulatory-associated protein of mTOR), regulates mammalian target of rapamycin complex 1 (mTORC1) activity which involves in cell growth, survival, and autophagy. It also has role in maintaining the size of the cell. It is mostly expressed in muscle cells including lungs. In a study that evaluate airway smooth muscle tissue with RNA sequencing to compare asthmatic and normal people, RPTOR was found one of the four genes which were associated with airway hyper-responsiveness.⁴⁷ mTOR (mechanistic target of rapamycin kinase), was also found to involve in airway remodeling models in mouse models as well as regulation of Th17/Treg and Th1/Th2 ratio.⁴⁸ Therefore, epigenetic regulation of mTOR and related pathways significantly influence asthma development.

RGS3 and HCK and asthma

RGS3 involves in inflammation and Tcell migration,⁴⁹ and HCK takes plays in MAPK signaling pathway which in turn related with airway smooth muscle proliferation.⁵⁰ Labonte et al found these two genes were significantly hypermethylated in the brain tissue of the traumatic children, but currently there is no data that shows the epigenetic change in the blood or other tissues in these two genes.

CXCL10 and asthma

C-X-C motif chemokine ligand 10 (IFN- γ -induced protein 10 (IP-10) or small inducible cytokine B10) is a small protein coded by *CXCL10* gene. It is expressed in monocytes, endothelial cells, and fibroblasts and has a variety of role in the inflammation. Therefore, the effects on the asthma pathogenesis is well studied. mRNA levels of *CXCL10* were found increased in bronchoalveolar fluid of asthmatic patients and data suggest that it has role in corticosteroid therapy-induced persistent type I inflammation.⁵¹ Kaufman et al., evaluated epigenetic influences on the obesity in adverse childhood experienced children and found decreased methylation of the promoter region of *CXCL10*. Therefore, inducing obesity or involving inflammation, epigenetic regulation of the gene may contribute to asthma development in the abused children.

STAT and asthma

STAT family of proteins (signal transducer and activator of transcription) act as transcription factors after extracellular cytokine or growth factor binding in the cell and lead to the expression of proliferation, differentiation, and apoptosis-related genes. *STAT1* was reported one of the asthma-ignorome gene together with *NR3C1*³⁰ and variants of *STAT6* found related with risk for asthma in a meta-analysis study.⁵² Epigenetic regulation of Jak-STAT pathway in childhood asthma has been recently reported.⁴¹

CREB and asthma

CREB (cAMP response element-binding protein) is another transcription factor that regulate expression or suppression of multiple genes after getting activated by upstream signaling pathways those use cAMP as secondary messenger. Therefore, it involves multiple pathways in Asthma signaling including therapy response.⁵³

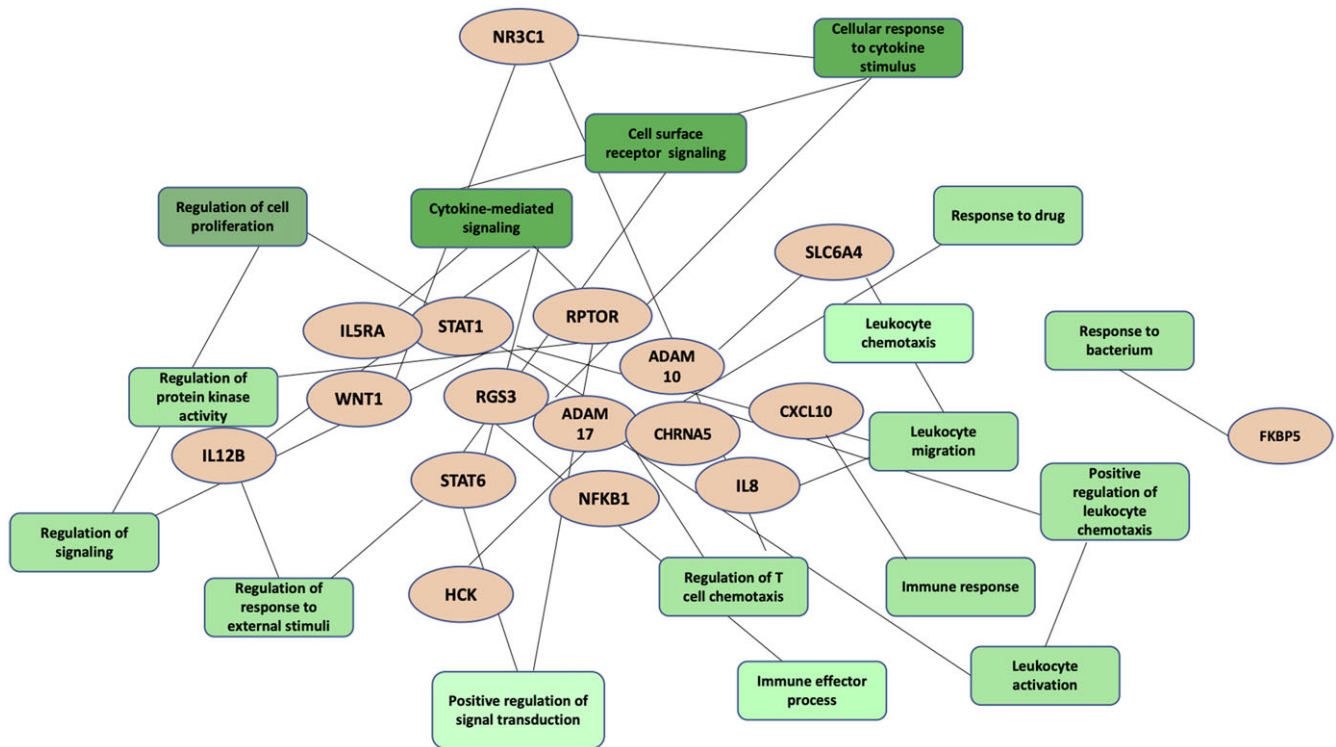


Fig. 2. Gene enrichment and pathway analysis of the affected genes. Differentially methylated genes in the adults who had childhood adversity, that have relationship with asthma are shown in orange color and significantly related pathways are in green. Dark green shows strongest relation. The full list of the pathways and corresponding full image are given in supplementary material.

NFKB, IKappaBR, and asthma

NF- κ B (nuclear factor kappa-light-chain enhancer of activated B cells), consists of protein to regulate transcription, cytokine production, and survival when the cell is under stress. Together with MAPK pathway, epigenetic regulation of NF- κ B signaling was found significantly affected in adult-onset asthma.⁵⁴ Researchers have been interested in targeting NF- κ B for asthma treatment for the last years. *IKappaBR* (*IKBR*) or *TONSL* gene codes for Tonsoku like DNA Repair protein that negatively regulate NFKB mediated transcription. Therefore, increased methylation of *IKBR* and demethylation of *NFKB* would worsen asthma phenotype in the maltreated children.

ALDH2 and asthma

ALDH2 (acetaldehyde dehydrogenase 2) is a member of ALDH family of proteins that acts as an antioxidant and its increased expression inhibits the harmful effects of oxidative stress. Polymorphisms of the gene may cause premature lung aging⁵⁵ and ALDH2-deficient mice had alcohol-induced asthma in an animal model study.⁵⁶ These and several other studies indicate that increased methylation of *ALDH2* may have role in the asthma pathogenesis.

MAPK pathway, NPY – FOXP4, and asthma

MAPK pathway is a series of signaling cascade pathways that induce distinct pathways in the cell regarding differentiation, proliferation, and cell survival. Its role in asthma pathogenesis has been shown and reviewed in the literature.⁵⁷

NPY (Neuropeptid Y) involves in multiple signaling pathways including MAPK pathway, and it also regulates intracellular

calcium levels. The expression levels of NPY on antigen presenting cells in allergic asthma has been shown, and its role on cytokine expression is not yet clear.⁵⁸ In a study, the loss of FOXP4 (Forkhead box protein 4) together with FOXP1, which are transcription factors located in airway cells, led to increased NYP expression resulting in airway hyperreactivity through activation of smooth muscle myosin light-chain phosphorylation.⁵⁹

ATP2A3 and asthma

ATP2A3 (Sarcoplasmic/Endoplasmic Reticulum Calcium ATPase 3), involving intracellular calcium levels, influence a variety of signaling pathways. the expression was shown to vary among different childhood asthma phenotypes.⁶⁰

Other relevant genes and gene ontology and pathway analysis

The rest of the genes listed in Table 1 are usually found in the literature that assessed obesity asthma relationship or that took place in the gene expression pathways in immune system.

After reviewing these genes in the literature, we further ran a GO analysis using two different online tools^{14,61} and visualized the pathways related with the evaluated genes (Fig. 2 and Fig. S1 and Table S3).

Conclusion and discussion

In this research, we demonstrated that childhood adversity, including sexual and physical abuse and neglect, effects the epigenetic regulation of the genes that are shown to play role in asthma pathogenesis, as a potential risk factor of the adulthood asthma.

These epigenetic changes focused on altered methylation of DNA, which has become convenient to study comparing to chromatin purification.⁶² The increased or decreased methylation of the genes found in the adults, who experienced adverse childhood events, were included well-studied asthma-related genes such as *ADAM10* & 17 and *CXCL10* as well as the genes that seemed less common in asthma pathogenesis regarded studies such as *ALDH2*, *FOXP4*, and *FANK1*. We would like to remind that the list of “significantly changed” genes might have caused a bias in the generation of our table in this article, which the cutoff points of each study were different, especially in the whole genome array-based studies. This might have led the ignorance of many other affected genes that may play role in asthma pathogenesis.

Up-to-date scientific investigations of asthma pathogenesis indicate a multi-complex pathophysiological process that might be responsible of the diverse phenotypes among asthma patients, and recent studies indicate a contribution of lifelong epigenetic influences to this pathophysiology.⁶³ The complexity increases while the type of the adversity and its relationship with asthma may differ from population to population, related to cultures and traditions, and therefore, one of the limitations of this systematic review is that the origin of most studies included the countries with similar socioeconomic background, decreasing the value of generalizability. This study also did not include in utero period of the children, which may also contribute to epigenetic alterations. This systematic review included Pubmed and Google Academic search due to their common usage in the field and therefore, studies in other databases might have been excluded due to this limitation. Although the genes listed in this study have been associated with asthma and a few of them are well studied, the underlying pathway for these associations may need deeper research for future discoveries of personalized medicine and to understand the pathogenesis of the relationship of these epigenetic influences with asthma.

Supplementary materials. For supplementary material for this article, please visit <https://doi.org/10.1017/S2040174422000083>

Acknowledgements. We thank to Cukurova University Library for helping to build the key words for database search.

Author contributions. YS designed the study analyzed the results and wrote the manuscript; HO and OB involved in literature search and data collection; OY critically reviewed the manuscript.

Financial support. None.

Conflict of interest. All authors declare no support, financial or otherwise, from any organization for the submitted work.

Ethical standards. The article contains only publicly available data and ethical approval is not applicable.

References

- Carraro S, Scheltema N, Bont L, Baraldi E. Early-life origins of chronic respiratory diseases: understanding and promoting healthy ageing. *Eur Respir J*. 2014; 44(6), 1682–1696. DOI [10.1183/09031936.00084114](https://doi.org/10.1183/09031936.00084114).
- Zhang Y, Kutateladze TG. Diet and the epigenome. *Nat Commun*. 2018; 9(1), 2376. DOI [10.1038/s41467-018-05778-1](https://doi.org/10.1038/s41467-018-05778-1).
- Shields ME, Hovdestad WE, Pelletier C, Dykxhoorn JL, O'Donnell SC, Tonmyr L. Childhood maltreatment as a risk factor for diabetes: findings from a population-based survey of Canadian adults. *BMC Public Health*. 2016; 16(1). DOI [10.1186/s12889-016-3491-1](https://doi.org/10.1186/s12889-016-3491-1).
- Bhan N, Glymour MM, Kawachi I, Subramanian SV. Childhood adversity and asthma prevalence: evidence from 10 US states (2009–2011). *BMJ Open Respir Res*. 2014; 1(1), e000016. DOI [10.1136/bmjresp-2013-000016](https://doi.org/10.1136/bmjresp-2013-000016).
- Ayaydin H, Abali O, Okumus Akdeniz N, et al. Immune system changes after sexual abuse in adolescents. *Pediatr Int*. 2016; 58(2), 105–112. DOI [10.1111/ped.12767](https://doi.org/10.1111/ped.12767).
- Remigio-Baker RA, Hayes DK, Reyes-Salvail F. Adverse childhood events are related to the prevalence of asthma and chronic obstructive pulmonary disorder among adult women in Hawaii. *Lung*. 2015; 193(6), 885–891. DOI [10.1007/s00408-015-9777-8](https://doi.org/10.1007/s00408-015-9777-8).
- Clemens V, Huber-Lang M, Plener PL, Brähler E, Brown RC, Fegert JM. Association of child maltreatment subtypes and long-term physical health in a German representative sample. *Eur J Psychotraumatol*. 2018; 9(1), 1510278. DOI [10.1080/20008198.2018.1510278](https://doi.org/10.1080/20008198.2018.1510278).
- Rosas-Salazar C, Han Y-Y, Brehm JM, et al. Gun violence, African ancestry, and asthma. *Chest*. 2016; 149(6), 1436–1444. DOI [10.1016/j.chest.2016.02.639](https://doi.org/10.1016/j.chest.2016.02.639).
- Bonfim CB, dos Santos DN, Barreto ML. The association of intrafamilial violence against children with symptoms of atopic and non-atopic asthma: a cross-sectional study in Salvador. *Brazil Child Abuse Neglect*. 2015; 50(2), 244–253. DOI [10.1016/j.chiabu.2015.05.021](https://doi.org/10.1016/j.chiabu.2015.05.021).
- Bellis MA, Hughes K, Leckenby N, Hardcastle KA, Perkins C, Lowey H. Measuring mortality and the burden of adult disease associated with adverse childhood experiences in England: a national survey. *J Public Health*. 2014; 37(3), 445–454. DOI [10.1093/pubmed/dfu065](https://doi.org/10.1093/pubmed/dfu065).
- Sinha R. Chronic stress, drug use, and vulnerability to addiction. *Ann NY Acad Sci*. 2008; 1141(1), 105–130. DOI [10.1196/annals.1441.030](https://doi.org/10.1196/annals.1441.030).
- Yang B-Z, Zhang H, Ge W, et al. Child abuse and epigenetic mechanisms of disease risk. *Am J Prev Med*. 2013; 44(2), 101–107. DOI [10.1016/j.amepre.2012.10.012](https://doi.org/10.1016/j.amepre.2012.10.012).
- Moher D, Liberati A, Tetzlaff J, Altman DG, PRISMA Group*. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *Ann Intern Med*. 2009; 151(4), 264–269.
- Pomaznoy M, Ha B, Peters B. GONet: a tool for interactive gene ontology analysis. *BMC Bioinform*. 2018; 19(1), 470. DOI [10.1186/s12859-018-2533-3](https://doi.org/10.1186/s12859-018-2533-3).
- Mehta D, Klengel T, Conneely KN, et al. Childhood maltreatment is associated with distinct genomic and epigenetic profiles in posttraumatic stress disorder. *Proc Natl Acad Sci USA*. 2013; 110(20), 8302–8307. DOI [10.1073/pnas.1217750110](https://doi.org/10.1073/pnas.1217750110).
- Provençal N, Suderman MJ, Caramaschi D, et al. Differential DNA methylation regions in cytokine and transcription factor genomic loci associate with childhood physical aggression. *PLoS ONE*. 2013; 8(8), e71691. DOI [10.1371/journal.pone.0071691](https://doi.org/10.1371/journal.pone.0071691).
- Guillemin C, Provençal N, Suderman M, et al. DNA methylation signature of childhood chronic physical aggression in T cells of both men and women. *PLoS ONE*. 2014; 9(1), e86822. DOI [10.1371/journal.pone.0086822](https://doi.org/10.1371/journal.pone.0086822).
- Provençal N, Suderman MJ, Guillemin C, et al. Association of childhood chronic physical aggression with a DNA methylation signature in adult human T cells. *PLoS ONE*. 2014; 9(4), e89839. DOI [10.1371/journal.pone.0089839](https://doi.org/10.1371/journal.pone.0089839).
- Suderman M, Borghol N, Pappas JJ, et al. Childhood abuse is associated with methylation of multiple loci in adult DNA. *BMC Med Genom*. 2014; 7(1), 68. DOI [10.1186/1755-8794-7-13](https://doi.org/10.1186/1755-8794-7-13).
- Labonté B, Turecki G. Epigenetic effects of childhood adversity in the brain and suicide risk (Chapter 13). In *The Neurobiological Basis of Suicide* (ed. Dwivedi Y), 2012, CRC Press/Taylor & Francis, Boca Raton, FL.
- Schwaiger M, Grinberg M, Moser D, et al. Altered stress-induced regulation of genes in monocytes in adults with a history of childhood adversity. *Neuropsychopharmacol*. 2016; 41(10), 2530–2540. DOI [10.1038/npp.2016.57](https://doi.org/10.1038/npp.2016.57).
- Cicchetti D, Hetzel S, Rogosch FA, Handley ED, Toth SL. An investigation of child maltreatment and epigenetic mechanisms of mental and physical health risk. *Dev Psychopathol*. 2016; 28(4pt2), 1305–1317. DOI [10.1017/S0954579416000869](https://doi.org/10.1017/S0954579416000869).

23. Cecil CAM, Smith RG, Walton E, Mill J, McCrory EJ, Viding E. Epigenetic signatures of childhood abuse and neglect: implications for psychiatric vulnerability. *J Psychiatr Res.* 2016; 83(7), 184–194. DOI [10.1016/j.jpsychires.2016.09.010](https://doi.org/10.1016/j.jpsychires.2016.09.010).
24. Kaufman J, Montalvo-Ortiz JL, Holbrook H, et al. Adverse childhood experiences, epigenetic measures, and obesity in youth. *J Pediatr.* 2018; 202, 150–156.e3. DOI [10.1016/j.jpeds.2018.06.051](https://doi.org/10.1016/j.jpeds.2018.06.051).
25. Marzi SJ, Sugden K, Arseneault L, et al. Analysis of DNA methylation in young people: limited evidence for an association between victimization stress and epigenetic variation in blood. *Am J Psychiatry.* 2018; 175(6), 517–529. DOI [10.1176/appi.ajp.2017.17060693](https://doi.org/10.1176/appi.ajp.2017.17060693).
26. Houtepen LC, Hardy R, Maddock J, et al. Childhood adversity and DNA methylation in two population-based cohorts. *Transl Psychiatry.* 2018; 8(1), 378. DOI [10.1038/s41398-018-0307-3](https://doi.org/10.1038/s41398-018-0307-3).
27. Marinova Z, Maercker A, Küffer A, et al. DNA methylation profiles of elderly individuals subjected to indentured childhood labor and trauma. *BMC Med Genet.* 2017; 18(1), 608. DOI [10.1186/s12881-017-0370-2](https://doi.org/10.1186/s12881-017-0370-2).
28. Khulan B, Manning JR, Dunbar DR, et al. Epigenomic profiling of men exposed to early-life stress reveals DNA methylation differences in association with current mental state. *Transl Psychiatry.* 2014; 4(9), e448. DOI [10.1038/tp.2014.94](https://doi.org/10.1038/tp.2014.94).
29. Panek M, Jonakowski M, ZioŁo J, et al. A novel approach to understanding the role of polymorphic forms of the NR3C1 and TGF-β1 genes in the modulation of the expression of IL-5 and IL-15 mRNA in asthmatic inflammation. *Mol Med Rep.* 2016; 13(6), 4879–4887. DOI [10.3892/mmr.2016.5104](https://doi.org/10.3892/mmr.2016.5104).
30. Riba M, Garcia Manteiga JM, Bošnjak B, et al. Revealing the acute asthma ignorome: characterization and validation of uninvestigated gene networks. *Sci Rep.* 2016; 6(1), 2055. DOI [10.1038/srep24647](https://doi.org/10.1038/srep24647).
31. Panek M, Pietras T, Fabijan A, et al. The NR3C1 glucocorticoid receptor gene polymorphisms may modulate the TGF-beta mRNA expression in asthma patients. *Inflammation.* 2015; 38(4), 1479–1492. DOI [10.1007/s10753-015-0123-3](https://doi.org/10.1007/s10753-015-0123-3).
32. Al-Hussainy A, Mohammed R. Consequences of maternal psychological stress during pregnancy for the risk of asthma in the offspring. *Scand J Immunol.* 2021; 93(1), e12919.
33. Singhania A, Rupani H, Jayasekera N, et al. Altered epithelial gene expression in peripheral airways of severe asthma. *PLoS ONE.* 2017; 12(1), e0168680. DOI [10.1371/journal.pone.0168680](https://doi.org/10.1371/journal.pone.0168680).
34. Zannas AS, Wiechmann T, Gassen NC, Binder EB. Gene-stress-epigenetic regulation of FKBP5: clinical and translational implications. *Neuropsychopharmacology.* 2015; 41(1), 261–274. DOI [10.1038/npp.2015.235](https://doi.org/10.1038/npp.2015.235).
35. Zannas AS, Jia M, Hafner K, et al. Epigenetic upregulation of FKBP5 by aging and stress contributes to NF-κB-driven inflammation and cardiovascular risk. *Proc Natl Acad Sci USA.* 2019; 116(23), 11370–11379. DOI [10.1073/pnas.1816847116](https://doi.org/10.1073/pnas.1816847116).
36. Wu C, Orozco C, Boyer J, et al. BioGPS: an extensible and customizable portal for querying and organizing gene annotation resources 2021. <http://biogps.org/#goto=genereport&id=92565>.
37. Tremblay K, Lemire M, Potvin C, et al. Genes to diseases (G2D) computational method to identify asthma candidate genes. *PLoS ONE.* 2008; 3(8), e2907. DOI [10.1371/journal.pone.0002907](https://doi.org/10.1371/journal.pone.0002907).
38. Wilk JB, Shrine NRG, Loehr LR, et al. Genome-wide association studies identify chrna5/3andHTR4 in the development of airflow obstruction. *Am J Respir Crit Care Med.* 2012; 186(7), 622–632. DOI [10.1164/rccm.201202-0366oc](https://doi.org/10.1164/rccm.201202-0366oc).
39. Kraiss AM, Hautefeuille AH, Cros M-P, et al. CHRNA5 as negative regulator of nicotine signaling in normal and cancer bronchial cells: effects on motility, migration and p63 expression. *Carcinogenesis.* 2011; 32(9), 1388–1395. DOI [10.1093/carcin/bgr090](https://doi.org/10.1093/carcin/bgr090).
40. Xie P, Kranzler HR, Zhang H, et al. Childhood adversity increases risk for nicotine dependence and interacts with α5 nicotinic acetylcholine receptor genotype specifically in males. *Neuropsychopharmacology.* 2011; 37(3), 669–676. DOI [10.1038/npp.2011.240](https://doi.org/10.1038/npp.2011.240).
41. Zhang N-Z, Chen X-J, Mu Y-H, Wang H. Identification of differentially expressed genes in childhood asthma. *Medicine.* 2018; 97(21), e10861. DOI [10.1097/md.00000000000010861](https://doi.org/10.1097/md.00000000000010861).
42. Dreymueller D, Uhlig S, Ludwig A. ADAM-family metalloproteinases in lung inflammation: potential therapeutic targets. *Am J Physiol Lung Cell Mol Physiol.* 2015; 308(4), L325–L343. DOI [10.1152/ajplung.00294.2014](https://doi.org/10.1152/ajplung.00294.2014).
43. Kang BN, Ha SG, Bahaie NS, et al. Regulation of serotonin-induced trafficking and migration of Eosinophils. *PLoS ONE.* 2013; 8(1), e54840. DOI [10.1371/journal.pone.0054840](https://doi.org/10.1371/journal.pone.0054840).
44. Flanagan TW, Sebastian MN, Battaglia DM, Foster TP, Cormier SA, Nichols CD. 5-HT2 receptor activation alleviates airway inflammation and structural remodeling in a chronic mouse asthma model. *Life Sci.* 2019; 236(Suppl 2), 116790. DOI [10.1016/j.lfs.2019.116790](https://doi.org/10.1016/j.lfs.2019.116790).
45. Farjadian S, Moghtaderi M, Fakhraei B, Nasiri M, Farjam M. Association between serotonin transporter gene polymorphisms and childhood asthma. *J Asthma.* 2013; 50(10), 1031–1035. DOI [10.3109/02770903.2013.834503](https://doi.org/10.3109/02770903.2013.834503).
46. Amrani Y, Syed F, Huang C, et al. Expression and activation of the oxytocin receptor in airway smooth muscle cells: regulation by TNFα and IL-13. *Respir Res.* 2010; 11(1), 629. DOI [10.1186/1465-9921-11-104](https://doi.org/10.1186/1465-9921-11-104).
47. Yick CY, Zwinderman AH, Kunst PW, et al. Gene expression profiling of laser microdissected airway smooth muscle tissue in asthma and atopy. *Allergy.* 2014; 69(9), 1233–1240. DOI [10.1111/all.12452](https://doi.org/10.1111/all.12452).
48. Zhang Y, Jing Y, Qiao J, et al. Activation of the mTOR signaling pathway is required for asthma onset. *Sci Rep.* 2017; 7(1), 143. DOI [10.1038/s41598-017-04826-y](https://doi.org/10.1038/s41598-017-04826-y).
49. Williams JW, Yau D, Sethakorn N, et al. RGS3 controls T lymphocyte migration in a model of Th2-mediated airway inflammation. *Am J Physiol Lung Cell Mol Physiol.* 2013; 305(10), L693–L701. DOI [10.1152/ajplung.00214.2013](https://doi.org/10.1152/ajplung.00214.2013).
50. Sakai H, Nishimura A, Watanabe Y, et al. Involvement of Src family kinase activation in angiotensin II-induced hyperresponsiveness of rat bronchial smooth muscle. *Peptides.* 2010; 31(12), 2216–2221. DOI [10.1016/j.peptides.2010.09.012](https://doi.org/10.1016/j.peptides.2010.09.012).
51. Gauthier M, Chakraborty K, Oriss TB, et al. Severe asthma in humans and mouse model suggests a CXCL10 signature underlies corticosteroid resistant Th1 bias. *JCI Insight.* 2017; 2(13), e94580. DOI [10.1172/jci.insight.94580](https://doi.org/10.1172/jci.insight.94580).
52. Qian X, Gao Y, Ye X, Lu M. Association of STAT6 variants with asthma risk: a systematic review and meta-analysis. *Hum Immunol.* 2014; 75(8), 847–853. DOI [10.1016/j.humimm.2014.06.007](https://doi.org/10.1016/j.humimm.2014.06.007).
53. Kim D, Cho S, Woo JA, Liggett SB. A CREB-mediated increase in miRNA let-7f during prolonged β-agonist exposure: a novel mechanism of β2-adrenergic receptor down-regulation in airway smooth muscle. *The FASEB J.* 2018; 32(7), 3680–3688. DOI [10.1096/fj.201701278r](https://doi.org/10.1096/fj.201701278r).
54. Jeong A, Imboden M, Ghantous A, et al. DNA methylation in inflammatory pathways modifies the association between BMI and adult-onset non-atopic asthma. *Int J Environ Res Public Health.* 2019; 16(4), 600. DOI [10.3390/ijerph16040600](https://doi.org/10.3390/ijerph16040600).
55. Kuroda A, Hegab AE, Jingtao G, et al. Effects of the common polymorphism in the human aldehyde dehydrogenase 2 (ALDH2) gene on the lung. *Respir Res.* 2017; 18(1), 583. DOI [10.1186/s12931-017-0554-5](https://doi.org/10.1186/s12931-017-0554-5).
56. Shimoda T, Obase Y, Matsuse H, Asai S, Iwanaga T. The pathogenesis of alcohol-induced airflow limitation in acetaldehyde dehydrogenase 2-deficient mice. *Int Arch Allergy Immunol.* 2016; 171(3-4), 276–284. DOI [10.1159/000452709](https://doi.org/10.1159/000452709).
57. Chung KF. p38 mitogen-activated protein kinase pathways in asthma and COPD. *Chest.* 2011; 139(6), 1470–1479. DOI [10.1378/chest.10-1914](https://doi.org/10.1378/chest.10-1914).
58. Makinde TO, Steininger R, Agrawal DK. NPY and NPY receptors in airway structural and inflammatory cells in allergic asthma. *Exp Mol Pathol.* 2013; 94(1), 45–50. DOI [10.1016/j.yexmp.2012.05.009](https://doi.org/10.1016/j.yexmp.2012.05.009).
59. Li S, Koziol-White C, Jude J, et al. Epithelium-generated neuropeptide Y induces smooth muscle contraction to promote airway hyperresponsiveness. *J Clin Invest.* 2016; 126(5), 1978–1982. DOI [10.1172/JCI81389](https://doi.org/10.1172/JCI81389).

60. Boeck A, Landgraf-Rauf K, Vogelsang V, *et al.* Ca^{2+} and innate immune pathways are activated and differentially expressed in childhood asthma phenotypes. *Pediatr Allergy Immunol.* 2018; 29(8), 823–833. DOI [10.1111/pai.12971](https://doi.org/10.1111/pai.12971).
61. Fabregat A, Sidiropoulos K, Viteri G, *et al.* Reactome pathway analysis: a high-performance in-memory approach. *BMC Bioinform.* 2017; 18(1), 383. DOI [10.1186/s12859-017-1559-2](https://doi.org/10.1186/s12859-017-1559-2).
62. DeVries A, Vercelli D. Epigenetic mechanisms in asthma. *Ann Am Thorac Soc.* 2016; 13(Suppl 1), S48–S50. DOI [10.1513/AnnalsATS.2015-07-420MG](https://doi.org/10.1513/AnnalsATS.2015-07-420MG).
63. Chogtu B, Bhattacharjee D, Magazine R. Epigenetics: the new frontier in the landscape of asthma. *Scientifica.* 2016; 2016(2), 4638949. DOI [10.1155/2016/4638949](https://doi.org/10.1155/2016/4638949).