British Journal of Nutrition

cambridge.org/bjn

Research Article

Cite this article: Obikawa K, Kitaguchi M, Kondo E, and Okamura K (2025). Relationship between the ratio of increase in lean tissue to body weight gain and energy required to gain body weight in growing rats. *British Journal of Nutrition*, page 1 of 9. doi: 10.1017/S000711452400326X

Received: 29 September 2023 Revised: 16 October 2024 Accepted: 3 December 2024

Keywords

Energy intake; Energy to store; Maintenance energy; Fat-free mass; Fat mass

Abbreviations:

AT, adipose tissue; BW, body weight; EX, energy expenditure; HF, high fat; HP, high protein; LT, lean tissue; ME, metabolisable energy; N, normal; PA, energy for physical activity; WG, energy for body weight gain

Corresponding author:

Koji Okamura;

Email: okamurakoji0002@gmail.com

© The Author(s), 2024. Published by Cambridge University Press on behalf of The Nutrition Society.





Relationship between the ratio of increase in lean tissue to body weight gain and energy required to gain body weight in growing rats

Kiyora Obikawa, Mizuki Kitaguchi, Emi Kondo 🗅 and Koji Okamura

Graduate School of Sport and Exercise Sciences, Osaka University of Health and Sport Sciences, 1-1 Asashirodai, Kumatori-cho, Sennan-gun, Osaka 590-0496, Japan

Abstract

Although the energy stored in the lean tissue (LT) and adipose tissue (AT) is well known, the energy required to synthesise these tissues is obscure. Theoretically, the energy at the point at which $\Delta LT/\Delta$ body weight (BW) reaches 100 % on a regression line, which indicates the relationship between $\Delta LT/\Delta BW$ and the energy required for BW gain, is considered to be the energy expended to synthesise LT. Therefore, we investigated this relationship in rats. Rats were fed diets with different ratios of protein, fat and carbohydrates because their $\Delta LT/\Delta BW$ values were expected to be different. Six-week-old male Sprague-Dawley rats had *ad libitum* access to normal (N, *n* 6), high-fat (HF, *n* 7) or high-protein (HP, *n* 8) diets for 4 weeks. The $\Delta LT/\Delta BW$ was 0-77 in the N, 0-70 in the HF and 0-87 in the HP groups, respectively. The average energy required to gain BW was 8-8 kJ/g in the N group, 7-0 kJ/g in the HF group and 11-3 kJ/g in the HP group. We observed a positive correlation between $\Delta LT/\Delta BW$ and energy required for BW gain. The regression line demonstrated that the energy expended to synthesise LT was 13-9 kJ/g and AT was -7.9 kJ/g. Therefore, combined with the energy stored in LT, the energy required to accumulate LT is approximately 19 kJ/g, whereas the energy to accumulate AT could not be elucidated.

Although the energy required to synthesise protein and fat has been demonstrated in rats⁽¹⁾ and humans^(2,3), the energy required to synthesise lean tissue (LT; e.g. skeletal muscle, internal organs and bone) and adipose tissue (AT; e.g. abdominal and subcutaneous adipose tissue) is obscure. Elucidation of the energy required for LT synthesis is important for individuals who aim to increase their skeletal muscle mass, such as athletes. Elucidating the energy required for AT synthesis may have implications for nutritional therapy for individuals such as anorexia nervosa and malnourished. In addition, AT not only stores fat but also has endocrine and immune functions⁽⁴⁾.

Spady *et al.* showed that the energy stored in the body is calculated by measuring the difference between energy intake (metabolisable energy, ME) and the energy expenditure $(EX)^{(5)}$. EX is the sum of the energy required to maintain the body (MT), for the synthesis of newly accumulated tissues and physical activity (PA). Thus, the energy required to synthesise newly accumulated tissue, that is the energy required for body weight (BW) gain (WG) is calculated by subtracting MT and PA from EX. During BW gain, energy is stored in the LT and AT. The energy stored in LT (Δ LT_E) and AT (Δ AT_E) are calculated by accreted LT (Δ LT) and AT (Δ AT) multiplied by their respective energy densities of 5·23 kJ/g LT and 30·96 kJ/g AT⁽⁶⁾. The sum of the energy stored in these tissues is the energy stored in the body (ST), which is the difference between ME and EX. The relationship of these energies is shown in Fig. 1.

The sum of the weights of Δ LT and Δ AT is Δ BW. Thus, Δ LT and Δ AT are calculated using the following simultaneous equations^(6,7):

$$\Delta LT(g) + \Delta AT(g) = \Delta BW(g)$$
 (1)

$$\Delta LT (g) \times 5.23 (kJ/g) + \Delta AT (g) \times 30.96 (kJ/g) = ST (kcal)$$
 (2)

Theoretically, the energy at the point at which $\Delta LT/\Delta BW$ reaches 100 % on the regression line, which indicates the relationship between $\Delta LT/\Delta BW$ and WG, is considered to be the energy expended to synthesise LT. Conversely, the energy at the point at which $\Delta LT/\Delta BW$ reaches 0 % is considered to be the energy expended to synthesise AT.

To determine this relationship, it was necessary to prepare the animals with different $\Delta LT/\Delta BW$ values. These animals can be prepared by feeding them diets containing different ratios of protein, fat and carbohydrates⁽⁸⁾. It has been shown that the energy required for protein synthesis is greater than that for fat⁽¹⁻³⁾. Bray *et al.*⁽⁹⁾ showed that resting energy expenditure and body protein (lean body mass) increased with a high-protein diet in humans. This greater resting



2 K. Obikawa et al.

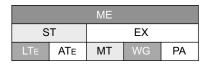


Fig. 1. Theoretical distribution of ME. ME, metabolisable energy; ST, stored energy; EX, expended energy; LT_E, energy stored in the lean tissue; AT_E, energy stored in the adipose tissue; MT, energy for body maintenance; WG, energy for body weight gain; PA, energy for physical activity.

energy expenditure may be associated with the greater energy expended for protein synthesis. It can be assumed that the energy required to synthesise LT is greater than that required to synthesise AT because the synthesis of proteins requires a large amount of energy. Thus, it is considered that WG differs in animals with different $\Delta LT/\Delta BW$ values.

Therefore, we investigated the effects of standard, high-protein and high-fat diets on $\Delta LT/\Delta BW$ and WG in growing rats to determine the energy required for the synthesis of LT and AT.

Materials and methods

Animals and outline of the procedure

Twenty-seven 5-week-old male Sprague-Dawley rats were obtained from CLEA Japan. The rats were divided into groups fed a standard diet (N, n 12), high-protein diet (HP, n 8) and high-fat diet (HF, n 7) and were individually housed in metabolic chambers. The rats were fed the respective diets for 7 d prior to the study to acclimatise them to the diets and metabolic chambers. Water and diet were provided ad libitum. Six rats in the N group were euthanised after the 7-d acclimatisation period to determine the weight of the gastrointestinal contents, which was used to calculate the BW gain, as described below. The remaining rats were used to measure EX for 4 weeks as described below. Water and diet were provided ad libitum. BW, food intake and EX were measured daily. The temperature of the animal room was $23 \pm 1^{\circ}$ C; the dark period was from 08.00 to 20.00, and the light period was from 20.00 to 08.00.

This study was conducted in accordance with the Guidelines for Proper Conduct of Animal Experiments of the Science Council of Japan and was approved by the Experimental Animal Committee of the Research Integrity Committee of Osaka University of Health and Sport Sciences (approval numbers 21-2 and 21-4).

Diet

Table 1 shows the composition of the diets used. A commercial standard diet CE-2 (CLEA Japan) was used for the N group.

Not all ingested energy is absorbed or metabolised. In the present study, we used the energy metabolised in the body. Thus, we used ME as energy intake for this study. The ME of the diets used in this study was determined based on reports by MaCraken⁽¹⁰⁾ and Raman *et al.*⁽¹¹⁾, and the values have been reported previously⁽¹²⁾. Briefly, 5–6 rats were individually housed in metabolic cages and fed each diet for 7 d. For the next 7 d, the rats were fed the same diet *ad libitum*, food intake was measured, and all faeces and urine were collected. The energy content of each diet, faeces and urine was measured using bomb calorimetry (Japan Food Research Center). The faeces were freeze-dried, and urine was dried in an oven at 60° C⁽¹³⁾ to avoid loss of short-chain fatty acids (SCFA) prior to bomb calorimetry analysis, whereas undried specimens were used for diet analysis. Samples weighing approximately 0·4–0·5 g were used. ME

Table 1. Dietary composition

	N*	HF	HP
Casein (g/kg)		264	379
Maize oil (g/kg)		345-5	70
Maize starch (g/kg)		182-986	289-986
α-maize starch (g/kg)		60	97
Sucrose (g/kg)		46.5	63
Cellulose (g/kg)		50	50
AIN-93G mineral-mix (g/kg)		35	35
AIN-93 vitamin-mix (g/kg)		10	10
L-cystine (g/kg)		3	3
Choline bitartrate (g/kg)		3	3
t-Butylhydroquinone (g/kg)		0.014	0.014
Protein (g/100 g)	25-10	22.78	32.71
Fat (g/100 g)	4.51	35-12	7.84
Carbohydrates (g/100 g)	49.72	27.37	41.45

N, normal; HF, high fat; HP, high protein.

*Information such as the raw materials and content of vitamins and minerals are available on the manufacturer's (CLEA Japan) website (https://www.clea-japan.com/en/products/genera l_diet/item_d0030).

was calculated by subtracting the energy excreted into the faeces and urine from the energy intake for the last 7 d. ME was 1323 kJ/100 g for the N diet, 2248 kJ/100 g for the HF diet and 1675 kJ/100 g for the HP diet.

Measurement of energy expenditure

EX was measured using an open-circuit system $^{(14)}$ in rats individually housed in metabolic chambers 22 cm \times 34 cm \times 14 cm (width \times depth \times height). The chamber was ventilated at 2100–4100 ml/min depending on the rat BW and oxygen consumption. During the experiment, a portion of the ventilated air (150 ml/min) was collected in a 250 L Douglas bag (Yagami) for 23 h and 45 min, and the oxygen concentration was measured using a portable gas monitor (VO2000, S & ME.). Oxygen consumption was calculated by multiplying the difference in oxygen concentration between the room air and sampled air by the ventilation rate of the chamber, and the EX was calculated as 20·08 kJ/l oxygen. The EX was converted per 24 h.

Sampling of organs and tissues, and whole-body biochemical analyses

The rats were euthanised under isoflurane anesthesia. Internal organs (heart, liver, kidneys, adrenal glands, spleen, pancreas and intestines), skeletal muscles (flexor hallucis longus, soleus, gastrocnemius and plantaris) and AT (perirenal, retroperitoneal, epididymal and mesenteric) were collected and weighed. After removing the intestinal contents, the intestines were weighed. The collected blood, internal organs, skeletal muscles and AT were returned to the abdominal cavity of the carcass and frozen for biochemical analysis.

The carcass was dried in an oven at 60°C⁽¹³⁾ to avoid loss of SCFA. The dried samples were pulverised into a powder using a mill (Vita-Max Absolute Blender, Osaka Chemical Co., Ltd).

Table 2. Body weight (Mean values with their standard errors)

	N		Н	HF)		One-way ANOVA		Cohen's d		
	Mean	SE		Mean	SE		Mean	SE		Р	N v. HF	N v. HP	HF v. HP
Initial (g)	144-4	3.6		144-4	2.8		147-1	2.9		0.801	0.001	0.290	0.313
Final (g)	349-3	6.6		385-1	12.5		378-9	6.8		0.054	1.235	1.519	0.219
Δ(g)	204-9	5.8	а	240-7	10-6	b	231.8	4.7	ab	0.020	1.449	1.834	0.387

N, normal; HF, high fat; HP, high protein.
Values with different letters differ significantly.

The total lipid content was determined using the Folch method. Approximately 1 g of the sample was homogenised in chloroform: methanol (2:1), the chloroform layer was dried and the weight of the residue was measured. Protein content was calculated as the nitrogen content of the sample, which was determined using the Kjeldahl method multiplied by 6·25. For glycogen, approximately 100 mg of the sample was decomposed with 30 % potassium hydroxide, ethanol was added to precipitate the glycogen, which was then dissolved in an appropriate amount of water and coloured using the phenol-sulphuric acid method, and the absorbance was measured⁽¹⁵⁾.

Theoretical distribution of metabolisable energy

Figure 1 shows the theoretical distribution of ME. Because the ME is expended or stored, the ST was calculated as the difference between the ME and EX.

EX consists of MT, WG and PA. In the present study, PA was considered minimal because rats were in the chamber; thus, WG was calculated by subtracting only MT from EX. In humans, MT is considered to be $1.5 \times$ basal metabolic rate (BMR)⁽⁵⁾. In the present study, as the rats were in the chamber, their PA was assumed to be minimal, as mentioned above. However, no data are available for the appropriate factor to multiply BMR to obtain the energy required to maintain the body in sedentary rats. According to Gleeson et al. (16) the lowest EX of the sedentary rats during the resting period was 1.66 kJ/kg/h which was considered to be their BMR, and the EX during the rats eating (2.88 kJ/kg/h) was $1.7 \times$ the lowest EX, and the EX during the active period while not eating (1.95 kJ/kg/h) was $1.2 \times$ the lowest EX. The average of 1.7and 1.2 was 1.45. In the present study, the rats had ad libitum access to food. Therefore, we set the MT as 1.5 x estimated BMR⁽¹⁷⁾.

Calculation for lean tissue and adipose tissue deposition

When animals grow, the sum of the increases in the weight of LT (Δ LT) and AT (Δ AT) is the BW gain (Δ BW). In addition, energy is stored in either LT or AT. Therefore, the energy stored in the body (ST) is the sum of the energy stored in LT (LT_E) and AT (AT_E). Thus, Δ LT and Δ AT can be calculated using the following simultaneous equation^(6,7):

$$\Delta LT (g) + \Delta AT (g) = \Delta BW (g)$$
 (1)

$$\Delta LT (g) \times 5.23 (kJ/g) + \Delta AT (g) \times 30.96 (kJ/g) = ST (kJ) (2)$$

Equation 1 indicates that the sum of the increases in LT and AT is the BW gain and Equation 2 indicates that the sum of LT_E and AT_E is the energy stored in the body.

The LT_E and AT_E were calculated by multiplying the energy density of each tissue by the accreted tissue weight obtained using this simultaneous equation.

The BW without gastrointestinal content weight was used to calculate the BW gain in Equation 1 because the gastrointestinal content was measured as BW, but this was not the body. The gastrointestinal content weight used for this calculation was obtained by sampling organs and tissues, as described above. The BW without the gastrointestinal contents of the rats at the start of the study was assumed to be $89.45\,\%$ of their BW because the gastrointestinal content accounted for $10.55\,\%$ (SE 0.78) of the BW of the rats that were euthanised before starting the study.

Statistics

The sample size was calculated from a statistical power $(1-\beta)$ of 0.8, α error of 0.05 and a significant minimum effect size (f) of 1.0. As there was no available information regarding changes in the energy required for BW gain due to differences in diets, we set the effect size to 1 to find a 1 SD difference. This power calculation determined that a minimum sample size of five animals was required to detect a statistically significant difference in the energy required for BW gain using G * Power 3·1. One-way ANOVA was used for comparisons among groups, and the Bonferroni test was used as a *post hoc* test (IBM SPSS Statistics version 27.0.1.0). Pearson's correlation was used to determine the relationship between Δ LT/ Δ BW and the energy required for BW gain, as the data passed the Shapiro–Wilk test. Statistical significance was set at P < 0.05.

Results

Table 2 shows that the BW gain in the HF was the highest, but not significantly differ from the HP group.

Figure 2 and online Supplementary Table S1 show the distribution of ME. The EX, WG and LT_E were higher in the HP group than in the N and HF groups. The ST and AT_E were the greatest in the HF group, which did not differ from that of the N group.

The energy density per gram of accumulated tissue in the HF group (12·9 kJ/g (se 0·8)) was significantly higher than that in the HP group (8·6 kJ kcal/g (se 0·8), P = 0.003, d = 1.813), whereas that in the N group (11·3 kJ/g (se 0·5)) was not different from either the HF (P = 0.620, d = 0.795) or HP groups (P = 0.087, d = 1.355).

Table 3 shows the increases in the weights of LT (Δ LT) and AT (Δ AT). Δ LT was significantly greater in the HP group than in the other two groups, whereas Δ AT was the greatest in the HF group, which did not differ from that in the N group. The ratio of Δ LT to

4 K. Obikawa *et al.*

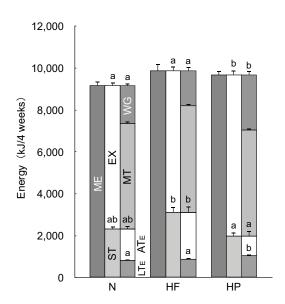


Fig. 2. Distribution of ME. Means and se. Values with different letters differed significantly. ME, metabolisable energy; ST, stored energy; EX, expended energy; MT, energy for body maintenance; WG, energy for body weight gain; LT_E, energy stored in the lean tissue; AT_E, energy stored in the adipose tissue; N, normal; HF, high fat; HP, high protein.

the increase in BW (Δ LT/ Δ BW) was the highest in the HP group but was not significantly different from that in the N group. The ratio of Δ AT to the increase in BW (Δ AT/ Δ BW) was higher in the HF group than that in the HP group, whereas the ratio in the N group did not differ from that in the HF or HP groups.

Table 4 shows organ and tissue weights. The skeletal muscle weight was significantly greater in the HP group than in the HF group. There was no significant difference in the skeletal muscle weight between the HP and the N group except for FHL. The AT weight except for perirenal was the highest in the HF group than the other groups, while the weight of retroperitoneal and mesenteric did not significantly differ from the HP group and the weight of epididymal did not differ from the N group. The weights of the kidneys, adrenal, pancreas and intestines were the lowest in the HF group, but the pancreas weight was not significantly different between the N group.

Table 5 shows the organ and tissue weights per 100 g of BW. The skeletal muscle weight was the lowest in the HF group. The weights of the retroperitoneal and epididymal AT were the highest in the HF group. Retroperitoneal AT did not differ between the HF and HP groups, and epididymal AT did not differ between the HF and N groups. The internal organ weights were lowest in the HF group, but the weights of the heart, spleen and pancreas did not differ from those in the N group.

Table 6 shows the whole-body protein, total lipid and glycogen contents. The protein content did not differ among the groups, whereas the total lipid content was the highest in the HF group. The glycogen content was higher in the HP group than in the other groups.

Table 7 shows the whole-body protein, total lipid and glycogen contents per 100 g of BW. The total lipid content was the highest in the HF group.

Figure 3 shows the positive correlation between $\Delta LT/\Delta BW$ and the energy required to gain 1 g of BW, which was calculated by dividing ΔBW by WG. The energy required to gain 1 g of BW was

significantly higher in the HP group (11.3 kJ/g (SE 0.6)) than in the HF group (7.0 kJ/g (SE 0.8), P < 0.01, d = 2.103), while there was no difference between the N group (8.8 kJ/g (SE 0.5)) and the HF group (P = 0.315, d = 0.966) or HP group (P = 0.074, d = 1.442). The regression line demonstrated that the energy required to gain 1 g of BW at the point of 100% on $\Delta \text{LT}/\Delta \text{BW}$ was 13.9 kJ/g, while the energy required to gain 1 g of BW at the point of 0% on $\Delta \text{LT}/\Delta \text{BW}$ was -7.9 kJ/g.

Discussion

In this study, no differences in ME were observed between the groups, but BW gain was the highest in the HF group among the groups. Thermic effect of food (TEF) of protein is greater than that of carbohydrates, which is greater than that of fat⁽¹⁸⁾. The higher BW gain in the HF group was presumably due to the smaller TEF of the HF diet, which resulted in less EX and more ST. This greater ST was thought to be associated with the higher BW gain in the HF group relative to the N group. In the HP group, it was assumed that the TEF of the HP diet was higher than that of the other diets. Therefore, the ST may be small in the HP group. However, LT accretion in the HP group was greater than in the other two groups. Because the energy density of LT is lower than that of AT, LT can accumulate with less ST. Therefore, it is considered that the increase in the BW of the HP group was not smaller than that of the other groups. The increase in LT was greatest in the HP diet, and the increase in AT was greatest in the HF diet. The AT increase accounted for 13-30 % of BW gain, while 47–72 % of the energy stored in the body was stored in AT. Thus, the accumulation of LT and AT differed among the groups fed different diets, whereas a large proportion of energy was stored in the AT, even though the weight increase in the AT was not very large.

Energy is required for several metabolic pathways⁽¹⁸⁾. Major macronutrient fluxes such as gluconeogenesis, de novo lipogenesis, triacylglycerol (TAG) synthesis and protein turnover require energy, and these flux rates can be influenced by both the energy content of the diet and its composition⁽¹⁸⁾. In the present study, the amount of energy required for BW gain differed depending on diet. It was higher in the HP diet group than in the N and HF diet groups. When TAG accumulates in AT, metabolic processes differ depending on the TAG substrate. Regarding carbohydrates, de novo lipogenesis is involved in TAG deposition. In the case of proteins, deamination and urea synthesis are involved, in addition to de novo lipogenesis. It is considered that the more metabolic processes involved in accumulating TAG, the more energy is expended. It is suggested that the energy expended to accumulate TAG was greater in the HP diet than in the other diets. Ingested amino acids are utilised for body protein synthesis, and proteins accumulate in the LT, which also requires energy. The amount of amino acids ingested during the study, as estimated by the ME and protein contents of the diets was 140.9 g for the N group, 94.0 g for the HF group and 175.7 g for the HP group. In addition, the accretion of LT was the greatest in the HP diet group. It is suggested that the energy expended to accumulate protein in the body is greater in the HP diet than in the other diets. Therefore, it seems reasonable that the energy required for BW gain in the HP diet group was highest among the three groups.

A positive correlation was observed between $\Delta LT/\Delta BW$ and energy required for BW gain. It is considered that the energy used for weight gain when $\Delta LT/\Delta BW$ is 100 % is the energy required for the synthesis of LT, which was 13.9 kJ/g in this study. In other

Table 3. Accretion of LT and AT (Mean values with their standard errors)

	N			Н	HF			P		One-way ANOVA	Cohen's d		
	Mean	SE		Mean	SE		Mean	SE		<i>P</i> -value	N v. HF	N v. HP	HF v. HP
ΔLT (g/4 weeks)	156-5	5.8	а	168-7	9.8	а	202-0	8.5	b	0.006	0.525	2.051	1.242
Δ AT (g/4 weeks)	48-4	4-2	ab	72-0	9-1	b	29.8	7.1	а	0.004	1.146	1.036	1.788
ΔLT/ΔBW (%)	76-4	1.97	ab	70-3	3-22	а	87.1	3.09	b	0.004	0.794	1.357	1.814
ΔΑΤ/ΔBW (%)	23-6	1.97	ab	29.7	3-22	b	12-9	3.09	а	0.004	0.794	1.357	1.814

LT, lean tissue; AT, adipose tissue; N, normal; HF, high fat; HP, high protein.

Values with different letters indicate significant differences. The Bonferroni test was used as a post hoc test.

reports of ours, the energy required to synthesise LT was 12.2 kJ/g⁽⁶⁾ and 12.6 kJ/g⁽⁷⁾, which are comparable to the energy observed in the present study. To our knowledge, no other studies have reported the energy required to synthesise LT. To increase LT, it is considered rational to add the energy required for LT synthesis and the energy accumulated in the LT. The energy density of LT is 5.2 kJ/g. Therefore, the additional energy intake required to increase LT was estimated to be approximately 19 kJ/g. However, the results of the present study could not elucidate the energy required for AT synthesis. Sekiguchi et al. (6) reported that the energy required for AT synthesis was 4.6 kJ/g, and that the energy required to accumulate AT, including the energy stored in AT, was approximately 35.6 kJ/g. In their study, the rats with smaller $\Delta LT/\Delta BW$ were included than the rats in the present study; therefore, the regression curve was different from that in the present study, and a positive value was obtained when $\Delta LT/\Delta BW = 0$. Rats with smaller $\Delta LT/\Delta BW$ are necessary to determine the energy required for AT synthesis.

In this study, growing rats were used. Inoue *et al.*⁽⁷⁾ reported that in a 2-week study that used rats of 4, 7, 9 and 14 weeks of age, the increase in AT was most of or more than the BW gain at 9 and 14 weeks of age, and some rats of 14 weeks of age showed a decrease in LT. In the present study, it was necessary to examine rats at an age when LT was increasing. Additionally, weight gain was small in 9- and 14-week-old rats. We considered it better to conduct this study during a period of large weight gain for the calculation. Therefore, 6-week-old rats were used in this study.

As discussed below, the values obtained in the present study may vary with age but may not differ significantly among species. When athletes attempt to increase their BW, their aim is to increase muscle mass (which accounts for a large part of the LT) without increasing the AT. To do this, they increase their energy intake and perform resistance exercise training. Garthe et al. (19) reported that increasing energy intake by approximately 2100 kJ per d and adding training to athletes for 8-12 weeks resulted in a 2.7 kg increase in BW with a 1.7 kg increase in fat-free mass and a 1.1 kg increase in fat mass. Miyauchi et al. (20) showed that when male college American football players increased their daily energy intake by 2100 to 4200 kJ and performed power training for 1 year, they gained 9.7 kg in BW, with an fat-free mass of 5.2 kg and an fat mass of 4.5 kg. When calculating the energy accumulated in the body from the increase in fat-free mass and fat mass, and the energy density of these components, approximately 80 % of the accumulated energy was stored in the fat mass. In addition, the energy density of skeletal muscle is 5200 kJ/kg^(6,7). Therefore, when 1 kg of skeletal muscle is accumulated, 5200 kJ/kg of energy should be accumulated.

However, skeletal muscle mass does not increase by 1 kg within a few days. Therefore, the added energy intakes in these studies may have been too high.

The present study has several limitations. The energy density of LT in the equation used in this study does not consider the energy density of the bone. In addition, energy is required for bone synthesis, which is particularly important during the growth period. However, to the best of our knowledge, this energy is not clear, and we are unable to describe this energy from the data of the present study. Rats are known to be coprophagous. In the experiments performed to measure oxygen consumption, we noticed that the amount of faeces was small or that faecal matter was sometimes not seen in the chamber when the rats were fasted or fed restricted diets. In the present study, there was a normal amount of faeces in the chamber. Therefore, the rats might have eaten their faeces, although it was assumed that the amount was not large. Regarding the influence of different animal species, it has been shown that the energy required for protein synthesis does not markedly different between different species (21-23). The BMR in species of different sizes is proportional to the BW raised to the 0.75 power⁽¹⁷⁾. It has been reported that the contribution of protein turnover to the resting metabolic rate is approximately 20 % in an average human⁽²¹⁾. Assuming that this contribution is comparable among species, differences in the energy required for LT synthesis may not be large among species. Regarding sex differences, there are no sex-related differences in the metabolic pathways involved in protein synthesis. Therefore, we presume that there are no sex differences in the energy required for LT synthesis. As animals age, it is assumed that the body needs to synthesise more tissue to gain weight due to increased breakdown compared with synthesis, leading to an increase in the energy required to gain BW. Therefore, the values obtained in this study may have differed according to age. We used the energy of $1.5 \times$ the estimated BMR as the energy for maintaining the body (MT). The BMR is assumed to be lower in animals with a higher proportion of body fat. In the present study, the total lipid content of the whole body was higher in the HF diet group. Animals with high body fat had less LT. The BMR depends on the amount of LT. Therefore, it can be inferred that the BMR of the HF group was low. Because WG was calculated by subtracting MT from EX in the present study, WG increased as MT decreased, leading to an increase in the energy required for BW gain.

In conclusion, the energy expended to synthesise LT was 13.9 kJ/g. Therefore, combined with the energy stored in LT, the energy required to accumulate LT is approximately 19 kJ/g in growing rats. However, the energy required for the synthesis of AT has not been elucidated.

 Table 4. Organ and tissue weight (Mean values with their standard errors)

	١	1		Н	F		Н	Р		One-way ANOVA		Cohen's d	
	Mean	SE		Mean	SE		Mean	SE		Р	N v. HF	N v. HP	HF v. HP
Skeletal muscle													
FHL* (g)	0.91	0.02	а	0.86	0.04	а	1.05	0.03	b	0.004	0.471	1.712	1.758
Soleus (g)	0-21	0.01	ab	0.17	0.01	а	0-25	0.01	b	0.003	1.131	1.147	1.905
Gastrocnemius (g)	3.80	0.15	ab	3.52	0.09	a	4-04	0.11	b	0.021	0.847	0.695	1.761
Plantaris (g)	0-66	0.08	ab	0.59	0.03	а	0.76	0.03	b	0.005	0.899	1.082	1.935
Adipose tissue													
Perirenal (g)	0.83	0.13		1.20	0.14		1.12	0.08		0-129	1.019	1.009	0.244
Retroperitoneal (g)	2.79	0-35	а	5.57	0.79	b	3-92	0.25	ab	0.011	1.558	1.355	1.003
Epididymal (g)	3.76	0-36	ab	5-22	0.53	b	3-47	0.16	a	0.012	1.128	0.395	1.610
Mesenteric (g)	2.99	0.31	a	4.97	0.70	b	3.47	0.19	ab	0.030	1.247	0.699	1.047
Organ													
Heart (g)	0.94	0.04		0.93	0.04		1.07	0.03		0.050	0.112	1.263	1.255
Liver (g)	13-20	0.50		13-28	0.52		14-41	0.42		0-193	0.063	0.929	0.820
Kidneys (g)	2.82	0.09	b	2.34	0.11	а	2.96	0.10	b	0.002	1.607	0.517	1.998
Adrenal (g)	0.028	0.006	b	0.015	0.004	a	0.028	0.007	b	0.008	1.617	0.252	1.831
Spleen (g)	0.73	0.04		0.63	0.04		0.80	0.05		0-060	0.842	0.518	1.289
Pancreas (g)	1.47	0-10	ab	1.22	0.05	а	1.60	0.10	b	0-023	1.202	0.488	1.623
Intestines (g)	6-10	0.21	b	4.16	0.19	а	5.44	0.25	b	< 0.001	3.504	0.960	1.907

N, normal; HF, high fat; HP, high protein.

Values with different letters indicate significant differences. Bonferroni test was used as a post hoc test.

^{*}FHL, flexor hallucis longus.

 Table 5. Organ and tissue weight per 100 g of body weight (Mean values with their standard errors)

	I	N		F	IF		Н	P		One-way ANOVA		Cohen's d	
	Mean	SE		Mean	SE		Mean	SE		Р	N v. HF	N v. HP	HF v. HP
Skeletal muscle													
FHL* (g)	0-26	0.01	b	0-22	0.01	a	0-28	0.01	b	< 0.001	1.456	0.839	2-610
Soleus (g)	0-06	0.00	b	0.04	0-00	а	0.06	0.00	b	< 0.001	1.982	0.707	2.504
Gastrocnemius (g)	1.09	0.07	b	0.92	0.03	а	1.07	0.02	b	0.008	1.451	0.219	2.605
Plantaris (g)	0.19	0.01	b	0.15	0.01	а	0.20	0.01	b	0.002	1.489	0.546	2.306
Adipose tissue													
Perirenal (g)	0.23	0.03		0.31	0.03		0.30	0.02		0.195	0.916	0.818	0.215
Retroperitoneal (g)	0.79	0.10	a	1.42	0.19	b	1.03	0.06	ab	0.011	1.549	1.147	1.062
Epididymal (g)	1.08	0-11	ab	1.34	0.11	b	0.91	0.03	а	0.009	0.909	0.840	2.008
Mesenteric (g)	0.85	0.09		1.27	0.17		0.91	0.04		0.034	1.157	0.392	1.127
Organ													
Heart (g)	0.27	0.01	ab	0.24	0.01	a	0.28	0.01	b	0.020	0.912	0.536	1.923
Liver (g)	3.77	0.09	b	3.45	0.09	a	3.80	0.07	b	0.011	1.472	0.132	1.626
Kidneys (g)	0.80	0.02	b	0.61	0.03	а	0.78	0.02	b	< 0.001	3.366	0.486	3.037
Adrenal (g)	0.008	0.001	b	0.004	0.000	a	0.007	0.001	b	0.004	1.982	0.463	1.400
Spleen (g)	0.21	0.01	ab	0.16	0.01	а	0.21	0.01	b	0.026	1.334	0.036	1.433
Pancreas (g)	0-42	0.04	ab	0-32	0-02	а	0-42	0.02	b	0.024	1.360	0.027	1.561
Intestines (g)	1.75	0.07	С	1.08	0.05	а	1.43	0.05	b	< 0.001	4.501	1.947	2.589

N, normal; HF, high fat; HP, high protein.

Values with different letters indicate significant differences. Bonferroni test was used as a post hoc test.

^{*}FHL, flexor hallucis longus.

https://doi.org/10.1017/S000711452400326X Published online by Cambridge University Press

8 K. Obikawa *et al.*

Table 6. Protein, total lipid and glycogen contents in the whole body (Mean values with their standard errors)

	N		Н	F		Н	HP		One-way ANOVA		Cohen's d			
	Mean	SE		Mean	SE		Mean	SE		Р	N v. HF	N v. HP	HF v. HP	
Protein (g)	71.1	1.4		75-9	3.4		76-2	1.6		0.342	0.618	1.141	-	
Total lipid (g)	26-1	1.8	а	50.0	5.5	b	31.7	1.0	а	< 0.001	1.985	1.416	1.684	
Glycogen (g)	0.07	0.00	а	0.07	0.00	а	0.10	0.01	b	0.002	0.137	1.863	1.583	

N, normal; HF, high fat; HP, high protein.

Values with different letters indicate significant differences. The Bonferroni test was used as a post hoc test.

Table 7. Weight of protein, total lipid and glycogen in 100 g of the whole-body components (Mean values with their standard errors)

	N		Н	=		HP			One-way ANOVA	Cohen's d			
	Mean	SE		Mean	SE		Mean	SE		Р	N v. HF	N v. HP	HF v. HP
Protein (g)	20-4	0.2		19.7	0.3		20.1	0.17		0.144	1.017	0.570	0.648
Total lipid (g)	7-4	0.4	a	12.8	1.2	b	8-3	0.2	а	< 0.001	2.187	1.138	2.013
Glycogen (g)	0.02	0.00		0.02	0.00		0.03	0.00		0-050	0.512	0.949	1.130

N, normal; HF, high fat; HP, high protein.

Values with different letters indicate significant differences. Bonferroni test was used as a post hoc test.

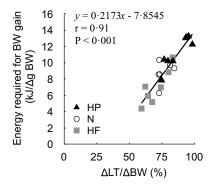


Fig. 3. Relationship between Δ LT/ Δ BW and the energy required for body weight gain. LT, lean tissue; BW, body weight; N, normal; HF, high fat; HP, high protein.

Supplementary material. For supplementary material/s referred to in this article, please visit https://doi.org/10.1017/S000711452400326X

Acknowledgement. This research received no specific grants from any funding agency, commercial sector or not-for-profit sectors.

K. Ob., M. K. and K. Ok. designed the research; K. Ob. and M. K. conducted the research; K. Ob. analysed the data; and K. Ob., M. K., E. K. and K. Ok. wrote the paper. K. Ok. was the primary responsibility for the final content. All the authors have read and approved the final manuscript.

The authors declare that they have no known competing financial interests or personal relationships that could influence the work reported in this study.

References

- Pullar JD & Webster AJ (1977) The energy cost of fat and protein deposition in the rat. Br J Nutr 37, 355–363.
- Roberts SB & Young VR (1988) Energy costs of fat and protein deposition in the human infant. Am J Clin Nutr 48, 951–955.
- Hall KD (2006) Computational model of *in vivo* human energy metabolism during semistarvation and refeeding. *Am J Physiol Endocrinol Metab* 291, E23–E37.

- Speakman JR & Hall KD (2023) Models of body weight and fatness regulation. Philos Trans R Soc Lond B Biol Sci 378, 20220231.
- 5. Spady DW, Payne PR & Picou D (1976) Energy balance during recovery from malnutrition. *Am J Clin Nutr* **29**, 1073–1088.
- Sekiguchi F, Kitaguchi M, Kondo E, et al. (2024) The energy required to synthesize lean and adipose tissue in rats. J Nutr Sci Vitaminol 70, 150–157.
- Inoue H, Maeda M, Fujii T, et al. (2024) Relationship between the increase rate of lean tissue in body weight gain and energy for tissue synthesis in rats. Osaka Taiiku Daigaku Kiyo. Bull Osaka Univ Health Sport Sci (In Press). (in Japanese).
- 8. Dulloo AG & Girardier L (1992) Influence of dietary composition on energy expenditure during recovery of body weight in the rat: implications for catch-up growth and obesity relapse. *Metabolism* **41**, 1336–1342.
- Bray GA, Smith SR, de Jonge L, et al. (2012) Effect of dietary protein content on weight gain, energy expenditure, and body composition during overeating: a randomized controlled trial. JAMA 307, 47–55.
- 10. McCracken KJ (1975) Effect of feeding pattern on the energy metabolism of rats given low-protein diets. *Br J Nutr* **33**, 277–289.
- Raman A, Baum ST, Colman RJ, et al. (2007) Metabolizable energy intake during long-term calorie restriction in rhesus monkeys. Exp Gerontol 42, 988–994.
- 12. Okamura K & Maeda M (2022) Metabolizable energy of standard, high-fat and high-protein diets in rats. Osaka Taiiku Daigaku Kiyo. *Bull Osaka Univ Health Sport Sci* **53**, 17–22. (in Japanese).
- Król E & Speakman JR (1999) Isotope dilution spaces of mice injected simultaneously with deuterium, tritium and oxygen-18. J Exp Biol 202, 2839–2849.
- Lim K, Murakami E, Lee S, et al. (1996) Effects of intermittent food restriction and refeeding on energy efficiency and body fat deposition in sedentary and exercised rats. J Nutr Sci Vitaminol (Tokyo) 42, 449–468.
- Lo S, Russell JC & Taylor AW (1970) Determination of glycogen in small tissue samples. J Appl Physiol 28, 234–236.
- Gleeson M, Brown JF, Waring JJ, et al. (1982) The effects of physical exercise on metabolic rate and dietary-induced thermogenesis. Br J Nutr 47, 173–181
- Terpstra AHM (2001) Differences between humans and mice in efficacy of the body fat lowering effect of conjugated linoleic acid: role of metabolic rate. J Nutr 131, 2067–2068.
- Hall KD & Guo J (2017) Obesity energetics: body weight regulation and the effects of diet composition. Gastroenterology 152, 1718–1727.

https://doi.org/10.1017/S000711452400326X Published online by Cambridge University Press

- 19. Garthe I, Raastad T, Refsnes PE, *et al.* (2013) Effect of nutritional intervention on body composition and performance in elite athletes. *Eur J Sport Sci* **13**, 295–303.
- 20. Miyauchi S, Oshima S, Asaka M, et al. (2013) Organ size increases with weight gain in power-trained athletes. Int J Sport Nutr Exerc Metab 23, 617–623.
- Welle S & Nair KS (1990) Relationship of resting metabolic rate to body composition and protein turnover. Am J Physiol 258, E990–E998.
- 22. Bernier JF, Calvert CC & Baldwin RL (1987) Energetics of protein synthesis in mice with a major gene for growth. *J Nutr* **117**, 2036–2045.
- 23. Barry TN, Davis SR & Hughson GA (1981) Protein synthesis in tissues of growing lambs. *Br J Nutr* **46**, 409–419.