cambridge.org/jhl

Research Paper

Cite this article: Cruz-Tamayo A, González-Garduño R, Torres-Hernández G, Becerril-Pérez C, Hernández-Mendo O, Ramírez-Bribiesca E, López-Arellano ME, Vargas-Magaña J, Hernández-Rueda E (2020). Artificial *Haemonchus contortus* infection as a strategy to induce protective immune response to natural infection in Pelibuey lambs. *Journal of Helminthology* **94**, e181, 1–8. https://doi.org/ 10.1017/S0022149X20000656

Received: 24 February 2020 Revised: 8 August 2020 Accepted: 28 July 2020

Key words:

Sheep; resistance; gastrointestinal nematodes; total plasma protein; eosinophils

Author for correspondence: R. González-Garduño, E-mail: robgardu@hotmail.com

© The Author(s), 2020. Published by Cambridge University Press



Artificial *Haemonchus contortus* infection as a strategy to induce protective immune response to natural infection in Pelibuey lambs

A. Cruz-Tamayo^{1,2}, R. González-Garduño³, G. Torres-Hernández¹,
C. Becerril-Pérez¹, O. Hernández-Mendo¹, E. Ramírez-Bribiesca¹,
M.E. López-Arellano⁴, J. Vargas-Magaña² and E. Hernández-Rueda²

¹Colegio de Postgraduados, Campus Montecillo, Montecillo, Estado de México, Mexico; ²Escuela Superior de Ciencias Agropecuarias, Universidad Autónoma de Campeche, Mexico; ³Universidad Autónoma Chapingo, Unidad Regional Universitaria Sursureste, Teapa, Tabasco, Mexico and ⁴Centro Nacional de Investigación Disciplinaria en Salud Animal e Inocuidad, INIFAP, Mexico

Abstract

The objective of this study was to evaluate the reduction in nematode faecal egg count (FEC) in Pelibuey lambs segregated as resistant (RES), susceptible (SUS) and intermediate (INT) to gastrointestinal nematodes. Twenty-nine weaned Pelibuey lambs, aged five months old, free of nematode infection, were used. Nine lambs were RES, six were SUS and 14 were INT lambs. The study consisted of two phases: in Phase 1 the lambs were infected experimentally with Haemonchus contortus. In Phase 2, the lambs were naturally infected by grazing. Faecal and blood samples were taken every week. The packed cell volume and total protein were quantified. The FEC value (FECmax) per lamb was recorded together with a natural reduction in FEC in the two phases. The data were analysed with a model of measures repeated over time. During Phase 1, the RES lambs showed the lowest FEC (1061 ± 1053) compared to the other groups (INT: 2385 ± 1794 eggs per gram of faeces (EPG); and SUS: 3958 ± 3037 EPG). However, in Phase 2 no significant differences (p > 0.05) were observed between the groups of lambs (RES: 275 ± 498 EPG; SUS: 504 ± 1036 EPG; and INT: 603 ± 1061 EPG). At the end of Phase 1, the FEC of RES lambs was naturally reduced by 75.5% in respect to FECmax (p < 0.05), and at the end of Phase 2 the reduction in FEC was 90% in respect to FECmax (p > 0.05); the same behaviour was observed in RES and SUS lambs. It is concluded that the artificial infection in the lambs induced a more rapid immune response in RES than SUS lambs, and all lambs developed high acquired resistance by continuous infection.

Introduction

Parasitism caused by Haemonchus contortus and other gastrointestinal nematodes (GIN) is responsible for significant economic losses in the production of grazing sheep due to stunted growth, mortality and the expense of anthelmintic treatments for their control. However, the great genetic diversity of these parasites has allowed them to develop resistance to anthelmintics through mutations established in the population, causing the phenomenon of anthelmintic resistance (AR) (Hodgkinson et al., 2019). Given this scenario, one alternative to avoid AR is the breeding of animals selected for their high resistance or low susceptibility to GIN. Genetic resistance to GIN has a heritability between 0.2 and 0.4 (Sréter et al., 1994). It represents the genetic capacity of the immune response to infections and produces in each generation a segregation of low- and high-resistant lambs with different abilities to develop the parasitic infection (Emery et al., 2016). This resistance to GIN is an individual characteristic that depends mainly with age and previous exposure to the parasite, which allows the animal to express innate immunity and develop acquired immunity that protects the host from infection (Greer & Hamie, 2016). The selection of individuals for their GIN resistance results in a reduction in the number of eggs in the environment, resulting in a decrease in the pasture – the source of parasitic infection (Sweeney et al., 2016). The feasibility of selecting sheep resistant to GIN has been widely demonstrated in various breeds and environments (Woolaston et al., 1990; Zvinorova et al., 2016; Moreno et al., 2017). In commercial flocks and experimental stations, GIN-resistant individuals have been selected using phenotypic markers such as faecal egg count (FEC) to assess resistance status and packed cell volume (PCV) as a measure of natural infection impact. Some breeds of hair sheep have been shown to have developed natural resistance to GIN infections, such as Blackbelly, St Croix, Florida Native, Gulf Cost Native, Thin Tail, Garole and Red Masai (Miller & Horohov, 2006; Shakya et al., 2009; Saddiqi et al., 2010; Karrow et al., 2014). In these breeds, a genetic variation of resistance to H. contortus has also been confirmed; therefore, the selection of resistant (RES) individuals

represents an option for the control of parasitic infections in tropical and subtropical environments where high prevalence of AR is reported (Torres-Acosta et al., 2012). In Mexico, the Pelibuev breed has been reported to have resistance to GIN, with significant variations within the breed (Morteo-Gómez et al., 2004; Castillo et al., 2011; González-Garduño et al., 2013; Zaragoza-Vera et al., 2019), which makes it a candidate for selection to genetically improve this trait in this breed. In order to select RES individuals, it is necessary to generate the infection and to monitor the development of the corresponding immunity. With many variants, oral challenge of lambs with infective larvae (L3) and their subsequent challenge with either artificial or natural infection is the most widely used protocol (Saccareau et al., 2017), where the level of antigenic stimulus is an important event in the speed of acquisition of acquired immunity. In a previous study, it is reported that in Pelibuey lambs initially infected orally with 7500 L3 of H. contortus, it was possible to successfully segregate RES and susceptible (SUS) individuals at the end of that infection according to their average FEC (Ojeda-Robertos et al., 2017).

The objective of the present study was to determine the impact on FEC due to the natural resistance presented by Pelibuey lambs segregated as RES, SUS and intermediate (INT) during two consecutive infections with GIN.

Materials and methods

Location of area of study

The study was conducted from February 4 to June 10, 2017, in the municipality of Salto de Agua, Chiapas, Mexico, which has a humid tropical climate (Kottek *et al.*, 2006); the average annual temperature there is 26.6°C and the precipitation is 3298 mm (CONAGUA, 2019).

Animal handling

The procedures were carried out in accordance with the normative terms of the 'Regulations for the use and care of animals intended for research in the Postgraduate College', approved by the General Academic Council (CGA) on November 18, 2016.

From a group of 140 Pelibuey lambs born subsequent to a 30-day breeding season at the Centre for Training and Reproduction of Minor Species (CECAREM) in Villahermosa, Tabasco, Mexico, 29 lambs were acquired after weaning. The age difference between them did not exceed 30 days. The lambs during lactation remained housed until weaning, which occurred after 60 days, so they did not present infection with GIN, which was confirmed with a coproparasitoscopic examination. The lambs were identified with a numbered collar and kept in elevated floor cages to keep them free of GIN infection. When they were approximately five months old (14 days before the start of the experiment) they were dewormed (levamisole 7.7 mg kg⁻¹ body weight (BW)) and treated for coccidia (toltrazuril 20 mg kg⁻¹ BW). The study consisted of two phases: Phase 1 (day 0-day 70) and Phase 2 (day 84-day 154). In Phase 1 the lambs remained in the elevated-floor cages, where they received ad libitum an integral balanced food with 14% crude protein and water. At the end of Phase 1 (day 70) the lambs were dewormed (albendazole 10 mg kg⁻¹ BW) and kept in the cages until the beginning of Phase 2 (day 84), at which time they were introduced to grasslands of African star grass (Cynodon plectostachyus). They grazed approximately ten hours per day, and were housed in a galley in

the afternoon where they received a dietary supplement with 14% crude protein at a rate of 300 g day⁻¹.

Artificial and natural infection

L3 were obtained by coproculture (Corticelli & Lai, 1963) from two donor lambs previously infected with an oral dose of 300 L3 from a local strain of *H. contortus*. The 29 lambs were trickle infected on days three, five and seven orally with 100 L3 of *H. contortus* per kg BW, receiving, in total, approximately 6000 L3 per lamb. The natural infection (Phase 2) began on day 84 when the lambs were withdrawn from the cages and placed in grassland of *C. plectostachyus* that was being used by a herd of Blackbelly sheep infected with GIN kept in the meadows.

Measurements

Eleven measurements were made in each phase (fig. 1): each week, faecal samples were taken directly from the rectum of the lambs and blood from the jugular vein, using vacuum tubes containing ethylene diamine tetra acetate (EDTA) as anticoagulant (Vacutainer, Becton Dickinson, USA). All samples were processed in the Animal Parasitology Laboratory of the South-East University Regional Unit (URUSSE) of the Chapingo Autonomous University (UACh) in Teapa, Tabasco, Mexico.

Parasitology

The FEC was determined in faeces by the McMaster method (Thienpont *et al.*, 2003) with sensitivity of 50 eggs per gram of faeces (EPG). In Phase 1, the presence of *H. contortus* was confirmed by L3 obtained by coprocultures, while the identification of GIN L3 genera during grazing (Phase 2) was performed according to available keys (van Wyk & Mayhew, 2013) after coproculture according to the methodology described by Corticelli & Lai (1963) of faecal samples on day 105.

Haematology

The PCV, total white blood cell count (WBC) and the differential count of three leukocyte fractions, consisting of lymphocytes (LYM), basophils, monocytes and eosinophils (MID) and neutrophils (GRAN), were determined by an automated haematological analyser based on electronic impedance (Medonic CA-620, Boule Medical AB, Stockholm, Sweden). Total plasma protein (TPP) was quantified by a manual refractometer (Atago, Tokio, Japan).

Lamb segregation model

At the end of Phase 1, the lambs were segregated into three groups based on the FEC results: RES lambs selected from mean EPG – three times the standard error; SUS from mean EPG + three times the EE; and INT, formed by the lambs not included in the other groups (Morteo-Gómez *et al.*, 2004). According to the segregation model, the RES group was composed of nine lambs, the SUS group of six and the INT group of 14.

Equation to calculate FEC reduction

In Phase 1, the maximum value of FEC (FECmax) was recorded together with the day this value occurred by each lamb. With the FECmax as a reference value, the reduction of the FEC was

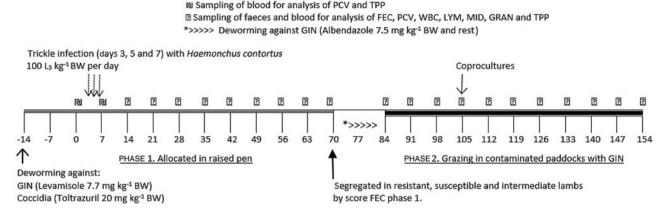


Fig. 1. Experimental design for gastrointestinal nematode resistant, susceptible and intermediate Pelibuey lambs under artificial Haemonchus contortus infection and natural pasture infection.

determined at the end of Phase 1 and at the end of Phase 2. The following formula was used for each lamb:

Reduction in FEC (%) = $100 - (FECx / FECmax \times 100)$

where FECx is the FEC at the end of Phase 1 (x = 1, day 70) and at the end of Phase 2 (x = 2, day 154).

Statistical analysis

Data were analysed between groups of segregated lambs such as RES, SUS and INT with a model of repeated measures over time using the MIXED procedure of the statistical package, and the means were compared by the Tukey test (SAS, 2004). To eliminate asymmetry and to approximate the data to a normal distribution, the EPG values were transformed (log EPG + 1). The following model was used:

$$Y_{ijkl} = \mu + \rho_i + \zeta_j + \gamma_{k(i)} + \delta(\zeta)_{il} + \rho^* \zeta_{ij} + \rho^* \delta(\zeta)_{ijl} + \varepsilon_{ijkl}$$

where Y_{ijkl} = variable response (EPG, PCV, WBC, LYM, MID, GRAN and TPP); μ = general mean; ρ_i = fixed effect of the type of animal (i = RES, SUS, INT); ζ_j = fixed effect of the phase (j = 1, 2); $\gamma_{k(i)}$ = random effect of lambs nested in the type of lamb, $\delta(\zeta)_{jl}$ = fixed effect of the sampling day nested in phase; $\rho^*\zeta_{ij}$ = interaction of type of animal with phase; $\rho^*\delta(\zeta)_{ijl}$ = interaction of the type of animal with the sampling day nested in phase; and ε_{ijkl} = random error associated with the l-th repeated measurement in the k-th lamb.

Results

Parasitology

FEC

During Phase 1, the RES lambs presented the lowest FEC (1061 ± 1053 EPG) with respect to the other groups (P < 0.05), but the INT and SUS (INT: 2385 ± 1794; and SUS: 3958 ± 3037) were no different (P > 0.05). However, in Phase 2, no significant differences (p > 0.05) were observed between the groups of lambs (RES: 275 ± 498; SUS: 504 ± 1036; and INT: 603 ± 1061). Figure 2 shows the dynamics of the FEC in the two phases of infection. It is noteworthy that the FEC reduction started from day 28 in the RES group, two weeks

before the SUS group (day 42) and three weeks before the INT group (day 49). During Phase 2, the anthelmintic was not effective and, therefore, the FEC remained high until day 91 and subsequently decreased 14 days after grazing began.

FEC reduction

Significant differences (p < 0.05) were observed in the FECmax between the segregated lambs in Phase 1, being 2883 EPG for the RES lambs on day 28, 8725 EPG for SUS lambs on day 42 and 5446 EPG for INT lambs on day 49. Table 1 shows the percentage reduction in FEC from the FECmax at the end of Phase 1 (day 70) and Phase 2 (day 154).

Significant differences were found in the reduction in FEC at the end of Phase 1 (p < 0.05), where the FEC of RES lambs was reduced by 75.5%, compared to 65.1% for SUS and 62.6% for INT lambs. No significant differences (p > 0.05) were found in the reduction in FEC at the end of Phase 2; the reduction in FEC as a response to acquired resistance provoked the highest reduction value in all groups of lambs.

Coprocultures

The GIN genera found in the coprocultures in the grazing period (day 105) were *Haemonchus* (72.0%), *Trichostrongylus* (13.8%), *Cooperia* (12.3%) and *Oesophagostomum* (1.9%).

Haematology

The RES group presented the highest values (p < 0.05) of PCV with 26.2 ± 4.0% in Phase 1 compared to the other groups (SUS 25.1 ± 4.8%; and INT 23.6 ± 3.6%). In Phase 2, significant differences were observed (p < 0.05) where the RES (27.0 ± 3.0%) and SUS lambs (26.7 ± 3.2%) presented higher values than the INT group (24.6 ± 3.4%).

Figure 3 shows the changes in the PCV of segregated SUS, RES and INT lambs in the two phases of infection. In Phase 1, the decrease in PCV in all groups of lambs was evident from infection with the hematophagous parasite *H. contortus*. However, RES lambs showed their lowest PCV value on day 21 (two weeks post-infection) with 22.0%. The lowest PCV values for SUS and INT lambs were recorded on day 35 (four weeks post-infection) with 20.0% and 21.3%, respectively. Subsequently, the PCV values increased in all groups until day 91. After the natural infection,

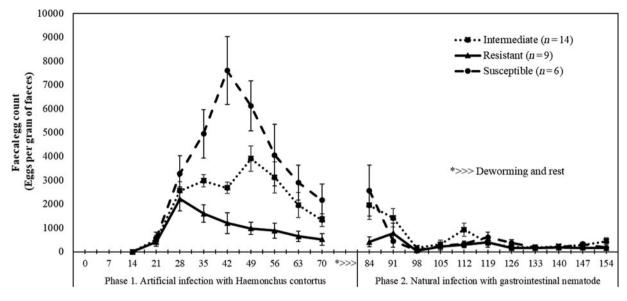


Fig. 2. Faecal egg count (least squares means ± standard error) for gastrointestinal nematode resistant, susceptible and intermediate Pelibuey lambs under artificial Haemonchus contortus infection and natural pasture infection.

Table 1. Faecal egg count (FEC) and FEC reduction with respect to the maximum FEC (FECmax) exhibited during artificial infection (Phase 1) and natural infection
(Phase 2) in Pelibuey lambs segregated into resistant, susceptible and intermediate groups in each infection phase.

Segregation	Experimental day	Mean (± SE) EPG	FEC reduction (± SE) %	
Resistant (n = 9)	28 (FECmax)	2883 (403)		
	70 (FEC end Phase 1)	706 (206)	75.5 (10.9) ^a	
	154 (FEC end Phase 2)	253 (83)		91.2 (3.8) ^a
Susceptible (n = 6)	42 (FECmax)	8725 (897)		
	70 (FEC end Phase 1)	3044 (667)	65.1(7.1) ^b	
	154 (FEC end Phase 2)	483 (211)		94.4(2.6) ^a
Intermediate (n = 14)	49 (FECmax)	5446 (385)		
	70 (FEC end Phase 1)	2035 (307)	62.6(5.0) ^b	
	154 (FEC end Phase 2)	592 (158)		89.1(2.9) ^a

 a,b Different letters in the same column indicate significant differences (p < 0.05). SE, standard error; EPG, eggs per gram of faeces.

there was a slight gradual decrease until day 126 and, again, a recovery until the end of Phase 2.

In Phase 1, the RES lambs showed the highest values (p < 0.05) for WBC, LYM, MID, GRAN and TPP. In Phase 2, the highest values (p < 0.05) for WBC, LYM, MID and GRAN were for INT lambs, and RES and SUS lambs presented higher TPP values (p < 0.05) than those of the INT group (table 2).

Discussion

Phase 1

In Phase 1, artificial infection with L3 of *H. contortus* over four weeks allowed the RES lambs to demonstrate their ability to control the establishment of this parasite in the abomasum by mounting a rapid immune response, which was reflected in the lowest FEC values (Stear *et al.*, 1999), compared to the INT and SUS lambs. In addition, the FECmax in the RES group occurred two weeks before the other lambs (28 days post-infection).

The high FEC during the GIN infection of Phase 1 has also long been observed in wool breeds (Stear et al., 1995) and it has been indicated that after the prepatent period (without the expulsion of nematode eggs), a peak occurs in the FEC around 6-8 weeks post-infection. However, in hair breeds the FEC peak occurs around the fourth week post-infection, as observed in the present study and as reported in an artificial infection with 10,000 L3 of H. contortus in the crossbred Barbados Blackbell and Virgin Islands White sheep (Notter et al., 2003) and St Croix lambs (Jacobs et al., 2015). However, in this study a different response was observed, and only the RES lambs showed a peak at four weeks, while the SUS and INT lambs showed a FEC peak at the sixth week, reflecting a late response, similar to that observed in wool sheep. In a previous study with Pelibuey sheep, FEC peaks were also affected by the dose used in the artificial infection. The FEC peak was observed at 35 days when the animals had resistance against GIN, while other groups showed the FEC peak on days 42 and 56, which was observed in SUS animals (González-Garduño et al., 2019).

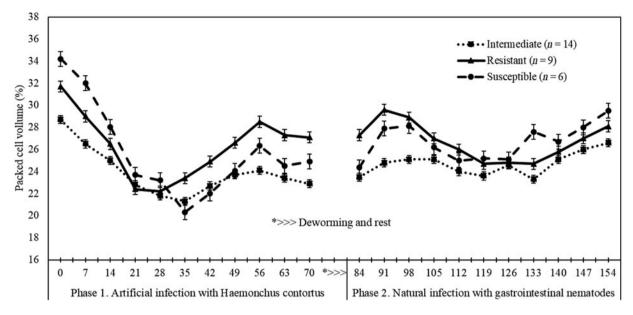


Fig. 3. Blood packed cell volume (least squares means ± standard error) for gastrointestinal nematode resistant, susceptible and intermediate Pelibuey lambs under artificial infection and natural pasture infection.

Segregation						
Variable	Resistant $(n = 9)$	Intermediate (n = 14)	Susceptible (n = 6)	Reference values*		
WBC (×10 ³ cell μ L ⁻¹	1)					
Phase 1	8.85 ± 0.76^{a}	8.25 ± 0.65^{b}	8.24 ± 0.85^{b}	4–12		
Phase 2	9.01 ± 0.66^{a}	9.33 ± 0.68^{a}	8.21 ± 0.67^{b}			
LYM (×10 ³ cell μ L ⁻¹))					
Phase 1	5.53 ± 0.59^{a}	5.28 ± 0.47^{a}	5.46 ± 0.69^{a}	2–9		
Phase 2	5.09 ± 0.44^{a}	5.36 ± 0.42^{a}	4.71 ± 0.77^{b}			
MID (×10 ³ cell μ L ⁻¹))					
Phase 1	2.49 ± 0.29^{a}	2.23 ± 0.22^{b}	2.07 ± 0.31^{b}	0–2		
Phase 2	2.64 ± 0.26^{a}	2.67 ± 0.31^{a}	2.30 ± 0.30^{b}			
GRAN (×10 ³ cell μ L	-1)					
Phase 1	$0.84 \pm 0.10^{\text{a}}$	0.73 ± 0.09^{b}	0.71 ± 0.11^{b}	0.7–6		
Phase 2	1.28 ± 0.23	1.29 ± 0.22	1.29 ± 0.30			
TPP (g dL^{-1})						
Phase 1	6.26 ± 0.18^{a}	$6.00 \pm 0.13^{\circ}$	6.11 ± 0.22^{b}	6–7.5		
Phase 2	6.47 ± 0.18^{a}	6.36 ± 0.14^{b}	6.49 ± 0.22^{a}			

Table 2. Means (± standard error) of haematological values in Pelibuey lambs that are resistant, susceptible and intermediate to gastrointestinal nematode infection.

a.bDifferent letters in the same row indicate significant differences (*p* < 0.05). *Byers & Kramer (2010). WBC, blood cell count; LYM, lymphocytes; MID, medium cells (monocytes; basophils and eosinophils); GRAN, neutrophils; TPP, total plasma protein.

The percentage of FEC reduction in RES lambs at the end of the artificial infection was 75.5% with a 706 EPG, while the other two groups (SUS and INT) reached the end of Phase 1 with FECs more than three times that of the RES lambs (3044 and 2035, respectively). However, the percentage reductions in FEC measured as the difference from FECmax were similar (65.1% and 57.6%, respectively) between the INT and SUS groups (P > 0.05). This indicates that these lambs had the same capacity to mount an immune response. The differences observed were from the day in which the peak of infection was observed, attributed to immune reaction in the host, and in the value of the FEC.

There is a report of St Croix lambs being re-infected with 10,000 L3 of *H. contortus* that indicates the development of an acquired immune response on day seven post-infection by presenting high levels of LYM, production of immunoglobulin A and the infiltration of immune cells into the mucosa of the

abomasum (Bowdridge *et al.*, 2015). After an earlier infection, the immune response may limit the establishment of adult parasites and induce morphological changes that reduce their fertility and, therefore, shorten the duration and severity of the infection (Bowdridge *et al.*, 2015; McRae *et al.*, 2015).

During Phase 1, higher cell counts of WBC, MID and GRAN were observed in RES lambs compared to the other two groups. This supports what has been observed in the immune response of mammals to infection with helminths, which is characterized by an increase in cellular immune responses such as in eosinophils, basophils, cell mast cells and macrophages in response to tissue damage. These immune cells respond, in turn, with an increase in cytokine production (Karrow et al., 2014). The rapid response of RES lambs may be due to an early increase in circulating CD4⁺ T LYM in response to the infiltration of *H. contortus* larvae into the abomasum. This release and excretion/secretion of products induces a combination of humoral immunity, eosinophilia and mast cell hyperplasia that leads to changes in the abomasum mucosa causing expulsion of L3 and length reduction of L₄, which may be related to the decrease in fertility in the adult stage (Karanu et al., 1997; Bowdridge et al., 2015). This early response to primary infection is genetically controlled and has been reported for RES sheep breeds in gene expression studies related to T LYM proliferation (Terefe et al., 2007; Yang et al., 2015) with complementary activation. Both are the first events in the host's immune responses to infection (Lee et al., 2011; Valilou et al., 2015; Guo et al., 2016). The same response occurs in the SUS and INT lambs, but at a later day, which results in the establishment of a greater number of adult parasites and, of course, a higher FEC.

The reduction in PCV associated with increasing FEC after infection with *H. contortus* is a reflection of infection with this parasite, whose hematophagous feeding habits directly affect the health of animals by reducing PCV; this negative relationship is reported in numerous studies (MacKinnon *et al.*, 2010; Castillo *et al.*, 2011). RES lambs showed the highest PCV values during Phase 1, which agrees with other studies, where higher levels of PCV have been found in RES compared to SUS sheep when infected with *H. contortus* (Mugambi *et al.*, 1997; Shakya *et al.*, 2009).

The TPP values of the SUS and INT lambs were significantly lower than those of the RES lambs. Decreased values of TPP in the SUS and INT lambs could be an important indicator because this reduction in TPP has been reported in sheep with clinical conditions such as hypoproteinaemia, diarrhoea, lack of appetite and malnutrition after infection with *H. contortus*, so this parameter might be considered for selecting animals with greater resistance to GIN (Zaros *et al.*, 2014).

Phase 2

At the beginning of Phase 2 (day 84), FEC were similar to day 70 of Phase 1, indicating that the anthelmintic was not effective. Thus, a second true prepatent period may not have occurred. The FEC reduction in Phase 2 may have been a continued response carried out from Phase 1 or could be attributed to an acquired immunological development by lambs leading to a FEC of almost 0. This latter explanation has occurred in RES breeds (Zaragoza-Vera *et al.*, 2019). In response to the continued infection, the development of acquired immunity was seen. The FEC reduction in Phase 2 was similar among resistance classifications of lambs (RES = 91.2%; SUS = 94.4%; and INT = 89.1%) with respect to the maximum value recorded in Phase 1.

A strong acquired immune response would not allow a new infection to establish. The specific acquired immunity mechanism is characterized by an increase in T_H^2 cytokines, the recruitment of eosinophils, mast cells and globular leukocytes, and an increase in the production of specific IgA (Balic *et al.*, 2002; Shakya *et al.*, 2009; Jacobs, 2013). This immune response intervenes in mechanisms to control the number of parasites either by reducing the establishment of larvae, inducing hypobiosis, stunting morphology or decreasing the fecundity of adult nematodes (Rowe *et al.*, 2008; Ortolani *et al.*, 2013).

Other studies indicate a reduction in FEC (Eady *et al.*, 2003) by acquired immunity, but also depend on the degree of acquired response (González-Garduño *et al.*, 2019). This represents an important alternative, especially when the nematodes in the flocks show AR, when the persistence of infection can be very long and the lambs have insufficient immune stimulation to achieve self-cure (González-Garduño *et al.*, 2016).

Acquired resistance has been characterized by a local and blood eosinophilia response, as well as an increase in T cells (CR⁺ $\gamma\delta$) in the first three days after challenge (Robinson *et al.*, 2010). In addition, a very rapid increase in CD4⁺ T cells occurs. All subpopulations of LYM and tissue and blood eosinophils are reduced on day seven, and subsequently increase at 28 days post-infection, suggesting a different immune response against larvae and adults (Robinson *et al.*, 2010).

During Phase 2, all groups of lambs showed similarities in cell values, PCV and TPP. At the same time that FEC was reduced, improvement in the haematological value was observed, keeping the average within the normal values for the breed (Byers & Kramer, 2010).

Conclusion

An artificial trickle infection of Pelibuey lambs with a dose of 6000 L3 of *H. contortus* induced a rapid immune response in the most RES animals, which could be identified and selected to develop a flock genetically resistant to GIN in the future.

Acknowledgements. The author Alvar Alonzo Cruz-Tamayo is a PhD student of the Program of Genetic Resources and Productivity – Livestock at the Graduate College (RGP-COLPOS), and thanks the Professional Teacher Development Program for the scholarship (PRODEP-DSA/103.5/16/5957 UNACAM-118).

Financial support. This work was supported by the Academic Body Strengthening Program (PRODEP-DSA/103.5/15/14473).

Conflicts of interest. None.

Ethical standards. The procedures were carried out in accordance with the normative terms of the 'Regulations for the use and care of animals intended for research in the Postgraduate College', approved by the CGA on November 18, 2016, in which the Mexican Official Laws, Regulations and Standards and other legal provisions are considered.

References

- Balic A, Bowles VM and Meeusen ENT (2002) Mechanisms of immunity to Haemonchus contortus infection in sheep. Parasite Immunology 24(1), 39–46.
- Bowdridge SA, Zajac AM and Notter DR (2015) St. Croix sheep produce a rapid and greater cellular immune response contributing to reduced establishment of *Haemonchus contortus*. Veterinary Parasitology 208(3–4), 204–210.
- Byers SR and Kramer JW (2010) Normal hematology of sheep and goats. pp. 836–842 *in* Weiss DJ, Wardrop KJ (*Eds*) *Schalm's veterinary hematology*. 6th edn. Iowa, USA, Wiley-Blackwell.

- Castillo JA, Medina RD, Villalobos JM, Gayosso-Vázquez A, Ulloa-Arvízu R, Rodríguez RA, Ramírez HP and Morales RA (2011) Association between major histocompatibility complex microsatellites, fecal egg count, blood packed cell volume and blood eosinophilia in Pelibuey sheep infected with *Haemonchus contortus*. Veterinary Parasitology 177(3–4), 339–344.
- CONAGUA (2019) Servicio Meteorológico Nacional. Normales climatológicas, https://smn.conagua.gob.mx/es/informacion-climatologicapor-estado?estado=tab (accessed August 13, 2020).
- **Corticelli B and Lai M** (1963) Studies on the technique of culture of infective larvae of gastrointestinal strongyles of cattle. *Acta de Medicina Veterinaria Napoli* **9**, 347–357.
- Eady SJ, Woolaston RR and Barger IA (2003) Comparison of genetic and nongenetic strategies for control of gastrointestinal nematodes of sheep. *Livestock Production Science* 81(1), 11–23.
- **Emery DL, Hunt PW and Le Jambre LF** (2016) *Haemonchus contortus*: the then and now, and where to from here? *International Journal for Parasitology* **46**, 755–769.
- **González-Garduño R, Gives PM and Torres-Hernández G** (2013) Variability in the fecal egg count and the parasitic burden of hair sheep after grazing in nematode infected paddocks. *Pesquisa Veterinaria Brasileira* **33**(4), 469-475.
- González-Garduño R, López-Arellano ME, Ojeda-Robertos N, Liébano-Hernández E and Mendoza-de Gives P (2014) Diagnóstico *in vitro* y en campo de resistencia antihelmíntica en nematodos gastrointestinales de pequeños rumiantes. *Archivos de Medicina Veterinaria* **46**(3), 399–405.
- González-Garduño R, López-Arellano ME, Mendoza-De-Gives P, Ojeda-Robertos NF and Chay-Canul AJ (2016) Temporary immunity of Blackbelly lambs reinfected with *Trichostrongylus colubriformis*. Acta Scientiae Veterinariae 44, 1–6.
- González Garduño R, López-Arellano ME, Torres-Hernández G, Oliva-Hernández J and Hinojosa-Cuéllar JA (2019) Assessment of acquired resistance in previously infected lambs with. *Indian Journal of Animal Research* 53(9), 1223–1228.
- Greer AW and Hamie JC (2016) Relative maturity and the development of immunity to gastrointestinal nematodes in sheep: an overlooked paradigm? *Parasite Immunology* 38(5), 263–272.
- Guo Z, González JF, Hernandez JN, McNelly TN, Corripio-Miyar Y, Frew D, Morrison T, Yu P and Li RW (2016) Possible mechanisms of host resistance to *Haemonchus contortus* infection in sheep breeds native to the Canary Islands. *Scientific Reports* 6, 1–14.
- Hodgkinson JE, Kaplan RM, Kenyon F, et al. (2019) Refugia and anthelmintic resistance: concepts and challenges. *International journal for parasit*ology. Drugs and drug resistance 10, 51–57.
- Jacobs JR (2013) Characterizing peripheral cellular and humoral immune responses to *Haemonchus contortus* in sheep. *Graduate Theses, Dissertations, and Problem Reports* 489. doi:10.33915/etd.489
- Jacobs JR, Greiner SP and Bowdridge SA (2015) Serum interleukin-4 (IL-4) production is associated with lower fecal egg count in parasite-resistant sheep. *Veterinary Parasitology* **211**(1–2), 102–105.
- Karanu FN, McGuire TC, Davis WC, Besser TE and Jasmer DP (1997) CD4 + T lymphocytes contribute to protective immunity induced in sheep and goats by *Haemonchus contortus* gut antigens. *Parasite Immunology* 19 (10), 435–445.
- Karrow NA, Goliboski K, Stonos N, Schenkel F and Peregrine A (2014) Review: genetics of helminth resistance in sheep. *Canadian Journal of Animal Science* **94**(1), 1–9.
- Kottek M, Grieser J, Beck C, Rudolf B and Rubel F (2006) World map of the Köppen-Geiger climate classification updated. *Meteorologische Zeitschrift* 15(3), 259–263.
- Lee CY, Munyard KA, Gregg K, Wetherall JD, Stear MJ and Groth DM (2011) The influence of MHC and immunoglobulins A and E on host resistance to gastrointestinal nematodes in sheep. *Journal of Parasitology Research* 2011, 101848.
- MacKinnon KM, Zajac AM, Kooyman FNJ and Notter DR (2010) Differences in immune parameters are associated with resistance to *Haemonchus contortus* in Caribbean hair sheep. *Parasite Immunology* **32** (7), 484–493.

- McRae KM, Stear MJ, Good B and Keane OM (2015) The host immune response to gastrointestinal nematode infection in sheep. *Parasite Immunology* **37**(12), 605–613.
- Miller JE and Horohov DW (2006) Immunological aspects of nematode parasite control in sheep. Journal of Animal Science 84(Suppl), 124–132.
- Moreno CR, Sallé G, Jacquiet P, et al. (2017) La résistance génétique aux infections par les nématodes gastro-intestinaux chez les petits ruminants: un enjeu de durabilité pour les productions à l'herbe. *INRA Productions Animales* **30**(1), 47–56.
- Morteo-Gómez R, González-Garduño R, Torres-Hernández G, Nuncio-Ochoa G, Becerril-Pérez C, Gallegos-Sánchez J and Aranda-Ibañez E (2004) Effect of the phenotypic variation in the resistance of Pelibuey lambs to the infestation with gastrointestinal nematodes. *Agrociencia* 38(4), 395–404.
- Mugambi JM, Bain RK, Wanyangu SW, Ihiga MA, Duncan JL, Murray M and Stear MJ (1997) Resistance of four sheep breeds to natural and subsequent artificial *Haemonchus contortus* infection. *Veterinary Parasitology* **69** (3–4), 265–273.
- Notter DR, Andrew SA and Zajac AM (2003) Responses of hair and wool sheep to a single fixed dose of infective larvae of *Haemonchus contortus*. *Small Ruminant Research* **47**(3), 221–225.
- Ojeda-Robertos NF, Torres-Acosta JFJ, González-Garduño R and Notter DR (2017) Phenotypic expression of parasite susceptibility to *Haemonchus contortus* in Pelibuey sheep. *Veterinary Parasitology* 239, 57–61.
- Ortolani EL, do Rego Leal ML, Minervino AH, Aires AR, Coop RL, Jackson F and Suttle NF (2013) Effects of parasitism on cellular immune response in sheep experimentally infected with *Haemonchus contortus*. Veterinary Parasitology 196(1–2), 230–234.
- Robinson N, PIedrafita D, Snibson K, Harrison P and Meeusen EN (2010) Immune cell kinetics in the ovine abomasal mucosa following hyperimmunization and challenge with *Haemonchus contortus*. Veterinary Research 41 (37), 1–10.
- Rowe A, McMaster K, Emery D and Sangster N (2008) Haemonchus contortus infection in sheep: parasite fecundity correlates with worm size and host lymphocyte counts. Veterinary Parasitology 153(3–4), 285–293.
- Saccareau M, Sallé G, Robert-Granié C, Duchemin T, Jacquiet P, Blanchard A, Cabaret J and Moreno CR (2017) Meta-analysis of the parasitic phase traits of *Haemonchus contortus* infection in sheep. *Parasites and Vectors* 10 (1), 1–14.
- Saddiqi HA, Iqbal Z, Khan MN and Muhammad G (2010) Comparative resistance of sheep breeds to *Haemonchus contortus* in a natural pasture infection. *International Journal of Agriculture and Biology* 12(5), 739–743.
- Saddiqi HA, Sarwar M, Iqbal Z, Nisa M and Shahzad MA (2012) Markers/ parameters for the evaluation of natural resistance status of small ruminants against gastrointestinal nematodes. *Animal* 6(6), 994–1004.
- SAS (2004) The SAS System for Windows, version 9. SAS Institute. Inc., Cary, North Carolina.
- Shakya KP, Miller JE and Horohov DW (2009) A Th2 type of immune response is associated with increased resistance to *Haemonchus contortus* in naturally infected Gulf Coast native lambs. *Veterinary Parasitology* 163 (1–2), 57–66.
- Sréter T, Kassai T and Takács E (1994) The heritability and specificity of responsiveness to infection with *Haemonchus contortus* in sheep. *International Journal for Parasitology* 24(6), 871–876.
- Stear MJ, Bairden K, Duncan JL and Murray M (1995) A comparison of the responses to repeated experimental infections with *Haemonchus contortus* among Scottish Blackface lambs. *Veterinary Parasitology* **60**(1–2), 69–81.
- Stear MJ, Strain S and Bishop SC (1999) Mechanisms underlying resistance to nematode infection. *International Journal for Parasitology* 29(1), 51–56.
- Sweeney T, Hanrahan JP, Ryan MT and Good B (2016) Immunogenomics of gastrointestinal nematode infection in ruminants – breeding for resistance to produce food sustainably and safely. *Parasite Immunology* 38 (9), 569–586.
- Terefe G, Lacroux C, Andreoletti O, et al. (2007) Immune response to Haemonchus contortus infection in susceptible (INRA 401) and resistant (Barbados Black Belly) breeds of lambs. Parasite Immunology 29(8), 415–424.

- Thienpont D, Rochette F and Vanparijs OFJ (2003) *Diagnosing helminthiasis* by coprological examination. 3rd edn. Beerse, Belgium, Janssen Research Foundation.
- Torres-Acosta JFJ, Mendoza-de-Gives P, Aguilar-Caballero AJ and Cuéllar-Ordaz JA (2012) Anthelmintic resistance in sheep farms: update of the situation in the American continent. *Veterinary Parasitology* 189 (1), 89–96.
- Valilou RH, Rafat SA, Notter DR, Shojda D, Moghaddam G and Nematollahi A (2015) Fecal egg counts for gastrointestinal nematodes are associated with a polymorphism in the MHC-DRB1 gene in the Iranian Ghezel sheep breed. *Frontiers in Genetics* 6, 1–11.
- van Wyk JA and Mayhew E (2013) Morphological identification of parasitic nematode infective larvae of small ruminants and cattle: a practical lab guide. Onderstepoort Journal of Veterinary Research 80(1), 1–14.
- Woolaston RR, Barger IA and Piper LR (1990) Response to helminth infection of sheep selected for resistance to *Haemonchus contortus*. International Journal for Parasitology 20(8), 1015–1018.

- Yang Y, Zhou QJ, Chen XQ, Yan BL, Guo XL, Zhang HL and Du AF (2015) Profiling of differentially expressed genes in sheep T lymphocytes response to an artificial primary *Haemonchus contortus* infection. *Parasites and Vectors* 8(1), 235.
- Zaragoza-Vera CV, Aguilar-Caballero AJ, González-Garduño R, Arjona-Jiménez G, Zaragoza-Vera M, Torres-Acosta J, Medina-Reynés JU and Berumen-Alatorre AC (2019) Variation in phenotypic resistance to gastrointestinal nematodes in hair sheep in the humid tropics of Mexico. Parasitology Research 118(2), 567–573.
- Zaros LG, Neves MRM, Benvenuti CL, Navarro AM, Sider LH, Coutinho LL and Vieira LS (2014) Response of resistant and susceptible Brazilian Somalis crossbreed sheep naturally infected by *Haemonchus contortus*. *Parasitology Research* **113**(3), 1155–1161.
- Zvinorova PI, Halimani TE, Muchadeyi FC, Matika O, Riggio V and Dzama K (2016) Breeding for resistance to gastrointestinal nematodes the potential in low-input/output small ruminant production systems. *Veterinary Parasitology* **225**, 19–28.