

Changes of ultrastructure and downregulation of heat shock protein 70 and fibroblast growth factor 2 in gastric mucosa of rats with depressive-like behaviour

Hou G, Tang M, Yan L, Wang M. Changes of ultrastructure and downregulation of heat shock protein 70 and fibroblast growth factor 2 in gastric mucosa of rats with depressive-like behaviour.

Objective: To understand the underlying mechanism of gastric symptoms in patients with depressive disorder.

Methods: This study tested in the rat depression model evoked with chronic mild stress whether the microstructure gastric mucosa is injured using scanning electronic microscopy and transmission electronic microscopy (TEM). In addition, the expression of heat shock protein 70 (HSP70) and fibroblast growth factor 2 (FGF2) proteins in the gastric mucosa were measured by Western blotting.

Results: We found that the gastric epithelial cells were ruptured and the gastric pits were widened in rats with depression. The amount of mucous granules was also reduced in the surface mucous cells. Moreover, parietal cells became active, and the secretory canaliculi were magnified. Expression of HSP70 and FGF2 was reduced in the gastric mucosa.

Conclusions: These findings suggested that gastric symptoms in rats with depressive-like behaviour were caused by the injury of the gastric mucosa, AQP1 and HSP70 and FGF2 may be key molecules in the pathogenesis.

**Gonglin Hou, Mingming Tang,
Linlin Yan, Mingming Wang**

Department of psychology, Zhejiang Sci-Tech University, Hangzhou 310018, China

Keywords: depression; electronic microscopy; fibroblast growth factor 2; gastric mucosa; heat shock protein 70; ultrastructure

Gonglin Hou, Department of psychology, Zhejiang Sci-Tech University, 579# Mailbox, Hangzhou 310018, China.

Tel: +86 57186843234;

Fax: +86-57188492242

E-mail: gonglinhou@163.com

Accepted for publication July 30, 2011

Significant outcomes and limitations

- The study showed the depression could cause injury to the gastric mucosa at the ultrastructure level, and some related signalling pathways were shown.
- The limitation is that currently there lacks a proper grading system to classify the injuries at the ultrastructure level, therefore the analysis of the correlation between molecular changes and cytology is difficult. We will be interesting to look at the underlying mechanisms in our future studies.

Introduction

Chronic stress (CS) is one of the most important factors contributing to clinical depression, and was sometimes considered as the cause of digestive symptoms in patients. These gastrointestinal symptoms were diagnosed as functional gastrointestinal disorder, FGID (1–3). Previous studies showed that psychological factors were critical in the development of functional dyspepsia and irritable

bowel syndrome (4,5). The potential underlying mechanism was suggested to be stress-ulcer response (6); however, whether organic lesions occurred in the gastrointestinal system during depression was not investigated. In addition, in recent years it has been found that stress could induce the expression of heat shock protein 70 (HSP70) and fibroblast growth factor 2 (FGF2) in of gastric mucosa (7,8), reflecting the significance of mucosal damage and the capacity

of regeneration. In this study, the authors set up to examine the organic changes in ultrastructure of gastric mucosa using electronic microscopy, then the expression and changes of HSP70 and FGF2 in gastric mucosa of a rat model of depression, to explore if the gastric symptoms in depression were associated with organic lesions of the mucosa.

Materials and methods

Animals

Sixteen male Sprague-Dawley rats (250 ± 10 g) were purchased from Experimental Animal Center of Chinese Academy of Sciences (Shanghai, China), and randomly assigned into two groups after 7 days of adaptation [control (CT) and CS groups].

The rat model of depression with chronic stress

The rat model of depression with CS was employed according to previous studies (9). The combination of solitary care (isolation stress) and chronic unpredictable mild stress (CUMS) was applied on CS group for 35 days. In the CT group, eight rats were raised in two cages; in CS group, each rat was raised in isolated caged and subjected to CUMS protocol as follows: fasting (24 h), water deprivation (24 h), tail clamp (1 min), foot shock (36-V AC, current intensity 1.0 mA, at intervals of 1 min per stimulation, each lasting 10 s, a total of 30 times), ice water swimming (4 °C, 5 min), wet bedding (lasting for 24 h, an addition of 100–300 ml of water was used to wet cage bedding), reversed light cycle; the seven types of stress stimuli were randomly generated to the rats with one each per day. All procedures with animal experiments were approved by Animal Experiment Ethic Committee at the Animal Experimental Center of Zhejiang University of Technology, Hangzhou, China.

Open field test

An open box with length and width of the 80 cm \times 80 cm and height at 40 cm was used. The bottom floor was divided into 64 black and white grids (10 cm \times 10 cm each). The 16 grids in the middle were considered as central, and the rests were peripheral grids. The animal behaviour was recorded at the active cycle of the animal under red light for 5 min. Noldus software was used for the analysis of the total travelling distance, the percentage of time the animal spent in central region and traces of movement.

Sucrose water consumption test

Animals were deprived of water for 12 h per day for 4 days before the sucrose water consumption

test (SWCT) for 3 h per day. The animals were provided with two choices between 1% sucrose water and water bottles, with their positions randomly assigned every day. The ratio of the amount of liquid consumption was calculated as the index of hedonia.

Scanning electronic microscopy and transmission electronic microscope

XL30 scanning electron microscope (Philips, Amsterdam, The Netherlands) and JEM-1230 transmission electron microscope (Japan JFOL Company, Tokyo, Japan) were used for the EM studies. The animals were subjected to a food deprivation of 24 h before sacrifice, and the stomach tissue was harvested and rinsed with 0.1 M phosphate buffer solution before fixation into 2.5% glutaraldehyde overnight at 4 °C.

For scanning electron microscopic (SEM) samples, the tissue was processed with 1% osmium tetroxide solution (1 h), gradient dehydration with alcohol, ethanol and amyl acetate resin mixture (1:1 by volume) before amyloacetate overnight, drying and coating for cutting slices.

For transmission electronic microscopic (TEM) samples, the tissues were treated with gradient dehydration with alcohol, pure acetone (20 min), embedding medium and acetone mixture (1 h, 1:1 by volume), embedding medium and acetone mixture (1 h, 3:1 by volume), pure embedding medium overnight, and infiltration at 70 °C overnight. The sections were cut on a Reichert slice machine at 70–90 nm. Then the slices were stained in the lead citrate solution and uranyl acetate saturated solution of 50% ethanol for 15 min before observation. All the EM studies were performed at the EM center of Zhejiang University, China.

Western blot

After the animals were sacrificed, the gastric tissues were harvested and stored in liquid nitrogen before protein extraction. The following reagents were used: RIPA lysis buffer (Shanghai Health Workers, Shanghai, China), BCA protein concentration determination Kit (Pik days, Shanghai, China), HSP70, FGF2 rabbit anti-rat primary antibody, IgG goat anti-rabbit secondary antibody (Wuhan Boster Biological, Wuhan, China) and ECL fluorescent indicator (U.S. Pirece company, San Francisco, US).

Statistics

SPSS 17.0 was used for all statistical analyses. The results were represented by indicators mean \pm standard deviation ($M \pm SD$), and the

Table 1. Behaviour results (M ± SD, n = 8)

Group	Travelling distance (cm) (OFT)	Percentage of time spent in central grids (%) (OFT)	Percentage of SWCT in total (%)
Control	2689.96 ± 520.79	44.93 ± 18.85	76.05 ± 5.18
Chronic stress	1946.26 ± 568.89*	73.61 ± 19.80*	65.68 ± 6.32**

p* < 0.05 and *p* < 0.01 when compared with control group.

p value was determined with independent sample *t* test.

Results

Rat model of depression with behaviour changes

In the open field test (OFT, the CS group showed decreased locomotor behaviour in the road travelling distance (*t*(14) = 2.46, *p* < 0.05), and increased time spent in the central wells (*t*(14) = -2.78, *p* < 0.05) (Table 1). In the SWCT, the CS group showed decreased intake of sucrose water (*t*(14) = 3.59, *p* < 0.01), suggesting the success of our rat model of depression with CS.

The changes of ultra structure in gastric mucosa layer

In the SEM pictures, the surface of gastric mucosa in CT group was covered with mucus layer, and rick in mucus. With rinsing treatment, the mucosal epithelial cell showed integrity, and the mucosal surface still showed mucus coverage, with clear visualisation of gastric pits (Fig. 1a–c). While in CS group, with

same cleaning procedure, more epithelial cells were exposed, and showing damaged structure, as well as expanded gastric pits (Fig. 1d–f).

In TEM pictures, the mucous cells in CT group were abundant with mucus granules, which showed dense black spots in the EM picture (white ellipse, Fig. 2a). In contrast, the CS group showed decreased amount of granules, and these granules were exposed directly on the cell surface (Fig. 2b). In addition, we examined the changes in ultrastructure of gastric parietal cells. In the CT group, these cells were found to be in the resting state, whereas in CS group these cells were activated, with the expansion of microtubule bubbles inside the cell, as well as a larger low-density area (Fig. 3).

Changes in HSP70 and FGF2 expression in gastric mucosa under depression

The Western blot results obtained through optical density analyses between bands showed that in the gastric mucosa of CS group animals, HSP expression significantly decreased (*t*(14) = 4.93, *p* < 0.001), and the FGF2 expression also significantly reduced (*t*(14) = 2.53, *p* < 0.05) (Fig. 4, Table 2).

Discussion

Chronic unpredictable stress is a validated experimental approach in the induction of depression behaviour in rats (9), which could be measured with sucrose water consumption behaviour experiment (10,11). In this study, the sucrose water intake was

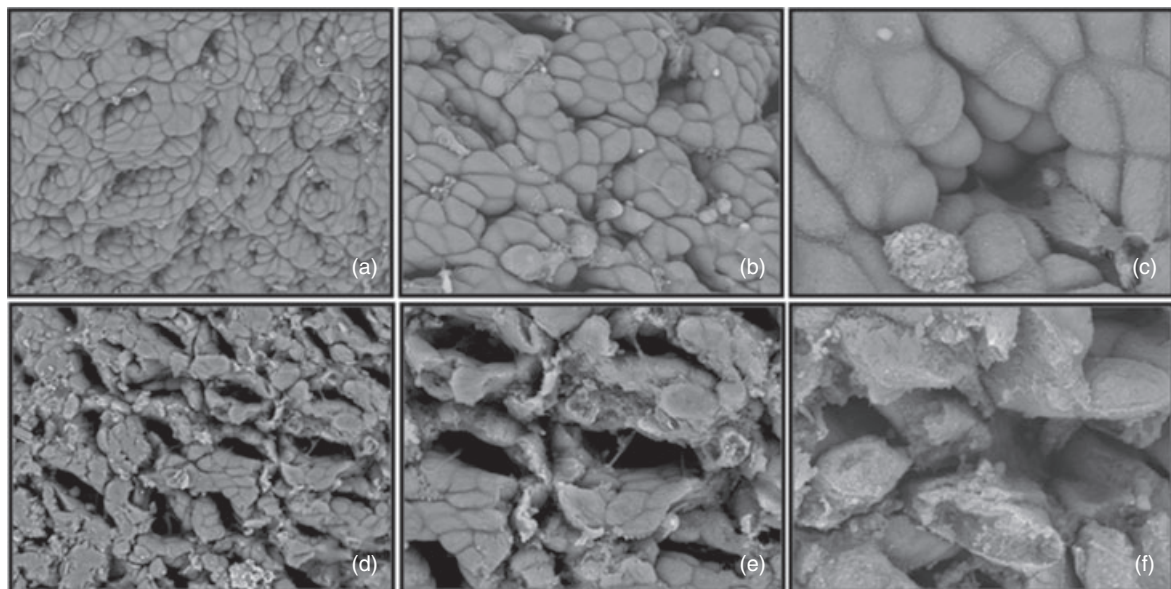


Fig. 1. SEM results for the control group (a–c) and chronic stress group (d–f). Magnification: (a, d) 1000×, (b, e) 2000×, (c, f) 5000×. In CS group, with same cleaning procedure, more epithelial cells were exposed, and showing damaged structure, as well as expanded gastric pits.

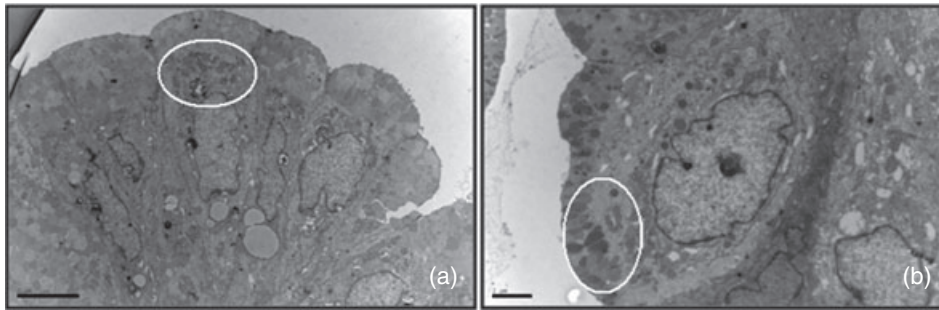


Fig. 2. TEM pictures: surface mucous cells; (a) 5000× magnification for the control group, (b) from chronic stress group 8000× magnification. The white ellipse highlighted the position of granules. The CS group showed decreased amount of granules, and these granules were exposed directly on the cell surface.

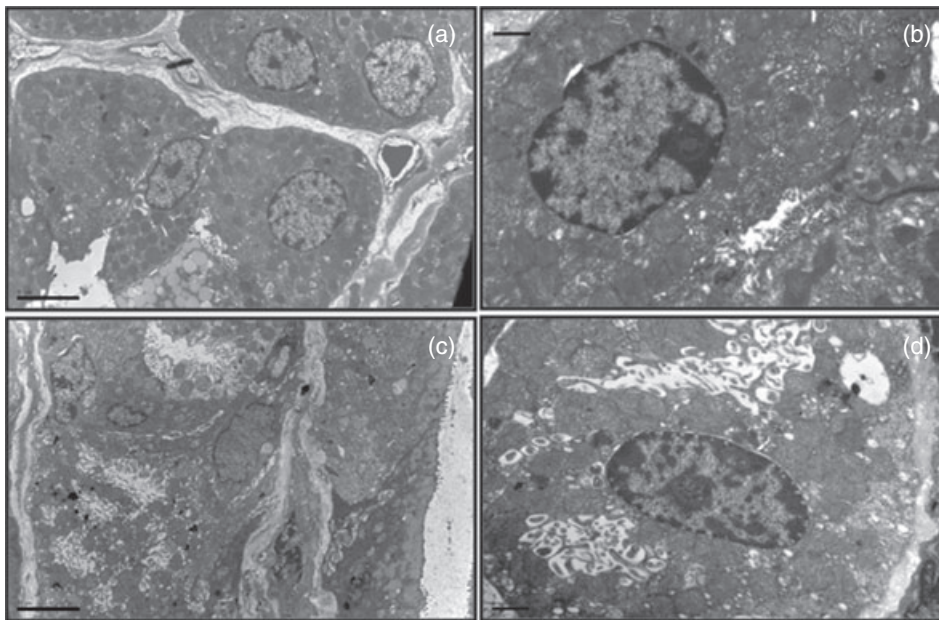


Fig. 3. Gastric parietal cells: (a, b) as the control group, (c, d) chronic stress (CS) group, showing that parietal cells in CS group were active, with the expansion of microtubules bubbles inside the cell, as well as a larger low-density area. (a, c) 5000× magnification, (b, d) 15000× magnification.

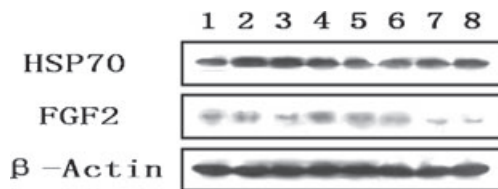


Fig. 4. Western blot results, 1234 for the control group, 5678 for the chronic stress group.

Table 2. The ratio of integrated optical density (IOD) of the target protein to internal reference protein (%) ($M \pm SD$, $n = 8$)

Group	HSP70	FGF2
Control	62.65 ± 10.78	26.32 ± 8.35
Chronic stress	35.26 ± 11.44***	17.45 ± 5.37*

* $p < 0.05$, *** $p < 0.001$ when compared to the control group.

significantly decreased in CS group. In addition, in the OFT, the CS group showed reduced avoidance behaviour or increased exposure in central grids, with decreased locomotion activity (12), both of which validated our model as a rat model of depression.

In stressful conditions, the hypothalamic pituitary adrenal axis was activated and corticotropin releasing

factor was released from hypothalamic paraventricular nucleus, stimulating the releasing of adrenocorticotrophic hormone, which in turn leads to the increase in adrenal glucocorticosteroid (GC) (13–15). GC was found to be involved in physiological activities of the gastric mucosa. For instance, Gama et al. treated rats *in vivo* and the cultured rat gastric tissue *in vitro* with GC, and the results showed decreased gastric epithelial cell proliferation as well as increased cell apoptosis (16). It was also found

that GC could regulate the expression of MUC5AC mRNA, which directly inhibits the synthesis of epithelial mucus layer (17). In addition, GC could regulate the glucocorticoid-regulated kinase SGK1 to enhance the secretion of gastric acid (18). We believed that the lesion we observed in rat model of depression of gastric mucosa was because of the abnormal level of GC, which disrupted the normal function of gastric mucosal cells and increased the secretion of gastric acid. Both factors would contribute to the destruction of the gastric mucosal layer, and other phenomenon.

HSP70 is widely expressed in mammalian cells under heat stress, environmental stress and psychological stress conditions with anti-apoptotic functions (19,20) through the maintenance of normal transmembrane transport (21). The upregulation of HSP70 could inhibit stress-induced JNK pathway, p38 and other apoptotic signalling pathways (22). It has also been shown that the HSP70 was protective in gastric mucosa against stress, and inhibited the apoptosis as well as necrosis, and heat shock cognate 70 (HSC70) (stress-induced isoform) was critical for this process (23–25). In our study, we found that after a period of stress, HSP70 decreased in depression group, suggesting the self-defence mechanism was weakened. Whether acute psychological stress would increase HSP70 in this model is yet unknown.

FGF2 could promote the growth of blood vessels, the regeneration of epithelial cells, and was found to be involved in gastric ulcer as well as other gastric diseases (8,26). It was shown that FGF2 expression would contribute to the healing after gastric ulcer (27), and during the healing process, in the early stage, FGF2 promotes the epithelial cell proliferation, while in the later stage the vascular endothelial cells promotes regrowth of blood vessels (28). Particularly, FGF was used to evaluate the treatment efficiency to gastric mucosal lesions in past studies (8,26,29). This study showed that FGF2 signalling was downregulated in animals of CS group, which might explain the fact that the lesions were not regenerated, which ultimately led to gastric dysfunction. Consistent with this, previous studies showed that FGF2 expression was reduced in different brain regions among several rodent models of depression (30,31), and the ventricle infusion of FGF2 could act as anti-depressant to some extent (31).

Taken together, this study reported that CS would result in depression-associated organic lesion of the gastrointestinal system, especially the gastric mucosal layer. The HSP70 and FGF2 were dysregulated, which might contribute to the failure in self-repair. The underlying mechanism is highly likely to be the increased release of adrenal GC under stressful conditions, which through the circulation

depressed the brain and hurt the stomach at the same time. The study implied that the gastrointestinal syndromes observed in patients with depression were more than psychological problems, and deserved better and focused treatments.

References

1. BUDAVARI AI, OLDEN KW. Psychosocial aspects of functional gastrointestinal disorders. *Gastroenterol Clin North Am* 2003;**32**:477–506.
2. LEVY RL, OLDEN KW, NALIBOFF BD et al. Psychosocial aspects of the functional gastrointestinal disorders. *Gastroenterology* 2006;**130**:1447–1458.
3. VAN OUDENHOVE L, AZIZ Q. Recent insights on central processing and psychological processes in functional gastrointestinal disorders. *Dig Liver Dis* 2009;**41**:781–787.
4. BARRY S, DINAN TG. Functional dyspepsia: are psychosocial factors of relevance? *World J Gastroenterol* 2006;**12**:2701–2707.
5. MAYER EA, NALIBOFF BD, CHANG L, COUTINHO SVV. Stress and irritable bowel syndrome. *Am J Physiol Gastrointest Liver Physiol* 2001;**280**:G519–G524.
6. OVERMIER JB, MURISON R. Anxiety and helplessness in the face of stress predisposes, precipitates, and sustains gastric ulceration. *Behav Brain Res* 2000;**110**:161–174.
7. KIANG JG, TSOKOS GC. Heat shock protein 70 kDa: molecular biology, biochemistry, and physiology. *Pharmacol Ther* 1998;**80**:183–201.
8. ESAKI M, AOYAGI K, MATSUMOTO T, KUWANO Y, SHIMIZU M, FUJISHIMA M. Effects of omeprazole and famotidine on fibroblast growth factor-2 during artificial gastric ulcer healing in humans. *Eur J Gastroenterol Hepatol* 2002;**14**:365–369.
9. WILLNER P. Chronic mild stress (CMS) revisited: consistency and behavioural-neurobiological concordance in the effects of CMS. *Neuropsychobiology* 2005;**52**:90–110.
10. MAR A, SPREEKMEESTER E, ROCHFORD J. Fluoxetine-induced increases in open-field habituation in the olfactory bulbectomized rat depend on test aversiveness but not on anxiety. *Pharmacol Biochem Behav* 2002;**73**:703–712.
11. BRENES SAENZ JC, VILLAGRA OR, FORNAGUERA TRIAS J. Factor analysis of forced swimming test, sucrose preference test and open field test on enriched, social and isolated reared rats. *Behav Brain Res* 2006;**169**:57–65.
12. SARKISOVA KY, KULIKOV MA. Behavioral characteristics of WAG/Rij rats susceptible and non-susceptible to audiogenic seizures. *Behav Brain Res* 2006;**166**:9–18.
13. KRISHNAN V, NESTLER EJ. The molecular neurobiology of depression. *Nature* 2008;**455**:894–902.
14. NESTLER EJ, BARROT M, DiLEONE RJ, EISCH AJ, GOLD SJ, MONTEGGIA LM. Neurobiology of depression. *Neuron* 2002;**34**:13–25.
15. SHELTON RC. The molecular neurobiology of depression. *Psychiatr Clin North Am* 2007;**30**:1–11.
16. GAMA P, GOLDFEDER EM, DE MORAES JC, ALVARES EP. Cell proliferation and death in the gastric epithelium of developing rats after glucocorticoid treatments. *Anat Rec* 2000;**260**:213–221.
17. TANAKA T, KOBAYASHI T, SUNAGA K, TANI S. Effect of glucocorticoid on expression of rat MUC5AC mRNA in rat gastric mucosa in vivo and in vitro. *Biol Pharm Bull* 2001;**24**:634–637.

18. SANDU C, ARTUNC F, GRAHAMMER F et al. Role of the serum and glucocorticoid inducible kinase SGK1 in glucocorticoid stimulation of gastric acid secretion. *Pflugers Arch* 2007;**455**:493–503.
19. YENARI MA, LIU J, ZHENG Z, VEXLER ZS, LEE JE, GIFFARD RG. Antiapoptotic and anti-inflammatory mechanisms of heat-shock protein protection. *Ann N Y Acad Sci* 2005;**1053**:74–83.
20. JOLLY C, MORIMOTO RI. Role of the heat shock response and molecular chaperones in oncogenesis and cell death. *J Natl Cancer Inst* 2000;**92**:1564–1572.
21. FEDER ME, HOFMANN GE. Heat-shock proteins, molecular chaperones, and the stress response: evolutionary and ecological physiology. *Annu Rev Physiol* 1999;**61**:243–282.
22. KUMAR Y, TATU U. Stress protein flux during recovery from simulated ischemia: induced heat shock protein 70 confers cytoprotection by suppressing JNK activation and inhibiting apoptotic cell death. *Proteomics* 2003;**3**:513–526.
23. OTAKA M, ODASHIMA M, WATANABE S. Role of heat shock proteins (molecular chaperones) in intestinal mucosal protection. *Biochem Biophys Res Commun* 2006;**348**:1–5.
24. OTAKA M, ODASHIMA M, IZUMI Y et al. Target molecules of molecular chaperone (HSP70 family) in injured gastric mucosa in vivo. *Life Sci* 2009;**84**:664–667.
25. OTAKA M, MATSUHASHI T, ODASHIMA M et al. Enhancement of cytoprotective ability and cell restoration in 70-kDa heat shock protein gene-transfected rat gastric mucosal cells. *Aliment Pharmacol Ther* 2006;**24**:272–277.
26. LUO JC, LIN HY, LU CL et al. Dexamethasone inhibits basic fibroblast growth factor-stimulated gastric epithelial cell proliferation. *Biochem Pharmacol* 2008;**76**:841–849.
27. HULL MA, BROUGH JL, POWE DG, CARTER GI, JENKINS D, HAWKEY CJ. Expression of basic fibroblast growth factor in intact and ulcerated human gastric mucosa. *Gut* 1998;**43**:525–536.
28. KIMURA T, NODA M, SUGIHARA H, KASHIMA K, HATTORI T. Expression of fibroblast growth factor-2 transcripts in the healing of acetic acid-induced gastric ulcers. *APMIS* 1999;**107**:767–772.
29. BECKER JC, BECKBAUER M, DOMSCHKE W, HERBST H, POHLE T. Fibrin glue, healing of gastric mucosal injury, and expression of growth factors: results from a human in vivo study. *Gastrointest Endosc* 2005;**61**:560–567.
30. KATO M, OKUGAWA G, WAKENO M et al. Effect of basic fibroblast growth factor (FGF2) gene polymorphisms on SSRIs treatment response and side effects. *Eur Neuropsychopharmacol* 2009;**19**:718–725.
31. TURNER CA, GULA EL, TAYLOR LP, WATSON SJ, AKIL H. Antidepressant-like effects of intracerebroventricular FGF2 in rats. *Brain Res* 2008;**1224**:63–68.